

# Abstract

DNA and RNA are very important building blocks to all life at the molecular level. All known organisms' genetic instructions for development, function, and reproduction are stored in DNA. RNA is also a nucleic acid and has similar structures to DNA, and is essential for protein synthesis. It acts as a messenger and copies genetic instructions from DNA to translate into proteins. RNA is especially important to most viruses because it carries the genetic information. In the current project we are working on West Nile Virus(WNV), a member of the Flavivirus family. Flaviviruses are a group of viruses that have single stranded RNA genomes that directly translate to viral proteins. These viruses are found to have uniquely folded RNA that prevents breakdown by exoribonucleases: enzymes that digest viral RNA in the body. A previous study done on Zika virus has shown exoribonuclease mediated digestion which leads to accumulation of smaller fragments in the host. The RNase enzymes that help defend the cell against invading viral RNA are used to generate fragments that help the virus infect the cell.These smaller fragments lead to apoptosis and cell death. My research involves studying West Nile Virus and it mutants to research the regions involved in exoribonuclease digestion. I did molecular biology work that involved introducing mutations in DNA, modifying DNA by putting promoter sites to make RNA, and finally RNA synthesis.

# **Background and Motivation**

- The unique structure in flaviviruses prevent complete degradation of viral genomic RNA
- This leaves small RNA fragments that attach anywhere in the body, which enhances viral replication and infection



**Goal** is to figure out which interactions in the xrRNA structure are essential for RNase resistance and be able to disrupt those interactions when developing potential drugs



# **Probing the origin of RNase resistance in viral xrRNAs** Onyinyechi Okoro, Sneha Munshi, Daniiar Zhaguparov, Ishrat J. Meghla, and Michael T. Woodside 00 Department of Physics, Faculty of Science, University of Alberta

Site Directed Mutagenesis(SDM) and Polymerase Chain Reaction (PCR) - To study the regions affected by the exoribonuclease digestion, a specific base in the DNA is changed (SDM) and many copies of the DNA are made



#### SDM and PCR

- The bands show the molecular weight of the sample and that the DNA has properly multiplied
- ✤ The two bands shown are between 7000 and 5000 base pair length (bp length) according to the marker

#### **Bacterial Transformation**



- ✤ After DNA is amplified, it is introduced into bacteria to grow colonies and clone itself
- Select colonies are picked up to purify to test the DNA sequence and for RNA synthesis

#### **DNA Sequencing**



- All the stars show that our DNA sequence exactly matches the parental strand except for the one spot with the mutation
- Once sequencing was correct, RNA was synthesized and purified

- This gel shows the result of exoribonuclease mediated digestion in these flaviviruses
- Two samples of each virus was run, is without enzyme digestion, + is with enzyme digestion
- The lower band shows positive results because the enzyme digested the virus into smaller fragments that migrate farther down the gel.
- My research is focused on WNV and its mutants
- Three single mutations introduced: G3C, G55U,
- One double mutation introduced G56U | G37C



### Summary

- regions involved in the digestion
- amplified
- and purified

# **Discussion - Further Work**

- digestion
- Due to the constriction of time, I did molecular biology work and generated WNV mutants and synthesised RNA which will be used to do gel based assay
- The gel based assay will show if the exoribonuclease will fully digest the virus or if it will still digest into smaller fragments
- This will provide more information on the structural regions responsible for the resistance

# References

Chapman, E. G., Moon, S. L., Wilusz, J., & Kieft, J. S. (2014). RNA structures that resist degradation by Xrn1 produce a pathogenic Dengue virus RNA. ELife, 3. https://doi.org/10.7554/elife.01892 Slonchak, A., Parry, R., Pullinger, B., Sng, J. D. J., Wang, X., Buck, T. F., Torres, F. J., Harrison, J. J., Colmant, A. M. G., Hobson-Peters, J., Hall, R. A., Tuplin, A., & Khromykh, A. A. (2022). Structural analysis of 3'UTRs in insect flaviviruses reveals novel determinants of sfRNA biogenesis and provides new insights into flavivirus evolution. Nature Communications, 13(1). https://doi.org/10.1038/s41467-022-28977-3 Zhao, M., & Woodside, M. T. (2021). Mechanical strength of RNA knot in Zika virus protects against cellular defenses. Nature Chemical Biology, 17(9), 975–981. https://doi.org/10.1038/s41589-021-00829-z Szucs, M. J., Nichols, P. J., Jones, R. A., Vicens, Q., & Kieft, J. (2020). A New Subclass of Exoribonuclease-Resistant RNA Found in Multiple Genera of Flaviviridae. MBio, 11(5). https://doi.org/10.1128/mbio.02352-20

# Acknowledgements

A special thanks to Professor Michael T. Woodside for the opportunity to work in your lab. Thanks to my supervisor Sneha Munshi and to all the people in the Woodside lab who helped me make this project possible. Thanks to CIHR for all the lab materials used for this research. I would also like to thank WAGE, AHC, and Syncrude for their sponsorship and WISEST for this incredible opportunity.



✤ The resistance of exoribonuclease in flaviviruses prompted the research on the

All four mutants, G3C, G55U, G56U, and G56U | G37C were successfully done and

The DNA of the bacterial colonies was purified and from that, RNA was synthesized

Zika, Dengue isoforms 2 and 4, and West Nile all undergo exoribonuclease mediated



Femmes et Égalité des genres Canada