## **Supplementary Information for:**

# Human neuraminidase isoenzymes show variable activities for 9-O-acetyl-sialoside substrates

Carmanah D. Hunter, Neha Khanna, Michele R. Richards, Reza Rezaei Darestani, Chunxia Zou, John S. Klassen, and Christopher W. Cairo

### **Table of Contents**

| Figure S1. Kinetics of hNEU cleaving 4MU-Neu5Ac (solid line) and 4MU-Neu5,9Ac2 (dashed line)3               |
|---|
| Figure S2. One-pot-two-enzyme synthesis of Neu5Ac (4,5) and Neu5,9Ac2 (8,9) octyl GM3 derivatives.3         |
| Figure S3. Synthetic route to Neu5Gc octyl GM3 derivatives (6,7)  |
| Experimental procedures for the synthesis of SI6, SI13, and SI14  |
| Figure S4. Control experiment showing that NEU1 does not cleave octyl sialyllactoside 4                     |
| Figure S5. Kinetics of NEU2 cleaving octyl sialyllactosides   |
| Figure S6. Kinetics of NEU3 cleaving octyl sialyllactosides   |
| Figure S7. Kinetics of NEU4 cleaving octyl sialyllactosides   |
| Table S1. Summary of kinetics assays showing the effect of sialic acid modification on hydrolysis with      |
| hNEU  |
| Figure S8. Ring conformations from 25-ns MD simulations for GM3 trisaccharides                              |
| Figure S9. Histogram of ring conformations (along $\theta$ ) from a 25-ns MD simulation for the GM3-analogs |
| <b>4</b> , <b>6</b> , and <b>8</b> bound to NEU2  |
| Table S2. Pyranose ring conformations and Cremer-Pople coordinates from MD simulations of three GM3-        |
| analogs in the NEU2 binding site  |

| Table S3. Pyranose ring conformations and Cremer-Pople coordinates from MD simulations of three GM3                               |
|---|
| analogs in the NEU3 binding site  |
| Figure S10. An overlay of 10 conformations of the trisaccharide of a) <b>4</b> b) <b>6</b> and c) <b>8</b> from the MD simulation |
| in the NEU2 active site   |
| Figure S11. An overlay of 10 conformations of the trisaccharide of a) <b>4</b> b) <b>6</b> and c) <b>8</b> from the MD simulation |
| in the NEU3 active site   |
| Figure S12. Negative control ESI-MS experiments done in the absence of enzyme   |



**Figure S1**. Kinetics of hNEU cleaving 4MU-Neu5Ac (solid line) and 4MU-Neu5,9Ac2 (dashed line) for a) NEU1, b) NEU2, c) NEU3, d) NEU4. Slopes were fit by linear regression forced through zero with triplicate experiments. Error bars denote standard error.



Figure S2. One-pot-two-enzyme synthesis of Neu5Ac (4,5) and Neu5,9Ac2 (8,9) octyl GM3 derivatives.



**Figure S3**. Synthetic route to Neu5Gc octyl GM3 derivatives (6,7). a) Generation of N-glycolyl-d-mannosamine. b) One-pot-three-enzyme synthesis of Neu5Gc octyl GM3 derivatives.

Experimental procedures for the synthesis of SI6, SI13, and SI14.



#### 5-Acetamido-9-*O*-acetyl-3,5-dideoxy-D-glycero-β-D-galacto-2-nonulopyranosonic acid (SI6):

Starting from *N*-acetylneuraminic acid, (SI6) was prepared in one step, as previously reported.<sup>1 <sup>1</sup>H and <sup>13</sup>C NMR data were consistent with previous reports.<sup>1, 2</sup></sup>



N-glycolyl-D-mannosamine (SI3):

*N*-glycolyl-D-mannosamine was prepared as previously reported, with minor modifications.<sup>3</sup> Briefly, glycolic acid (100 mg, 1.31 mmol) was added to a mixture of mannosamine hydrochloride (**SI2**) (338.5 mg, 1.57 mmol) and triethylamine (0.22 mL, 1.57 mmol) in anhydrous DMF. The reaction was cooled to 0 °C and DIC (0.24 mL, 1.57 mmol) and HOBt (194.7 mg, 1.44 mmol) were added sequentially. After stirring overnight the reaction mixture was filtered and concentrated under reduced pressure. Crude product was purified with iatrobeads (CHCl<sub>3</sub>:MeOH, 2:1) to yield 238 mg (77 %) of (**SI3**) as an off-white solid. <sup>1</sup>H and <sup>13</sup>C NMR data were consistent with previous reports.<sup>3</sup>



#### Octyl $\beta$ -D-galactopyranosyl-(1 $\rightarrow$ 4)- $\beta$ -D-glucopyranoside (SI4):

Starting from  $\beta$ -lactose, octyl  $\beta$ -D-galactopyranosyl-(1 $\rightarrow$ 4)- $\beta$ -D-glycopyranoside was prepared as previously reported.<sup>4</sup> <sup>1</sup>H and <sup>13</sup>C NMR data were consistent with previous reports.<sup>4</sup>



**Figure S4**. Control experiment showing that NEU1 does not cleave octyl sialyllactoside **4**. 4MU-Neu5Ac is a positive control. The result is consistent with previous reports that NEU1 does not cleave glycolipid substrates,<sup>5</sup> and is consistent with the majority of the neuraminidase activity in the crude HEK293E cell lysate being NEU1.



**Figure S5.** Kinetics of NEU2 cleaving octyl sialyllactosides (5-9 – dashed line) in reference to  $\alpha(2\rightarrow 3)$  Neu5Ac octyl sialyllactoside (4 – solid line, matched control replotted on every panel). Slopes were fit by linear regression forced through zero of triplicate experiments, error bars denote standard error.



**Figure S6.** Kinetics of NEU3 cleaving octyl sialyllactosides (5-9 – dashed line) in reference to  $\alpha(2\rightarrow 3)$  Neu5Ac octyl sialyllactoside (4 – solid line, matched control replotted on every panel). Slopes were obtained by linear regression forced through zero of triplicate experiments, error bars denote standard error.



**Figure S7.** Kinetics of NEU4 cleaving octyl sialyllactosides (5-9 – dashed line) in reference to  $\alpha(2\rightarrow 3)$  Neu5Ac octyl sialyllactoside (4 – solid line, matched control replotted on every panel). Slopes were obtained by linear regression forced through zero of triplicate experiments, error bars denote standard error.

**Table S1.** Summary of kinetics assays showing the effect of sialic acid modification on hydrolysis with hNEU. Rates were obtained by taking points every 10 min for 40 min (or every 1 min for 4 min compounds 4 and 6 only); followed by a linear regression of triplicate experiments. Data is in the form of relative rates,  $\pm$  standard error, normalized to  $\alpha(2\rightarrow 3)$  Neu5Ac.

| Assay    | 4               | MU  |                 |                   | Octyl si        | l sialyllactosides                            |                   |                 |  |
|----------|-----------------|---|-----------------|-------------------|-----------------|---|-------------------|-----------------|--|
| Compound | 1               | 2   | 4               | 5                 | 6               | 7   | 8                 | 9               |  |
| NEU1     | $1.00\pm0.05$   | $0.56 \hspace{0.2cm} \pm \hspace{0.2cm} 0.01$ |                 |                   |                 |   |                   |                 |  |
| NEU2     | $1.0 \pm 0.2$   | $0.026\pm0.006$                               | $1.0 \pm 0.1$   | $0.011 \pm 0.001$ | $0.6\pm0.1$     | $0.052\pm0.004$                               | $0.008 \pm 0.001$ | $0.006\pm0.003$ |  |
| NEU3     | $1.00\pm0.04$   | $0.17\pm0.01$                                 | $1.00\pm0.04$   | $1.1 \pm 0.2$     | $0.8\pm0.1$     | $0.35 \hspace{0.2cm} \pm \hspace{0.2cm} 0.07$ | $0.8 \pm 0.1$     | $0.11 \pm 0.04$ |  |
| NEU4     | $1.00 \pm 0.03$ | $1.97 \pm 0.04$                               | $1.00 \pm 0.09$ | $0.85 \pm 0.08$   | $0.51 \pm 0.04$ | $0.28 \pm 0.04$                               | $0.5 \pm 0.1$     | $0.08 \pm 0.02$ |  |



**Figure S8.** Ring conformations from 25-ns MD simulations for GM3 trisaccharides of 4, 6, and 8 bound to a) NEU2 and b) NEU3. See also Table S2 and Table S3.



**Figure S9.** Histogram of ring conformations (along  $\theta$ ) from a 25-ns MD simulation for the GM3-analogs 4, 6, and 8 bound to NEU2. Neu5Ac in 4 is shown as a solid line and has two populations, one at  $\theta = 99^{\circ}$  (OS3, 18%) and a second at  $\theta = 170^{\circ}$  (2C5, 82%). Neu5Gc of 6 is shown as a dotted line and has one population at  $\theta = 166^{\circ}$  (2C5). Neu5,9Ac2 in 8 is shown as a line with long dashes and has one population at  $\theta = 93^{\circ}$  (B2,5). See also Figure S8.

| CM2                        | <b>D</b> .    | Ring                    |                   | $\theta$ (degrees) <sup>a</sup> |  |
|----------------------------|---------------|-------------------------|-------------------|---------------------------------|--|
| GM3 analog                 | Pyranose ring | conformation            | $\phi$ (degrees)" |                                 |  |
| with Neu5Ac                | Glc           | ${}^{4}C_{1}$           | $217 \pm 117$     | 11 ± 6                          |  |
|                            | Gal           | ${}^{4}C_{1}$           | $208 \pm 100$     | $10\pm 6$                       |  |
|                            | Neu           | $^{2}C_{5}$             | $191 \pm 119$     | $158\pm26$                      |  |
| with Neu5Gc                | Glc           | ${}^{4}C_{1}$           | $204 \pm 120$     | $11 \pm 7$                      |  |
|                            | Gal           | ${}^{4}C_{1}$           | $208\pm90$        | $9\pm5$                         |  |
|                            | Neu           | $^{2}C_{5}$             | $247 \pm 124$     | $166 \pm 6$                     |  |
|                            | Glc           | ${}^{4}C_{1}$           | $189 \pm 122$     | $11 \pm 6$                      |  |
| with Neu5,9Ac <sub>2</sub> | Gal           | ${}^{4}C_{1}$           | $215\pm93$        | $9\pm5$                         |  |
|                            | Neu           | <i>B</i> <sub>2,5</sub> | $556\pm13$        | $93\pm5$                        |  |
|                            |               |                         |                   |                                 |  |

**Table S2.** Pyranose ring conformations<sup>6</sup> and Cremer-Pople coordinates<sup>7</sup> from MD simulations of three GM3analogs in the NEU2 binding site. See also Figure S8, Figure S9, and Figure S10.

<sup>*a*</sup> The average value  $\pm$  the standard deviation over the 25-ns MD trajectory are shown.

| Table S3. Pyranose ring conformations <sup>6</sup> and Cremer-Pople coordinates <sup>7</sup> from MD simulations of three GM |
|--|
| analogs in the NEU3 binding site. See also Figure S8 and Figure S11  |

| CM2 analog                 | Drugonogo ving | Ring          | h (dogroog)4         | $\theta$ (degrees) <sup><i>a</i></sup> |  |
|----------------------------|----------------|---------------|----------------------|--|--|
| GM3 analog                 | Pyranose ring  | conformation  | $\varphi$ (degrees)* |  |  |
| with Neu5Ac                | Glc            | ${}^{4}C_{1}$ | $223.55 \pm 101.25$  | $10.91\pm5.89$                         |  |
|                            | Gal            | ${}^{4}C_{1}$ | $223.88 \pm 112.5$   | $11.09\pm 6.32$                        |  |
|                            | Neu            | $^{2}C_{5}$   | $200.04\pm128.42$    | $168.09\pm6.16$                        |  |
|                            | Glc            | ${}^{4}C_{1}$ | $228.12\pm93.71$     | $10.43\pm5.59$                         |  |
| with Neu5Gc                | Gal            | ${}^{4}C_{1}$ | $215.59\pm119.55$    | $11.09\pm5.88$                         |  |
|                            | Neu            | $^{2}C_{5}$   | $111.68\pm125.29$    | $166.13\pm7.26$                        |  |
|                            | Glc            | ${}^{4}C_{1}$ | $240.92\pm78.38$     | $11.58 \pm 6.44$                       |  |
| with Neu5,9Ac <sub>2</sub> | Gal            | ${}^{4}C_{1}$ | $212.76\pm121.08$    | $11.25\pm7.3$                          |  |
|                            | Neu            | $^{2}C_{5}$   | $206.28\pm130.93$    | $166.02\pm7.76$                        |  |

<sup>*a*</sup> The average value  $\pm$  the standard deviation over the 25-ns MD trajectory are shown.



**Figure S10.** An overlay of 10 conformations of the trisaccharide of a) 4 b) 6 and c) 8 from the MD simulation in the NEU2 active site. The protein is omitted for clarity.



**Figure S11**. An overlay of 10 conformations of the trisaccharide of a) 4 b) 6 and c) 8 from the MD simulation in the NEU3 active site. The protein is omitted for clarity.



Figure S12. Negative control ESI-MS experiments done in the absence of enzyme. Data shows that sialic acid cleavage cannot be attributed to fragmentation of the substrate.

#### References:

- 1. Ogura, H., Furuhata, K., Sato, S., Anazawa, K., Itoh, M., and Shitori, Y. (1987) Synthesis of 9-O-acyl- and 4-O-acetyl-sialic acids, *Carbohydr. Res.* 167, 77-86.
- Kiefel, M. J., Wilson, J. C., Bennett, S., Gredley, M., and von Itzstein, M. (2000) Synthesis and evaluation of C-9 modified N-acetylneuraminic acid derivatives as substrates for Nacetylneuraminic acid aldolase, *Bioorganic Med. Chem.* 8, 657-664.
- Yu, H., Huang, S., Chokhawala, H., Sun, M., Zheng, H., and Chen, X. (2006) Highly Efficient Chemoenzymatic Synthesis of Naturally Occurring and Unnatural α2,6-Linked Sialosides: A P. damsela α2,6-Sialyltransferase with Extremely Flexible Donor Substrate Specificity, *Angew. Chem. Int. Ed.* 45, 3938-3944.
- 4. Sandbhor, M. S., Soya, N., Albohy, A., Zheng, R. B., Cartmell, J., Bundle, D. R., Klassen, J. S., and Cairo, C. W. (2011) Substrate recognition of the membrane-associated sialidase NEU3 requires a hydrophobic aglycone, *Biochemistry* 50, 6753-6762.
- Seyrantepe, V., Landry, K., Trudel, S., Hassan, J. A., Morales, C. R., and Pshezhetsky, A. V. (2004) Neu4, a novel human lysosomal lumen sialidase, confers normal phenotype to sialidosis and galactosialidosis cells, *J. Biol. Chem.* 279, 37021-37029.
- 6. Nomenclature, I.-I. J. C. o. B. (1980) Conformational Nomenclature for Five and Six-Membered Ring Forms of Monosaccharides and Their Derivatives, *Eur. J. Biochem.* 111, 295–298.
- 7. Cremer, D., and Pople, J. A. (1975) General definition of ring puckering coordinates, J. Am. Chem. Soc. 97, 1354–1358.