

An oxidation–amidation approach for the synthesis of glycuronamides

Sicheng Lin,^{+[a]} Roger A. Ashmus,^{+[a]} and Todd L. Lowary^{*[a]}

Abstract: A route for the synthesis of glycuronamides via the intermediacy of 6-S-tolyl-glycosides and uronic acid thioesters, is reported. The route, which is compatible with a variety of carbohydrate residues and protecting groups, was used to synthesize the repeating unit of the *E. coli* K54 capsular polysaccharide.

Introduction

Glycuronamides, amidated uronic acid derivatives, are commonly found in bacterial lipopolysaccharides (LPS) and capsular polysaccharides (CPS).^[1–5] These structures frequently consist of glucuronic acid, galacturonic acid, mannuronic acid, or 2-acetamido-2-deoxy-D-galacturonic acid amidated with a range of different amines including ammonia,^[1] ethanolamine,^[6] L-alanine,^[2] L-serine,^[3] L-lysine,^[7] L-threonine,^[4] and serinol.^[5] Examples include *Escherichia coli* K54 CPS, which has a repeating unit possessing a L-threonine-containing glycuronamide^[9] and *Proteus mirabilis* O28 LPS, which contains two galacturonamide residues, one incorporating L-lysine and the other L-serine (Figure 1).^[7] Polysaccharides containing glycuronamide residues play essential roles in the organisms that produce them. For instance, the L-threonine-containing galacturonamide in the *Colwellia psychrerythraea* 34H CPS has been suggested to be important for the survival of the organism in subfreezing marine environments.^[4] Other studies have shown that CPS possessing uronic acids amidated with amino acids are important in the induction of protective immunity against abscess formation.^[10] In addition, molecules containing these structures

have often been targeted in the development of vaccines.^[7,11–13] Consequently, the chemical synthesis of glycans possessing glycuronamides is of interest.

The conventional approach for glycuronamide synthesis involves converting a uronic acid to a reactive intermediate (e.g., an activated ester or acyl halide) and then addition of an amine (Scheme 1, **1**→**2**→**6**).^[14–20] Recently, Crich and Sasaki have developed an alternate method in which uronic acid thiocarboxylates made in situ react with isocyanates or isothiocyanates to produce glycuronamides (**1**→**3**→**6**).^[21] In another alternate approach, treatment of 1,6- and 3,6-carbohydrate-based lactones with amines leads to the corresponding glycuronamides (**1**→**4/5**→**6**).^[22–24] These alternative approaches, however have only been demonstrated on glucuronic acid residues. In addition, they are difficult to implement if the glycan of interest contains more than one glycuronamide (e.g., the repeating unit found in *P. mirabilis* O28 LPS, Figure 1).

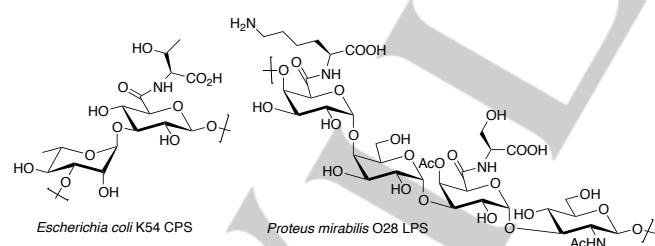
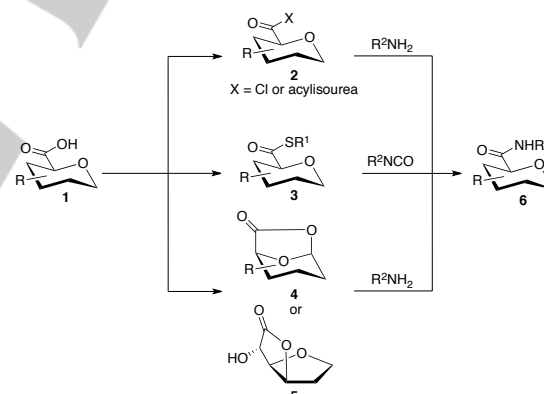


Figure 1. Examples of polysaccharide repeating units containing glycuronamide residues



Scheme 1. Approaches for glycuronamide synthesis.

In recent efforts to synthesize glycoconjugates containing glycuronamide residues using the conventional approach described above (**1**→**2**→**6**), we encountered an unexpected problem: The generation of a glucuronic acid derivative, either by hydrolysis of an ester or oxidation of a primary alcohol, led to decomposition of the molecule. Thus, we required a method for the formation of a glycuronamide that did not involve a carboxylic acid intermediate. Although methods for the direct conversion of primary alcohols to amides have been described,^[25–36] in our hands these were unsuccessful.

We then turned our attention to a report by Yu and coworkers, which described the preparation of uronic acid esters from 6-S-phenyl-glycosides (Scheme 2a).^[37,38] In this approach, the first step involves treatment of the sulfide precursor **7** with

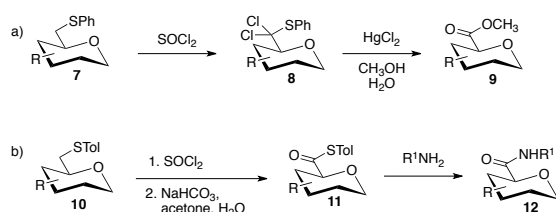
[a] Department of Chemistry and Alberta Glycomics Centre, The University of Alberta
Edmonton, Alberta T6G 2G2, Canada
E-mail: tlowary@ualberta.ca

[+] These authors contributed equally to this work.

Supporting information for this article is given via a link at the end of the document.

FULL PAPER

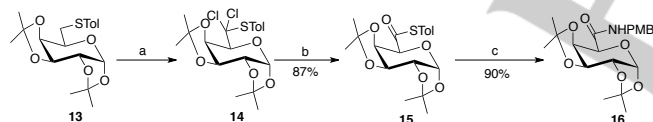
sulfonyl chloride to generate the corresponding α,α -dichlorosulfide (**8**), which in a second step can be converted to the uronic acid methyl ester (**9**). In other previous work, Fortes and coworkers showed that α,α -dichlorosulfides, when treated with water and acetone in the presence of Na_2CO_3 , generate thioesters.^[39,40] These investigations prompted us to explore the possibility of preparing glycuronamides from **10** via the formation of the uronic acid thioesters (e.g., **11**) and then treatment with amines (Scheme 2b).



Scheme 2. a) Synthesis of uronic acid methyl esters by Yu and coworkers.^[37,38] b) proposed route to glycuronamides.

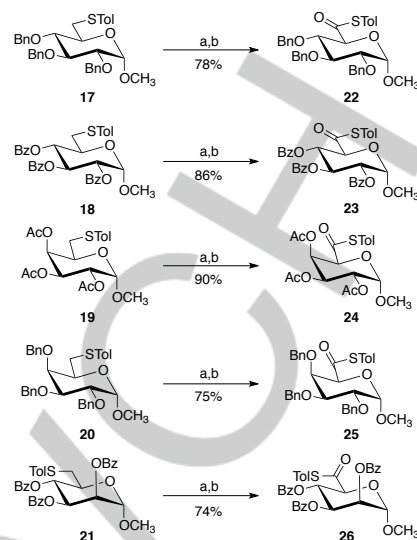
Results and Discussion

In a first series of experiments, we treated 6-S-tolyl-galactose derivative **13** with sulfonyl chloride to produce the dichlorosulfide intermediate **14** (Scheme 3). Without purification, this adduct was stirred with water in acetone in the presence of NaHCO_3 , leading to the formation of thioester **15**, which could be isolated in 87% yield. Subsequent treatment of **15** with *p*-methoxybenzylamine in dichloromethane furnished the glucuronamide **16** in 90% isolated yield.



Scheme 3. Synthesis of galacturonamide **16** from thioether **13**. *Reagents and conditions:* (a) pyridine, SO_2Cl_2 , CCl_4 , 0°C , 5 h; (b) NaHCO_3 , acetone– H_2O (3:2), rt, 18 h; (c) *p*-methoxybenzylamine, CH_2Cl_2 , r.t., 10 h.

Having succeeded in applying this approach to **13**, we then examined the oxidation of various 6-S-tolyl-glycosides (**17–21**)^[41] to the corresponding thioesters (**22–26**) as illustrated in Scheme 4. In each case, the two-step conversion afforded the desired product in good to excellent yield. Neither the electron-withdrawing nature of the protecting groups, nor the carbohydrate stereochemistry had a significant influence on the reaction yield or reaction time. In addition, the reaction could also be carried out both milligram and gram scale without difficulty. With the thioesters in hand, we explored their amidation with various amines (Table 1).



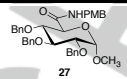

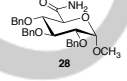
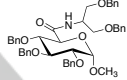
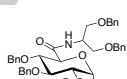
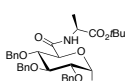
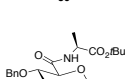
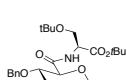
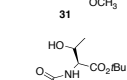
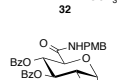
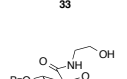
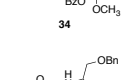
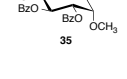
Scheme 4. Synthesis of uronic acid thioesters from 6-S-tolyl-glycosides. *Reagents and conditions:* (a) pyridine, SO_2Cl_2 , CCl_4 , 0°C , 5 h; (b) NaHCO_3 , acetone– H_2O (3:2), r.t., 18 h.

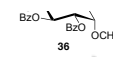
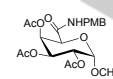
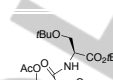
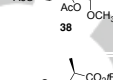
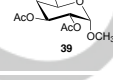
In exploring the scope of the amidation reaction (Table 1), we focused on the preparation of naturally occurring glycuronamides in their protected form. We found that varying the carbohydrate residue and their electronic properties (i.e., the electron withdrawing or donating ability of the protecting groups) did not affect the amidation yield or reaction rate.

When simple amines such as *p*-methoxybenzylamine (Entries 1, 10, 15, 18, 19) and ethanolamine (Entry 11) were used, the amidation proceeded in good yields (> 82%). Ammonia from concentrated ammonium hydroxide could also be used as the amine, albeit this afforded the glycuronamide **28** in somewhat lower (66%) yield and longer reactions times were required (Entry 3). The use of amines derived from L-alanine, L-serine, L-threonine as well as serinol, provided the corresponding glycuronamides in 63–73% yields.

The data in Table 1 also supports a general trend for the reaction rate: simple amines (e.g. *p*-methoxybenzylamine, ethanolamine) > moderately bulky amines (e.g. protected L-alanine derivatives) > more sterically demanding amines (the serinol analogue, L-threonine derivatives, L-serine derivatives). It was discovered that for L-threonine *t*-butyl ester changing the solvent from CH_2Cl_2 (Entry 13) to DMF (Entry 14) resulted in the reaction rate increasing nearly 2-fold and a significantly improved product yield. The same trend is observed with other amino acid-derived amines (Entries 8, 16). In attempts to increase the reaction rate, triethylamine was added (Entries 4, 5); however, no increase in the rate or improvement of the yield was observed. We also explored the use of 2,6-di-*t*-butyl-4-methyl-pyridine, but again no improvement in rate or product yield was seen. Indeed, for substrates possessing acyl protecting groups at O-4, the yield decreased (see next paragraph). It was also evident that the free amine (Entry 7) is a substantially better substrate than the corresponding hydrochloric salt, even in the presence of an excess of triethylamine (Entry 6). Not surprisingly, the data in

Table 1. Amidation of uronic acid thioesters with amines

Entry	Thioester	Amines ^[a]	Solvent	Time	Product	Yield ^[b]
1	22	PMB-NH ₂	CH ₂ Cl ₂	10 h		85%
2	22	PMB-NH ₂	DMF	10 h		82%
3	22	NH ₃ OH (aq.)	DMF	40 h		66%
4	22	1,3-Di-O-benzyl-serinol	CH ₂ Cl ₂	72 h		64%
5	22	1,3-Di-O-benzyl-serinol	CH ₂ Cl ₂	72 h		67% ^[c]
6	22	HCl·H ₂ N-L-Ala-tBu	CH ₂ Cl ₂	24 h		33% ^[c]
7	22	H ₂ N-L-Ala-tBu	CH ₂ Cl ₂	24 h		71%
8	22	H ₂ N-L-Ser(tBu)-tBu	CH ₂ Cl ₂	72 h		65%
9	22	H ₂ N-L-Thr-tBu	CH ₂ Cl ₂	72 h		63%
10	23	PMB-NH ₂	CH ₂ Cl ₂	10 h		86%
11	23	Ethanolamine	CH ₂ Cl ₂	17 h		86%
12	23	Serinol	CH ₂ Cl ₂	40 h		73%
13	23	H ₂ N-L-Thr-tBu	CH ₂ Cl ₂	87 h		41%
14	23	H ₂ N-L-Thr-tBu	DMF	38 h		71%

15	24	PMB-NH ₂	CH ₂ Cl ₂	8 h		91%
16	24	H ₂ N-L-Ser(<i>t</i> Bu)- <i>t</i> Bu	DMF	42 h		64%
17	24	H ₂ N-L-Ala- <i>t</i> Bu	DMF	18 h		63%
18	25	PMB-NH ₂	CH ₂ Cl ₂	10 h		83%
19	26	PMB-NH ₂	CH ₂ Cl ₂	10 h		98%

[a] 1.5 equiv. of amine except NH₄OH (aq.). [b] Yield of isolated product. [c] Triethylamine was added to the reaction.

entries 9, 11, and 13 demonstrate that amidation will occur over esterification when competing alcohol nucleophiles are present.

It is important to note that when O-4 in the carbohydrate is protected with an ester, significant elimination can occur in the presence of an excess amine nucleophile or when triethylamine is added to the reaction (Figure 2). Small amounts of the elimination adduct were also observed when a sterically demanding amine was used in the amidation step. However, changing the solvent from dichloromethane to DMF in the amidation of **23** with L-threonine *t*-butyl ester (Entries 13, 14), led to less elimination. In addition, by carefully controlling the amount of amine in the reaction and avoiding the addition of triethylamine, the formation of the elimination product with O-4 acylated substrates could be largely avoided (e.g., Table 1, Entry 10).

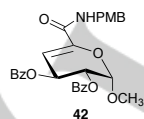
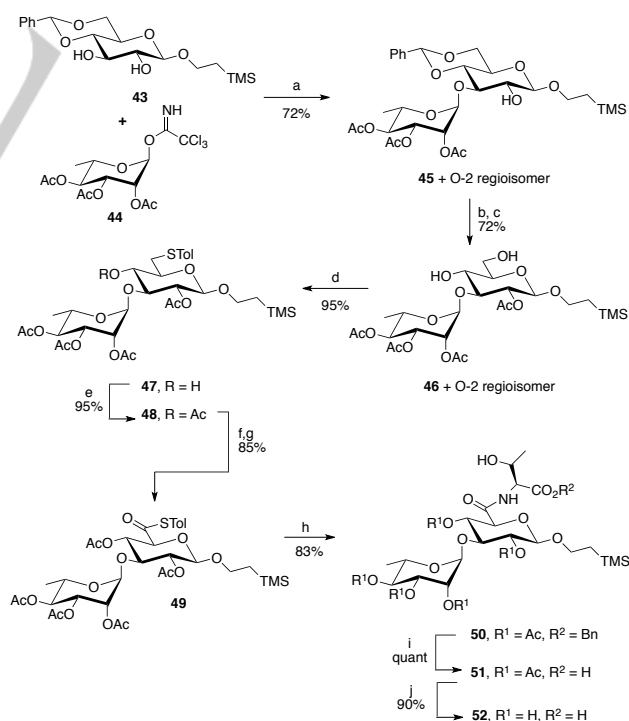


Figure 2. Example of the elimination product formed from O-4 acylated substrates (e.g., **23**) when an triethylamine is used in the amidation reaction.

To demonstrate the applicability of the approach to glycan synthesis, we used it to prepare the repeating unit of the *E. coli* K54 CPS, a disaccharide possessing a glucuronamide residue linked to L-threonine (**52**, Scheme 5).^[9] The synthesis started with a TBSOTf-catalyzed glycosylation of glucosyl acceptor **43**^[42] with rhamnosyl donor **44**,^[43] which led to a 2:1 inseparable mixture of

45 and its O-2 regioisomer in favor of the desired adduct. Subsequent acetylation with acetic anhydride in pyridine and



Scheme 5. Regents and conditions: (a) 4 Å MS, TBSOTf, CH₂Cl₂, -78 °C, 20 min (inseparable, **46**:**46a** = 2:1); (b) Ac₂O, pyridine, DMAP, 3 h; (c) 80% CH₃COOH-H₂O, 80 °C, 2 h (separable, **47**:**47a** = 2:1); (d) (TolS)₂, Me₃P in THF,

FULL PAPER

pyridine, r.t., overnight; (e) Ac₂O, pyridine, DMAP, 3 h; (f) pyridine, SO₂Cl₂, CCl₄, 0 °C, 5 h; (g) NaHCO₃, acetone:H₂O (3:2), r.t., 18 h; (h) L-threonine benzyl ester, CH₂Cl₂, r.t., 44 h; (i) 5% Pd/C, CH₃OH, rt, overnight; (j) 50 mM NaOCH₃-CH₃OH, r.t., 4 h.

removal of the benzylidene acetal afforded disaccharide **46** and its regioisomer **46a** in 34% and 17% yield, respectively, over three steps. Treatment of **46** with tolyl disulfide and trimethylphosphine in pyridine selectively afforded the 6-S-tolyl sulfide **47** in 95% yield. Subsequent acetylation of **47** with acetic anhydride in pyridine afforded an 85% yield of **48**. Oxidation of **48** into thioester **49** proceeded in 85% yield upon treatment with sulfuryl chloride in carbon tetrachloride and then with water in the presence of NaHCO₃. The amidation of thioester **49** with L-threonine benzyl ester afforded glycuronamide **50** in 83% yield. With the disaccharide prepared, it was deprotected by first hydrogenolysis of the benzyl ester over 5% Pd/C, which afforded the free acid **51**. Subsequent treatment of **51** with sodium methoxide in methanol produced the disaccharide **52** in 90% yield.

Conclusions

In conclusion, an effective strategy for the preparation of glycuronamides from 6-S-tolyl-glycosides was developed. The method was compatible with a range of carbohydrate residues and amines and the approach could be easily applied to the synthesis of the *E. coli* K54 CPS repeating unit. To the best of our knowledge, the route described here is the first of its kind to completely avoid a carboxylic acid intermediate in the synthesis of complex glycans containing glycuronamides. Moreover, staged introductions of 6-S-tolyl-moieties into oligosaccharides should allow the preparation of molecules containing more than one uronic acid derivatives amidated to different amines.

Experimental Section

General Experimental Methods: All reagents were purchased from commercial sources and were used without further purification unless noted. Reaction solvents were purified by successive passage through columns of alumina and copper under argon. Unless stated otherwise, all reactions were carried out at room temperature and under a positive pressure of argon and were monitored by TLC on Silica Gel G-25 F254 (0.25 mm). Visualization of the reaction components was achieved using UV fluorescence (254 nm) and/or by charring with acidified anisaldehyde solution in ethanol, acetic acid and sulfuric acid. Organic solvents were evaporated under reduced pressure, and the products were purified by column chromatography on silica gel (230–400 mesh). Optical rotations were measured in a microcell (1 cm, 1 mL) at 21 °C and are in units of degree·mL/(g·dm). ¹H NMR spectra were recorded at 400 MHz or 500 MHz and chemical shifts are referenced to residual CHCl₃ (7.26 ppm, CDCl₃) or CHD₂OD (3.30 ppm, CD₃OD). ¹³C NMR spectra were recorded at 125 MHz, and chemical shifts are referenced to CDCl₃ (77.0 ppm) or CD₃OD (49.3 ppm). Reported splitting patterns are abbreviated as s = singlet, d = doublet, t = triplet, m = multiplet, br = broad, app = apparent. Assignments of NMR spectra were based on two-dimensional experiments (¹H–¹H COSY, HSQC and HMBC). High-resolution ESI-MS spectra (time-of-flight analyzer) were recorded on samples suspended in THF or CH₃OH and with added NaCl.

General Method 1: Preparation of 6-S-tolyl glycosides^[37]

To a stirred solution of the 6-hydroxy glycopyranoside (1.0 mmol) in dry pyridine (5.0 mL) under argon were added (ToIS)₂ (2.0 mmol) and PMe₃ (2.0 mmol, 1.0 M in THF). The reaction mixture was stirred overnight at room temperature. Then, the reaction mixture was concentrated, diluted with EtOAc (30 mL) and washed with brine. The organic layer was dried over MgSO₄, filtered and concentrated.

General Method 2: Preparation of uronic acid thioesters^[39,40]

To a solution of the thioether (1.0 mmol) in dry CCl₄ (15 mL) at 0 °C under argon were added anhydrous pyridine (2.0 mmol) and then SO₂Cl₂ (2.0 mmol). The reaction mixture was stirred for 5 h at 0 °C, then diluted with CH₂Cl₂ and the mixture was washed with brine. The organic layer dried over MgSO₄, filtered and concentrated. The crude residue was dissolved in 3:2 acetone–H₂O (20 mL), and solid NaHCO₃ (4.0 mmol) was added. The reaction mixture was stirred for 18 h at room temperature, diluted with brine, and then extracted with EtOAc (3 × 30 mL). The organic layer was dried over MgSO₄, filtered and concentrated.

General Method 3: Preparation of uronamides.

To a solution of the thioester (0.1 mmol) in CH₂Cl₂ (or DMF) (1 mL) was added the amine (0.15 mmol). The reaction mixture was monitored by TLC until the consumption of starting material and then diluted with CH₂Cl₂ (5 mL), washed with sat. NH₄Cl (aq.) and brine. The organic layer dried over MgSO₄, filtered and concentrated.

1,2:3,4-Di-O-isopropylidene-6-thio-6-S-p-tolyl-α-D-galactopyranose (13): Scale 1.0 mmol. Prepared as described in general method 1. The crude residue was purified by flash chromatography (5:1→4:1 hexane–EtOAc) to afford **13** (330 mg, 90%) as a clear viscous oil. *R*_f 0.39 (4:1 hexane–EtOAc); [α]_D –84.3 (0.4 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 7.33–7.31 (m, 2H, Ar), 7.11–7.09 (m, 2H, Ar), 5.53 (d, 1H, *J* = 5.0 Hz, H-1), 4.61 (dd, 1H, *J* = 7.9, 2.4 Hz, H-3), 4.39 (dd, 1H, *J* = 7.9, 1.8 Hz, H-4), 4.29 (dd, 1H, *J* = 5.0, 2.4 Hz, H-2), 3.83 (td, 1H, *J* = 7.0, 1.7 Hz, H-5), 3.14 (d, 2H, *J* = 7.0 Hz, H-6), 2.31 (s, 3H, ArCH₃), 1.47 (s, 3H, C(CH₃)₂), 1.36 (s, 3H, C(CH₃)₂), 1.30 (s, 3H, C(CH₃)₂), 1.26 (s, 3H, C(CH₃)₂); ¹³C NMR (125 MHz; CDCl₃): δ 136.4 (Ar), 131.9 (Ar), 130.4 (Ar), 129.7 (Ar), 109.3 (C(CH₃)₂), 108.6 (C(CH₃)₂), 96.7 (C-1), 71.3 (C-4), 70.9 (C-3), 70.6 (C-2), 66.2 (C-5), 34.0 (C-6), 26.0 (C(CH₃)₂), 25.6 (C(CH₃)₂), 25.0 (C(CH₃)₂), 24.5 (C(CH₃)₂), 21.0 (ArCH₃); HRMS (ESI) Calc. for (M + Na) C₁₉H₂₆NaO₅S: 389.1393; Found 389.1397.

S-p-Tolyl thio(1,2:3,4-Di-O-isopropylidene-α-D-galactopyranose)-uronate (15): Scale: 1.0 mmol. Prepared as described in general method 2. The crude residue was purified by flash chromatography (4:1 hexane–EtOAc) to afford **15** (331 mg, 87%) as a white amorphous solid. *R*_f 0.29 (4:1 hexane–EtOAc); [α]_D –179.5 (0.7 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 7.35–7.32 (m, 2H, Ar), 7.24–7.22 (m, 2H, Ar), 5.75 (d, 1H, *J* = 4.9 Hz, H-1), 4.69 (dd, 1H, *J* = 7.7, 2.6 Hz, H-3), 4.65 (dd, 1H, *J* = 7.7, 2.2 Hz, H-4), 4.49 (d, 1H, *J* = 1.9 Hz, H-5), 4.43 (dd, 1H, *J* = 4.9, 2.5 Hz, H-2), 2.39 (s, 3H, ArCH₃), 1.56 (s, 3H, C(CH₃)₂), 1.55 (s, 3H, C(CH₃)₂), 1.38 (s, 3H, C(CH₃)₂), 1.36 (s, 3H, C(CH₃)₂); ¹³C NMR (125 MHz; CDCl₃): δ 197.9 (C-6), 139.5 (Ar), 134.7 (Ar), 130.0 (Ar), 123.6 (Ar), 110.1 (C(CH₃)₂), 109.3 (C(CH₃)₂), 96.7 (C-1), 74.2 (C-5), 72.3 (C-4), 70.8 (C-3), 70.7 (C-2), 26.1 (C(CH₃)₂), 26.0 (C(CH₃)₂), 24.8 (C(CH₃)₂), 24.6 (C(CH₃)₂), 21.4 (ArCH₃); HRMS (ESI) Calc. for (M + Na) C₁₉H₂₄NaO₆S: 403.1186; Found 403.1186.

N-(4-Methoxybenzyl) 1,2:3,4-Di-O-isopropylidene-α-D-galactopyranosiduronamide (16): Scale 0.1 mmol. Prepared as described in general method 3. The crude residue was purified by flash chromatography (2:1 hexane–EtOAc) to afford **16** (35 mg, 90%) as a white solid. *R*_f 0.18 (2:1 hexane–EtOAc); [α]_D –78.2 (0.5 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 7.24–7.21 (m, 2H, Ar), 6.85–6.82 (m, 2H, Ar), 6.78 (app t, 1H, *J* = 5.5, 5.5 Hz, CONHCH₂), 5.54 (d, 1H, *J* = 4.9 Hz, H-1), 4.72 (dd, 1H, *J* = 7.9, 2.1 Hz, H-3), 4.68–4.64 (m, 2H, H-4, NHCH₂Ar), 4.34 (m, 2H,

FULL PAPER

H-2, H-5), 4.23 (dd, 1H, $J = 14.8, 4.7$ Hz, NHCH_2Ar), 3.78 (s, 3H, ArOCH_3), 1.51 (s, 3H, $\text{C}(\text{CH}_3)_2$), 1.39 (s, 3H, $\text{C}(\text{CH}_3)_2$), 1.35 (s, 3H, $\text{C}(\text{CH}_3)_2$), 1.32 (s, 3H, $\text{C}(\text{CH}_3)_2$); ^{13}C NMR (175 MHz; CDCl_3): δ 168.2 (C-6), 158.9 (Ar), 130.0 (Ar), 128.9 (Ar), 113.9 (Ar), 109.4 ($\text{C}(\text{CH}_3)_2$), 109.2 ($\text{C}(\text{CH}_3)_2$), 96.2 (C-1), 71.6 (C-3), 70.7 (C-2), 70.4 (C-4), 68.8 (C-5), 55.3 (ArOCH_3), 42.2 (NHCH_2Ar), 26.0 ($\text{C}(\text{CH}_3)_2$), 25.9 ($\text{C}(\text{CH}_3)_2$), 24.8 ($\text{C}(\text{CH}_3)_2$), 24.2 ($\text{C}(\text{CH}_3)_2$); HRMS (ESI) Calc. for (M + Na) $\text{C}_{20}\text{H}_{27}\text{NNaO}_7$: 416.1680; Found 416.1678.

Methyl 2,3,4-tri-*O*-benzyl-6-thio-6-*S*-*p*-tolyl- α -*D*-glucopyranoside (17): Scale 1.0 mmol. Prepared from methyl 2,3,4-tri-*O*-benzyl- α -*D*-glucopyranoside^[44] as described in general method 1. The crude residue was purified by flash chromatography (5:1→4:1 hexane–EtOAc) to afford **17** (542 mg, 95%) as a clear viscous oil. The ^1H NMR data for the product was consistent with that reported in the literature.^[45]

Methyl 2,3,4-tri-*O*-benzoyl-6-thio-6-*S*-*p*-tolyl- α -*D*-glucopyranoside (18): Scale 1.0 mmol. Prepared from methyl 2,3,4-tri-*O*-benzoyl- α -*D*-glucopyranoside^[46] as described in general method 1. The crude residue was purified by flash chromatography (5:1→4:1 hexane–EtOAc) to afford **18** (576 mg, 94%) as a clear viscous oil. R_f 0.44 (4:1 hexane–EtOAc); $[\alpha]_D^{25} +40.8$ (1.0 c, CHCl_3); ^1H NMR (500 MHz; CDCl_3): δ 8.00–7.98 (m, 2H, Ar), 7.96–7.94 (m, 2H, Ar), 7.88–7.86 (m, 2H, Ar), 7.57–7.51 (m, 2H, Ar), 7.46–7.38 (m, 5H, Ar), 7.32–7.26 (m, 4H, Ar), 7.06 (d, 2H, $J = 7.9$ Hz, Ar), 6.11 (app t, 1H, $J = 9.8$ Hz, H-3), 5.49 (app t, 1H, $J = 9.7$ Hz, H-4), 5.29 (dd, 1H, $J = 10.2, 3.7$ Hz, H-2), 5.24 (d, 1H, $J = 3.7$ Hz, H-1), 4.24 (ddd, 1H, $J = 9.5, 9.4, 2.5$ Hz, H-5), 3.50 (s, 3H, OCH_3), 3.23 (dd, 1H, $J = 14.0, 2.5$ Hz, H-6a), 3.13 (dd, 1H, $J = 14.0, 9.1$ Hz, H-6b), 2.33 (s, 3H, ArCH_3); ^{13}C NMR (125 MHz; CDCl_3): δ 165.8 (2 × C=O), 165.5 (C=O), 136.6 (Ar), 133.5 (Ar), 133.4 (Ar), 133.1 (Ar), 132.3 (Ar), 130.2 (Ar), 130.0 (Ar), 129.9 (Ar), 129.8 (Ar), 129.7 (Ar), 129.3 (Ar), 129.1 (Ar), 128.0 (Ar), 128.4 (2 × Ar), 128.3 (Ar), 96.8 (C-1), 72.3 (C-4), 72.2 (C-2), 70.4 (C-3), 68.9 (C-5), 55.5 (OCH_3), 36.6 (C-6), 21.0 (ArCH_3); HRMS (ESI) Calc. for (M + Na) $\text{C}_{25}\text{H}_{32}\text{NaO}_8\text{S}$: 635.1710; Found 635.1707.

Methyl 2,3,4-tri-*O*-acetyl-6-thio-6-*S*-*p*-tolyl- α -*D*-galactopyranoside (19): Scale 1.0 mmol. Prepared from methyl 2,3,4-tri-*O*-acetyl- α -*D*-galactopyranoside^[47] as described in general method 1. The crude residue was purified by flash chromatography (3:2→2:1 hexane–EtOAc) to afford **19** (363 mg, 85%) as a clear viscous oil. R_f 0.44 (2:1 hexane–EtOAc); $[\alpha]_D^{25} +82.7$ (1.0 c, CHCl_3); ^1H NMR (500 MHz; CDCl_3): δ 7.29 (d, 2H, $J = 8.1$ Hz, Ar), 7.13 (d, 2H, $J = 7.9$ Hz, Ar), 5.49 (dd, 1H, $J = 3.4, 1.2$ Hz, H-4), 5.33 (dd, 1H, $J = 10.9, 3.4$ Hz, H-3), 5.15 (dd, 1H, $J = 10.9, 3.6$ Hz, H-2), 5.00 (d, 1H, $J = 3.6$ Hz, H-1), 4.05 (ddd, 1H, $J = 7.8, 5.5, 1.0$ Hz, H-5), 3.39 (s, 3H, OCH_3), 3.07 (dd, 1H, $J = 13.8, 8.0$ Hz, H-6a), 2.90 (dd, 1H, $J = 13.8, 5.4$ Hz, H-6b), 2.35 (s, 3H, ArCH_3), 2.15 (s, 3H, COCH_3), 2.09 (s, 3H, COCH_3), 2.00 (s, 3H, COCH_3); ^{13}C NMR (125 MHz; CDCl_3): δ 170.4 (C=O), 170.3 (C=O), 169.9 (C=O), 137.0 (Ar), 131.6 (Ar), 130.6 (Ar), 129.9 (Ar), 97.2 (C-1), 69.6 (C-3), 68.2 (C-2), 67.8 (C-4), 67.5 (C-5), 55.4 (OCH_3), 35.2 (C-6), 21.0 (COCH_3), 20.8 (COCH_3), 20.7 (COCH_3); HRMS (ESI) Calc. for (M + NH_4) $\text{C}_{20}\text{H}_{30}\text{NO}_8\text{S}$: 444.1687; Found 444.1679.

Methyl 2,3,4-tri-*O*-benzyl-6-thio-6-*S*-*p*-tolyl- α -*D*-galactopyranoside (20): Scale 1.0 mmol. Prepared from methyl 2,3,4-tri-*O*-benzyl- α -*D*-galactopyranoside^[48] as described in general method 1. The crude residue was purified by flash chromatography (6:1→4:1 hexane–EtOAc) to afford **20** (479 mg, 82%) as a clear viscous oil. R_f 0.44 (4:1 hexane–EtOAc); $[\alpha]_D^{25} -13.0$ (0.3 c, CHCl_3); ^1H NMR (400 MHz; CDCl_3): δ 7.40–7.26 (m, 15H, Ar), 7.18–7.16 (m, 2H, Ar), 7.07 (d, 2H, $J = 8.3$ Hz, Ar), 5.00 (d, 1H, $J = 11.4$ Hz, PhCH_2O), 4.87 (d, 1H, $J = 11.7$ Hz, PhCH_2O), 4.82 (d, 1H, $J = 12.1$ Hz, PhCH_2O), 4.75 (d, 1H, $J = 11.7$ Hz, PhCH_2O), 4.69–4.65 (m, 2H, PhCH_2O , H-1), 4.55 (d, 1H, $J = 11.4$ Hz, PhCH_2O), 4.02 (dd, 1H, $J = 10.0, 3.6$ Hz, H-2), 3.97 (d, 1H, $J = 2.5$ Hz, H-4), 3.89 (dd, 1H, $J = 10.0, 2.5$ Hz, H-3), 3.77–3.73 (m, 1H, H-5), 3.32 (s, 3H, OCH_3), 3.07 (dd, 1H, $J = 13.4, 6.5$ Hz, H-6a), 2.89 (dd, 1H, $J = 13.4, 7.1$ Hz, H-6b), 2.31 (s, 3H, ArCH_3); ^{13}C NMR (125 MHz; CDCl_3): δ 138.8 (Ar), 138.5 (2 × Ar), 136.2 (Ar), 132.4 (Ar), 129.8 (Ar), 129.6 (Ar), 128.4 (3 × Ar), 128.3 (Ar), 128.1 (Ar), 127.7 (2 × Ar)

127.6 (Ar), 127.5 (Ar), 98.9 (C-1), 79.4 (C-3), 76.3 (C-2), 75.7 (C-4), 74.9 (PhCH_2O), 73.5 (2 × PhCH_2O), 69.4 (C-5), 55.3 (OCH_3), 34.7 (C-6), 21.0 (ArCH_3); HRMS (ESI) Calc. for (M + NH_4) $\text{C}_{35}\text{H}_{42}\text{NO}_5\text{S}$: 588.2778; Found 588.2764.

Methyl 2,3,4-tri-*O*-benzoyl-6-thio-6-*S*-*p*-tolyl- α -*D*-mannopyranoside (21): Scale: 1.0 mmol. Prepared from methyl 2,3,4-tri-*O*-benzoyl- α -*D*-mannopyranoside^[9] as described in general method 1. The crude residue was purified by flash chromatography (6:1→2:1 hexane–EtOAc) to afford **21** (594 mg, 97%) as a white amorphous solid. R_f 0.42 (4:1 hexane–EtOAc); $[\alpha]_D^{25} -112.0$ (1.0 c, CHCl_3); ^1H NMR (400 MHz; CDCl_3): δ 8.13–8.11 (m, 2H, Ar), 7.96–7.94 (m, 2H, Ar), 7.84–7.82 (m, 2H, Ar), 7.64–7.38 (m, 7H, Ar), 7.28–7.24 (m, 4H, Ar), 7.05 (d, 2H, $J = 8.0$ Hz, Ar), 5.87–5.80 (m, 2H, H3, H4), 5.67 (app t, 1H, $J = 1.7$ Hz, H-2), 4.99 (d, 1H, $J = 1.7$ Hz, H-1), 4.27–4.22 (m, 1H, H-5), 3.53 (s, 3H, OCH_3), 3.29 (dd, 1H, $J = 13.8, 2.5$ Hz, H-6a), 3.20 (dd, 1H, $J = 13.8, 8.7$ Hz, H-6b), 2.31 (s, 3H, ArCH_3); ^{13}C NMR (125 MHz; CDCl_3): δ 165.7 (C=O), 165.6 (C=O), 165.4 (C=O), 136.5 (Ar), 133.5 (2 × Ar), 133.1 (Ar), 132.3 (Ar), 130.2 (Ar), 130.0 (Ar), 129.9 (2x, Ar), 129.8 (Ar), 129.4 (Ar), 129.2 (Ar), 129.0 (Ar), 128.6 (Ar), 128.5 (Ar), 128.2 (Ar), 98.5 (C-1), 70.6 (C-2), 70.0 (2 × C3 & C4), 69.7 (C5), 55.4 (OCH_3), 36.8 (C-6), 21.0 (ArCH_3); HRMS (ESI) Calc. for (M + Na) $\text{C}_{35}\text{H}_{32}\text{NaO}_8\text{S}$: 635.1710; Found 635.1706.

***S*-*p*-Tolyl thio(methyl 2,3,4-tri-*O*-benzyl- α -*D*-glucopyranosid)uronate (22):** Scale 1.0 mmol. Prepared as described in general method 2. The crude residue was purified by flash chromatography (6:1→4:1 hexane–EtOAc) to afford **22** (456 mg, 78%) as a colorless oil. R_f 0.40 (4:1 hexane–EtOAc); $[\alpha]_D^{25} +15.5$ (1.0 c, CHCl_3); ^1H NMR (500 MHz; CDCl_3): δ 7.38–7.25 (m, 19H, Ar), 4.97 (d, 1H, $J = 10.9$ Hz, PhCH_2O), 4.86 (d, 1H, $J = 10.9$ Hz, PhCH_2O), 4.85 (d, 1H, $J = 12.4$ Hz, PhCH_2O), 4.78 (d, 1H, $J = 10.4$ Hz, PhCH_2O), 4.73 (d, 1H, $J = 3.5$ Hz, H-1), 4.69 (d, 1H, $J = 12.4$, PhCH_2O), 4.68 (d, 1H, $J = 10.4$, PhCH_2O), 4.37 (d, 1H, $J = 9.7$ Hz, H-5), 4.05 (app t, 1H, $J = 9.3$ Hz, H-3), 3.82 (dd, 1H, $J = 9.7, 9.3$ Hz, H-4), 3.62 (dd, 1H, $J = 9.3, 3.5$ Hz, H-2), 3.49 (s, 3H, OCH_3), 2.40 (s, 3H, ArCH_3); ^{13}C NMR (125 MHz; CDCl_3): δ 195.4 (C-6), 140.0 (Ar), 138.6 (Ar), 138.0 (2 × Ar), 134.5 (Ar), 130.2 (Ar), 128.6 (Ar), 128.4 (2 × Ar), 128.2 (Ar), 128.1 (2 × Ar), 128.0 (Ar), 127.8 (Ar), 127.7 (Ar), 123.1 (Ar), 98.8 (C-1), 81.5 (C-3), 79.9 (C-4), 79.3 (C-2), 76.0 (PhCH_2O), 75.5 (C5), 75.3 (PhCH_2O), 73.7 (PhCH_2O), 55.9 (OCH_3), 21.4 (ArCH_3); HRMS (ESI) Calc. for (M + NH_4) $\text{C}_{35}\text{H}_{40}\text{O}_6\text{S}$: 602.2571; Found 602.2572.

***S*-*p*-Tolyl thio(methyl 2,3,4-tri-*O*-benzoyl- α -*D*-glucopyranosid)uronate (23):** Scale 1.0 mmol. Prepared as described in general method 2. The crude residue was purified by flash chromatography (dry loading) (4:1→3:1 hexane–EtOAc) to afford **23** (539 mg, 86%) as a white amorphous solid. R_f 0.31 (4:1 hexane–EtOAc); $[\alpha]_D^{25} +52.9$ (1.0 c, CHCl_3); ^1H NMR (500 MHz; CDCl_3): δ 8.03–8.01 (m, 2H, Ar), 7.97–7.95 (m, 2H, Ar), 7.93–7.90 (m, 2H, Ar), 7.57–7.30 (m, 11H, Ar), 7.22 (dd, 2H, $J = 8.5, 0.6$ Hz, Ar), 6.21 (app t, 1H, $J = 9.8$ Hz, H-3), 5.85 (app t, 1H, $J = 9.7$ Hz, H-4), 5.44 (d, 1H, $J = 3.6$ Hz, H-1), 5.38 (dd, $J = 10.1, 3.6$ Hz, H-2), 4.71 (d, 1H, $J = 9.7$ Hz, H-5), 3.59 (s, 3H, OCH_3), 2.37 (s, 3H, ArCH_3); ^{13}C NMR (125 MHz; CDCl_3): δ 194.7 (C-6), 165.8 (C=O), 165.6 (C=O), 165.1 (C=O), 140.0 (Ar), 134.8 (Ar), 133.5 (Ar), 133.3 (Ar), 133.2 (Ar), 130.2 (Ar), 130.0 (Ar), 129.9 (Ar), 129.7 (Ar), 129.1 (Ar), 129.0 (Ar), 128.9 (Ar), 128.5 (Ar), 128.3 (2 × Ar), 122.6 (Ar), 97.6 (C-1), 74.0 (C-5), 71.6 (C-2), 70.1 (C-4), 69.9 (C-3), 56.4 (OCH_3), 21.4 (ArCH_3); HRMS (ESI) Calc. for (M + Na) $\text{C}_{35}\text{H}_{30}\text{NaO}_9\text{S}$: 649.1503; Found 649.1497.

***S*-*p*-Tolyl thio(methyl 2,3,4-tri-*O*-acetyl- α -*D*-galactopyranosid)uronate (24):** Scale 1.0 mmol. Prepared as described in general method 2. The crude residue was purified by flash chromatography (2:1 hexane–EtOAc) to afford **24** (396 mg, 90%) as a clear viscous oil. R_f 0.36 (2:1 hexane–EtOAc); $[\alpha]_D^{25} +72.8$ (1.0 c, CHCl_3); ^1H NMR (500 MHz; CDCl_3): δ 7.27–7.23 (m, 4H, Ar), 5.82 (dd, 1H, $J = 3.3, 1.6$ Hz, H-4), 5.43–5.39 (m, 1H, H-3), 5.27–5.24 (m, 2H, H-1, H-2), 4.66 (d, 1H, $J = 1.6$ Hz, H-5), 3.55 (s, 3H, OCH_3), 2.39 (s, 3H, ArCH_3), 2.12 (s, 6H, COCH_3 , COCH_3), 2.00 (s, 3H, COCH_3); ^{13}C NMR (125 MHz; CDCl_3): δ 195.0 (C-6), 170.3 (C=O), 169.8

FULL PAPER

(C=O), 169.5 (C=O), 140.1 (Ar), 134.5 (Ar), 130.2 (Ar), 122.6 (Ar), 97.8 (C-1), 74.2 (C-5), 68.8 (C-4), 67.9 (C-2), 67.2 (C-3), 56.3 (OCH₃), 21.4 (COCH₃), 20.8 (COCH₃), 20.6 (COCH₃), 20.5 (ArCH₃); HRMS (ESI) Calc. for (M + NH₄) C₂₀H₂₈NO₉S: 458.1479; Found 458.1472.

S-p-Tolyl thio(methyl 2,3,4-tri-O-benzyl- α -D-galactopyranosid)uronate (25): Scale 1.0 mmol. Prepared as described in general method 2. The crude residue was purified by flash chromatography (4:1 hexane–EtOAc) to afford **25** (439 mg, 75%) as a clear viscous oil. *R*_f 0.42 (4:1 hexane–EtOAc); [α]_D –41.0 (0.2 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 7.40–7.29 (m, 15H, Ar), 7.23–7.21 (m, 4H, Ar), 4.92 (d, 1H, *J* = 11.1 Hz, PhCH₂O), 4.91 (d, 1H, *J* = 3.6 Hz, H-1), 4.88 (d, 1H, *J* = 12.0 Hz, PhCH₂O), 4.84 (d, 1H, *J* = 11.7 Hz, PhCH₂O), 4.75 (d, 1H, *J* = 11.7 Hz, PhCH₂O), 4.71 (d, 1H, *J* = 12.0 Hz, PhCH₂O), 4.56 (d, 1H, *J* = 11.1 Hz, H-1), 4.43 (d, 1H, *J* = 1.4 Hz, H-5), 4.39 (dd, 1H, *J* = 2.8, 1.4 Hz, H-4), 4.16 (dd, 1H, *J* = 10.1, 3.6 Hz, H-2), 3.97 (dd, 1H, *J* = 10.1, 2.8 Hz, H-3), 3.49 (s, 3H, OCH₃), 2.40 (s, 3H, ArCH₃); ¹³C NMR (125 MHz, CDCl₃): δ 198.2 (C-6), 139.8 (Ar), 138.5 (Ar), 138.4 (2 \times Ar), 134.8 (Ar), 130.1 (Ar), 128.4 (Ar), 128.1 (2 \times Ar), 127.8 (Ar), 127.6 (Ar), 127.5 (2 \times Ar), 123.4 (Ar), 99.4 (C-1), 78.3 (C-3), 76.4 (C-4), 76.1 (2 \times C-5, C-2), 75.6 (PhCH₂O), 73.9 (PhCH₂O), 73.2 (PhCH₂O), 56.1 (OCH₃), 21.4 (ArCH₃); HRMS (ESI) Calc. for (M + NH₄) C₃₅H₄₀NO₆S: 602.2571; Found 602.2557.

S-p-Tolyl thio(methyl 2,3,4-tri-O-benzoyl- α -D-mannopyranosid)uronate (26): Scale 1.0 mmol. Prepared as described in general method 2. The crude residue was purified by flash chromatography (4:1–3:1 hexane–EtOAc) to afford **26** (464 mg, 74%) as a white amorphous solid. *R*_f 0.31 (4:1 hexane–EtOAc); [α]_D –100.5 (1.0 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 8.21–8.19 (m, 2H, Ar), 8.01–7.99 (m, 2H, Ar), 7.90–7.88 (m, 2H, Ar), 7.66–7.45 (m, 5H, Ar), 7.40–7.28 (m, 6H, Ar), 7.24 (d, 2H, *J* = 8.0 Hz, Ar), 6.20 (app t, 1H, *J* = 9.7 Hz, H-4), 5.96 (dd, 1H, *J* = 9.7, 3.2 Hz, H-3), 5.77 (dd, 1H, *J* = 3.2, 2.2 Hz, H-2), 5.21 (d, 1H, *J* = 2.2 Hz, H-1), 4.76 (d, 1H, *J* = 9.7 Hz, H-5), 3.65 (s, 3H, OCH₃), 2.38 (s, 3H, ArCH₃); ¹³C NMR (125 MHz; CDCl₃): δ 194.9 (C-6), 165.50 (C=O), 165.35 (C=O), 165.29 (C=O), 140.0 (Ar), 134.8 (Ar), 133.7 (Ar), 133.3 (2 \times Ar), 130.2 (Ar), 130.1 (Ar), 129.9 (Ar), 129.8 (Ar), 129.2 (2 \times Ar), 129.0 (Ar), 128.7 (Ar), 128.4 (Ar), 122.7 (Ar), 99.1 (C-1), 75.4 (C-5), 69.9 (C-2), 69.4 (C-3), 67.8 (C-4), 56.3 (OCH₃), 21.4 (ArCH₃); HRMS (ESI) Calc. for (M + NH₄) C₃₅H₃₈NO₉S: 644.1949; Found 644.1935.

N-(4-Methoxybenzyl) (methyl 2,3,4-tri-O-benzyl- α -D-glucopyranosid)uronamide (27): Scale 0.1 mmol. Prepared as described in general method 3. The crude residue was purified by flash chromatography (2:1 hexane–EtOAc) to afford **27** (51 mg, 85%) as a white solid. *R*_f 0.18 (2:1 hexane–EtOAc); [α]_D –6.1 (1.0 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 7.38–7.25 (m, 15H, Ar), 7.15 (d, 2H, *J* = 8.6 Hz, Ar), 6.82 (d, 2H, *J* = 8.6 Hz, Ar), 6.33 (app t, 1H, *J* = 5.5 Hz, CONHCH₂), 4.97 (d, 1H, *J* = 10.9 Hz, PhCH₂O), 4.85 (d, 1H, *J* = 10.9 Hz, PhCH₂O), 4.82 (d, 1H, *J* = 12.0 Hz, PhCH₂O), 4.78 (d, 1H, *J* = 10.5 Hz, PhCH₂O), 4.67 (d, 1H, *J* = 12.0 Hz, PhCH₂O), 4.64–4.61 (m, 2H, H-1, PhCH₂O), 4.43 (dd, 1H, *J* = 14.4, 5.5 Hz, NHCH₂Ar), 4.34 (dd, 1H, *J* = 14.4, 5.5 Hz, NHCH₂Ar), 4.10 (d, 1H, *J* = 9.9 Hz, H-5), 4.02 (app t, 1H, *J* = 9.3 Hz, H-3), 3.79 (s, 3H, ArOCH₃), 3.63 (dd, 1H, *J* = 9.9, 9.3 Hz, H-4), 3.57 (dd, 1H, *J* = 9.3, 3.5 Hz, H-2), 3.40 (s, 3H, OCH₃); ¹³C NMR (125 MHz; CDCl₃): δ 168.6 (C-6), 159.1 (Ar), 138.6 (Ar), 138.0 (Ar), 137.9 (Ar), 129.8 (Ar), 129.3 (Ar), 128.5 (Ar), 128.4 (3 \times Ar), 128.2 (Ar), 128.1 (2 \times Ar), 128.0 (Ar), 127.8 (Ar), 127.7 (Ar), 114.1 (Ar), 98.5 (C-1), 81.5 (C-3), 80.3 (C-4), 79.2 (C-2), 75.9 (PhCH₂O), 75.2 (PhCH₂O), 73.6 (PhCH₂O), 70.8 (C-5), 55.8 (OCH₃), 55.3 (ArOCH₃), 43.0 (NHCH₂Ar); HRMS (ESI) Calc. for (M + H) C₃₆H₄₀NO₇: 598.2799; Found 598.2790.

Methyl 2,3,4-tri-O-benzyl- α -D-glucopyranosiduronamide (28): Scale 0.1 mmol. Prepared as described in general method 3. The crude residue was purified by flash chromatography (1:2 hexane–EtOAc) to afford **28** (32 mg, 66%) as a white solid. *R*_f 0.24 (1:2 hexane–EtOAc); [α]_D –8.2 (1.0 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 7.39–7.28 (m, 15H), 6.12 (br s, 1H,

CONH₂), 5.79 (br s, 1H, CONH₂), 4.98 (d, 1H, *J* = 10.9 Hz, PhCH₂O), 4.87–4.81 (m, 3H, PhCH₂O), 4.71–4.65 (m, 3H, PhCH₂O, H-1), 4.12 (d, 1H, *J* = 9.7 Hz, H-5), 4.02 (app t, 1H, *J* = 9.5 Hz, H-3), 3.61 (dd, 1H, *J* = 9.7, 9.5 Hz, H-4), 3.57 (dd, 1H, *J* = 9.5, 3.5 Hz, H-2), 3.41 (s, 3H, OCH₃); ¹³C NMR (125 MHz; CDCl₃): δ 171.4 (C-6), 138.5 (Ar), 138.0 (Ar), 137.7 (Ar), 128.6 (Ar), 128.5 (2 \times Ar), 128.4 (2 \times Ar), 128.3 (Ar), 128.2 (3 \times Ar), 128.1 (Ar), 128.0 (Ar), 127.9 (Ar), 127.7 (Ar), 98.4 (C-1), 81.6 (C-3), 80.2 (C-4), 79.2 (C-2), 75.9 (PhCH₂O), 75.4 (PhCH₂O), 73.6 (PhCH₂O), 72.0 (C-5), 55.8 (OCH₃); HRMS (ESI) Calc. for (M + H) C₂₈H₃₂NO₆: 478.2224; Found 478.2217.

2-N-(Methyl 2,3,4-tri-O-benzyl- α -D-glucopyranosiduronoyl)-1,3-di-O-benzyl-2-deoxy-glycerol (29): Scale 0.1 mmol. Prepared as described in general method 3. The crude residue was purified by flash chromatography (4:1–2:1 hexane–EtOAc) to afford **29** (47 mg, 64%) as a white solid. *R*_f 0.24 (2:1 hexane–EtOAc); [α]_D –6.3 (1.0 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 7.40–7.26 (m, 25H, Ar), 6.49 (d, 1H, *J* = 8.4 Hz, CONHCH₂), 4.96 (d, 1H, *J* = 10.9 Hz, PhCH₂O), 4.85–4.82 (m, 2H, PhCH₂O), 4.74 (d, 1H, *J* = 10.6 Hz, PhCH₂O), 4.69–4.64 (m, 3H, PhCH₂O, H-1), 4.52–4.44 (m, 4H, PhCH₂O), 4.37–4.33 (m, 1H, NHCH₂), 4.07 (d, 1H, *J* = 9.9 Hz, H-5), 4.01 (app t, 1H, *J* = 9.3 Hz, H-3), 3.65–3.56 (m, 4H, CHCH₂OBn, H-2, H-4), 3.53 (dd, 1H, *J* = 9.3, 5.9 Hz, CHCH₂OBn), 3.46 (dd, 1H, *J* = 9.3, 5.9 Hz, CHCH₂OBn), 3.40 (s, 3H, OCH₃); ¹³C NMR (125 MHz; CDCl₃): δ 168.7 (C-6), 138.7 (Ar), 138.1 (2 \times Ar), 138.0 (2 \times Ar), 128.5 (Ar), 128.4 (2 \times Ar), 128.3 (Ar), 128.2 (Ar), 128.0 (3 \times Ar), 127.7 (4 \times Ar), 127.6 (Ar), 98.5 (C-1), 81.5 (C-3), 80.4 (C-4), 79.2 (C-2), 75.9 (PhCH₂O), 75.1 (PhCH₂O), 73.6 (PhCH₂O), 73.1 (2 \times PhCH₂O), 70.9 (C-5), 68.3 (CHCH₂OBn), 68.2 (CHCH₂OBn), 55.8 (OCH₃), 48.4 (NHCH₂); HRMS (ESI) Calc. for (M + H) C₄₅H₅₀NO₈: 732.3531; Found 732.3516.

N-(Methyl 2,3,4-tri-O-benzyl- α -D-glucopyranosiduronoyl)-L-alanine tert-butyl ester (30): Scale 0.1 mmol. Prepared as described in general method 3. The crude residue was purified by flash chromatography (2:1 hexane–EtOAc) to afford **30** (43 mg, 71%) as a viscous oil. *R*_f 0.27 (2:1 hexane–EtOAc); [α]_D –4.64 (1.0 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 7.39–7.27 (m, 15H, Ar), 6.83 (d, 1H, *J* = 7.0 Hz, CONHCH₂), 4.96 (d, 1H, *J* = 10.9 Hz, PhCH₂O), 4.85 (d, 1H, *J* = 11.8 Hz, PhCH₂O), 4.83 (d, 1H, *J* = 12.7 Hz, PhCH₂O), 4.76 (d, 1H, *J* = 10.3 Hz, PhCH₂O), 4.73 (d, 1H, *J* = 10.4 Hz, PhCH₂O), 4.67 (d, 1H, *J* = 12.1 Hz, PhCH₂O), 4.66 (d, 1H, *J* = 3.5 Hz, H-1), 4.46 (dt, 1H, *J* = 14.2, 7.2 Hz, NHCHCO₂C(CH₃)₃), 4.11 (d, 1H, *J* = 9.9 Hz, H-5), 4.02 (app t, 1H, *J* = 9.3 Hz, H-3), 3.61 (dd, 1H, *J* = 9.9, 8.9 Hz, H-4), 3.56 (dd, 1H, *J* = 9.7, 3.5 Hz, H-2), 3.41 (s, 3H, OCH₃), 1.48 (s, 9H, NHCHCO₂C(CH₃)₃), 1.36 (d, 3H, *J* = 7.1 Hz, CHCH₃); ¹³C NMR (125 MHz; CDCl₃): δ 172.0 (C=O), 168.5 (C=O), 138.7 (Ar), 138.0 (Ar), 138.0 (Ar), 128.5 (Ar), 128.4 (Ar), 128.3 (Ar), 128.3 (Ar), 128.2 (Ar), 128.0 (Ar), 127.9 (Ar), 127.7 (Ar), 127.6 (Ar), 98.4 (C-1), 82.2 (NHCHCO₂C(CH₃)₃), 81.5 (C-3), 80.4 (C-4), 79.2 (C-2), 75.9 (PhCH₂O), 75.2 (PhCH₂O), 73.6 (PhCH₂O), 70.7 (C-5), 55.8 (OCH₃), 48.5 (NHCHCO₂C(CH₃)₃), 28.0 (NHCHCO₂C(CH₃)₃), 18.6 (CHCH₃); HRMS (ESI) Calc. for (M + Na) C₃₅H₄₃NNaO₈: 628.2881; Found 628.2876.

N-(Methyl 2,3,4-tri-O-benzyl- α -D-glucopyranosiduronoyl)-O-tert-butyl-L-serine tert-butyl ester (31): Scale 0.1 mmol. Prepared as described in general method 3. The crude residue was purified by flash chromatography (4:1–2:1 hexane–EtOAc) to afford **31** (42 mg, 70%) as a clear viscous oil. *R*_f 0.47 (2:1 hexane–EtOAc); [α]_D +11.3 (1.0 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 7.39–7.25 (m, 15H, Ar), 7.02 (d, 1H, *J* = 7.8 Hz, CONHCH₂), 4.94 (d, 1H, *J* = 10.8 Hz, PhCH₂O), 4.83 (d, 1H, *J* = 10.8 Hz, PhCH₂O), 4.83 (d, 1H, *J* = 12.3 Hz, PhCH₂O), 4.78 (d, 1H, *J* = 10.2 Hz, PhCH₂O), 4.73 (d, 1H, *J* = 10.3 Hz, PhCH₂O), 4.68 (d, 1H, *J* = 3.5 Hz, H-1), 4.68 (d, 1H, *J* = 12.3 Hz, PhCH₂O), 4.65–4.62 (m, 1H, NHCHCH₂), 4.15 (d, 1H, *J* = 9.9 Hz, H-5), 4.02 (app t, 1H, *J* = 9.3 Hz, H-3), 3.77 (dd, 1H, *J* = 8.8, 2.9 Hz, CHCH₂O), 3.63 (app t, 1H, *J* = 9.4 Hz, H-4), 3.59–3.57 (m, 2H, H-2, CHCH₂O), 3.42 (s, 3H, OCH₃), 1.48 (s, 9H, C(CH₃)₃), 1.11 (s, 9H, C(CH₃)₃); ¹³C NMR (125 MHz; CDCl₃): δ 169.2 (C=O), 168.7 (C=O), 138.7 (Ar), 138.1 (2 \times Ar), 128.5 (Ar), 128.4 (Ar), 128.3 (2 \times Ar), 128.2 (Ar), 128.0 (2 \times Ar), 127.6 (Ar), 98.4 (C-1), 82.0 (C(CH₃)₃), 81.5 (C-3), 80.6 (C-3), 79.3

FULL PAPER

(C-2), 76.0 (PhCH₂O), 75.1 (PhCH₂O), 73.6 (PhCH₂O), 73.0 (C(CH₃)₃), 70.9 (C-5), 62.1 (CH₂CH₂O), 55.7 (OCH₃), 53.1 (NHCH), 28.1 (C(CH₃)₃), 27.4 (C(CH₃)₃); HRMS (ESI) Calc. for (M + H) C₃₉H₅₂NO₉: 678.3637; Found 678.3631.

N-(Methyl 2,3,4-tri-O-benzoyl- α -D-glucopyranosiduronoyl)-L-threonine tert-butyl ester (32): Scale 0.1 mmol. Prepared as described in general method 3. The crude residue was purified by flash chromatography (2:1 hexane–EtOAc) to afford **32** (40 mg, 63%) as a oil. *R*_f 0.22 (2:1 hexane–EtOAc); [α]_D –14.6 (1.0 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 7.39–7.25 (m, 15H, Ar), 6.97 (d, 1H, *J* = 8.6 Hz, CONHCH), 4.96 (d, 1H, *J* = 10.8 Hz, PhCH₂O), 4.84 (d, 1H, *J* = 10.8 Hz, PhCH₂O), 4.83 (d, 1H, *J* = 12.4 Hz, PhCH₂O), 4.78–4.74 (m, 2H, PhCH₂O), 4.68 (d, 1H, *J* = 12.4 Hz, PhCH₂O), 4.68 (d, 1H, *J* = 3.5 Hz, H-1), 4.54 (dd, 1H, *J* = 8.6, 2.9 Hz, NHCH), 4.30–4.29 (m, 1H, CHCH₂OH), 4.20 (d, 1H, *J* = 10.0 Hz, H-5), 4.04 (app t, 1H, *J* = 9.2 Hz, H-3), 3.65 (dd, 1H, *J* = 9.8, 9.0 Hz, H-4), 3.58 (dd, 1H, *J* = 9.7, 3.5 Hz, H-2), 3.41 (s, 3H, OCH₃), 2.05 (s, 1H, CHO), 1.49 (s, 9H, C(CH₃)₃), 1.21 (d, 3H, *J* = 6.4 Hz, CHCH₃); ¹³C NMR (125 MHz; CDCl₃): δ 169.8 (C=O), 169.7 (C=O), 138.6 (Ar), 138.0 (2 × Ar), 128.6 (Ar), 128.4 (Ar), 128.3 (Ar), 128.2 (2 × Ar), 128.1 (Ar), 127.9 (Ar), 127.7 (Ar), 127.6 (Ar), 98.4 (C-1), 82.7 (C(CH₃)₃), 81.5 (C-3), 80.6 (C-4), 79.2 (C-2), 76.0 (PhCH₂O), 75.2 (PhCH₂O), 73.6 (PhCH₂O), 70.6 (C-5), 68.4 (CHCH₂OH), 57.5 (NHCH), 55.8 (OCH₃), 28.0 (C(CH₃)₃), 20.0 (CHCH₃); HRMS (ESI) Calc. for (M + H) C₃₆H₄₆NO₉: 636.3167; Found 636.3161.

N-(4-Methoxybenzyl) (methyl 2,3,4-tri-O-benzoyl- α -D-glucopyranosiduronamide (33): Scale 0.1 mmol. Prepared as described in general method 3. The crude residue was purified by flash chromatography (3:2→1:1 hexane–EtOAc) to afford **33** (55 mg, 86%) as a clear viscous oil. *R*_f 0.43 (1:1 hexane–EtOAc); [α]_D +99.7 (1.0 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 8.04–7.99 (m, 4H, Ar), 7.91–7.89 (m, 2H, Ar), 7.56–7.50 (m, 2H, Ar), 7.48–7.38 (m, 5H, Ar), 7.34–7.30 (m, 2H, Ar), 7.29–7.26 (m, 2H, Ar), 6.93–6.91 (m, 2H, Ar), 6.71 (app t, 1H, *J* = 5.6 Hz, CONHCH₂), 6.20 (app t, 1H, *J* = 9.9 Hz, H-3), 5.68 (app t, 1H, *J* = 9.9 Hz, H-4), 5.30 (d, 1H, *J* = 3.6 Hz, H-1), 5.24 (dd, 1H, *J* = 10.2, 3.6 Hz, H-2), 4.56 (d, 1H, *J* = 10.2 Hz, H-5), 4.49 (dd, 1H, *J* = 14.5, 6.0 Hz, NHCH₂Ar), 4.35 (dd, 1H, *J* = 14.5, 5.4 Hz, NHCH₂Ar), 3.84 (s, 3H, ArOCH₃), 3.50 (s, 3H, OCH₃); ¹³C NMR (125 MHz; CDCl₃): δ 166.9 (C=O), 165.9 (C=O), 165.6 (C=O), 165.5 (C=O), 159.2 (Ar), 133.5 (Ar), 133.2 (Ar), 133.1 (Ar), 130.0 (2 × Ar), 129.8 (Ar), 129.7 (Ar), 129.4 (Ar), 129.3 (Ar), 129.2 (Ar), 128.9 (Ar), 128.5 (Ar), 128.3 (2 × Ar), 114.2 (Ar), 97.2 (C-1), 71.9 (C-2), 70.4 (C-4), 69.8 (C-3), 68.7 (C-5), 56.2 (OCH₃), 55.3 (ArOCH₃), 42.7 (NHCH₂Ar); HRMS (ESI) Calc. for (M + H) C₃₆H₃₄NO₁₀: 640.2168; Found 640.2177.

N-(2-Hydroxyethyl) (methyl 2,3,4-tri-O-benzoyl- α -D-glucopyranosiduronamide (34): Scale 0.1 mmol. Prepared as described in general method 3. The crude residue was purified by flash chromatography (20:1 CH₂Cl₂–CH₃OH) to afford **34** (48 mg, 86%) as a clear viscous oil. *R*_f 0.31 (1:1 CH₂Cl₂–CH₃OH); [α]_D +75.0 (1.0 c, CHCl₃); ¹H NMR (600 MHz; CDCl₃): δ 8.01–7.99 (m, 4H, Ar), 7.90–7.88 (m, 2H, Ar), 7.56–7.50 (m, 2H, Ar), 7.46–7.38 (m, 5H, Ar), 7.32–7.29 (m, 2H, Ar), 6.89 (app t, 1H, *J* = 5.8 Hz, CONHCH₂), 6.22 (app t, 1H, *J* = 9.9 Hz, H-3), 5.62 (app t, 1H, *J* = 9.9 Hz, H-4), 5.34 (d, 1H, *J* = 3.6 Hz, H-1), 5.29 (dd, 1H, *J* = 10.1, 3.6 Hz, H-2), 4.50 (d, 1H, *J* = 10.2 Hz, H-5), 3.84–3.81 (m, 1H, CH₂CH₂OH), 3.71–3.69 (m, 1H, CH₂CH₂O), 3.60 (dddd, 1H, *J* = 14.2, 5.8, 5.8, 3.2 Hz, NHCH₂CH₂), 3.50 (s, 3H, OCH₃), 3.30 (dddd, 1H, *J* = 13.9, 8.1, 5.6, 3.4 Hz, NHCH₂CH₂), 2.76 (s, 1H, CH₂OH); ¹³C NMR (125 MHz; CDCl₃): δ 168.2 (C=O), 166.1 (C=O), 165.9 (C=O), 165.6 (C=O), 133.6 (Ar), 133.4 (Ar), 133.2 (Ar), 130.0 (2 × Ar), 129.7 (Ar), 129.1 (Ar), 128.9 (Ar), 128.8 (Ar), 128.5 (Ar), 128.4 (Ar), 128.3 (Ar), 97.3 (C-1), 71.9 (C-2), 71.0 (C-4), 69.5 (C-3), 69.1 (C-5), 61.2 (CH₂CH₂O), 56.2 (OCH₃), 42.2 (NHCH₂CH₂); HRMS (ESI) Calc. for (M + H) C₃₀H₃₀NO₁₀: 564.1864; Found 564.1859.

2-N-(Methyl 2,3,4-tri-O-benzoyl- α -D-glucopyranosiduronoyl)-1,3-di-O-benzyl-2-deoxy-glycerol (35): Scale 0.1 mmol. Prepared as described in

general method 3. The crude residue was purified by flash chromatography (2:1 hexane–EtOAc) to afford **35** (56 mg, 73%) as a clear viscous oil. *R*_f 0.23 (2:1 hexane–EtOAc); [α]_D +70.6 (1.0 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 8.03–7.99 (m, 4H, Ar), 7.92–7.90 (m, 2H, Ar), 7.57–7.30 (m, 19H, Ar), 6.88 (d, 1H, *J* = 8.5 Hz, CONHCH), 6.20 (app t, 1H, *J* = 9.9 Hz, H-3), 5.63 (app t, 1H, *J* = 9.9 Hz, H-4), 5.34 (d, 1H, *J* = 3.6 Hz, H-1), 5.29 (dd, 1H, *J* = 10.2, 3.6 Hz, H-2), 4.62 (d, 1H, *J* = 12.0 Hz, PhCH₂O), 4.59 (d, 1H, *J* = 12.0 Hz, PhCH₂O), 4.54 (d, 1H, *J* = 12.0 Hz, PhCH₂O), 4.51 (d, 1H, *J* = 12.0 Hz, PhCH₂O), 4.50 (d, 1H, *J* = 10.1 Hz, H-5), 4.31–4.28 (m, 1H, NHCH), 3.73 (dd, 1H, *J* = 9.3, 3.9 Hz, CHCH₂OBn), 3.69 (dd, 1H, *J* = 9.4, 4.3 Hz, CHCH₂OBn), 3.65 (dd, 1H, *J* = 9.3, 6.2 Hz, CHCH₂OBn), 3.56 (dd, 1H, *J* = 9.4, 5.9 Hz, CHCH₂OBn), 3.50 (s, 3H, OCH₃); ¹³C NMR (125 MHz; CDCl₃): δ 166.9 (C=O), 165.9 (C=O), 165.6 (C=O), 165.4 (C=O), 138.2 (Ar), 138.1 (Ar), 133.5 (Ar), 133.2 (Ar), 133.1 (Ar), 130.0 (2 × Ar), 129.7 (Ar), 129.3 (Ar), 129.2 (Ar), 128.9 (Ar), 128.5 (Ar), 128.4 (2 × Ar), 128.3 (2 × Ar), 127.8 (Ar), 127.7 (2 × Ar), 127.6 (Ar), 97.2 (C-1), 73.2 (PhCH₂O), 73.1 (PhCH₂O), 71.9 (C-2), 70.4 (C-4), 69.9 (C-3), 68.8 (C-5), 68.2 (CHCH₂OBn), 68.1 (CHCH₂OBn), 56.2 (OCH₃), 48.2 (NHCH); HRMS (ESI) Calc. for (M + H) C₄₅H₄₄NO₁₁: 774.2909; Found 774.2893.

N-(Methyl 2,3,4-tri-O-benzoyl- α -D-glucopyranosiduronoyl)-L-threonine tert-butyl ester (36): Scale 0.1 mmol. Following the general method 3, the crude residue was purified by flash chromatography (3:2→1:1 hexane–EtOAc) to afford **36** (28 mg, 41%) as a clear viscous oil. *R*_f 0.52 (1:1 hexane–EtOAc); [α]_D +67.1 (1.0 c, CHCl₃); ¹H NMR (600 MHz; CDCl₃): δ 8.02–7.97 (m, 4H, Ar), 7.90–7.88 (m, 2H, Ar), 7.56–7.36 (m, 7H, Ar), 7.32–7.29 (m, 2H, Ar), 7.23 (d, 1H, *J* = 8.7 Hz, CONHCH), 6.21 (app t, 1H, *J* = 9.9 Hz, H-3), 5.68 (app t, 1H, *J* = 9.9 Hz, H-4), 5.37 (d, 1H, *J* = 3.6 Hz, H-1), 5.31 (dd, 1H, *J* = 10.2, 3.6 Hz, H-2), 4.56 (d, 1H, *J* = 10.2 Hz, H-5), 4.47 (dd, 1H, *J* = 8.7, 3.2 Hz, NHCH), 4.31–4.30 (m, 1H, CHCH₂OH), 3.52 (s, 3H, OCH₃), 2.27 (s, 1H, CHO), 1.50 (s, 9H, C(CH₃)₃), 1.31 (d, 3H, *J* = 6.5 Hz, CHCH₃); ¹³C NMR (125 MHz; CDCl₃): δ 169.6 (C=O), 167.7 (C=O), 165.8 (C=O), 165.7 (C=O), 165.6 (C=O), 133.5 (Ar), 133.2 (2 × Ar), 130.0 (Ar), 129.7 (Ar), 129.1 (2 × Ar), 128.9 (Ar), 128.5 (Ar), 128.4 (Ar), 128.3 (2 × Ar), 97.3 (C-1), 82.8 (C(CH₃)₃), 71.8 (C-2), 70.7 (C-4), 69.8 (C-3), 68.9 (2 × C-5, CHCH₂OH), 57.5 (NHCH), 56.3 (OCH₃), 28.0 (C(CH₃)₃), 20.1 (CHCH₃); HRMS (ESI) Calc. for (M + H) C₃₆H₄₀NO₁₂: 678.2545; Found 678.2531.

N-(4-Methoxybenzyl) (methyl 2,3,4-tri-O-acetyl- α -D-galactopyranosiduronamide (37): Scale 0.1 mmol. Prepared as described in general method 3. The crude residue was purified by flash chromatography (3:2 EtOAc–hexane) to afford **37** (41 mg, 91%) as a clear viscous oil. *R*_f 0.25 (3:2 EtOAc–hexane); [α]_D +116.5 (1.0 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 7.24–7.21 (m, 2H, Ar), 6.90–6.87 (m, 2H, Ar), 6.81 (app t, 1H, *J* = 5.8 Hz, CONHCH₂), 5.88 (dd, 1H, *J* = 3.4, 1.7 Hz, H-4), 5.44 (dd, 1H, *J* = 10.5, 3.4 Hz, H-3), 5.12–5.08 (m, 2H, H-1, H-2), 4.62 (dd, 1H, *J* = 14.6, 7.0 Hz, NHCH₂Ar), 4.51 (d, 1H, *J* = 1.6 Hz, H-5), 4.22 (dd, 1H, *J* = 14.6, 5.0 Hz, NHCH₂), 3.82 (s, 3H, ArOCH₃), 3.43 (s, 3H, OCH₃), 2.10 (s, 3H, COCH₃), 1.99 (s, 3H, COCH₃), 1.95 (s, 3H, COCH₃); ¹³C NMR (125 MHz; CDCl₃): δ 170.5 (C=O), 169.7 (C=O), 169.3 (C=O), 166.3 (C=O), 159.2 (Ar), 130.1 (Ar), 129.2 (Ar), 114.1 (Ar), 97.5 (C-1), 69.5 (C-5), 69.0 (C-4), 68.0 (C-2), 67.1 (C-3), 56.1 (OCH₃), 55.3 (ArOCH₃), 42.5 (NHCH₂), 20.8 (COCH₃), 20.6 (COCH₃), 20.4 (COCH₃); HRMS (ESI) Calc. for (M + Na) C₂₁H₂₇NNaO₁₀: 476.1527; Found 476.1516.

N-(Methyl 2,3,4-tri-O-acetyl- α -D-galactopyranosiduronoyl)-O-tert-butyl-L-serine tert-butyl ester (38): Scale 0.1 mmol. Prepared as described in general method 3. The crude residue was purified by flash chromatography (3:2→1:1 hexane–EtOAc) to afford **38** (34 mg, 64%) as a clear viscous oil. *R*_f 0.48 (1:1 hexane–EtOAc); [α]_D +101.8 (1.0 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 7.29 (d, 1H, *J* = 8.1 Hz, CONHCH), 5.87 (dd, 1H, *J* = 3.4, 1.6 Hz, H-4), 5.44 (dd, 1H, *J* = 10.7, 3.4 Hz, H-3), 5.18 (dd, 1H, *J* = 10.7, 3.6 Hz, H-2), 5.15 (d, 1H, *J* = 3.6 Hz, H-1), 4.63 (app t, 1H, *J* = 8.6, 3.0 Hz, NHCH₂CH₂), 4.51 (d, 1H, *J* = 1.5 Hz, H-5), 3.82 (dd, 1H, *J* = 8.8, 3.1 Hz, CHCH₂O), 3.48 (dd, 1H, *J* = 8.8, 3.0 Hz, CHCH₂O), 3.47 (s,

FULL PAPER

3H, OCH₃), 2.12 (s, 3H, COCH₃), 2.08 (s, 3H, COCH₃), 2.00 (s, 3H, COCH₃), 1.49 (s, 9H, C(CH₃)₃), 1.19 (s, 9H, C(CH₃)₃); ¹³C NMR (125 MHz; CDCl₃): δ 170.5 (C=O), 169.7 (C=O), 169.3 (C=O), 169.0 (C=O), 166.2 (C=O), 97.4 (C-1), 82.0 (OC(CH₃)₃), 73.2 (OC(CH₃)₃), 69.3 (C-5), 68.9 (C-4), 68.0 (C-2), 67.3 (C-3), 62.5 (CHCH₂O), 56.1 (OCH₃), 52.9 (NHCH₂CH₂), 28.0 (C(CH₃)₃), 27.4 (C(CH₃)₃), 20.8 (COCH₃), 20.7 (COCH₃), 20.6 (COCH₃); HRMS (ESI) Calc. for (M + H) C₂₄H₄₀NO₁₂: 534.2545; Found 534.2538.

N-(Methyl 2,3,4-tri-O-acetyl-α-D-galactopyranosiduronoyl)-L-alanine tert-butyl ester (39): Scale 0.1 mmol. Prepared as described in general method 3. The crude residue was purified by flash chromatography (3:2→1:1 hexane–EtOAc) to afford **39** (29 mg, 63%) as a clear viscous oil. *R*_f 0.20 (3:2 hexane–EtOAc); [α]_D +92.7 (1.0 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 7.15 (d, 1H, *J* = 7.6 Hz, CONHCH), 5.84 (dd, 1H, *J* = 3.4, 1.7 Hz, H-4), 5.44 (dd, 1H, *J* = 10.1, 3.4 Hz, H-3), 5.16–5.12 (m, 2H, H-1, H-2), 4.52–4.46 (m, 2H, H-5, NHCH₂CH₃), 3.45 (s, 3H, OCH₃), 2.11 (s, 3H, COCH₃), 2.07 (s, 3H, COCH₃), 1.99 (s, 3H, COCH₃), 1.49 (s, 9H, C(CH₃)₃), 1.38 (d, 3H, *J* = 7.1 Hz, CHCH₃); ¹³C NMR (125 MHz; CDCl₃): δ 171.7 (C=O), 170.5 (C=O), 169.7 (C=O), 169.2 (C=O), 165.9 (C=O), 97.5 (C-1), 82.3 (C(CH₃)₃), 69.4 (C-5), 69.0 (C-4), 68.0 (C-2), 67.1 (C-3), 56.1 (OCH₃), 48.3 (NHCH₂CH₃), 28.0 (C(CH₃)₃), 20.8 (COCH₃), 20.6 (COCH₃), 20.4 (COCH₃), 18.7 (CHCH₃); HRMS (ESI) Calc. for (M + Na) C₂₀H₃₁NNaO₁₁: 484.1789 Found 484.1779.

N-(4-Methoxybenzyl) (2,3,4-tri-O-benzyl-α-D-galactopyranosid)uronamide (40): Scale 0.1 mmol. Prepared as described in general method 3. The crude residue was purified by flash chromatography (2:1 hexane–EtOAc) to afford **40** (50 mg, 83%) as a clear viscous oil. *R*_f 0.20 (2:1 hexane–EtOAc); [α]_D +60.1 (1.0 c, CHCl₃); ¹H NMR (400 MHz; CDCl₃): δ 7.39–7.26 (m, 15H, Ar), 7.11 (d, 2H, *J* = 8.5 Hz, Ar), 6.79 (app t, 1H, *J* = 5.6 Hz, CONHCH₂), 6.73 (d, 2H, *J* = 8.6 Hz, Ar), 4.92 (d, 1H, *J* = 10.9 Hz, PhCH₂O), 4.85 (d, 1H, *J* = 12.1 Hz, PhCH₂O), 4.81 (d, 1H, *J* = 11.8 Hz, PhCH₂O), 4.76 (d, 1H, *J* = 11.8 Hz, PhCH₂O), 4.70–4.67 (m, 2H, PhCH₂O, H-1), 4.58 (d, 1H, *J* = 10.9 Hz, PhCH₂O), 4.48–4.42 (m, 2H, H-3, NHCH₂Ar), 4.35–4.29 (m, 2H, NHCH₂Ar, H-5), 4.05–3.98 (m, 2H, H-2, H-4), 3.76 (s, 3H, ArOCH₃), 3.36 (s, 3H, OCH₃); ¹³C NMR (125 MHz; CDCl₃): δ 168.6 (C-6), 159.0 (Ar), 138.7 (Ar), 138.5 (2 × Ar), 129.7 (Ar), 129.1 (Ar), 128.4 (2 × Ar), 128.2 (Ar), 128.1 (Ar), 128.0 (Ar), 127.8 (Ar), 127.6 (Ar), 127.5 (2 × Ar), 114.0 (Ar), 99.2 (C-1), 78.4 (C-4), 76.6 (C-3), 75.7 (C-2), 75.4 (PhCH₂O), 73.7 (PhCH₂O), 73.0 (PhCH₂O), 71.3 (C-5), 55.9 (OCH₃), 55.3 (ArOCH₃), 42.6 (NHCH₂Ar); HRMS (ESI) Calc. for (M + H) C₃₆H₄₆NO₇: 598.2799; Found 598.2790.

N-(4-Methoxybenzyl) (2,3,4-tri-O-benzoyl-α-D-mannopyranosid)uronamide (41): Scale 0.1 mmol. Prepared as described in general method 3. The crude residue was purified by flash chromatography (3:2→1:1 hexane–EtOAc) to afford **41** (63 mg, 98%) as a clear viscous oil. *R*_f 0.43 (1:1 hexane–EtOAc); [α]_D –19.0 (1.0 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 8.07–8.05 (m, 4H, Ar), 7.87 (d, 2H, *J* = 7.4 Hz, Ar), 7.64–7.39 (m, 7H, Ar), 7.31–7.26 (m, 4H, Ar), 6.89 (d, 2H, *J* = 8.5 Hz, Ar), 6.80 (app t, 1H, *J* = 5.5 Hz, CONHCH₂), 6.04 (dd, 1H, *J* = 10.0, 10.0 Hz, H-4), 5.93 (dd, 1H, *J* = 10.1, 3.2 Hz, H-3), 5.69 (dd, 1H, *J* = 2.9, 1.9 Hz, H-2), 5.04 (d, 1H, *J* = 1.3 Hz, H-1), 4.58 (d, 1H, *J* = 10.0 Hz, H-5), 4.50 (dd, 1H, *J* = 14.5, 6.0 Hz, NHCH₂Ar), 4.38 (dd, 1H, *J* = 14.6, 5.4 Hz, NHCH₂Ar), 3.82 (s, 3H, ArOCH₃), 3.55 (s, 3H, OCH₃); ¹³C NMR (125 MHz; CDCl₃): δ 167.3 (C=O), 165.7 (C=O), 165.41 (C=O), 165.37 (C=O), 159.1 (Ar), 133.6 (Ar), 133.3 (Ar), 133.1 (Ar), 130.0 (Ar), 129.9 (2 × Ar), 129.8 (Ar), 129.5 (Ar), 129.2 (2 × Ar), 129.1 (Ar), 128.7 (Ar), 128.4 (Ar), 128.3 (Ar), 114.2 (Ar), 98.8 (C-1), 70.2 (C-2), 69.7 (C-5), 69.5 (C-3), 67.8 (C-4), 56.1 (OCH₃), 55.3 (ArOCH₃), 42.6 (NHCH₂Ar); HRMS (ESI) Calc. for (M + H) C₃₆H₃₄NO₁₀: 640.2177 Found 640.2163.

N-(4-Methoxybenzyl) (methyl 2,3-di-O-benzoyl-4-deoxy-β-L-threo-hex-4-enopyranosid)uronamide (42): To a solution of **23** (63 mg, 0.1 mmol) in CH₂Cl₂ (1 mL) was added *p*-methoxybenzylamine (19 μL, 0.15 mmol), followed by triethylamine (62 μL, 0.45 mmol). The reaction mixture

was stirred for 10 h at room temperature, then diluted with CH₂Cl₂ (10 mL), washed with sat. NH₄Cl (aq.), brine, dried over MgSO₄ and concentrated. The crude residue was purified by flash chromatography (2:1→3:2 hexane–EtOAc) to afford **42** (39 mg, 75%) as a clear viscous oil. *R*_f 0.19 (2:1 hexane–EtOAc); [α]_D +75.3 (1.2 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 8.06–8.01 (m, 4H, Ar), 7.59–7.54 (m, 2H, Ar), 7.46–7.41 (m, 4H, Ar), 7.29–7.26 (m, 2H, Ar), 6.93–6.90 (m, 2H, Ar), 6.80 (app t, 1H, *J* = 5.8 Hz, CONHCH₂), 6.30 (d, 1H, *J* = 3.0 Hz, H-4), 6.13 (dd, 1H, *J* = 8.0, 3.0 Hz, H-3), 5.54 (dd, 1H, *J* = 8.0, 2.5 Hz, H-2), 5.35 (d, 1H, *J* = 2.6 Hz, H-1), 4.57 (dd, 1H, *J* = 14.6, 6.1 Hz, NHCH₂Ar), 4.46 (dd, 1H, *J* = 14.6, 5.5 Hz, NHCH₂Ar), 3.83 (s, 3H, ArOCH₃), 3.54 (s, 3H, OCH₃); ¹³C NMR (125 MHz; CDCl₃): δ 165.8 (C=O), 165.6 (C=O), 160.5 (C=O), 159.2 (Ar), 143.7 (C-5), 133.6 (Ar), 133.3 (Ar), 130.0 (Ar), 129.8 (Ar), 129.8 (Ar), 129.5 (Ar), 129.3 (Ar), 129.0 (Ar), 128.5 (Ar), 128.4 (Ar), 114.2 (Ar), 104.9 (C-4), 98.9 (C-1), 69.9 (C-2), 66.8 (C-3), 57.1 (OCH₃), 55.3 (ArOCH₃), 43.0 (NHCH₂Ar); HRMS (ESI) Calc. for (M + Na) C₂₉H₂₇NNaO₈: 540.1629; Found 540.1633.

2-(Trimethylsilyl)ethyl 2,3,4-tri-O-acetyl-α-L-rhamnopyranosyl-(1→3)-4,6-O-benzylidene-β-D-glucopyranoside and 2-(Trimethylsilyl)ethyl 2,3,4-tri-O-acetyl-α-L-rhamnopyranosyl-(1→2)-4,6-O-benzylidene-β-D-glucopyranoside (45/45a): A mixture of trichloroacetimidate **44**^[43] (854 mg, 1.96 mmol), alcohol **43**^[42] (869 mg, 2.36 mmol), and 4 Å molecular sieves (4 g) in CH₂Cl₂ (80 mL) was stirred for 30 min at room temperature under an argon atmosphere. The mixture was cooled to –78 °C, then TBSOTf (46 μL, 0.2 mmol) was added. The reaction mixture was stirred for 20 min at –78 °C, then the TBSOTf was quenched by the addition of Et₃N (28 μL). The mixture was filtered through Celite and concentrated. The crude residue was purified by flash chromatography (2:1 hexane–EtOAc) to afford **45/45a** (904 mg, 72%, **45:45a** = 2:1) as a white solid, which was an inseparable mixture. *R*_f 0.27 (2:1 hexane–EtOAc); ¹H NMR (400 MHz; CDCl₃): δ 7.49–7.45 (m, 6H), 7.39–7.36 (m, 3H), 7.34–7.31 (m, 6H), 5.55 (s, 2H), 5.51 (s, 1H), 5.35–5.27 (m, 9H), 5.20 (d, 2H, *J* = 1.5 Hz, H-1' of **45**), 5.07 (t, 1H, *J* = 10.0 Hz), 4.97 (t, 2H, *J* = 10.0 Hz), 4.49 (d, 1H, *J* = 7.7 Hz, H-1 of **45a**), 4.40–4.32 (m, 7H), 4.20 (dq, 2H, *J* = 9.9, 6.3 Hz), 4.03–3.74 (m, 9H), 3.66–3.54 (m, 9H), 3.51–3.38 (m, 4H), 2.71 (d, 1H, *J* = 2.8 Hz), 2.41 (d, 1H, *J* = 2.7 Hz), 2.14 (s, 3H), 2.13 (s, 6H), 2.03 (s, 3H), 1.99 (s, 6H), 1.97 (s, 3H), 1.95 (s, 6H), 1.20 (d, 3H, *J* = 6.3 Hz), 1.06–0.92 (m, 6H), 0.85 (d, 6H, *J* = 6.2 Hz), 0.04 (s, 9H), 0.03 (s, 18H); ¹³C NMR (125 MHz; CDCl₃): δ 170.2, 170.2, 170.2, 170.0, 169.9, 137.2, 137.0, 129.3, 129.0, 128.4, 128.0, 126.3, 126.2, 103.0 (C-1 of **45**), 101.8, 101.7 (C-1 of **45a**), 101.6, 97.7 (C-1' of **45**), 97.6 (C-1' of **45a**), 80.7, 79.0, 77.1, 76.4, 75.7, 74.9, 71.3, 71.17, 69.7, 69.7, 69.3, 69.3, 68.8, 68.7, 68.2, 68.0, 66.8, 66.4, 66.0, 65.9, 21.0, 21.0, 20.8, 20.8, 18.5, 18.3, 17.3, 16.8, –1.4, –1.5; HRMS (ESI) Calc. for (M + Na) C₃₀H₄₄NaO₁₃Si: 663.2443; Found 663.2440.

2-(Trimethylsilyl)ethyl 2,3,4-tri-O-acetyl-α-L-rhamnopyranosyl-(1→3)-2-O-acetyl-4,6-O-benzylidene-β-D-glucopyranoside (S1) and 2-(Trimethylsilyl)ethyl 2,3,4-tri-O-acetyl-α-L-rhamnopyranosyl-(1→2)-3-O-acetyl-4,6-O-benzylidene-β-D-glucopyranoside (S1a): To a solution of **45/45a** (904 mg, 1.41 mmol) in pyridine (14 mL) and Ac₂O (7 mL) was added DMAP (17 mg, 0.14 mmol). The reaction mixture was stirred for 3 h at room temperature, then the Ac₂O was quenched by addition of CH₃OH (7 mL). Then, the mixture was co-concentrated with toluene (3 ×). The crude residue was purified by flash chromatography (1:1 hexane–EtOAc) to afford **S1/S1a** (934 mg, 97%, **S1:S1a** = 2:1) as a white solid, which was an inseparable mixture. *R*_f 0.56 (1:1 hexane–EtOAc); ¹H NMR (500 MHz; CDCl₃): δ 7.50–7.48 (m, 5H), 7.45–7.43 (m, 2H), 7.38–7.33 (m, 11H), 5.57 (s, 2H), 5.48 (s, 1H), 5.40 (t, 1H, *J* = 9.4 Hz), 5.34–5.29 (m, 4H), 5.11–5.05 (m, 5H), 5.04 (d, 1H, *J* = 1.9 Hz, H-1' of **S1a**), 4.99 (dd, 2H, *J* = 3.4, 1.8 Hz), 4.95 (t, 2H, *J* = 9.9 Hz), 4.91 (d, 2H, *J* = 1.7 Hz, H-1' of **S1**), 4.57 (d, 1H, *J* = 7.6 Hz, H-1 of **S1a**), 4.49 (d, 2H, *J* = 8.0 Hz, H-1 of **S1**), 4.41–4.37 (m, 3H), 4.32 (dq, 1H, *J* = 10.2, 6.3 Hz), 4.10 (dq, 2H, *J* = 10.0, 6.2 Hz), 4.04–3.88 (m, 5H), 3.87–3.75 (m, 3H), 3.75–3.62 (m, 4H), 3.62–3.44 (m, 6H), 2.15 (s, 3H), 2.14 (s, 6H), 2.14 (s, 3H), 2.12 (s, 6H), 2.05 (s, 3H), 2.02 (s, 3H), 2.00 (s, 6H), 1.98 (s, 6H), 1.21 (d, 3H, *J* = 6.2 Hz), 1.05–0.89 (m, 6H), 0.68 (d, 6H, *J* = 6.2 Hz), 0.06 (s, 9H), 0.03 (s, 18H); ¹³C NMR (125

FULL PAPER

MHz; CDCl₃): δ 170.0, 170.0, 169.8, 169.4, 137.1, 136.9, 129.2, 129.0, 128.2, 128.1, 126.4, 126.1, 102.0, 101.7 (C-1 of **S1a**), 101.4, 101.0 (C-1 of **S1**), 97.7 (C-1' of **S1a**), 97.6 (C-1' of **S1**), 79.1, 78.7, 73.9, 73.5, 71.4, 71.4, 70.6, 70.3, 68.8, 68.7, 68.6, 68.4, 68.1, 67.6, 66.6, 66.6, 66.3, 66.2, 25.0, 20.9, 20.9, 20.8, 20.8, 20.8, 18.4, 18.1, 17.3, 16.5, -1.42, -1.49; HRMS (ESI) Calc. for (M + Na) C₃₂H₄₆NaO₁₄Si: 705.2549; Found 705.2541.

2-(Trimethylsilyl)ethyl 2,3,4-tri-O-acetyl- α -L-rhamnopyranosyl-(1 \rightarrow 3)-2-O-acetyl- β -D-glucopyranoside (46) and 2-(Trimethylsilyl)ethyl 2,3,4-tri-O-acetyl- α -L-rhamnopyranosyl-(1 \rightarrow 3)-3-O-acetyl- β -D-glucopyranoside (46a):

To a solution of **S1/S1a** (767 mg, 1.12 mmol) in 80% CH₃COOH–H₂O (10 mL) was heated at 80 °C for 2 h and then cooled to room temperature. The mixture was concentrated and then co-evaporated with toluene (3 \times). The crude residue was purified by flash chromatography (1:1 \rightarrow 1:2 hexane–EtOAc) to afford **46** (326 mg, 49%) as a white amorphous solid and **46a** (166 mg, 25%) as a white amorphous solid. Data for **46**: *R*_f 0.45 (1:2 hexane–EtOAc); [α]_D –17.7 (1.5 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 5.22 (dd, 1H, *J* = 10.1, 3.3 Hz, H-3'), 5.15 (dd, 1H, *J* = 3.3, 2.1 Hz, H-2'), 5.11 (app t, 1H, *J* = 9.9 Hz, H-4'), 4.96 (dd, 1H, *J* = 9.5, 8.1 Hz, H-2), 4.90 (d, 1H, *J* = 2.0 Hz, H-1'), 4.47 (d, 1H, *J* = 8.1 Hz, H-1), 4.15 (dq, 1H, *J* = 9.8, 6.5 Hz, H-5'), 4.00–3.93 (m, 2H, H-6a, OCH₂CH₂Si(CH₃)₃), 3.84 (ddd, 1H, *J* = 11.7, 7.4, 4.8 Hz, H-6b), 3.65 (app td, 1H, *J* = 9.0, 2.4 Hz, H-4), 3.61–3.54 (m, 3H, H-3, OCH₂CH₂Si(CH₃)₃, 4-OH), 3.38 (ddd, 1H, *J* = 8.2, 4.6, 3.4 Hz, H-5'), 2.16 (s, 3H, COCH₃), 2.16 (s, 3H, COCH₃), 2.09 (t, 1H, *J* = 7.2 Hz, 6-OH), 2.06 (s, 3H, COCH₃), 2.00 (s, 3H, COCH₃), 1.26 (d, 3H, *J* = 6.3 Hz, H-6'), 0.98 (ddd, 1H, *J* = 14.1, 10.6, 6.7 Hz, OCH₂CH₂Si(CH₃)₃), 0.91 (ddd, 1H, *J* = 14.0, 10.2, 5.6 Hz, OCH₂CH₂Si(CH₃)₃), 0.03 (s, 9H, OCH₂CH₂Si(CH₃)₃); ¹³C NMR (125 MHz; CDCl₃): δ 170.1 (C=O), 169.9 (C=O), 169.6 (C=O), 169.4 (C=O), 100.3 (C-1), 99.1 (C-1'), 85.9 (C-3), 75.1 (C-5), 71.3 (C-2), 70.6 (C-4'), 70.2 (C-4), 69.8 (C-2'), 68.6 (C-3'), 67.9 (C-5'), 67.4 (OCH₂CH₂Si(CH₃)₃), 62.4 (C-6), 20.9 (COCH₃), 20.8 (COCH₃), 20.7 (COCH₃), 20.7 (COCH₃), 18.0 (OCH₂CH₂Si(CH₃)₃), 17.5 (C-6'), -1.4 (OCH₂CH₂Si(CH₃)₃); HRMS (ESI) Calc. for (M + Na) C₂₅H₄₂NaO₁₄Si: 617.2236 Found 617.2228. Data for **46a**: *R*_f 0.42 (1:2 hexane–EtOAc); [α]_D –28.0 (3.2 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 5.27 (dd, 1H, *J* = 10.1, 3.4 Hz, H-3'), 5.08 (app t, 1H, *J* = 10.1 Hz, H-4'), 5.06 (dd, 1H, *J* = 3.7, 1.8 Hz, H-2'), 5.02 (app t, 1H, *J* = 9.4 Hz, H-3), 5.00 (d, 1H, *J* = 1.8 Hz, H-1'), 4.48 (d, 1H, *J* = 7.8 Hz, H-1), 4.33 (dq, 1H, *J* = 10.1, 6.2 Hz, H-5'), 3.98 (td, 1H, *J* = 9.8, 8.1 Hz, OCH₂CH₂Si(CH₃)₃), 3.93 (dd, 1H, *J* = 11.9, 3.5 Hz, H-6a), 3.83 (dd, 1H, *J* = 11.9, 4.7 Hz, H-6b), 3.65–3.60 (m, 3H, H-2, H-4, OCH₂CH₂Si(CH₃)₃), 3.41 (ddd, 1H, *J* = 9.5, 4.6, 3.7 Hz, H-5), 3.16 (br s, 1H, 4-OH) 2.18–2.14 (m, 7H, 2 \times COCH₃, 6-OH), 2.04 (s, 3H, COCH₃), 2.01 (s, 3H, COCH₃), 1.19 (d, 3H, *J* = 6.2 Hz, H-6'), 1.04–0.98 (m, 2H, OCH₂CH₂Si(CH₃)₃), 0.04 (s, 9H, OCH₂CH₂Si(CH₃)₃); ¹³C NMR (125 MHz; CDCl₃): δ 172.7 (C=O), 170.1 (C=O), 170.0 (C=O), 169.9 (C=O), 101.0 (C-1), 97.6 (C-1'), 78.8 (C-3), 76.1 (C-2), 75.6 (C-5), 71.3 (C-4'), 70.2 (C-2'), 70.2 (C-4), 68.7 (C-3'), 67.9 (OCH₂CH₂Si(CH₃)₃), 66.5 (C-5'), 62.3 (C-6), 20.9 (COCH₃), 20.9 (COCH₃), 20.8 (COCH₃), 20.7 (COCH₃), 18.4 (OCH₂CH₂Si(CH₃)₃), 17.3 (C-6'), -1.5 (OCH₂CH₂Si(CH₃)₃); HRMS (ESI) Calc. for (M + Na) C₂₅H₄₂NaO₁₄Si: 617.2236; Found 617.2230.

2-(Trimethylsilyl)ethyl 2,3,4-tri-O-acetyl- α -L-rhamnopyranosyl-(1 \rightarrow 3)-2-O-acetyl-6-thio-6-S-p-tolyl- β -D-glucopyranoside (47):

To a stirred solution of **46** (128 mg, 0.216 mmol) in dry pyridine (1.1 mL) were added (TolS)₂ (108 mg, 0.44 mmol) and P(CH₃)₃ (0.44 mL, 0.44 mmol, 1.0 M in THF) under argon. The reaction mixture was stirred overnight at room temperature. Then, the reaction mixture was concentrated, diluted with EtOAc (30 mL) and washed with brine. The organic layer was dried over MgSO₄, filtered and concentrated. The crude residue was purified by flash chromatography (2:1 hexane–EtOAc) to afford **47** (144 mg, 95%) as a viscous oil. *R*_f 0.18 (2:1 hexane–EtOAc); [α]_D +1.7 (0.3 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 7.34–7.32 (m, 2H, Ar), 7.12–7.09 (m, 2H, Ar), 5.22 (dd, 1H, *J* = 10.0, 3.3 Hz, H-3'), 5.14 (dd, 1H, *J* = 3.2, 2.1 Hz, H-2'), 5.10 (app t, 1H, *J* = 9.9 Hz, H-4'), 4.98 (dd, 1H, *J* = 9.1, 8.4 Hz, H-2), 4.87 (d, 1H, *J* = 2.1 Hz, H-1'), 4.39 (d, 1H, *J* = 8.1 Hz, H-1), 4.12 (dq, 1H, *J* = 9.8, 6.3 Hz, H-5'), 3.92 (td, 1H, *J* = 9.9, 5.7 Hz, OCH₂CH₂Si(CH₃)₃), 3.57–3.41

(m, 6H, OCH₂CH₂Si(CH₃)₃, H-3, H-4, H-5, H-6a, 4-OH), 3.09 (dd, 1H, *J* = 13.7, 7.7 Hz, H-6b), 2.33 (s, 3H, Ar-CH₃), 2.16 (s, 3H, COCH₃), 2.15 (s, 3H, COCH₃), 2.07 (s, 3H, COCH₃), 2.00 (s, 3H, COCH₃), 1.25 (d, 3H, *J* = 6.3 Hz, H-6'), 0.96 (ddd, 1H, *J* = 14.0, 10.3, 6.8 Hz, OCH₂CH₂Si(CH₃)₃), 0.90 (ddd, 1H, *J* = 13.7, 9.9, 5.6 Hz, OCH₂CH₂Si(CH₃)₃), 0.02 (s, 9H, OCH₂CH₂Si(CH₃)₃); ¹³C NMR (125 MHz; CDCl₃): δ 170.0 (C=O), 169.9 (C=O), 169.5 (C=O), 169.4 (C=O), 136.1 (Ar), 132.9 (Ar), 129.8 (Ar), 129.7 (Ar), 100.0 (C-1), 99.2 (C-1'), 86.3 (C-3), 74.7 (C-5), 72.7 (C-4), 71.4 (C-2), 70.6 (C-4'), 69.8 (C-2'), 68.6 (C-3'), 67.9 (C-5'), 67.0 (OCH₂CH₂Si(CH₃)₃), 36.2 (C-6), 21.0 (Ar-CH₃), 20.9 (COCH₃), 20.8 (COCH₃), 20.7 (COCH₃), 20.7 (COCH₃), 17.9 (OCH₂CH₂Si(CH₃)₃), 17.5 (C-6'), -1.4 (OCH₂CH₂Si(CH₃)₃); HRMS (ESI) Calc. for (M + Na) C₃₂H₄₈NaO₁₃SSi: 723.2477; Found 723.2472.

2-(Trimethylsilyl)ethyl 2,3,4-tri-O-acetyl- α -L-rhamnopyranosyl-(1 \rightarrow 3)-2,4-di-O-acetyl-6-thio-6-S-p-tolyl- β -D-glucopyranoside (48):

To a solution of **47** (143.6 mg, 0.205 mmol) in pyridine (2 mL) and Ac₂O (1 mL) was added DMAP (2.6 mg, 0.021 mmol). The reaction mixture was stirred for 3 h at room temperature, then the Ac₂O was quenched by addition of CH₃OH (1 mL). Then the mixture was co-concentrated with toluene (3 \times). The crude residue was purified by flash chromatography (2:1 hexane–EtOAc) to afford **48** (145 mg, 95%) as a white amorphous solid. *R*_f 0.19 (2:1 hexane–EtOAc); [α]_D –10.7 (0.3 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 7.28–7.26 (m, 2H, Ar), 7.12–7.10 (m, 2H, Ar), 5.12–5.09 (m, 2H, H-3', H-2'), 5.03 (dd, 1H, *J* = 9.7, 8.1 Hz, H-2), 5.02 (app t, 1H, *J* = 9.8 Hz, H-4'), 4.98 (app t, 1H, *J* = 9.4 Hz, H-4), 4.80 (d, 1H, *J* = 1.2 Hz, H-1'), 4.38 (d, 1H, *J* = 8.0 Hz, H-1), 3.93–3.86 (m, 2H, H-5', OCH₂CH₂Si(CH₃)₃), 3.75 (app t, 1H, *J* = 9.3 Hz, H-3), 3.55 (td, 1H, *J* = 9.9, 6.6 Hz, OCH₂CH₂Si(CH₃)₃), 3.46 (ddd, 1H, *J* = 9.7, 8.3, 3.4 Hz, H-5), 3.05 (dd, 1H, *J* = 14.0, 8.2 Hz, H-6a), 2.99 (dd, 1H, *J* = 13.9, 3.4 Hz, H-6b), 2.33 (s, 3H, Ar-CH₃), 2.16 (s, 3H, COCH₃), 2.15 (s, 3H, COCH₃), 2.11 (s, 3H, COCH₃), 2.05 (s, 3H, COCH₃), 1.96 (s, 3H, COCH₃), 1.16 (d, 3H, *J* = 6.2 Hz, H-6'), 0.96 (ddd, 1H, *J* = 13.9, 10.3, 6.8 Hz, OCH₂CH₂Si(CH₃)₃), 0.90 (ddd, 1H, *J* = 14.0, 10.0, 5.8 Hz, OCH₂CH₂Si(CH₃)₃), 0.02 (s, 9H, OCH₂CH₂Si(CH₃)₃); ¹³C NMR (125 MHz; CDCl₃): δ 170.2 (C=O), 170.1 (C=O), 169.8 (C=O), 169.5 (C=O), 169.4 (C=O), 136.5 (Ar), 132.5 (Ar), 130.1 (Ar), 129.7 (Ar), 100.1 (C-1), 99.5 (C-1'), 81.9 (C-3), 73.5 (C-5), 73.2 (C-4), 71.9 (C-4'), 70.6 (C-2), 69.9 (C-2'), 68.9 (C-3'), 67.5 (C-5'), 67.1 (OCH₂CH₂Si(CH₃)₃), 36.5 (C-6), 21.3 (COCH₃), 21.0 (Ar-CH₃), 20.9 (COCH₃), 20.8 (COCH₃), 20.8 (COCH₃), 20.6 (COCH₃), 17.9 (OCH₂CH₂Si(CH₃)₃), 17.2 (C-6'), -1.4 (OCH₂CH₂Si(CH₃)₃); HRMS (ESI) Calc. for (M + Na) C₃₄H₅₀NaO₁₄SSi: 765.2583; Found 765.2580.

2-(Trimethylsilyl)ethyl 2,3,4-tri-O-acetyl- α -L-rhamnopyranosyl-(1 \rightarrow 3)-S-p-tolyl thio(2,4-di-O-acetyl- β -D-glucopyranosid)uronate (49):

To a solution of **48** (136 mg, 0.18 mmol) in dry CCl₄ (2.7 mL) at 0 °C were added anhydrous pyridine (29 μ L, 0.36 mmol) followed by the addition of SO₂Cl₂ (29 μ L, 0.36 mmol) under argon. The reaction mixture was stirred for 5 h at 0 °C and then diluted with CH₂Cl₂ and was washed with brine. The organic layer was dried over MgSO₄, filtered and concentrated. The crude residue was dissolved in 3:2 acetone–H₂O (3.6 mL), and solid NaHCO₃ (60.5 mg, 0.72 mmol) was added. The reaction mixture was stirred overnight at room temperature, diluted with brine and then extracted with EtOAc (3 \times 5 mL). The organic layer was dried over MgSO₄, filtered and concentrated. The crude residue was purified by flash chromatography (2:1 hexane–EtOAc) to afford **49** (116 mg, 85%) as a white amorphous solid. *R*_f 0.45 (1:1 hexane–EtOAc); [α]_D –66.8 (0.6 c, CHCl₃); ¹H NMR (500 MHz; CDCl₃): δ 7.31–7.29 (m, 2H, Ar), 7.25–7.23 (m, 2H, Ar), 5.32 (dd, 1H, *J* = 9.7, 9.3 Hz, H-4), 5.16 (dd, 1H, *J* = 9.3, 7.9 Hz, H-2), 5.13–5.11 (m, 2H, H-2', H-3'), 5.04 (app t, 1H, *J* = 9.8 Hz, H-4'), 4.86 (d, 1H, *J* = 1.5 Hz, H-1'), 4.56 (d, 1H, *J* = 7.9 Hz, H-1), 4.08 (td, 1H, *J* = 9.6, 6.7 Hz, OCH₂CH₂Si(CH₃)₃), 3.99 (d, 1H, *J* = 9.9 Hz, H-5), 3.92 (dq, 1H, *J* = 9.7, 6.1 Hz, H-5'), 3.86 (app t, 1H, *J* = 9.2 Hz, H-3), 3.68 (td, 1H, *J* = 9.5, 6.8 Hz, OCH₂CH₂Si(CH₃)₃), 2.39 (s, 3H, Ar-CH₃), 2.19 (s, 3H, COCH₃), 2.15 (s, 3H, COCH₃), 2.06 (s, 3H, COCH₃), 2.05 (s, 3H, COCH₃), 1.98 (s, 3H, COCH₃), 1.16 (d, 3H, *J* = 6.2 Hz, H-6'), 1.03 (ddd, 1H, *J* = 14.0, 9.4, 6.8 Hz, OCH₂CH₂Si(CH₃)₃), 0.99 (ddd, 1H, *J* = 14.0, 9.4, 6.8 Hz,

FULL PAPER

$\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$, 0.06 (s, 9H, $\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$); ^{13}C NMR (125 MHz; CDCl_3): δ 195.5 (C-6), 170.2 (C=O), 170.1 (C=O), 169.5 (C=O), 169.4 (C=O), 169.3 (C=O), 139.9 (Ar), 134.8 (Ar), 130.2 (Ar), 122.8 (Ar), 100.2 (C-1), 99.4 (C-1'), 81.1 (C-3), 77.7 (C-5), 71.4 (C-2), 70.8 (C-4), 70.6 (C-4'), 69.9 (C-2'), 68.8 (C-3'), 67.5 ($\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 67.5 (C-5'), 21.4 (Ar-CH₃), 21.0 (COCH_3), 21.0 (COCH_3), 20.8 (COCH_3), 20.8 (COCH_3), 20.7 (COCH_3), 18.0 ($\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 17.3 (C-6'), -1.3 ($\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$); HRMS (ESI) Calc. for (M + Na) $\text{C}_{34}\text{H}_{52}\text{NO}_{15}\text{Si}$: 774.2807 Found 774.2821.

N-{2-(Trimethylsilyl)ethyl 2,3,4-tri-O-acetyl- α -L-rhamnopyranosyl-(1 \rightarrow 3)-(2,4-di-O-acetyl- β -D-glucopyranosiduronoyl)}-L-threonine benzyl ester (50):

To a solution of **49** (79.8 mg, 0.105 mmol) in CH_2Cl_2 (2.1 mL) was added L-threonine benzyl ester (33.1 mg, 0.158 mmol). The reaction mixture was stirred for 44 h at room temperature, then diluted with CH_2Cl_2 (10 mL), washed with sat. NH_4Cl (aq.) and brine. The organic layer was dried over MgSO_4 , filtered and concentrated. The crude residue was purified by flash chromatography (2:1 \rightarrow 1:1 hexane-EtOAc) to afford **50** (73.5 mg, 83%) as a white amorphous solid. R_f 0.29 (1:1 hexane-EtOAc); $[\alpha]_D -23.1$ (0.6 c, CHCl_3); ^1H NMR (400 MHz; CDCl_3): δ 7.38–7.31 (m, 5H, Ar), 7.19 (d, 1H, $J = 8.8$ Hz, NHCHCO_2Bn), 5.22 (d, 1H, $J = 12.3$ Hz, CH_2Ph), 5.17 (d, 1H, $J = 12.3$ Hz, CH_2Ph), 5.14 (app t, 1H, $J = 9.4$ Hz, H-4), 5.11–5.06 (m, 3H, H-2, H-2', H-3'), 5.01 (app t, 1H, $J = 9.7$ Hz, H-4'), 4.86 (d, 1H, $J = 1.2$ Hz, H-1'), 4.55 (dd, 1H, $J = 8.7, 3.3$ Hz, NHCHCO_2Bn), 4.50 (d, 1H, $J = 8.0$ Hz, H-1), 4.29 (ddq, 1H, $J = 6.3, 5.5, 3.7$ Hz, HOCHCH_3), 3.97 (td, 1H, $J = 9.7, 6.4$ Hz, $\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 3.91–3.84 (m, 3H, H-5', H-3, H-5), 3.58 (td, 1H, $J = 9.6, 6.8$ Hz, $\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 2.16 (s, 3H, COCH_3), 2.13 (s, 3H, COCH_3), 2.10 (s, 3H, COCH_3), 2.06 (d, 1H, $J = 5.5$ Hz, HOCHCH_3), 2.02 (s, 3H, COCH_3), 1.95 (s, 3H, COCH_3), 1.24 (d, 3H, $J = 6.4$ Hz, HOCHCH_3), 1.14 (d, 3H, $J = 6.2$ Hz, H-6'), 0.97 (ddd, $J = 14.2, 9.7, 6.7$ Hz, $\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 0.92 (ddd, $J = 14.4, 10.0, 6.6$ Hz, $\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 0.02 (s, 9H, $\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$); ^{13}C NMR (125 MHz; CDCl_3): δ 170.1 (C=O), 170.1 (C=O), 170.0 (C=O), 169.9 (C=O), 169.6 (C=O), 169.4 (C=O), 167.6 (C-6), 135.2 (Ar), 128.65 (Ar), 128.50 (Ar), 128.2 (Ar), 100.2 (C-1), 99.1 (C-1'), 80.4 (C-3), 72.8 (C-5), 71.7 (C-2), 71.1 (C-4), 70.6 (C-4'), 69.9 (C-2'), 68.8 (HOCHCH_3), 68.5 (C-3'), 68.0 ($\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 67.4 (C-5'), 67.3 (CH_2Ph), 57.2 (NHCHCO_2Bn), 21.0 (COCH_3), 20.9 (COCH_3), 20.8 (COCH_3), 20.8 (COCH_3), 20.7 (COCH_3), 19.9 (HOCHCH_3), 18.0 ($\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 17.3 (C-6'), -1.4 ($\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$); HRMS (ESI) Calc. for (M + Na) $\text{C}_{38}\text{H}_{55}\text{NNaO}_{18}\text{Si}$: 864.3081; Found 864.3076.

N-{2-(Trimethylsilyl)ethyl 2,3,4-tri-O-acetyl- α -L-rhamnopyranosyl-(1 \rightarrow 3)-(2,4-di-O-acetyl- β -D-glucopyranosiduronoyl)}-L-threonine (51):

To a solution of **50** (67.4 mg, 0.08 mmol) in CH_3OH was added palladium on charcoal (5%, 10 mg). The flask was flushed with H_2 gas, and then the reaction mixture was stirred at room temperature overnight under a H_2 atmosphere. The mixture was filtered through Celite, and the filtrate was concentrated to afford **51** (60.2 mg, quant.) as a white solid. R_f 0.30 (5:1 CH_2Cl_2 - CH_3OH); $[\alpha]_D -30.2$ (1.6 c, CHCl_3); ^1H NMR (500 MHz; CD_3OD): δ 5.14 (app t, 1H, $J = 9.6$ Hz, H-4), 5.08 (dd, 1H, $J = 3.3, 2.0$ Hz, H-2'), 5.04 (dd, 1H, $J = 10.1, 3.3$ Hz, H-3'), 5.00 (dd, 1H, $J = 9.6, 8.1$ Hz, H-2), 4.96 (app t, 1H, $J = 9.6$ Hz, H-4) 4.92 (d, 1H, $J = 1.9$ Hz, H-1'), 4.68 (d, 1H, $J = 8.1$ Hz, H-1), 4.31–4.28 (m, 2H, NHCHCO_2H , HOCHCH_3), 4.07 (app t, 1H, $J = 9.5$ Hz, H-3), 4.04 (td, 1H, $J = 9.8, 5.9$ Hz, $\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 4.02 (d, 1H, $J = 9.9$ Hz, H-5), 3.85 (dq, 1H, $J = 9.7, 6.2$ Hz, H-5'), 3.64 (td, 1H, $J = 9.6, 6.7$ Hz, $\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 2.12 (s, 3H, COCH_3), 2.12 (s, 3H, COCH_3), 2.06 (s, 3H, s, 3H, COCH_3), 2.02 (s, 3H, s, 3H, COCH_3), 1.93 (s, 3H, s, 3H, COCH_3), 1.19 (d, 3H, $J = 6.3$ Hz, HOCHCH_3), 1.12 (d, 3H, $J = 6.2$ Hz, H-6'), 0.97 (ddd, 1H, $J = 14.0, 9.9, 6.7$ Hz, $\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 0.89 (ddd, 1H, $J = 14.0, 9.6, 5.8$ Hz, $\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 0.02 (s, 9H, $\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$); ^{13}C NMR (125 MHz; CD_3OD): δ 174.5 (C=O), 171.7 (C=O), 171.7 (C=O), 171.4 (C=O), 171.4 (C=O), 171.3 (C=O), 169.6 (C-6), 101.3 (C-1), 100.6 (C-1'), 81.9 (C-3), 73.7 (C-5), 73.4 (C-2), 72.5 (C-4), 71.9 (C-4'), 71.1 (C-2'), 70.3 (C-3'), 68.7 (HOCHCH_3), 68.6 ($\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 58.9 (NHCHCO_2H), 21.4 (COCH_3), 21.1 (COCH_3), 20.7 (COCH_3), 20.6 (COCH_3), 20.6 (COCH_3), 20.3 (HOCHCH_3), 18.9

($\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 17.7 (C-6'), -1.3 ($\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$); HRMS (ESI) Calc. for (M + Na) $\text{C}_{31}\text{H}_{49}\text{NNaO}_{16}\text{Si}$: 774.2611; Found 774.2600.

N-{2-(Trimethylsilyl)ethyl α -L-rhamnopyranosyl-(1 \rightarrow 3)- β -D-glucopyranosiduronoyl}-L-threonine (52):

To a solution of **51** (60.2 mg, 0.08 mmol) in CH_3OH (1 mL) was added a solution of NaOCH_3 in CH_3OH (1 mL, 0.1 M). The reaction mixture was stirred for 4 h at room temperature and then neutralized by addition of Amberlite® IR-120 (H^+) cation exchange resin. The mixture was filtered and the filtrate was concentrated. The residue was dissolved in water and then lyophilized to afford **52** (39.0 mg, 90%) as a white solid. R_f 0.24 (1:2:4 H_2O -2-propanol-EtOAc); $[\alpha]_D -53.2$ (0.3 c, CH_3OH); ^1H NMR (500 MHz; CD_3OD): δ 5.18 (d, 1H, $J = 1.6$ Hz, H-1'), 4.45 (br s, 1H, NHCHCO_2H), 4.39 (d, 1H, $J = 7.9$ Hz, H-1), 4.36 (br s, 1H, HOCHCH_3), 4.07 (dq, 1H, $J = 9.6, 6.2$ Hz, H-5'), 4.00 (ddd, 1H, $J = 11.0, 9.7, 6.0$ Hz, $\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 3.92 (dd, 1H, $J = 3.3, 1.7$ Hz, H-2'), 3.87 (d, 1H, $J = 9.3$ Hz, H-5), 3.71–3.65 (m, 2H, H-3', $\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 3.59 (app t, 1H, $J = 8.7$ Hz, H-3), 3.54 (app t, 1H, $J = 9.0$ Hz, H-4), 3.37 (app t, 1H, $J = 9.7$ Hz, H-4'), 3.34 (dd, 1H, $J = 8.5$ Hz, H-2), 1.23 (d, 3H, $J = 6.2$ Hz, H-6'), 1.19 (d, 3H, $J = 6.3$ Hz, HOCHCH_3), 1.05 (ddd, 1H, $J = 13.8, 11.0, 5.8$ Hz, $\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 0.99 (ddd, 1H, $J = 13.8, 11.0, 6.0$ Hz, $\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 0.03 (s, 9H, $\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$); ^{13}C NMR (125 MHz; CD_3OD): δ 173.4 (C=O), 172.2 (C-6), 103.8 (C-1), 102.5 (C-1'), 83.0 (C-3), 76.0 (C-5), 75.5 (C-2), 74.1 (C-4'), 72.4 (C-4), 72.4 (C-2'), 72.3 (C-3'), 69.9 (C-5'), 68.6 ($\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 68.4 (HOCHCH_3), 58.6 (NHCHCO_2H), 20.6 (HOCHCH_3), 19.2 ($\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$), 17.9 (C-6'), -1.4 ($\text{OCH}_2\text{CH}_2\text{Si}(\text{CH}_3)_3$); HRMS (ESI) Calc. for (M - H) $\text{C}_{21}\text{H}_{38}\text{NO}_{13}\text{Si}$: 540.2118; Found 540.2113

Acknowledgements

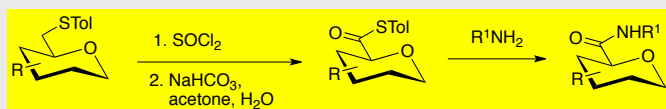
This work was supported by the Natural Sciences and Engineering Research Council of Canada and the Alberta Glycomics Centre. RAA thanks Alberta Innovates Health Solutions for a studentship award.

Keywords: glycuronamide • oxidation–amidation • synthesis • uronic acid • thioester

- [1] E. Vinogradov, W. J. Conlan, J. S. Gunn, M. B. Perry, *Carbohydr. Res.* **2004**, *339*, 649–654.
- [2] E. V. Vinogradov, D. Krajewska-Pietrasik, W. Kaca, A. S. Shashkov, Y. A. Knirel, N. K. Kochetkov, *Eur. J. Biochem.* **1989**, *185*, 645–650.
- [3] M. Siwińska, E. A. Levina, O. G. Ovchinnikova, D. Drzewiecka, A. S. Shashkov, A. Różalski and Y. A. Knirel, *Carbohydr. Res.* **2015**, *407*, 131–136.
- [4] S. Carillo, A. Casillo, G. Pieretti, E. Parrilli, F. Sannino, M. Bayer-Giraldi, S. Cosconati, E. Novellino, M. Ewert, J. W. Deming, R. Lanzetta, G. Marino, M. Parrilli, A. Randazzo, M. L. Tutino, M. M. Corsaro, *J. Am. Chem. Soc.* **2015**, *137*, 179–189.
- [5] G. O. Aspinall, S. Fujimoto, A. G. McDonald, H. Pang, L. A. Kurjanczyk, J. L. Penner, *Infect. Immun.* **1994**, *62*, 2122–2125.
- [6] Y. A. Knirel, E. V. Vinogradov, A. S. Shashkov, Z. Sidorczyk, A. Rozalski, J. Radziejewska-Lebrecht, W. Kaca, *J. Carbohydr. Chem.* **1993**, *12*, 379–414.
- [7] J. Radziejewska-Lebrecht, A. S. Shashkov, E. V. Vinogradov, H. Grosskurth, B. Bartodziejska, A. Rozalska, W. Kaca, L. O. Kononov, A. Y. Chernyak, H. Mayer, Y. A. Knirel, N. K. Kochetkov, *Eur. J. Biochem.* **1995**, *230*, 705–712.
- [8] F. S. Michael, C. M. Szymanski, J. Li, K. H. Chan, N. H. Khieu, S. Laroque, W. W. Wakarchuk, J.-R. Brisson, M. A. Monteiro, *Eur. J. Biochem.* **2002**, *269*, 5119–5136.
- [9] P. Hofmann, B. Jann, K. Jann, *Carbohydr. Res.* **1985**, *139*, 261–271.

FULL PAPER

- [10] A. O. Tzianabos, A. B. Onderdonk, D. F. Zaleznik, R. S. Smith, D. L. Kasper, *Infect. Immun.* **1994**, *62*, 4881–4886.
- [11] W. Gromska, H. Mayer, *Eur. J. Biochem.* **1976**, *62*, 391–399.
- [12] W. Kaca, Y.A. Knirel, E. V. Vinogradov, K. Kotelko, *Arch. Immunol. Ther. Exp. (Warsz)*. **1987**, *35*, 431–437.
- [13] Z. Sidorczyk, A. Swierzko, Y. A. Knirel, E. V. Vinogradov, A. Y. Chernyak, L. O. Kononov, M. Cedzynski, A. Rozalski, W. Kaca, A. S. Shashkov, N. K. Kochetkov, *Eur. J. Biochem.* **1995**, *230*, 713–721.
- [14] A. Y. Chernyak, L. O. Kononov, N. K. Kochetkov, *Carbohydr. Res.* **1991**, *216*, 381–398.
- [15] N. Röckendorf, T. K. Lindhorst, *J. Org. Chem.* **2004**, *69*, 4441–4445.
- [16] M. Tosin, P. V. Murphy, *Org. Lett.* **2002**, *4*, 3675–3678.
- [17] J. Wang, B. Elchert, Y. Hui, J. Y. Takemoto, M. Bensaci, J. Wennergren, H. Chang, R. Rai, C.-W. T. Chang, *Bioorg. Med. Chem.* **2004**, *12*, 6397–6413.
- [18] J. G. Taylor, X. Li, M. Oberthür, W. Zhu, D. E. Kahne, *J. Am. Chem. Soc.* **2006**, *128*, 15084–15085.
- [19] D. Doyle, P. V. Murphy, *Carbohydr. Res.* **2008**, *343*, 2535–2544.
- [20] R. Leyden, T. Velasco-Torrijos, S. André, S. Gouin, H.-J. Gabius, P. V. Murphy, *J. Org. Chem.* **2009**, *74*, 9010–9026.
- [21] D. Crich, K. Sasaki, *Org. Lett.* **2009**, *11*, 3514–3517.
- [22] M. Bosco, S. Rat, J. Kovensky, A. Wadouachi, *Tetrahedron Lett.* **2010**, *51*, 2553–2556.
- [23] N. M. Xavier, S. Schwarz, P. D. Vaz, R. Csuk, A. P. Rauter, *Eur. J. Org. Chem.* **2014**, *2014*, 2770–2779.
- [24] N. Xavier, S. Lucas, R. Jorda, S. Schwarz, A. Loesche, R. Csuk, M. C. Oliveira, *Synlett* **2015**, *26*, 2663–2672.
- [25] C. Gunanathan, Y. Ben-David, D. Milstein, *Science* **2007**, *317*, 790–792.
- [26] L. U. Nordstrøm, H. Vogt, R. Madsen, *J. Am. Chem. Soc.* **2008**, *130*, 17672–17673.
- [27] J. H. Dam, G. Osztrovszky, L. U. Nordstrøm, R. Madsen, *Chem. Eur. J.* **2010**, *16*, 6820–6827.
- [28] K. S. Goh, C.-H. Tan, *RSC Adv.* **2012**, *2*, 5536–5538.
- [29] S. C. Ghosh, J. S. Y. Ngiam, C. L. L. Chai, A. M. Seayad, T. T. Dang, A. Chen, *Adv. Synth. Catal.* **2012**, *354*, 1407–1412.
- [30] C. Chen, Y. Zhang, S. H. Hong, *J. Org. Chem.* **2011**, *76*, 10005–10010.
- [31] S. Muthaiah, S. C. Ghosh, J.-E. Jee, C. Chen, J. Zhang, S. H. Hong, *J. Org. Chem.* **2010**, *75*, 3002–3006.
- [32] A. J. A. Waston, A. C. Maxwell, J. M. J. Williams, *Org. Lett.* **2009**, *11*, 2667–2670.
- [33] G. Wang, Q.-Y. Yu, J. Wang, S. Wang, S.-Y. Chen, X.-Q. Yu, *RSC Adv.* **2013**, *3*, 21306–21310.
- [34] H. Yao, Y. Tang, K. Yamamoto, *Tetrahedron Lett.* **2012**, *53*, 5094–5098.
- [35] A. Schulze, A. Giannis, *Adv. Synth. Catal.* **2004**, *346*, 252–256.
- [36] B. A. Granger, I. T. Jewett, J. D. Butler, B. Hua, C. E. Knezevic, E. I. Pakinson, P. J. Hergenrother, S. F. Martin, *J. Am. Chem. Soc.* **2013**, *135*, 12984–12986.
- [37] B. Yu, X. Zhu, Y. Hui, *Org. Lett.* **2000**, *2*, 2539–2541.
- [38] B. Yu, X. Zhu, Y. Hui, *Tetrahedron* **2001**, *57*, 9403–9413.
- [39] C. C. Fortes, H. C. Fortes, D. C. R. G. Goncalves, *J. Chem. Soc. Chem. Commun.* **1982**, 857–858.
- [40] C. C. Fortes, C. R. O. Souto, E. A. Okino, *Synth. Commun.* **1991**, *21*, 2045–2052.
- [41] The synthesis of **17–21** was carried out from the corresponding alcohol by treatment with *p*-tolyl disulfide and trimethylphosphine.
- [42] H. Tanaka, R. Takeuchi, M. Jimbo, N. Kuniya, T. Takahashi, *Chem. Eur. J.* **2013**, *19*, 3177–3187.
- [43] C. G. Francisco, A. J. Herrera, A. R. Kennedy, A. Martín, D. Melián, I. Pérez-Martín, L. M. Quintanal, E. Suárez, *Chem. Eur. J.* **2008**, *14*, 10369–10381.
- [44] M. Matwiejuk, J. Thiem, *Eur. J. Org. Chem.* **2011**, 5860–5878.
- [45] K. Umemura, H. Matsuyama, M. Kobayashi, N. Kamigata, *Bull. Chem. Soc. Jpn.* **1989**, *62*, 3026–3028.
- [46] E. I. Balmond, D. M. Coe, M. C. Galan, E. M. McGarrigle, *Angew. Chem. Int. Ed.* **2012**, *51*, 9152–9155.
- [47] R. Sundell, L. T. Kanerva, *Eur. J. Org. Chem.* **2013**, 4971–4978.
- [48] A. Noel, B. Delpéch, D. Crich, *Org. Lett.* **2012**, *14*, 4138–4141.
- [49] A. M. Esmurziev, N. Simic, B. H. Hoff, E. Sundby, *J. Carbohydr. Chem.* **2010**, *29*, 348–367.

**Glycuronamide Synthesis**

Sicheng Lin, Roger A. Ashmus Todd L. Lowary*

Page No. – Page No.

An oxidation–amidation approach for the synthesis of glycuronamides

A route for the synthesis of glycuronamides via the intermediacy of 6-S-tolylglycosides and uronic acid thioesters, is reported. The route, which is compatible with a variety of carbohydrate residues and protecting groups, was used to synthesize the repeating unit of the *E. coli* K54 capsular polysaccharide.