Bacillus species in food fermentations: an under-appreciated group of organisms for safe use in food fermentations

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Abstract

Fermented foods are consumed throughout the world and substantially contribute to food security. Many fermented products rely on the participation of *Bacillus* species but their contribution to food quality is not well documented. The purpose of this review is to highlight metabolic properties that relate to food quality and food safety. Most fermented foods that include *Bacillus* species use legumes or tubers as substrate and are traditionally prepared in East Asia, Africa and South America. In food fermentations, *Bacillus* species produce amylases and proteases, extracellular polysaccharides and polypeptides, and lipopeptides with antimicrobial activity. The metabolic traits of bacilli also provide opportunities for use in food fermentations in which they do not traditionally occur.

Keywords: *Bacillus*; food fermentations; microbial enzymes; antimicrobial lipopeptides; probiotic; *Bacillus cereus*

1 Introduction

2 Food fermentations utilize the activity of microorganisms for conversion of food components 3 to improve shelf-life and safety, as well as sensory and nutritional properties of food. In 4 addition, the presence of live microorganisms in fermented foods is increasingly recognized to 5 improve gastrointestinal health [1–3]. The assembly of communities of fermentation microbes 6 is determined by the fermentation conditions, i.e., the ingredients, temperature, pH, moisture 7 content, and using back-slopping or starter cultures (5). Most fermentations include yeasts 8 and/or lactic acid bacteria (LAB) as major fermentation microbes but mycelial molds, acetic 9 acid bacteria, staphylococci, propionibacteria, Enterobacteriaceae and bacilli also play 10 dominant roles in some fermented foods [4]. The contribution of *Bacillus* to food quality is not 11 nearly as well documented as the contribution of yeasts and lactobacilli.

12 *Bacillus* spp. are isolated from soil, water and plants, and are almost ubiquitously present in 13 foods [5]. They form endospores as a key element of their ecology [6]. Bacillus endospores 14 resist adverse conditions including heat, pressure, desiccation, and chemical assaults [6,7]. The resistance of spores to high temperature and high pressure and thus the survival in ingredients 15 that are cooked or steamed is further increased by the $spoVA^{2mob}$ operon [8,9]. The ecology of 16 17 Bacillus spp. and their relevance for plant agriculture and food production is shown in Figure 18 1. The presence of *Bacillus* spores in plants including cereals, pulses and cassava relates to 19 their occurrence as endophytes [10–13]. Because spores survive even cooking steps [14], 20 cooked or uncooked cereal grains, pulses and cassava generally harbor endospores of Bacillus species [15,16]. 21

Bacillus species are recruited for traditional food fermentations of pulses, tubers or cereals in
 Asia, Africa and South America (Figure 2) [3,17–19]. This review aims to provide an overview
 on the role of *Bacillus* in food fermentations by presenting traditional fermented foods that

include *Bacillus* species as major fermentation organisms, their specific contribution to food
 quality and safety, and explore the potential of using these organisms in non-conventional
 fermentation processes.

28 The presence of *Bacillus* in food fermentations

An overview on food fermentations that include *Bacillus* species as major fermentation organisms is shown in Table 1. Major processing and fermentation steps are also indicated in Table 1 to inform on major processing steps and fermentation conditions that select for *Bacillus* species. The overview is likely incomplete because not all fermented foods are documented in the scientific literature. Moreover, the overview in Table 1 does not reflect that fermented foods are produced in numerous varieties that share major processing and fermentation steps but may differ in flavor and appearance [3].

36 Fermented products from soybean and pulses

Consumption of soybean and soybean derived fermented products has a long history in many
East and South Asian countries (Table 1) [19]. *Bacillus* spp. frequently occur in alkaline food
fermentations, which use soybeans as substrate [17]; Several examples are presented in more
detail below.

41 Natto

Natto is a traditional Japanese fermented food produced from soybeans fermented by *natto* starter strains of *Bacillus subtilis* var. *natto* [20]. To make *natto*, soybeans are soaked, steamed, inoculated with *B. subtilis* var. *natto* and fermented at 40 °C for 24 h. The *natto* is cooled and aged in refrigerator for up to 1 week allowing the development of stringiness (Table 1). The fermentation process supports secretion of proteases by bacilli to degrade soy proteins, the production and racemization of L- and D-glutamic acids, and the synthesis and secretion of poly-γ-glutamate [21].

49 Cheonggukjang

50 *Cheonggukjang* is a Korean soybean paste made from cooked soybeans fermented with 51 *Bacillus* species at around 40 °C for 2-3 days (Table 1) [22]. Fermentation with *Bacillus* 52 hydrolyses soybean proteins and polysaccharides. In addition, isoflavones are converted to the 53 corresponding aglycones during fermentation [22].

54 Doenjang

55 *Doenjang* is traditionally manufactured by fermentation of *meju*, the fermented soybean block. 56 Meju is traditionally prepared by soaking, steaming, crushing, and then fermented for 2 to 3 57 months with B. subtilis, Rhizopus spp., and Aspergillus spp. The fermented meju is separated 58 into two parts; the supernatant liquid part is filtered to prepare soy sauce and precipitated solid 59 part is used for further ripening for over 2 months to make *doenjang* (Table 1) [23]. In a 60 modified, commercialized method, meju fermentation is solely inoculated by B. subtilis to 61 make koji, and koji is further ripened by Aspergillus oryzae, additional grains (e.g., wheat and 62 barley), and/or other flavor enhancing materials. Therefore, the use of controlled 63 microorganisms and standardized processes can boost the production efficiency while maintaining the flavor consistency of *doenjang* [23]. 64

65 Gochujang

Gochujang is a traditional Korean fermented paste made from *meju*, red pepper powder, and
glutinous rice. The characteristic flavor of *gochujang* is a combination of hot taste from red
pepper, sweet taste from sugars, umami taste from amino acids, and salty taste from NaCl. *Gochujang* is fermented with *Aspergillus* spp., *Bacillus* spp. and *Rhizopus* spp. for several years
in large earthen pots by mixing glutinous rice powder, salt, and red pepper powder with *meju*powder (Table 1) [24].

72 Hawaijar

73 Hawaijar is a fermented soybean product in Manipur, India, with a distinct flavour and 74 stickiness [25]. It is consumed commonly in the local diet as a staple source of high protein food [26]. Whole soybeans are soaked overnight, washed and boiled. After draining excess 75 76 water, the cooked soybeans are washed with hot water, wrapped with cotton cloth or banana 77 leaves and packed tightly in a basket with a lid. The basket is wrapped with cloth and kept in 78 the sun, near a stove, or buried in paddy straw for fermentation for 4 to 5 days (Table 1). The 79 final product has a pH of 8.0 to 8.2 and is of brown colour with a sticky slimy white appearance 80 and a light ammonia odour [26,27]. In hawaijar fermentation, no starter culture is added during 81 its preparation. Bacillus strains that dominate the fermentation originate from raw soybeans 82 [11] or are acquired from other materials used for fermentation [27].

83 Sufu

84 Sufu is a Chinese fermented made from cubes of soybean curd (tofu) by mold ripening [28]. 85 Four steps are involved in *sufu*-making; preparing tofu from boiled soymilk; brining; preparing 86 *pehtze* (soybean curd overgrown with fungal mycelium) with a fungal starter culture; salting; 87 and ripening for 2-3 months in a dressing mixture. During brining, the tofu adsorbs the salt 88 until the salt content of tofu reaches about 6.5 %, which takes about 2 days. Pehtze is prepared 89 by inoculating pure culture of *Bacillus* spp. or *Micrococcus* spp. to tofu and incubated at 30-90 38 °C for about 1 week. Pehtze is dried at 50-60 °C for 12 h prior to subsequent fermentation 91 in the dressing mixture. The most common types are mold-fermented sufu, and bacteria-92 fermented sufu, which includes Bacillus and/or Micrococcus as major members of fermentation 93 microbiota (Table 1) [29].

94 **Roots, tubers, and vanilla fermentation products**

95 Tubers including cassava, yams and potatoes are second only to cereals in importance as a 96 global source of carbohydrates. In most African and Asian countries, the traditional diets of a majority of people rely largely on cassava and yam, which are poor in other nutrients, 97 particularly proteins, essential amino acids, vitamins and minerals, but contain significant 98 99 amounts of starch and dietary fiber [30]. Cassava also contains the cyanogenic glycosides 100 linamarin and lotaustralin that release cyanide during digestion unless the β-glucosidic bonds 101 are hydrolysed during food processing [31]. An overview on fermented tubers and the 102 organisms that occur in the fermentation is provided in Table 1; several examples are presented 103 in more detail below.

104 Ntoba Mbodi

105 Ntoba Mbodi is a popular alkaline fermented food in Congo [31]. It is made by fermenting 106 cassava leaves in the following way: the leaves are harvested, wilted for 2-3 days, cleaned, cut 107 into small pieces, wrapped in small portions into large leaves, and allowed to ferment at 108 ambient temperature for 2-4 days (Table 1). Fermentation also eliminates cyanogenic 109 glucosides [31,32]. A rise in pH of up to 10 is observed during the process. The main 110 microorganisms responsible for the fermentation are *Bacillus* spp. [31].

111 Cassava tape

112 Cassava tape (fermented cassava) is an Indonesian traditional food made by fermentation of 113 cassava. It is made from steamed cassava mixed with a starter commonly known as "ragi tape". 114 In general, the cleaned and peeled cassava tubers are steamed, cooled, and placed in basket. 115 Powdered ragi is sprinkled over the cassava. The cassava is covered with banana leaves and 116 incubated at room temperature for 2-3 d (Table 2.1). The quality of cassava tape depends on 117 many conditions, including the quality of cassava, preparation method, and microbes [33]. Several *Bacillus* spp. including *B. subtilis*, *B. amyloliquefaciens* and *B. thuringiensis* have been
isolated from cassava tapes [33].

120 *Elubo*

121 Yams (Dioscorea spp.) are processed and fermented to the traditional West African dried yam flour elubo. Yam slices are blanched at for 10 min 60 °C and fermented for 24 h at 30 °C. 122 123 During the first 24 h of spontaneous fermentation, the microbial population grows as the pH 124 falls from 6.2 to 5.4. Back-slopping at a rate of 10% (w/v) is used to accelerate natural lactic 125 fermentation by using the preceding fermentation batch as an inoculum (Table 1). Fermentation 126 experiment of blanched samples with pure cultures of the isolates indicated that 127 Lactiplantibacillus plantarum, Levilactobacillus brevis and B. subtilis are the main species in 128 the reconstituted flour paste [34].

129 *Taruba*

Indigenous populations in the Amazon region of South America produce fermented cassava
beverages for daily consumption from cassava alone, or from cassava with addition of corn or
potatoes. Cassava roots are generally washed and soaked, followed by crushing and sieving,
cooking or toasting, and fermentation for one or several days. Bacilli including *B. subtilis* and *B. amyloliquefaciens* were consistently identified as members of fermentation microbiota,
which also includes *Lp. plantarum*, *Lv. brevis* and pediococci [35,36].

136 Vanilla

Vanilla flavoring obtained from cured *Vanilla planifolia* (Andrews) beans is widely used in food, beverages, and cosmetics [37]. The characteristic vanilla flavor is formed during a curing process that yields the character impact compound vanillin. The conventional curing processes involves four steps: blanching, sweating, drying, and conditioning. Mature fresh vanilla beans are blanched and then stored at high humidity and temperature. Sweating retains a sufficiently high moisture content for the enzyme-catalyzed reaction. The vanilla beans are dried by sun or air to inhibit mold growth and stored in a closed box for several months for formation of vanillin [38,39]. Thermophilic bacilli develop during sweating (Table 1) [40]. *Bacillus* isolated from vanilla beans produced β-D-glucosidase, which hydrolyses glucovanillin to vanillin [39].

146 Cereal fermented product

147 **Daqu**

148 Daqu is a spontaneous solid-state cereal fermentation that is used as a saccharification starter 149 to initiate the alcoholic mash fermentation for production of cereal liquors and vinegars in 150 China (Figure 3) [41,42]. The liquor starter is prepared with different grains including wheat, 151 rice, sorghum, that are shaped in blocks and fermented with controlled temperature and 152 humidity [43]. The bacterial microbiota composition of *daqu* is diverse and includes *Bacillus* 153 spp., Enterobacteriaceae and LAB (Table 1) [4,44]. Fungal organisms include Aspergillus spp., 154 *Mucor* spp., and *Penicillium* spp. [44,45]; the most frequently isolated yeasts belong to the 155 genus Saccharomyces [44]. In China, daqu is also used as a starter culture for sourdough 156 fermentation but bacilli were not identified as members of microbiota of the corresponding 157 sourdough [46].

158 Fish sauce

Fish sauce is a condiment that is traditionally consumed in Southeast Asia but becomes increasingly popular in other places of the world [47]. It is produced by mixing fish, such as anchovies, with salts and fermenting for 6 to 12 months at room temperature [48]. Fish proteins are hydrolyzed by proteases from the fish and from halotolerant/halophile microorganisms, resulting in umami-tasting peptides and amino acids [49,50]. The fermentation organisms in fish sauces predominantly include halophilic or salt tolerant bacteria including *Bacillus* species (Table 1) [51,52].

166 The role of *Bacillus* in food fermentation

Bacillus species have a variety of desirable characteristics, including their ability to form endospores, and several strains or species have been awarded GRAS (Generally Regarded as Safe) status by the Food and Drug Administration [53] and QPS (Qualified Presumption of Safety) by the European Food Safety Authority [54]. Figure 4 provides an overview on the presence of desirable or undesirable metabolic traits in type strains of the genus *Bacillus*. The role of *Bacillus* as source of industrial enzymes has been well reviewed elsewhere [55,56]. In this section, the role of enzymes produced by *Bacillus* during food fermentations are discussed.

174 **Role of amylases**

175 The amylolytic system of *Bacillus* species includes glucan branching enzymes, extracellular 176 amylases including α -amylases, β -amylases, pullulanases and glucoamylases, and intracellular 177 oligosaccharide hydrolases. Most type strains of the *B. cereus* and *B. subtilis* groups produce 178 multiple amylases (Figure 4). Yams and cassava have a low content of fermentable 179 carbohydrates and low amylase activity [57]. Therefore, extracellular amylases produced by 180 *Bacillus* spp. are particularly important for the hydrolysis of starch [58].

181 Thermostable amylases of *Bacillus* are also a key element of *daqu* fermentations [59]. Strains 182 of the *B. subtilis* group that are present in *daqu* fermentations have a high starch degrading 183 ability including thermotolerant α -amylase activities [53]. Amylase activity in *daqu* was 184 significantly increased by inoculation with *B. licheniformis* [59]. Amylases produced by 185 *B. licheniformis* in *daqu* fermentation yielded maltose, maltotriose, and maltodextrins as major 186 products from starch [60].

187 Role of β-glucosidases, phytase and α-galactosidase

β-Glucosidase hydrolyzes β-glucosidic linkages between carbohydrate residues in aryl-aminoor alkyl-β-D-glucosides, cyanogenic glucosides, short chain oligosaccharides and

disaccharides under different physiological conditions [61]. The gene that produce BglA isencoded by most type strains of the *B. subtilis* group (Figure 4).

Isoflavones are abundant in soybeans and have various health benefits related to their
oestrogenic activities [62,63]. They have bioactivity only when the glycosides that are present
in soybeans are hydrolysed to the isoflavone aglycones [62,63]. During the fermentation of soy
with *Bacillus*, glucoside conjugates of isoflavones are converted into aglycones by *Bacillus*β-glucosidase [64].

197 The β-glucosidase of *Bacillus* species also hydrolyzes cyanogenic glycosides in cassava. 198 Linamarin accounts for 80 % of the cyanide content of cassava and is known to cause severe 199 disease upon continual consumption [65]. β-Glucosidase activity of bacilli detoxifies cassava 200 cyanides without compromising other nutrients [66].

Vanillin is the main aromatic constituent in cured vanilla pods [67]. In fresh vanilla beans, vanillin is present as glucovanillin, an odorless β -D-glucoside of vanillin. One of the most important aspects of curing is the conversion of glucovanillin to vanillin by β -glucosidase [68]. *Bacillus* in vanilla fermentation produce β -glucosidase which hydrolyse glucovanillin [39]. Its formation may be due to the action of thermo-tolerant bacteria such as *B. subtilis* because of their dominance in vanilla curing up to 65 °C.

Fermentation is also considered a necessary and accepted method to reduce the anti-nutritive phytic acid and raffinose family oligosaccharides in pulses (73). Most of total seed phosphorus (P) in legumes and cereal grains is present as phytate, which chelates minerals and decrease their bioavailability to humans and monogastric animals (74). Most fermented soy products involve a cooking step prior to fermentation. This cooking step inactivates plant enzymes and phytate hydrolysis during fermentation is dependent on microbial enzymes. Species of the *B. subtilis* group are effective producers of phytases, which hydrolyze phytate and increase the bioavailability of minerals in cereals and legumes (Figure 4) (75).

Raffinose family oligosaccharides (RFOs), i.e., raffinose, stachyose, verbascose, are abundant in soybean and legumes. They are composed of one or more galactose residues joined by α -(1 \rightarrow 6)-glycosidic bonds to sucrose. These indigestible carbohydrates can lead to flatulence and other gastrointestinal disorders [69]. α -Galactosidases degrade RFOs and improve the nutritional value of soy and legumes [70]. *Bacillus* species, particularly species of the *B. subtilis* group also encode for α -galactosidases, which hydrolyse raffinose-family oligosaccharides in soybean and legumes (Figure 4).

222 Role of proteases

Type strains of the *B. subtilis* group produce multiple extracellular proteinases (Figure 4). Because of their broad pH and temperature activity and stability range, proteases from *Bacillus* play an essential role in soybean and fish fermentations as they are tolerant to alkaline environments and high temperature.

227 Proteases are secreted by B. subtilis during food fermentations. Degradation of proteins to 228 peptides and amino acids impacts the texture and flavor of the products, may generate bioactive 229 peptides, and provides precursor amino acids for conversion to flavor volatiles [1,71–73]. 230 Fermented fish sauces are particularly rich in soluble taste-active peptides, and amino acids 231 [49,50]. They are produced during proteolytic degradation by proteases in the muscles or 232 digestive tracts of fish, and various microorganisms existing in the fermentation broth [74]. 233 Proteases produced by *Bacillus* play an essential role in the degradation process. A halotolerant 234 protease produced by *B. licheniformis* has been identified in Thai fish sauce with a high 235 enzymatic activity, even under high salt conditions (30 % NaCl) [48].

236 **Role of polyglutamate and glutaminase**

Glutaminase of *Bacillus* convert glutamine to the umami tastant glutamate. Glutaminases of *Bacillus* spp. are tolerant to high temperature and high salt conditions; at alkaline conditions,

glucaminases of *Bacillus* also synthesize kokumi-active γ -glutamylpeptides [71,75,76].

Glutamate accumulates in the food matrix and contributes to the umami taste but is also converted to poly- γ -glutamic acid (γ -PGA). PGA is synthesized by polymerizing D- and L-

glutamate to an anionic biopolymer comprised of only glutamic acid residues. It is water soluble, biodegradable, and has good thickening capacity and excellent absorbability [77]. γ -PGA is produced by *Bacillus* spp. to form part of the mucilage of *natto* and *cheonggukjang* (Figure 4) [78,79]. The stringy consistency of *natto* is mainly due to γ -PGA produced by *B*. *subtilis* [79]. The production of γ -PGA from *natto* starters were reduced in medium containing NaCl greater than 3 % [80]. A salt tolerant *B. subtilis* strain was found in a *cheonggukjang*

248 fermentation, which also produces γ -PGA [78].

249 Role of antimicrobial lipopeptides

239

Bacillus species produce lipopeptides, including surfactins, fengycins and iturins [81,82] with a broad-spectrum antimicrobial activity [82,83]. The non-ribosomal peptide synthases that synthesize these lipopeptides are present almost exclusively in strains of the *B. subtilis* group

253 (Figure 4).

Surfactins show antibacterial activity but, with some exceptions, no marked antifungal activity [81]. Iturins display a strong antifungal action against a wide variety of yeast and fungi but only limited antibacterial activity. For example, iturin from *B. pumilus* HY1 inhibited *A. flavus* and *A. parasiticus* with an minimum inhibitory concentration of 50 mg/L [84]. This fungitoxicity of iturins has been attributed to membrane permeabilization [82]. Fengycins also have a strong antifungal activity, specifically against filamentous fungi [81]. *In vitro* studies indicate that the ability of *B. subtilis* inhibiting aflatoxin-producing fungi was substantially
greater than the inhibitory effect of lactic acid bacteria [85]. Lipopeptides produced by *Bacillus*were detected in *cheonggukjang*, *daqu*, and other solid-state fermentations [86–88]. *In situ*antifungal activity of lipopeptides produced by *B. velezensis* and *B. amyloliquefaciens* was
demonstrated in model *daqu* fermentations [89]

265 Safety and health benefits of *Bacillus* species

266 Assessment of the safety of *Bacillus* species and their toxins

267 Bacillus species are also of concern for food safety. Toxins are produced by certain strains of 268 the B. cereus group (Figure 4). The B. cereus group includes B. cereus, B. anthracis, 269 B. thuringiensis, B. mycoides, which includes strains previously designated as 270 B. weihenstephanensis, B. pseudomycoides, B. cytotoxicus and B. toyonensis (Figure 4) [90]. 271 Toxins produced by these organisms such as non-haemolytic enterotoxin (*nhe*); haemolysin 272 BL (hbl); cytotoxin K (cytK), cereulide, and the lethal anthrax toxin (Figure 4). The 273 lipopeptides surfactin and lichenysin are biosurfactants with *in vitro* hemolytic and cytolytic 274 activity [82] but are not known to contribute to human (foodborne) disease [90], and are not 275 produced by pathogenic Bacillus species (Figure 4). The non-haemolytic enterotoxin, 276 haemolysin BL, and single protein cytotoxin K have been linked to the diarrheal type of B. 277 cereus food poisoning [91]. Nearly all B. cereus strains harbor the nhe genes, while hbl and 278 *cytK* are detected in about 30-70 % of isolates. The emetic toxin cereulide is produced is 279 produced by a non-ribosomal peptide synthase, which is present only in *B. cereus* and related 280 species (Figure 4) [92]. Cereulide produced in food results in rapid onset of vomiting after 281 ingestion but is rarely lethal. B. anthracis is the causative agent of anthrax, an acute lethal 282 disease in animals and in humans [93]. Bipartite exotoxins produced by B. anthracis include 283 protective antigen-lethal factor (PA-LF) and PA-edema factor (PA-EF) [90].

Because *Bacillus* strains other than strains of the *B. cereus* group rarely produce toxins, bacilli were included in the QPS list, with the qualification of "absence of toxigenic potential" [54,94]. Figure 4 documents that the potential for toxin production is closely linked to phylogeny.

287 Use of *Bacillus* species as probiotics

288 Probiotics are defined as "live microorganisms which confer a health benefit on the host when 289 administered in adequate amounts" [95]. The role of LAB as animal and human probiotics is 290 well documented [96,97]. Endospores of Bacillus species have been proposed as heat resistant 291 alternatives [98] and their probiotic activity is increasingly documented in clinical trials [99]. 292 Several clinical trials support claims that *Bacillus* probiotics reduce chronic gastrointestinal 293 symptoms, and increase the resistance to infections [99]. The occurrence of pathogenic strains 294 or species in the genus *Bacillus*, however, impedes consumer acceptance of *Bacillus* probiotics 295 [99]. Moreover, it remains unclear whether *Bacillus* fermented foods deliver a sufficient dose 296 to viable bacterial cells or spores to exert probiotic effects [100].

297 Conclusions

298 Bacillus species are an under-appreciated group of bacteria that are prevalent in numerous food 299 fermentations, particularly in Asia and Africa. Strains of the Bacillus subtilis group produce 300 extracellular hydrolytic enzymes, potent antifungal lipopeptides and extracellular polymeric 301 compounds. These properties that are only rarely found in other dominant groups of food 302 fermenting bacteria such as lactic acid bacteria, acetic acid bacteria, or propionic acid bacteria. 303 Toxigenic bacilli and those species that possess functional traits cluster in two separate 304 phylogenetic groups. With the better understanding and further study of Bacillus, the 305 fundamentally positive characteristics of the Bacillus species will make them the preferred 306 starter cultures for many novel and improved fermented foods.

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662 **Figure legends**

Figure 1. A simplified view of the lifestyle of *Bacillus* in different environmental niches. 663 664 Mutualistic interactions of *Bacillus* with plants or humans are highlighted in red letters; pathogenic interactions are in blue letters. Soil is the largest reservoir of Bacillus. Bacillus spp. 665 666 form stable symbiotic associations with plants and occur in the rhizosphere or as endophytes 667 [12,13]. The plants provide nutrients to *Bacillus*; in turn, *Bacillus* strains help the plant defend 668 against harmful microbes by production of antimicrobial peptides [101]. B. thuringiensis 669 additionally produces an insecticidal spore protein, which is used commercially as biopesticide 670 [102]. Bacillus spp. are generally present in plant foods and may cause food spoilage [14] or 671 improves food quality as a food fermenting organism [3,17]. In vertebrate hosts, pathogenic 672 Bacillus species of the B. cereus group cause emetic or diarrheal illness cause illness or death 673 [90], whereas some non-pathogenic Bacillus spp. are used as probiotics to promote host health 674 [99].

Figure 2. Geographic location of *Bacillus* species associated food fermentation. Panel A,
cereal fermented foods; Panel B, fermented soybeans and legumes; Panel C, fermented
cassava or yams. The maps were generated with a template file from www.freeworldmaps.net.

Figure 3. Flow chart of traditional Chinese liquor brewing process including the process of the *daqu* production (left) and the liquor production (right). The stacking fermentation is used for
production of few but not all liquors [103,104].

Figure 4 Core genome phylogenetic tree of type strains in the genus *Bacillus* (left) and genes coding for useful enzymes, or toxins, in these type strains of *Bacillus* spp. (right). Genome sequences of type strains of the genus *Bacillus* were downloaded from NCBI Genome database (Supplementary Table S1) using NCBI-genome-download (version 0.3.1). Gene annotion was performed by Prokka (version 1.14.6) in PGCGAP module (version 1.0.20) to obtain the GFF format annotion. The pan and core protein clusters of all these genomes were constructed using FastOrtho. The phylogenetic tree of all *Bacillus* type strains was established by IQ-TREE(version 2.0.3) with the model selection of MFP.

Query protein sequences were downloaded from UniprotKB database according to the Entry ID and searched against the Prokka-annotated translations of *Bacillus* genomes by BLASTP with an e-value cutoff of 10⁻⁵. Futhermore, shell script was used to screen the blastp results with identiy more than 0.60 and R script was used to create the heatmap matrix. Finally, the phylogenetic tree and the blastp results were depicted in iTOL.

Shown in different colors are (from left to right): the amylolytic system, other glycosyl
hydrolases, glutaminases, polyglutamate synthesis, proteases, antimicrobial lipopeptides,
enterotoxins of *B. cereus*, and the anthrax toxin.

Amylolytic system: GlgB (B7JDF1), branching enzyme; AmyE (P00691), α-amylase; AmyS
(P06279), heat stable α-amylase; BbmA (O06988), β-amylase; SpoII (P36924), β-amylase;
AmyX (C0SPA0), pullulanase or debranching enzyme; MdxE (WP_163131464), maltose
binding proteins.

Other glycosyl hydrolases: BglA (P22073), β-glucosidase; GanA and GanB (O07012 and
O07013), exo- and endo-β-galactosidases/β-galactanases; YesZ (O31529), β-galactosidase;
LplD (P39130), α-galactosidase; AguA (Q09LY5), thermostable xylanase; PhyC (O31097),
phytase.

705 **Glutaminases:** GlsA1 and GlsA2 (O31465 and O07637), glutaminases.

Polyglutamate synthesis: PghL (A0A6M3ZBI1), poly-γ-glutamate hydrolase, PgsABCR
(A0A6M3ZGL0, E0U4Z3, A0A6M4JPT9, and Q45449), poly-γ-glutamate synthetase and
regulatory genes.

Proteolytic system: HtrA and HtrB (O34358 and Q9R9I1), envelope-associated serine
proteases; YyxA (P39668), uncharacterized serine protease.

711 Antimicrobial lipopeptides: FenABCDE (A0A7G7U8D7, H9TE69, A0A7G7U8D5,

A0A7T5SHH4 and A0A7G7U8D6), fengycin synthase; ItuABCD (Q93I56, Q93I55, Q93I54

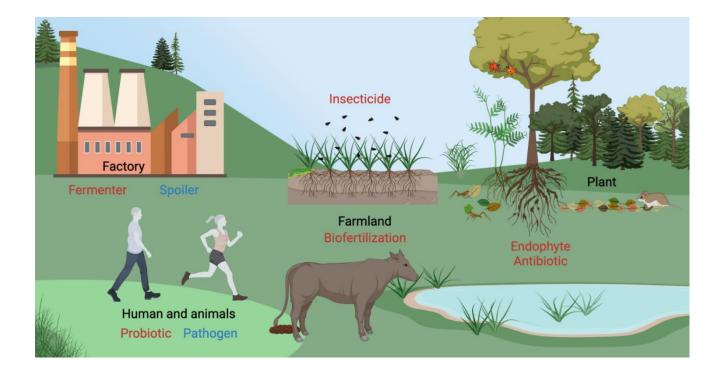
and X5F8R7), iturin synthase; MycABC (Q9R9J1, Q9R9J0 and Q9R9I9), mycosubtilin

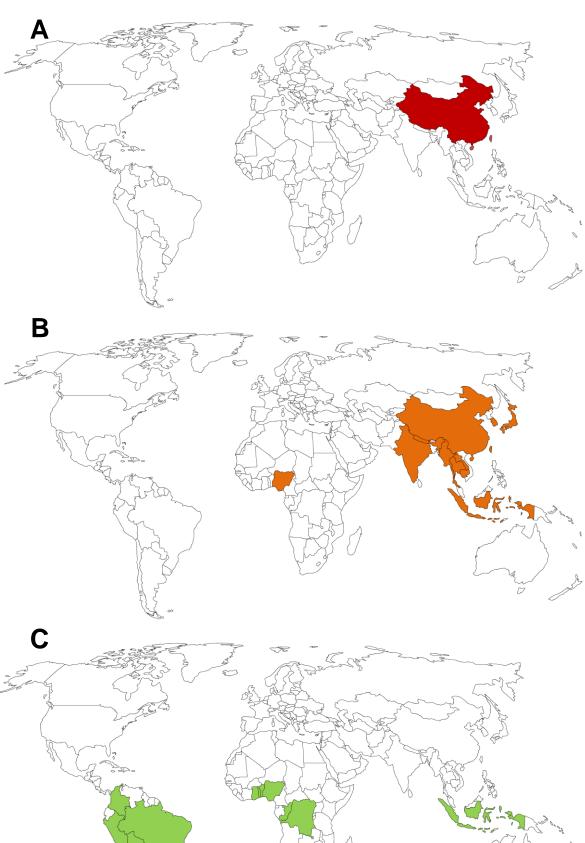
- 714 synthase; PpsBDE (A0A6H2JR43, A0A6H2JR81 and O31827), plipastatin synthase (doi:
- 715 10.6026/97320630007384); SrfAA SrfAB, SrfAC and SrfAD (P27206, Q04747, Q08787 and
- 716 Q08788), surfactin synthase; *sfp* (P39135), surfactin synthetase-activating enzyme.
- 717 Toxins: CesH (Q20CJ2), cereulide synthetase; CytK (Q63EQ2), cytotoxin K; HblACD
- 718 (A0A2A8M4Y0, Q9L4L8 and Q9REG7), Hemolysin BL; NheABC (A0A8F1XQI7,
- 719 A0A8I1GLU0 and A0A8F1XT31), Nhe enterotoxin; Anthrax toxin (TYC44981).
- The genome sequences of type strains that were used to generate this heatmap is provided as
- 721 Online Supplementary Table 1.

Product	Substrate	Main microorganisms	Ingredients and main processing steps. Fermentations are spontaneous fermentations unless back-slopping or use of starter culture is indicated.	Region of origin; food use
		Soy	beans or soy protein and legumes	
Natto	Soybeans	B. subtilis var. natto	Soybeans or pulses are soaked and steamed, inoculated with spores of <i>B. subtilis</i> var. <i>natto</i> and fermented for 15-24 h [20].	Japan; main course as meat substitute
Cheonggukjang	Soybeans, local pulses	B. subtilis, B. amyloliquefaciens and Rhizopus oligosporus	Soybeans or pulses are soaked and steamed, inoculated with spores of <i>Bacillus</i> and fermented for 1-3 d [22].	Korea; main course as meat substitute
Doenjang	Soybeans, local pulses	B. subtilis, Rhizopus spp., and Aspergillus spp.	<i>Meju</i> is made by soaking, steaming and crushing pulses and fermentation for 2 to 3 months. The solid fraction of <i>meju</i> is used for further ripening for over 2 months to make <i>doenjang</i> [23].	Korea; condiment
Gochujang	Soybeans, rice, pepper	Bacillus spp., including B. subtilis, B. amyloliquefaciens; Aspergillus spp.; and Rhizopus spp.	The ingredients, 25 % red pepper powder, 22.2 % glutinous rice, 5.5 % <i>meju</i> , 12.8 % salt, 5 % malt and 29 % water, are fermented for 6 months to 1 year [24].	Korea; condiment
Hawaijar	Whole soybeans	B. subtilis, B. licheniformis, S. sciuri, Alkaligenes spp., Providencia rettgeri	The soybeans are soaked, boiled, washed, wrapped with clean cotton cloth/healthy leaves, and packed tightly in a bamboo basket with a lid. The basket is kept warm for fermentation for 4-5 d [27].	India up (Asia); staple
Sufu	Tofu (soybean curd)	Bacillus spp. or Micrococcus spp.	Sufu is produced in four steps: preparation of tofu; brining, inoculation with a fungal starter culture to prepare <i>pehtze</i> ; and ripening in dressing mixture [29].	China up (Asia); side dish
Dawadawa	Locust bean and local pulses	Bacillus spp., including B. subtilis, B. pumilus, B. licheniformis and B. subtilis var. natto	The locust beans are cleaned, boiled, pounded and separated the seed coat from the cotyledons. The cotyledons are re-boiled and packed into baskets or perforated pots and allowed to ferment spontaneously for about 48 h. Before fermentation, ash, maize, or millet flour is sprinkled on the cotyledons [105].	West and central Africa; meat substitute
Ugba	Locust bean and local pulses	Bacillus spp., including B. subtilis, B. licheniformis, B. megaterium, B. pumilus; Staphylococcus spp.; and Micrococcus spp.	Soybeans or pulses are boiled, the cotyledons are sliced off. The sliced cotyledons are washed in water, soaked overnight and fermented for 3 to 5 d [106].	Nigeria; side dish
			Tubers and roots	
Ntoba Mbodi	Cassava leaves	 Bacillus spp., including B. subtilis, B. licheniformis, B. amyloliquefaciens, B. pumilis, B. sphaericus, and B. xylanilyticus 	Cassava leaves are wilted for 2-3 days, cleaned, cut into small pieces, washed with water, distributed into small portions, and wrapped in large leaves, and allowed to ferment at ambient temperature for 2-4 d [32,107].	Congo

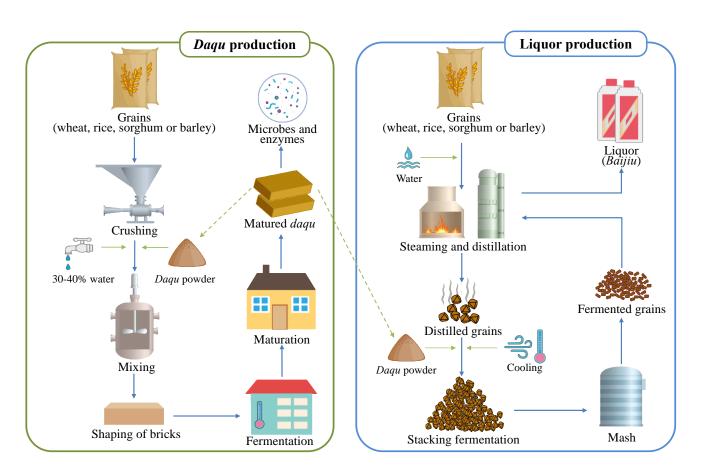
Table 1. Bacillus species associated fermented foods. With exception of fish sauces, all fermentations are solid state fermentations.

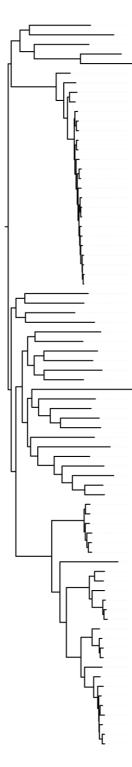
Product	Substrate	Main microorganisms	Ingredients and main processing steps. Fermentations are spontaneous fermentations unless back-slopping or use of starter culture is indicated.	Region of origin; food use
Agbelima	Cassava	Bacillus spp., including B. subtilis, B. licheniformis and B. pumilus; LAB; and yeasts.	Cassava root are peeled, grated and mixed with a traditional inoculum which is also prepared from cassava roots. The grated mash is packed into plastic sacks and fermented with weights placed on top of the sacks to dewater the mash [108,109].	West Africa; staple
Tape	Cassava	Bacillus spp., including B. subtilis, B. amyloliquefacie, and B. thuringiensis	Cassava roots are cleaned, peeled steamed, cooled, and placed in a basket. Powdered ragi is sprinkled over the cassava. The cassava is covered with banana leaves and incubated at room temperature for 2-3 d [33].	Indonesia; dessert or ingredient for baking
Elubo (yams)	Yams	Lactiplantibacillus, Levilactobacillus, B. subtilis	Yam slices are blanched at 60 °C for 10 min and then fermented at 30 °C for 24 h [34].	West African, staple
Taruba	Cassava	L. plantarum, L. brevis and B. amyloliquefaciens	Cassava is crushed and pressed, toasted for 30 min and fermented at ambient temperature for 12 d [35,36].	Amazon, beverage
			Vanilla	
Vanilla	Seed pods of Vanilla planifolia	Bacillus spp., including B. subtilis, B. fusiformis, and B. pumilus	Mature vanilla beans are blanched by immersing in hot water for 3-5 min to destroy the cell tissue structure. Then, the blanching vanilla beans are treated under conditions of high humidity and temperature. The sweating vanilla beans are sun- or air dried to inhibit mold growth and stored in a closed box for few months [38,39].	Madagascar, Indonesia, Mexico, others; condiment
			Cereals	
Daqu	Grains (wheat, rice, sorghum and barley)	Bacillus spp., including B. subtilis, B. amyloliquefaciens, B. velezensis, and B. licheniformis; LAB; Enterobacteriaceae; Aspergillus spp.; Rhizopus spp.; Saccharomyces; and Saccharomycopsis	Grains are ground and mixed with water to ~35 %. The mixture is shaped to bricks and then fermented for 2 months and matured for 2 months [41,42].	China; starter culture for liquor and vinegar
			Fish	
Fish sauce	Fish	Filobacillus, Bacillus, Micrococcus, Virgibacillus, Halobacillus, Halococcus; Tetragenococcus halophilus	Fish are mixed with salts and fermented for 6 to 12 months at room temperature [51,52].	Southeast Asia up; condiment











Bacillus solimangrovi Bacillus marinisedimentorum Bacillus taeanensis Bacillus shivajii Bacillus tamaricis Bacillus manliponensis Bacillus cytotoxicus Bacillus gaemokensis Bacillus pseudomycoides Bacillus paramycoides Bacillus hominis Bacillus mycoides Bacillus nitratireducens Bacillus proteolyticus Bacillus toyonensis Bacillus cereus Bacillus thuringiensis Bacillus luti Bacillus wiedmannii Bacillus paramobilis Bacillus mobilis Bacillus sanguinis Bacillus albus Bacillus fungorum Bacillus anthracis Bacillus tropicus Bacillus pacificus Bacillus paranthracis Bacillus mesophilus Bacillus tianshenii Bacillus sinesaloumensis Bacillus salinus Bacillus oleivorans Bacillus tepidiphilus Bacillus benzoevorans Bacillus dielmonensis Bacillus aquiflavi Bacillus methanolicus Bacillus ndiopicus Bacillus coahuilensis Bacillus pakistanensis Bacillus enclensis Bacillus salacetis Bacillus acidicola Bacillus smithii Bacillus ectoiniformans Bacillus xiapuensis Bacillus thermotolerans Bacillus wudalianchiensis Bacillus badius Bacillus altitudinis Bacillus xiamenensis Bacillus zhangzhouensis Bacillus pumilus Bacillus safensis Bacillus australimaris Bacillus gobiensis Bacillus glycinifermentans Bacillus sonorensis Bacillus swezeyi Bacillus licheniformis Bacillus haynesii Bacillus paralicheniformis Bacillus nakamurai Bacillus velezensis Bacillus siamensis Bacillus amyloliquefaciens Bacillus atrophaeus Bacillus halotolerans Bacillus vallismortis Bacillus spizizenii Bacillus cabrialesii Bacillus tequilensis Bacillus inaquosorun Bacillus subtilis Bacillus stercoris

