



ELSEVIER

Available online at www.sciencedirect.com



European Journal of Pharmaceutics and Biopharmaceutics xxx (2006) xxx–xxx

European
Journal of
Pharmaceutics and
Biopharmaceutics

www.elsevier.com/locate/ejpb

Research paper

Effervescent dry powder for respiratory drug delivery

Leticia Ely ^a, Wilson Roa ^b, Warren H. Finlay ^c, Raimar Löbenberg ^{a,*}

^a Faculty of Pharmacy, University of Alberta, Edmonton, AB, Canada

^b Department of Oncology, University of Alberta, Edmonton, AB, Canada

^c Department of Mechanical Engineering, University of Alberta, Edmonton, AB, Canada

Received 4 July 2006; accepted in revised form 24 October 2006

10 Abstract

11 The objective of this work was to develop a new type of respiratory drug delivery carrier particle that incorporates an active release
12 mechanism. Spray drying was used to manufacture inhalable powders containing polybutylcyanoacrylate nanoparticles and ciprofloxacin
13 as model substances for pulmonary delivery. The carrier particles incorporated effervescent technology, thereby adding an active
14 release mechanism to their pulmonary route of administration. Effervescent activity of the carrier particles was observed when the carrier
15 particles were exposed to humidity. Gas bubbles caused by the effervescent reaction were visualized by confocal laser scanning micro-
16 scopy. The images showed that nanoparticles were distributed throughout the gas bubble. For the effervescent formulation the average
17 mass median aerodynamic diameter (MMAD) was $2.17 \mu\text{m} \pm 0.42$, fine particle fraction (FPF_{≤5.6 μm}) was $46.7\% \pm 15\%$ and the GSD
18 was 2.00 ± 0.06 . The results also showed that the effervescent carrier particles released $56 \pm 8\%$ ciprofloxacin into solution compared
19 with $32 \pm 3\%$ when lactose carrier particles were used. The mean nanoparticle size did not significantly change upon release when the
20 nanoparticles were incorporated into an effervescent formulation. However, the mean size significantly increased upon release when only
21 lactose was used as carrier particle matrix. In conclusion, effervescent carrier particles can be synthesized with an adequate particle size
22 for deep lung deposition. This opens the door for future research to explore this technology for delivery of a large range of substances to
23 the lungs with possible improved release compared to conventional carrier particles.

24 © 2006 Elsevier B.V. All rights reserved.

25 **Keywords:** Effervescent; Inhalable dry powders; Nanoparticles; Pulmonary delivery; Aerosol; Drug delivery; Ciprofloxacin; Spray drying

27 1. Introduction

28 The pulmonary route of administration has been used
29 for many years for the local treatment of lung diseases.
30 More recently, systemic drug absorption has been investi-
31 gated, e.g., for the treatment of diabetes mellitus and pain
32 relief [1]. In addition, major areas of pulmonary research
33 are aimed at asthma [2], cystic fibrosis [3], lung cancer [4]
34 and tuberculosis [5,6]. Drug delivery to the lungs requires
35 an aerosol vehicle, which consists of either aerosol droplets
36 containing the drug, or powder particles of appropriate size

for lung delivery [7]. Dry powder delivery to the lungs 37
remains challenging due to powder aggregation that 38
increases the particle size above the optimal particle diam- 39
eter, which in general terms for deep lung deposition is 40
between about 1 and 5 μm [8–10]. Spray drying is one 41
technique to manufacture inhalable powders [8,10]. 42

Nanomedicine is an emerging field in the biomedical 43
sciences. Drug delivery systems involving nanoparticles 44
have been investigated for different routes of administra- 45
tion. The first nanoparticle-containing intravenous drug 46
delivery system was recently approved as medicine in 47
the United States under the name Abraxane[®]. It contains 48
albumin-bound paclitaxel for the treatment of metastatic 49
breast cancer [11]. Nanoparticles have been proposed 50
for pulmonary administration to utilize their advantages 51
in drug delivery to the lungs [12]. Furthermore, 52

* Corresponding author. 3118 Dentistry/Pharmacy Centre, Faculty of
Pharmacy, University of Alberta, Edmonton, AB, Canada T6G 2G8. Tel.:
+1 780 492 1255; fax: +1 780 492 1217.

E-mail address: rloebenberg@pharmacy.ualberta.ca (R. Löbenberg).

nanoparticles exhibit certain characteristics that make them ideal for pulmonary drug delivery and for treating lung specific diseases like lung cancer. Research has shown that nanoparticles avoid unwanted mucociliary clearance and in some cases phagocytic clearance [13] by remaining in the lung lining fluid until dissolution [14] or translocation by the epithelium cells [15]. One issue with pulmonary nanoparticle delivery is that their small size limits their lung deposition. Aerosolized nanoparticles have only very limited sedimentation, inertial impaction or diffusion, which causes them to be predominantly exhaled from the lungs after inhalation [7,13,16]. However, Sham et al. have shown that nanoparticles can be incorporated into carrier particles to produce the appropriate size for pulmonary drug delivery [12].

Effervescent preparations have been utilized in oral drug delivery for more than 200 years. Since that time, a large number of preparations utilizing effervescent technology have been produced including stomach distress medications, vitamin supplements, and analgesics [17]. However, effervescent powders have not previously been used for the pulmonary route of administration. In the present study, we investigated and optimized carrier particles for respiratory drug delivery that incorporate effervescent technology. The effervescent reaction adds an active release mechanism to the pulmonary route of administration. In this study, polybutylcyanoacrylate nanoparticles and ciprofloxacin hydrochloride hydrate were used as two different model substances for pulmonary delivery. Ciprofloxacin is a powerful antibiotic that is used orally to treat cystic fibrosis. However, currently there are no commercial dosage forms available for the pulmonary delivery of this antibiotic. Drug release and dispersion of nanoparticles were separately compared using lactose carrier particles that dissolve without effervescent reaction, to the new effervescent carrier particles.

2. Materials and methods

2.1. Chemicals

Butylcyanoacrylate was a gift from Loctite Ltd (Dublin, Ireland). Dextran 70 (~70 kDa), L-Leucine, ammonium hydroxide, Rhodamine G8, citric acid and fluorescein isothiocyanate-dextran (FITC-Dextran) were obtained from Sigma Chemical Co. (St. Louis, MO, USA). Lactose monohydrate was obtained from Wyndale (Kapuni, New Zealand). Sodium carbonate anhydrous was obtained from BDH Inc. (Toronto, ON, Canada). Polyethylene glycol (PEG) 6000 was obtained from Fluka Chemika-Biochemika (Buchs, Switzerland). Polysorbate 80 was from BASF (Ludwigshafen, Germany). 316 Silicone Release Spray was purchased from Dow Corning (Midland, MI, USA). Ciprofloxacin hydrochloride hydrate was obtained from US Biological (Swampscott, MA, USA). Cargille, oil, Type DF, SPI, was obtained from West Chester, PA. All chemicals were of analytical grade and used as received.

2.2. Preparation of poly (Butylcyanoacrylate) nanoparticles (PBCN)

Butylcyanoacrylate nanoparticles were prepared by a standard procedure [18]. In brief, 100 µl of the monomer was slowly added by pipette to an HCl 0.01 N solution, containing 0.0900 g Dextran 70.000 and 0.01 g of fluorescein isothiocyanate-dextran 70,000 (FITC). The polymerization was carried out under stirring (600 rpm) at room temperature for 4 h. The pH of the resulting colloidal suspension was neutralized using sodium hydroxide and did not exceed pH 7. Nanoparticles were protected from light through the polymerization process. The nanoparticles were purified from unbound dye by centrifugation at 20,000g (Beckman Model J2-21) for 10 min. The particles were purified by three cycles of centrifugation and redispersion in fresh water. After centrifugation the supernatant was removed and the nanoparticles were resuspended in 1 mL of sterilized water.

2.3. Preparation of empty carrier particles and ciprofloxacin carriers

Seven grams of lactose monohydrate was used to prepare the spray-dried samples. Lactose was added to 100 mL of distilled water. To produce the new carrier particles, different formulations that were used in effervescent tablets were tested. Sodium carbonate and citric acid were tested using different concentrations (see Table 1). Additionally lactose, ammonia water and excipients including L-leucine, PEG 6000 and polysorbate 80 were used. 0.6 mg of methanol solution of Rhodamine G8 was added to the 100 mL of effervescent solution to stain the carrier matrix. The lubricants were chosen from a selection of excipients considered suitable for inhalation [8,16,19] or proven to be safe for human use. Solid ingredients were weighed and added to an aqueous ammonia solution. 10 ml of ammonia was used to increase the pH of the solution to inhibit an effervescent reaction prior to spray drying. The pH was maintained at approximately 8.0. Carrier particles containing ciprofloxacin were prepared using 100 mg of ciprofloxacin hydrochloride hydrate. The drug was first dissolved in HCl 0.01 N and then added to the ammonia-carbonate solution and to the lactose solution. A Büchi 190 Mini-Spray Dryer (Büchi AG, Flawil, Switzerland) was used to produce the carrier

Table 1
Ingredients used for the formulations

	Ingredients used	Concentration tested (%)
Carbonates	Sodium carbonate	0.75–1.5
Acid	Citric acid	1.2
Lubricants	L-Leucine	0.8–1
	Polyethylene glycol 6000	0.8–1
Alcohols	Ethanol	10–30
Surfactants	Polysorbate 80	1
	Sodium lauryl sulfate	1

150 particles. The diameter of the nozzle was 0.7 mm. In each
 151 experiment, 100 mL of either lactose solution or efferves-
 152 cent solution was spray dried at an inlet temperature
 153 120–160 °C, and an aspirator setting of 15 (out of 20),
 154 the air flow in the nozzle was 800 NormL/h and a feed rate
 155 of 2 mL/min was used (see Table 2) The spray-dried pow-
 156 ders were collected in vials. Immediately after their collec-
 157 tion, the powders were stored in a desiccator over silica gel.

158 2.4. Determination of ciprofloxacin loading efficiency

159 Fifteen milligrams of the effervescent or lactose powders
 160 was dissolved in 100 ml of water. Before the measurements,
 161 the samples were filtered (0.22 µm). The dissolved cipro-
 162 floxacin content was analyzed using UV spectroscopy at
 163 $\lambda = 271$ nm (SPECTRONIC 3000 ARRAY – Milton
 164 Ray). A calibration curve was established, the correlation
 165 coefficient for the calculated linear regression was 0.9999
 166 and the correlation equation was used to determine the
 167 dissolved drug content.

168 2.5. Incorporation of the nanoparticles into carrier particles 169 and fluorescent labeling

170 Seven milliliters of a suspension containing poly-
 171 butylcyanoacrylate nanoparticles was added to either a
 172 7% lactose solution, or to a 7% lactose solution contain-
 173 ing PEG 6000 and L-leucine, or to an effervescent
 174 formulation solution or to an effervescent formulation
 175 solution containing PEG 6000 and L-leucine (5 mL of
 176 2.5% L-leucine and 2.5% PEG). The lactose solution was
 177 spray dried at temperatures between 150 and 160 °C
 178 and the corresponding outlet temperature was 130 °C.
 179 The effervescent formulation was spray dried at tempera-
 180 tures between 125 and 130 °C and the outlet temperature
 181 was approximately 110 °C.

182 2.6. Physico-chemical characterization of the nanoparticles 183 and the carrier particles

184 2.6.1. Particle size

185 The particle size was measured using photon correlation
 186 spectroscopy (HSA3000, Malvern Instruments, UK).
 187 Three milliliters of fresh filtered (0.45 µm) water was filled
 188 into a disposable cuvette. An aliquot of approximately
 189 100 µl nanoparticle suspensions was added to the cuvette.

Table 2
 Spray drying parameters

Parameters	Effervescent powders	Lactose powders
Inlet temperature	125–130 °C	155–160 °C
Outlet temperature	110 °C	130 °C
Atomizer air flow rate	800 NormL/h	800 NormL/h
Feed rate (pump)	2 mL/min	2 mL/min
Air flow rate (dial setting)	15	15
Heating rate (dial setting)	10	15

190 Samples were sonicated for 1 min immediately prior to
 191 measurement. To measure the size of nanoparticles after
 192 spray drying, an adequate amount of both lactose and
 193 effervescent powder containing nanoparticles was dissolved
 194 in distilled and filtered water and sonicated immediately
 195 prior to measurement.

2.6.2. Mass median aerodynamic diameter (MMAD)

196 The MMAD was measured using a Mark II Andersen
 197 Cascade Impactor (Thermo Andersen, Smyrna, GA) in
 198 combination with a new high efficiency inhaler [20] as
 199 shown in Fig. 1. This inhaler deagglomerates powders to
 200 a higher percentage compared to conventional inhalers. It
 201 utilizes a cyclone action and mechanical impaction to
 202 disperse powder particles [20]. The flow rate used was
 203 60 l/min. Calibration of the Andersen at the higher flow
 204 rate of 60 l/min has been published by Nichols et al. [21]
 205 and this calibration is used here. The MMAD was calculat-
 206 ed by a nonlinear regression fit of a log-normal function to
 207 the data.
 208

2.6.3. Fine Particle Fraction (FPF), Geometrical Standard Deviation (GSD) and Emitted Dose (ED)

209 In this study, fine particle fraction (FPF) was defined
 210 as the fraction of loaded powder that was collected on
 211 plates 1–6 (i.e., aerodynamic diameter ≤ 5.6 µm, at a
 212 flow rate of 60 l/min). The Mark II Andersen Cascade
 213 Impactor was used to determine the fine particle fraction.
 214 Geometric standard deviation is a measure of the
 215 variability of the particle diameter within the aerosol
 216 [7]. It is defined by the ratio of the diameters of particles
 217 from aerosols corresponding to 84% and 50% on the
 218 cumulative distribution curve of the weights of particles.
 219 To calculate the GSD, a nonlinear least squares analysis
 220 with a log-normal function was used. The emitted
 221 dose was calculated as the amount of loaded powder
 222 minus the amount collected in the Andersen Cascade
 223 Impactor.
 224
 225

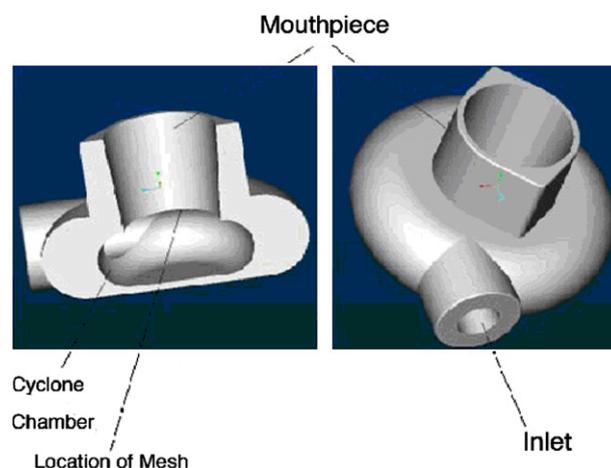


Fig. 1. Dry powder inhaler.

226 2.6.4. Scanning-electron microscopy (SEM)

227 The lactose and effervescent powders were sprinkled
228 onto a stub with silicon from a sticky tab. The unbound
229 powders were dusted out by an air gun. The samples were
230 coated with gold sputter using a S150B Sputter Coater
231 (BOC Edwards, Crawley, West Sussex, UK) and examined
232 by scanning electron microscope (S2500 SEM, Hitachi,
233 Tokyo, Japan).

234 2.6.5. Confocal laser scanning microscopy (CLSM)

235 The geometric diameter of the spray-dried powders, the
236 distribution of the nanoparticles through the carrier parti-
237 cles and effervescent effect of the carrier particles were
238 investigated using a Zeiss LSM 510 confocal laser-scanning
239 microscope (Oberkochen, Germany). The LSM 510 Soft-
240 ware, version 2.0 was used to control the microscope and
241 to analyze the images. The carrier particles were labeled
242 with a red fluorescent label and the nanoparticles with a
243 green fluorescent label. Small amounts of the powders were
244 dispersed in immersion oil on glass slides and visually
245 observed. The samples were observed before and after
246 being exposed to humidity. The oil phase prevented any
247 contact of humidity with the particles during the observa-
248 tion of the images. The particle morphology (porous vs.
249 solid) was investigated by imaging different layers of the
250 carrier particles.

251 3. Results

252 Different powder compositions were tested in order to
253 produce carrier particles with an appropriate size. Lactose
254 is the most common type of excipient used for dry powder
255 lung delivery and is well documented in the literature
256 [22,23]. Blank lactose carrier particles were spray dried at
257 inlet temperatures between 140 and 160 °C. The mass
258 median aerodynamic diameter of the carrier particles was
259 analyzed using the Andersen cascade impactor. Ten sam-
260 ples of lactose carrier particles were analyzed and the mean
261 MMAD was found to be 10 μm or larger. The fine particle
262 fraction (FPF) was found to be 13.86% ± 5.56 (n = 8).
263 Fig. 2 shows a lactose powder that was made without the
264 presence of any other solvent or excipients. The image
265 shows that the majority of the particles in this powder were
266 spherical.

267 In oral tablet formulations, effervescent formulations
268 use a mixture of acids such as citric acid and carbonates.
269 A typical ratio that generally achieves a fast effervescent
270 reaction and acceptable stability uses a mixture of 50%
271 sodium carbonate and 50% sodium bicarbonate [24]. How-
272 ever, sodium bicarbonate decomposes at temperatures
273 above 50 °C and for this reason it is not recommended to
274 be used for spray-drying procedures which use tempera-
275 tures that will exceed this value in the powders. The effe-
276 rescent reaction is pH dependent. Two components react
277 in an aqueous environment as shown in the formula.

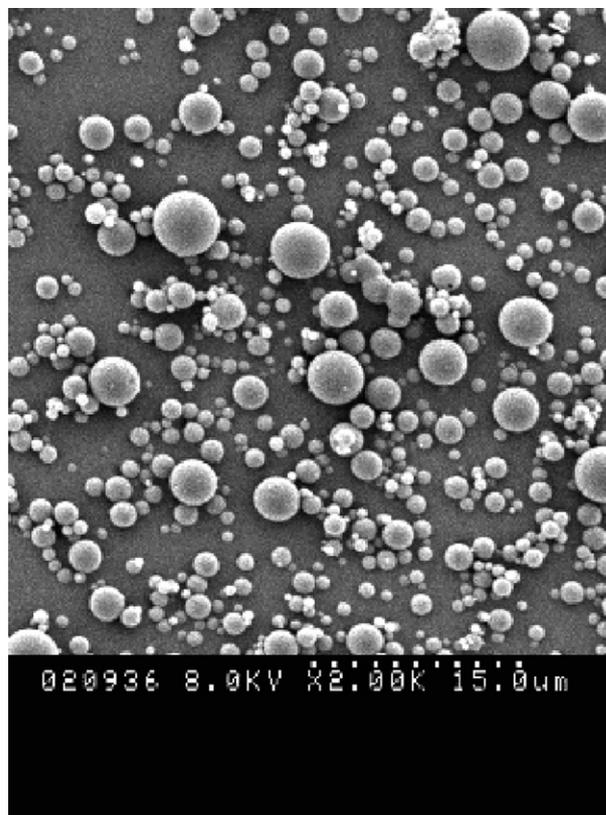
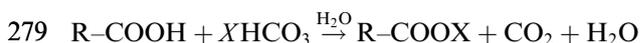


Fig. 2. SEM image of typical lactose particles observed after spray drying of a 7% of lactose solution.

280 As shown, this reaction releases carbon dioxide. The phase
281 transition from a solid to a gas phase increases the volume
282 and this is used in tablets to increase tablet disintegration
283 and drug dissolution [25].

284 To produce inhalable effervescent powders, the first step
285 was to establish an effervescent formulation. The basic for-
286 mulation contained sodium carbonate, citric acid and
287 water. However in order to prevent an effervescent reaction
288 from happening before spray drying, the pH of the solution
289 was increased using ammonia in order to maintain the pH
290 at 8.0. The ammonia evaporates in the spray-drying
291 process and the resulting powders contain citric acid and
292 carbonate in the solid state.

293 Different ingredients such as ethanol, polysorbate 80,
294 L-leucine and PEG 6000 were added to the basic formula-
295 tion to improve the particle size and to achieve an
296 appropriate MMAD. In addition, different concentrations
297 of lactose were tested. Different amounts of lactose had a
298 large impact on the size and morphology of the carrier parti-
299 cles. Increasing the amount of lactose led to smaller and
300 denser particles and also produced particles with more
301 spherical shape. These results are in agreement with results
302 reported by Vanbever et al. [19]. Fig. 3 shows a sequence of
303 SEM pictures with increasing concentrations of lactose and
304 consequently increasing the MMAD of the carrier parti-
305 cles. The MMAD of the particles, measured by cascade
306 impaction, were 3.85, 5.22, 8.3, and 10 μm, respectively.

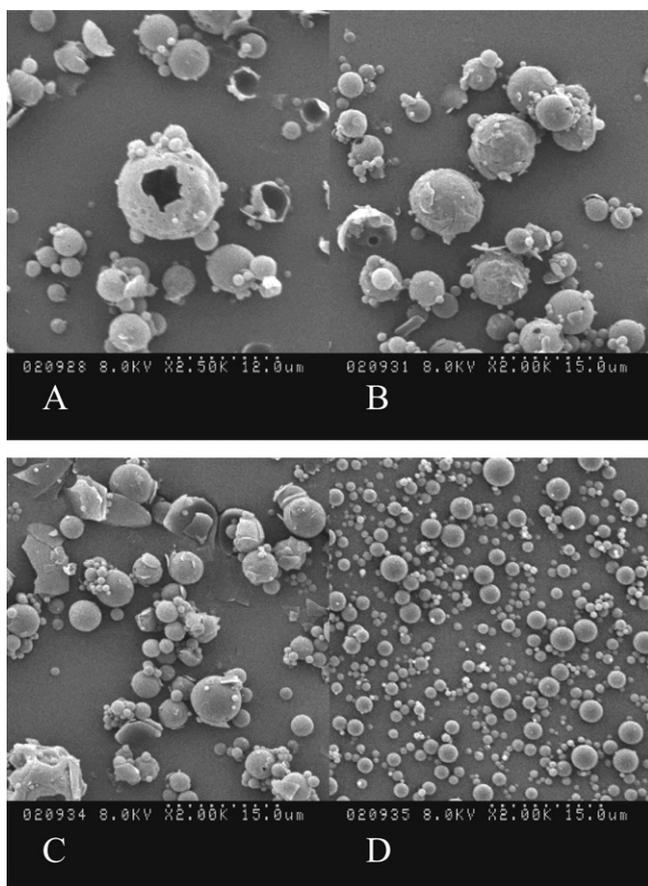


Fig. 3. SEM images of spray-dried effervescent formulations containing (A) 1.2% lactose, (B) 2.4% lactose, (C) 3.5% lactose, and (D) 10% lactose.

307 Ethanol was added to the formulation with the objective of
 308 producing larger porous particles with a lower density.
 309 However our results showed that when ethanol was used
 310 it did not improve the particle size or the morphology of
 311 the carrier particles. The MMAD was still approximately
 312 8.5 μm and not suitable for deep lung deposition. This
 313 might be due to the relatively low ethanol concentrations
 314 used, which were between 10 and 30% v/v. Other studies
 315 have reported using up to 70% of the total volume of eth-
 316 anol to produce porous particles [16]. Polysorbate 80 also
 317 did not show any improvement in particle size when com-
 318 pared to the lactose formulation. A large improvement in
 319 particle size and MMAD was observed when 5 mL of both
 320 solutions containing 2.5% L-leucine and 2.5% PEG 6000
 321 was added to the formulation. For these effervescent carrier
 322 particles the average MMAD were $2.17 \pm 0.42 \mu\text{m}$, FPF
 323 was approximately $46.47 \pm 15\%$ and the GSD was
 324 $2.00 \pm 0.06\%$ (see Table 3). The emitted dose for powders
 325 made just of lactose was found to be $73.38 \pm 13\%$ and
 326 for powders containing the L-leucine/PEG 6000 efferves-
 327 cent formulation was $68.55 \pm 23.90\%$.

328 Using L-leucine and PEG 6000 (powder 5) in the effer-
 329 vescent formulation, it was possible to obtain inhalable
 330 particles as indicated by the SEM pictures in Fig. 4. These
 331 particles show a more irregular morphology when com-

pared to the highly spherical lactose carrier particles. For
 all the following experiments this formulation was used.

3.1. Comparisons of drug release from effervescent and conventional carrier particles

Ciprofloxacin is poorly water soluble at physiological
 pH. Its drug release from conventional lactose particles
 was compared with the effervescent formulation. The
 results show that the effervescent carrier particles released
 $56 \pm 8\%$ ciprofloxacin into solution compared with
 $32 \pm 3\%$ when lactose particles were used, which is a signif-
 icant difference (*t*-test, $P < 0.05$). The remaining drug was
 visual as precipitate before filtering the solution.

3.2. Carrier particles containing PBC nanoparticles

Polybutylcyanoacrylate nanoparticles were spray dried
 in an aqueous solution containing lactose as well as the
 effervescent preparation. In order to compare the different
 formulations and the effects of effervescent reaction and
 excipients, four types of powders were produced. The
 amount of L-leucine and PEG was kept constant in all for-
 mulations. The particle diameter of the nanoparticles was
 measured before and after spray drying. A *t*-test was per-
 formed to compare the sizes of the samples before and after
 spray drying. When only lactose was used, the nanoparti-
 cles had a size of $126.17 \pm 20.20 \text{ nm}$ before and
 $259.00 \pm 52.70 \text{ nm}$ after spray drying. This is a statistically
 significant increase in particle size at a P value < 0.05 . For
 lactose containing PEG and L-leucine the particle size
 before was $247.5 \pm 13.4 \text{ nm}$ and $225 \pm 11.17 \text{ nm}$ after-
 wards. For the effervescent particles the results were
 $244 \pm 26.8 \text{ nm}$ and $252 \pm 29 \text{ nm}$ before and afterwards,
 respectively. Using the effervescent preparations containing
 L-leucine and PEG 6000, the size before spray drying was
 $149.9 \pm 26.46 \text{ nm}$ and the size after spray drying was
 $176.83 \pm 15.45 \text{ nm}$. For the last three formulations a *t*-test
 did not indicate a statistical difference between the nano-
 particles before and after the spray drying process.

3.3. Effervescent properties of the carrier particles containing nanoparticles

The effervescent properties of the carrier particles were
 observed when the carrier particles were exposed to water,
 aqueous surfaces or moist air. Fig. 5 shows a carrier parti-
 cle, which was under 5 μm in diameter with spherical shape
 (Fig. 5A–D). The nanoparticles were distributed
 continuously throughout the carrier particle matrix.
 Fig. 6 shows the swelling and dissolution of the carrier parti-
 cles after exposure to humid air (Fig. 6A–C). The matrix
 of the particles dissolves (red channel) while a green bubble
 of more than 30 μm is filled with nanoparticles. This indi-
 cates that the nanoparticles were actively distributed
 throughout the gas bubble. If the effervescent powder

Table 3
Particle size measurements are given for a variety of spray-dried powders

Powder	MMAD (μm)	FPF (%)	GSD
Blank lactose (7% lactose)	10	13.86% \pm 5.56	–
Ethanol 30%	8.5 \pm 1.8	17.87 \pm 4	–
Lactose 10%	10	17.60 \pm 3.5	–
Polysorbate 80	10	12.50 \pm 2	–
Five milliliters of a 2.5% solution of both L-leucine and PEG 6000	2.17 \pm 0.42	46.47% \pm 15	2.00 \pm 0.06

Powder 1 was spray drying only with lactose. Powder formulations from 2 to 5 were all effervescent formulations that included the listed excipients.



Fig. 4. Scanning electronic micrograph of inhalable effervescent particles containing L-leucine and PEG 6000 as excipients. Some asperities are present on the surface of the carriers due to the presence of PEG 6000.

382 was dispersed in water, small gas bubbles were visible
383 immediately after dispersion.

384 4. Discussion

385 In this paper, different powder compositions were produced
386 in order to develop and partially optimize an effervescent
387 aerosol carrier particle formulation. To improve particle size,
388 the addition of L-leucine, PEG 6000, polysorbate 80 and ethanol
389 was examined. The most pronounced effect on particle size
390 occurred with the addition of L-leucine and PEG 6000, which
391 improved the aerodynamic characteristics of the powder particles.
392 The in vitro results indicated that the particles were suitable for
393 deposition throughout the lungs. These results are also in agreement
394 with other studies [8,26]. Gliński et al. found that when
395

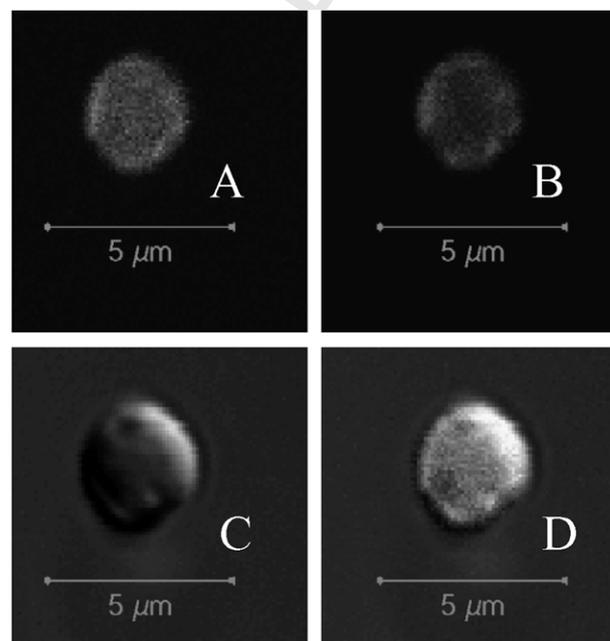


Fig. 5. Confocal microscopy pictures of an effervescent carrier particle. (A) Green channel shows the nanoparticles distributed continuously through out the carrier particle. (B) Red channel shows the matrix of the carrier particle. (C) Particle in normal light. (D) Super imposed picture of the red and green channels.

L-leucine was added to a water solution it caused a rapid
decrease in the surface tension [26]. In addition, L-leucine
allows the preparation of powders with better aerolization
properties [27]. Corrigan et al. and Gilane et al. investigated
the use of PEG in their formulations [28,29]. They found
that polyethylene glycol had a major impact on the size and
morphology of carrier particles. In addition, the presence
of PEG 6000 changed the surface texture of the carrier particles
from a smooth surface to a more aspirated surface. Similar
effects were observed in our study using the effervescent
formulation. In addition the presence of polyethylene glycol
might also influence the crystalline and polymorphic form
of spray-dried lactose and presumably of incorporated drugs
[28,29].

The results reported for the emitted dose are satisfactory
for both the dry powder formulation containing just lactose
and the dry powder L-leucine/PEG 6000 formulation containing
effervescent carrier particles. However the emitted dose
achieved for the lactose carrier particles mostly contained
powder particles which were too large

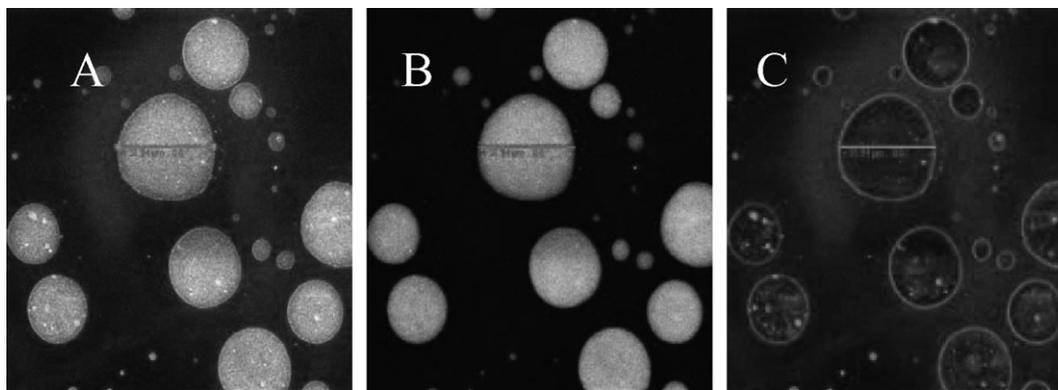


Fig. 6. Confocal microscopy picture of effervescent particles exposed to humidity. (A) Super imposed red and green channel showing gas bubbles of different diameters. (B) Green channel showing the nanoparticles distributed throughout the gas bubble. (C) Red channel showing the dissolved carrier matrix.

416 for deep lung deposition, since this powder mainly
417 deposited on plate zero of the cascade impactor while
418 most of the effervescent powder was deposited on plates
419 2, 3, and 4, as shown by the FPF of $13.86 \pm 5.56\%$ for
420 the lactose formulation and $46.46 \pm 15\%$ for the efferves-
421 cent formulation.

422 The formulations containing effervescent release mecha-
423 nisms and the lactose formulations that contained L-leucine
424 and PEG 6000 were able to release nanoparticles with less
425 agglomeration compared to the carrier particles made just
426 of lactose which lack an active release. The results show
427 that both the effervescent formulation and the choice of
428 excipients had a major effect on the release of nanoparti-
429 cles. The effervescent reaction of the carrier particles gener-
430 ates forces that helped the nanoparticles to disperse more
431 efficiently and avoid particle aggregation. Sham et al. [12]
432 conducted a study using lactose carrier particles containing
433 nanoparticles. In the cited study it was found that some
434 clusters of nanoparticles were observed in the carrier parti-
435 cles, which increased the nanoparticle size after spray-dry-
436 ing. Our results showed a significant increase in the size of
437 the released nanoparticles when lactose alone was used as
438 carrier. However, when effervescent carrier particles were
439 used, no statistically significant difference was observed.
440 These findings indicate that the effervescent reaction
441 appears to improve the dispersion of the nanoparticles
442 from the carrier particle.

443 Ciprofloxacin was used as a model drug in order to evalu-
444 ate the effect of active drug release from the carrier parti-
445 cles compared to passive release and dissolution. It was
446 found that the effervescent carrier particles were able to
447 increase the drug dissolution. Rygnestad et al. reported
448 that effervescent paracetamol tablets were absorbed signifi-
449 cantly faster compared to conventional tablets [30]. Dos-
450 age form disintegration and drug dissolution are typically
451 increased when effervescent formulations are used. Howev-
452 er, more studies are needed to evaluate if an effervescent
453 inhalable powder can increase the absorption and bioavail-
454 ability of drugs in the lungs due to improved drug dissolu-
455 tion properties.

5. Conclusion

456

457 A new formulation was established for the use in the
458 pulmonary route of administration. The new formulation
459 contained effervescent and lubricant excipients. The active
460 release mechanism increased drug dissolution and
461 enhanced the dispersion of nanoparticles over the efferves-
462 cent gas bubble interface. These carrier particles can be
463 synthesized with an adequate particle size for deep lung
464 deposition. Furthermore, effervescent carrier particles can
465 be used to deliver a large range of substances to the lungs
466 with possibly a faster release compared to conventional
467 carrier particles. However further studies are required to
468 evaluate how the effervescent particles will behave at the
469 lung surfactant air interface.

Acknowledgement

470

471 This work was partially supported financially by a
472 NSERC Strategic grant.

References

473

- 474 [1] N.-R. Labiris, M.-B. Dolovich, Pulmonary drug delivery. Part I:
475 physiological factors affecting therapeutic effectiveness of aerosolized
476 medications, *J. Clin. Pharmacol.* 56 (2003) 588–599.
- 477 [2] J.-G. Hardy, T.-S. Chadawick, Sustained release drug delivery to the
478 lungs, *Clin. Pharmacokinet.* 39 (2000) 1–4.
- 479 [3] L. Garcia-Contreras, A.-J. Hickey, Pharmaceutical and biotechno-
480 logical aerosols for cystic fibrosis therapy, *Adv. Drug Deliv. Rev.* 54
481 (2002) 1491–1504.
- 482 [4] R. Rao, S. Markovic, P. Anderson, Aerosol therapy for malignancy
483 involving the lungs, *Curr. Cancer Drug Targets* 3 (2003) 239–250.
- 484 [5] R. Pandey, G.-K. Khuller, Antitubercular inhaled therapy: opportu-
485 nities, progress and challenges, *J. Antimicrob. Chemother.* 55 (2005)
486 430–435.
- 487 [6] A. Zahoor, S. Sharma, G.-K. Khuller, Inhalable alginate nanopar-
488 ticles as antitubercular drug carriers against experimental tuberculosis,
489 *Int. J. Antimicrob. Agents* 26 (2005) 298–303.
- 490 [7] W.H. Finlay, *Mechanics of Inhaled Pharmaceutical Aerosols: An*
491 *Introduction*, Academic Press, New York, 2001.
- 492 [8] C. Bosquillon, C. Lombry, V. Preat, R. Vanbever, Influence of
493 formulation excipients and physical characteristics of inhalation dry

- 494 powders on their aerolization performance, *J. Control. Release* 70
 495 (2001) 329–339. 530
- 496 [9] L.-A. Dailey, T. Schmehl, T. Gessler, M. Wittmar, F. Griminger, W. 531
 497 Seeger, T. Kissel, Nebulization of biodegradable nanoparticles: 532
 498 impact of nebulizer technology and nanoparticle characteristics on 533
 499 aerosol features, *J. Control. Release* 86 (2003) 131–144. 534
- 500 [10] P. Lucas, K. Anderson, U.-J. Potter, J.-N. Staniforth, Enhancement 535
 501 of small particle size dry powder aerosol formulations using an ultra 536
 502 low density additive, *Pharm. Res.* 16 (1999) 1643–1647. 537
- 503 [11] Abraxane [prescribing information]. Schaumburg, Ill: Abraxis Oncolo- 538
 504 gy, A Division of American Pharmaceutical Partners, Inc., January 539
 505 2005. 540
- 506 [12] J.-O. Sham, Y. Zhang, W.-H. Finlay, W.-H. Roa, L. Raimar, 541
 507 Formulation and characterization of spray dried powders containing 542
 508 nanoparticles for aerosol delivery to the lung, *Int. J. Pharm.* 269 543
 509 (2004) 457–467. 544
- 510 [13] A. Grenha, B. Seijo, C. Remuñán-López, Microencapsulated 545
 511 chitosan nanoparticles for lung protein delivery, *Eur. J. Pharm. Sci.* 546
 512 25 (2005) 427–437. 547
- 513 [14] S. Schürch, M. Geiser, m.-M. Lee, P. Gehr, Particles at the airway 548
 514 interfaces of the lung, *Colloids Surf. B: Biointerfaces* 15 (1999) 549
 515 339–353. 550
- 516 [15] G. Oberdörster, E. Oberdörster, J. Oberdörster, Nanotoxicology: an 551
 517 emerging discipline evolving from studies of ultrafine particles, 552
 518 *Environ. Health Perspect.* 113 (2005) 823–839. 553
- 519 [16] N. Tsapis, D. Bennet, B. Jackson, D.-A. Weitz, D.-A. Edwards, 554
 520 Trojan particles: large porous carrier of nanoparticles for drug 555
 521 delivery, *Proc. Natl. Acad. Sci.* 99 (2002) 12001–12005. 556
- 522 [17] J.-D. Eichman, J.-R. Robinson, Mechanistic studies on efferves- 557
 523 cent-induced permeability enhancement, *Pharm. Res.* 15 (1998) 558
 524 925–930. 559
- 525 [18] P. Sommerfeld, U. Schroeder, A.-S. Bernhard, Sterilization of 560
 526 unloaded polybutylcyanoacrylate nanoparticles, *Int. J. Pharm.* 164 561
 527 (1998) 113–118. 562
- 528 [19] R. Vanbever, J.-D. Mintzes, J. Wnag, J. Nice, D. Chen, R. Batycky, 563
 529 R. Langer, D.-A. Edwards, Formulation and physical characteriza- 564
 tion of large porous particles for inhalation, *Pharm. Res.* 16 (1999) 565
 1735–1742. 566
- [20] Z. Wang, B. Grgic, W.-H. Finlay, A dry powder inhaler with reduced 532
 mouth-throat deposition, *J. Aerosol Med.*, in press. 533
- [21] S.C. Nichols, D.R. Brown, M. Smurthwaite, New concept for the 534
 variable flow rate Anderson impactor and calibration data, *J. Aerosol 535
 Med.* 11 (1998) 133–138. 536
- [22] J. Elversson, A. Millqvist-Fureby, G. Alderborn, U. Elofsson, 537
 Droplet and particle size relationship and shell thickness of inhalable 538
 lactose particles during spray drying, *J. Pharm. Sci.* 92 (2003) 539
 900–910. 540
- [23] M. Karhu, J. Kuikka, T. Kauppinen, K. Bergström, M. Vidgren, 541
 Pulmonary deposition of lactose carrier used in inhalation powders, 542
Int. J. Pharm. 19 (2000) 95–103. 543
- [24] A. Rau, Multisensory technologies for today's effervescent bath and 544
 shower products, *Cosmet. Toiletries* 49 (2001). 545
- [25] M. Otsuka, M. Sato, Y. Matsuda, Comparative evaluation of 546
 tableting compression behaviors by methods of internal and external 547
 lubricant addition: inhibition of enzymatic activity of trypsin 548
 preparation by using external lubricant addition during the tableting 549
 compression process, *AAPS Pharm. Sci.* 3 (2001) 1–11. 550
- [26] J. Gliński, G. Chavepeyer, J.-K. Platten, Surface properties of 551
 aqueous solutions of L-leucine, *Biophys. Chem.* 84 (2000) 99–103. 552
- [27] N.R. Rabbani, P.-C. Seville, The influence of formulation compo- 553
 nents on the aerosolisation properties of spray dried powders, *J. 554
 Control. Release* 110 (2005) 130–140. 555
- [28] O.-D. Corrigan, A.- M. Healy, O.-I. Corrigan, The effect of spray 556
 drying solutions of polyethylene glycol (PEG) and lactose/PEG on 557
 their physicochemical properties, *Int. J. Pharm.* 235 (2002) 193–205. 558
- [29] K. Gilane, A.-B. Najafabadi, M. Barghi, M. Rafiee, Therani, 559
 Aerolization of beclomethasone diprionate using spray dried lac- 560
 tose/polyethylene glycol carriers, *Eur. J. Pharm. Biopharm.* 58 (2004) 561
 596–606. 562
- [30] T. Rygnestad, K. Zahlsen, F.- A. Samdal, Absorption of effervescent 563
 paracetamol tablets relative to ordinary paracetamol tablets in 564
 healthy volunteers, *Eur. J. Clin. Pharmacol.* 56 (2000) 41–143. 565
 566