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THE UNIVERSITY OF ALBERTA

STUDIES ON THE SUB-UNIT STRUCTURE OF THE

INNER MITOCHONDRIAL MEMBRANE

by

(C) SANTOSH S. SIKERWAR

A THESIS

SUBMITTED TO THE FACULTY OF GRADUATE STUDIES AND RESEARCH

IN PARTIAL FULFILMENT OF THE REQUIREMENTS FOR THE DEGREE OF

Master of Science

in

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THE UNIVERSITY OF ALBERTA

FACULTY OF GRADUATE STUDIES AND RESEARCH

The undersigned certify that they have read, and re-
commend to the Faculty of Graduate Studies and Research, for ac-
ceptance, a thesis entitled, "Studies on the Sub-unit Struc-
ture of the Inner Mitochondrial Membrane" submitted by
Santosh S. Sikerwar in partial fulfilment of the requirements
for the degree of Master of Science in Cell Biology.

Supervisor

Date: 8th December, 1977.

Dedicated to my parents

and my brother

ABSTRACT

Structural organization of the inner mitochondrial membrane, isolated in the form of the inside-out vesicles called the sub-mitochondrial particles (SMP), has been investigated in the present work. The appearance of F_1-ATPase molecules in the negatively stained preparations of isolated mitochondrial membrane has led many workers to suggest that there is a regularly repeating structure in the inner mitochondrial membrane. In the present work experiments have been carried out to explore the periodic two-dimensional structure within the plane of inner mitochondrial membrane in SMP. SMP prepared from beef heart mitochondria have been negatively stained with 2% PTA (pH 7.1), and subsequently photographed in an electron microscope suitably modified to reduce the specimen damage due to the electron beam. To enhance the high resolution components in the image, the electron micrographs were taken at underfocus settings. Such micrographs of negatively stained SMP have been subjected to the optical diffraction analysis, and the resulting optical Fourier transforms have revealed an absence of any periodic two-dimensional structure within the plane of the membrane, thereby negating the possibility of a sub-unit structure in the membrane as discerned in SMP.

Spatial arrangement of the intramembranous proteins in the hydrophobic interior of the membrane has been examined by freeze-etching of the SMP preparations suspended in glycerol or water. This technique provides the direct visualization of membrane proteins present both on the surface and in the hydrophobic zone of the membrane. The concave (PF) and convex (EF) fracture faces display the presence of typical intramembranous particles ranging in diameter from 4nm to 16nm. An asymmetry in the distribution of these particles between the two fracture faces has been noted. The packing density of particles in the concave fracture face (PF) is 2000 ± 400 particles/μm^2 and in the convex fracture face (EF) is 1300 ± 300 particles/μm^2. Most of the particles fall into the size range 10-12nm, followed by 5-7nm range. On comparing the frequency distribution of particle sizes with frequency distribution of the sizes of the members of respiratory chain calculated from the available data, it can be inferred that there is no direct correspondence between the intramembranous particles and individual members of respiratory chain. It appears therefore that the components of respiratory chain are involved in molecular associations, which may occur between members of the respiratory chain. Such associations would be distinctly advantageous as they may facilitate cooperative interactions.

The optical diffraction experiments on the electron micrographs of freeze-fracture replicas display.

a diffuse ring extending from about 9 to 12nm in the optical Fourier transforms. Had there been a regular two-dimensional lattice of intramembranous particles in the hydrophobic interior of the membrane of SMP, the optical Fourier transforms should have displayed sharp diffraction spots or ring(s). Therefore, it appears that there is a lack of any regular two-dimensional periodic arrangement of the particles in the hydrophobic interior of the membrane of SMP. This may reflect the fluid matrix of the membrane, as most biological membranes are currently thought to have.

An attempt has also been made to visualize the F_1-ATPase knobs on the surface of SMP by freeze-etching SMP suspended in water. The resulting freeze-etch replicas have revealed a smooth etched surface (PS). This observation is contradictory to what is expected on the basis of negative staining experiments on SMP which have displayed the presence of ~10nm F_1-ATPase knobs attached to the surface of SMP by the stalks. Consequently the in vivo organization of F_1-ATPase particles in relation to the rest of the membrane remains uncertain.

ACKNOWLEDGEMENT

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LIST OF ABBREVIATIONS

$$SMP = \text{sub-mitochondrial particles}$$
$$TRU = \text{tripartite repeating unit}$$
$$P.E.P. = \text{peak enveloping power}$$
$$F_1\text{-ATPase} = \text{coupling factor } F_1$$
$$OSCP = \text{oligomycin sensitivity conferring protein}$$
$$EF = \text{exoplasmic fracture face}$$
$$PF = \text{protoplasmic fracture face}$$
$$PS = \text{protoplasmic surface}$$
$$ES = \text{exoplasmic surface}$$
$$HBHM = \text{heavy beef heart mitochondria}$$

INTRODUCTION

The last decade has witnessed a rapid growth in
our understanding of the structure of biomembranes and
ideas about lipid-protein interactions and the diversity of
protein types in biomembranes have undergone a consider-
able change. Biological membranes are currently thought
to be fluid mosaics (Singer, 1971; Singer and Nicolson,
1972), with globular proteins intercalated into an inter-
rupted lipid bilayer. As with all generalised models of
membrane structure, such a picture should be viewed with
caution. Especially it should be borne in mind that al-
though the existence of a lipid bilayer may be considered
to be well established in some membranes, precise informa-
tion about the membrane proteins and their mode of inter-
action with lipids is very scarce (Cherry, 1976). How-
ever, irrespective of the particular arrangement of pro-
teins in a membrane, an obvious consequence of fluidity
in the bilayer matrix leads to the possibility that the
membrane components may be mobile. Lipids may diffuse
laterally in the plane of the membrane (Kornberg and
McConnell, 1971a; Devaux and McConnell, 1972; Träuble
and Sackmann, 1972; Scandella et al., 1972) or by a flip
flop process cross from one side of the bilayer to the
other (Deamer and Branton, 1967; Kornberg and McConnell,

1971b; Kennedy and Rothfield, 1977). Proteins can also be expected to show both lateral (Frye and Edidin, 1970) and rotational (Wahl et al., 1971; Vanderkooi and Martonosi, 1971) diffusion if they are not held in their place by interactions with each other or with other cellular components, e.g., microtubules and microfilaments.

It is evident from the recent developments in our understanding of the structure of biomembranes that biophysical and biochemical approaches have been rewarding in providing knowledge about their structure. Techniques of nuclear magnetic resonance spectroscopy (Lee et al., 1974; Levine, 1972), electron spin resonance spectroscopy (Likhtenshtein, 1976; Berliner, 1976; Seelig, 1976; Smith and Butler, 1976; Griffith and Jost, 1976; McConnell, 1976), X-ray diffraction (Worthington, 1976; Luzzati, 1976a; Luzzati, 1976b; Luzzati, 1968; Levine, 1973), flourescence spectroscopy (Radda, 1975), neutron beam studies (Worcester, 1976), differential scanning calorimetry and differential thermal analysis (Chapman, 1973; Sturtevant, 1974; Melchoir and Steim, 1976), electron microscopy (Unwin and Henderson, 1975; Zingsheim and Plattner, 1976) and classic techniques of surface science (Sears and Stark, 1973) have provided a wealth of information. Such studies have led to the acceptance of the generalized membrane model described above.

One of the functions of biomembranes is to serve as the reversible transducers of several forms of energy (Mueller, 1969; Chance et al., 1971; Clayton, 1973; Trebst, 1974; Hodgkin, 1963; Haggins, 1972; Inesi, 1972; Rang, 1974;

Kaback, 1974; Skou, 1975; Montal, 1976). One of the major unsolved problems of bioenergetics is to obtain a complete working knowledge of the structure of the membranes of energy transducing organelles. The process of oxidative phosphorylation in which ATP is synthesized by the energy liberated during the substrate oxidation (Slater, 1976; Hatefi *et al.*, 1976; Slater, 1971) takes place in the inner mitochrondrial membrane in eukaryotes. Therefore the study of the structural organization of inner mitochondrial membrane becomes necessary as it would be important in elucidating the mechanism of oxidative phosphorylation.

The inner membrane of mitochondria contains approximately 80% proteins and 20% lipids (Lehninger, 1975). It is composed of many kinds of proteins and phospholipids. Functionally the proteins present in the inner mitochondrial membrane can be divided into three groups:

1. electron transport system, such as flavoproteins and cytochromes (Keilin, 1930; Yakushiji and Okunuki, 1940; Hatefi, 1966);

2. energy transfer system, including mitochondrial ATPase (Pullman *et al.*, 1960) and oligomycin sensitivity conferring proteins (Kagawa and Racker, 1966a; MacLennan and Tzagoloff, 1968); and

3. other proteins, such as carrier proteins of ions and nucleotides.

The inner mitochondrial membrane is functionally and structurally asymmetric and has a specific vectorial organization (Racker, 1970). It is intrinsically permeable to H^+, OH^-, Cl^- and most other simple cations and anions, but has a highly selective permeability for specific metabolites and certain mineral ions because of the presence of specific transport systems (Lehninger, 1975).

Distinct arrays of polyhedral or spherical substructure were observed by Fernández-Morán in 1962, on the surface of "electron transport particles". Later on Fernández-Morán *et al.* (1964) published their detailed studies on these particles and concluded that the cristae of beef heart mitochondria were made up of thousands of particles, each of which had three parts:

1.   a spherical or polyhedral head piece (8 to 10 nm in diameter);

2.   a cylindrical stalk (about 5 nm long and 3-4 nm in diameter); and

3.   a base piece (4 x 11 nm).

These authors thought of base-pieces as forming the integral part of the outer dense layers of the cristae (of the surface that faced away from the matrix). These authors also claimed that negative staining with phosphotungstate was one of the several methods which could be used for the reproducible demonstration of the isolated particle. Isolated particle, when stained with phosphotungstate or shadow-cast, was shown as being a studded ellipsoid appro-

ximately 12 nm in its short dimension and 18 nm in its long
dimension (Blair *et al.*, 1963)..

Same particles (8.5 nm in diameter) were observed
by Stoeckenius (1963) in the negatively stained preparations
of mitochondria from *Neurospora crassa*, while a few experi-
ments with the mitochondria from the organs of mammals gave
the same results. The stain used was usually phospho-
tungstic acid (pH 6.8-7.2) and the results obtained with
uranyl acetate (pH 7.2) in the presence of ethylenediamine-.
tetraacetic acid (EDTA) were almost identical, although in
general less satisfactory. In his electron micrographs,
usually, the particles appeared to be separated from the
edge of the membrane by a little empty space, although in
some preparations at higher magnifications, a narrow
stalk, 4.0 to 5.0 nm long, was seen. Chemical fixation by
glutaraldehyde destroys at least 95% of the stalked mem-
brane bound particles (Stoeckenius, 1963) (Keyhani, 1972).
When the mitochondria were negatively stained after fixa-
tion with $KMnO_4$, OsO_4 vapors (Stoeckenius, 1963), or
OsO_4 solutions (Keyhani, 1972), the particles were complete-
ly absent. In a study conducted by Parsons (1963), pro-
jecting subunits were found on the cristae of mitochondria
from eight types of mammalian tissue. Malhotra and Eakin
(1967) concluded from their study on the mitochondria iso-
lated from the wild type *Neurospora crassa* that the knobs
were seen in the negatively stained preparations only when
the sucrose isolation medium did not contain EDTA. These

authors suggested that the presence of knobs may have been

dependent on the physiological state of mitochondria and

procedure of negative staining, and their appearance in

the micrographs did not reflect on their capacity for

oxidative phosphorylation. Racker and his collaborators

soon demonstrated that the inner mitochondrial membrane

knobs were ATPase molecules ("coupling factor F_1";

Racker et $al.$, 1965; Racker, 1970; Racker, 1976) and iden-

tified the stalk as a factor for conferring sensitivity

for a certain antibiotic to the ATPase knobs ("OSCP" =

oligomycin sensitivity conferring protein; Racker, 1970;

Racker, 1976).

Another coupling factor, F_6 (Fessenden-Raden,

1972) has also been identified, and this in addition to

OSCP is needed for the attachment of F_1-ATPase to the mem-

brane (Racker, 1976; Kanner et $al.$, 1976). A hydrophobic

protein factor F_0 has been obtained in the form of a mem-

branous preparation from mitochondria (Kagawa et $al.$,

1973). It confers oligomycin sensitivity to F_1 and con-

sists of a proteolipid. It is thought to be responsible

for the transmembranous movement of protons (Racker, 1976).

OSCP and F_6 are made up of the single polypeptide chains

of molecular weights 18,000 and 8,000 respectively (Racker,

1976). Another coupling factor F_2 or factor B (Racker,

1970) has also been described, but it still needs further

characterization.

As for the F_1-ATPase knobs, Sjöstrand suggested that these "lollipop"-like structures are artifacts (Sjöstrand, 1968). He proposed that the procedure of negative staining would induce these "lollipop"-like structures because the ATPase molecules would "stick out" due to osmotic shock during specimen drying. However even when the osmotic shock was minimized by negative staining in ammonium molybdate (Muscatello and Horne, 1968; Muscatello and Carafoli, 1969); these "lollipops" were still present. These stalked globules were later on demonstrated in the ultrathin sections of mitochondria and submitochondrial particles, after gluteraldehyde fixation and staining in uranyl acetate (Methanolic) (Talford and Racker, 1973).

The concept that there is a basic repeating unit in the inner mitochondrial membrane came originally from Green and his colleagues (Fernández-Morán, et al., 1964). These repeating units were described as tripartite structures, each consisting of a polyhedral or spherical headpiece (8.0 to 10.0 nm in diameter), a cylindrical stalk (about 5.0 nm long and 3-4 nm wide), and a base-piece (4 x 11 nm). However, later on Green proposed the electromechanochemical model (EMC model) for the energy transduction in mitochondria, and again invoked the existence of an inner membrane repeat unit, in which the component proteins assume metastable conformations (Green, 1972; Green, 1974). Green also proposed a mitochondrial super-

molecule (for his EMC model) which consists of five complexes (Green, 1974): the four complexes of the electron transfer chain and a central unit, the tripartite repeating unit (TRU), which contains systems involved in both ATP synthesis (the F_1-ATPase) and active transport. The complexes including the headpiece of TRU, all span the membrane from matrix to the intracristal side (Green, 1974).

In the present work Green's hypothesis has been tested by investigating the sub-unit structure within the plane of the inner mitochondrial membrane (isolated in the form of the vesicular SMP), obtained from beef heart mitochondria. The theoretical basis underlying the experiments that were carried out to achieve the above-mentioned objective, is described below.

A membrane is a three-dimensional structure and a schematic drawing of the single membrane is given in Figure 1. The electron density distribution of this membrane can be represented by t(r), where r represents real space coordinates (Worthington, 1976). This t(r) can be written as

$$t(r) = t(x)t(y,z),$$

(Worthington, 1976), where t(x) and t(y,z) are lameller and sub-unit structure respectively. The one-dimensional electron density distribution in a direction at right

angles to the membrane surface is referred to as the lameller structure. The sub-unit structure refers to the structure within the plane of the membrane (Worthington, 1976).

In an X-ray diffraction experiment, diffractions arising from t(x) and t(y,z) can be distinguished as the X-ray beam parallel to the surface of the membrane gives rise to lameller diffraction, whereas when the X-ray beam is perpendicular to the surface of the membrane, it will give rise to sub-unit diffraction.

In the case of the inner mitochondrial membrane, if there is any regularly repeating structure within the plane of the membrane as has been proposed by the workers cited above, this regular surface lattice must be represented in the optical Fourier transform of the image of SMP in electron micrographs. This situation is analogous to the X-ray diffraction experiment, designed to obtain the sub-unit diffraction from the membrane. In the optical diffraction, a laser is used instead of X-rays and the membrane specimen is replaced by its electron micrograph. In the present study, the technique of optical diffraction was employed to reveal all the periodicities present in the electron micrographs, as it provides the optical Fourier transforms of the micrographs. Klug and Berger (1964) first suggested the application of optical diffraction for the analysis of periodic structures in electron micrographs. Later on Klug and his coworkers (Klug and

DeRosier, 1968; DeRosier and Klug, 1968) developed two
novel methods which enable an electron microscopist to ex-
tract phase information in the micrographs. (Optical dif-
fraction pattern records only the intensity $|\psi|^2$ of the
Fourier components of the specimen and not the phases).
The information that can be obtained from the optical dif-
fraction pattern (optical Fourier transform) of an object
is quite extensive. Unit-cell dimensions can be measured
much more accurately from an optical diffractogram than
from the direct measurements on the micrograph (Berger,
1969; Sternlieb and Berger, 1969). By employing optical
diffraction one can get separate details of periodic
character from the substrate noise (phase contrast induced
granulation). Symmetry elements are revealed in the opti-
cal diffraction pattern of the micrograph (Klug and Berger,
1964).

The electron micrographs of the freeze-etch re-
plicas were also processed for optical diffraction, to
investigate the arrangement of the "intramembranous parti-
cles" on the fracture faces of SMP. Purpose of this exer-
cise was to find information regarding the order of arrange-
ment of these particles in the hydrophobic interior of the
inner mitochondrial membrane (SMP).

MATERIAL AND METHODS

Beef heart was chosen as the source for obtaining the
mitochondria because (1) it is available in bulk quantities and
is cheaper, and (2) beef heart mitochondria are more stable over
long storage periods. All the chemicals other than potassium
succinate and PTA (which were purchased from Fisher Scientific
Co.) were obtained from Sigma Chemical Co.

Isolation of Mitochondria

The mitochondria were isolated from the beef heart by
the method given by Smith (1967). Two beef hearts were obtained
from a slaughter house within one to two hours after the animals
were sacrificed. To ensure the cooling of the tissue for transport
to the laboratory, the hearts were placed in ice. All subsequent
operations were carried out at 2-4°C. The fat and the connective
tissue were trimmed from the heart and then the tissue was chopped
into small pieces. Approximately 300 grams of the chopped tissue
was passed through the meat grinder which was maintained at 4°C.
The meat grinder had plate holes approximately 4-5mm. in diameter.
Thus obtained mince was placed in 400 ml. of 0.25M sucrose and
0.01M Tris-HCl, pH 7.8. The pH of the suspension was adjusted to
7.5 as rapidly as possible with 2M Tris (pH 10.8, unneutralized).
This neutralized ground heart mince was placed in a double

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layer of cheesecloth and was squeezed free of the sucrose solution. About two hundred grams of this ground, neutralized heart tissue were suspended in 400 ml. of 0.25M sucrose, 0.01M Tris-HCl, pH 7.8; 1 mMTris-succinate, and 0.2 mM EDTA (this solution will be referred to as the sucrose solution). In a glass homogenizing vessel, fifty milliliters of the suspension was placed, and a loose-fitting pestle driven by the heavy-duty drill at 1400 rpm was inserted into the vessel for one pass of 10 seconds and two passes of about 5 seconds each. The rest of the suspension was homogenized in a similar way, and the final homogenate was adjusted to pH 7.8 with the addition of 1 M KOH. The homogenate was centrifuged for 20 minutes at 1200 g (Sorvall centrifuge RC2-B, rotor SS34 at a setting of 3250 rpm) in order to separate unruptured muscle tissue and nuclei. The supernatant solution was slowly decanted, taking care not to disturb the loosely-packed fluffy layer. To remove lipid granules, it was filtered through two layers of cheesecloth, and the pH of the suspension was again brought to 7.8 with 1M KOH. This suspension was centrifuged for 15 minutes in a type 40 rotor of the Beckman L3-40 centrifuge at 26,000 g (22,000 rpm). A pellet resulted, which consisted of three distinct layers: (1) a light, loosely-packed buff-colored layer (light beef heart mitochondria), (2) a dark brown layer (heavy beef heart mitochondria), and (3) a tiny brown-black button at the bottom of the tube. The top layer, which consists of dam-

aged mitochondria was discarded by decanting about 20-25 ml. of the supernatant solution and then gently shaking the sucrose solution in the centrifuge tube. Thus the loosely packed damaged mitochondria were dislodged and the mixture was decanted out. Some light beef heart mitochondria adhered to the sides of the tube, which were removed with the aid of a glass stirring rod. The dark brown layer of the heavy beef heart mitochondria was then dislodged by a glass stirring rod, mixed with 10 ml. of sucrose solution, and decanted, leaving behind the brown-black pellet. This mitochondrial suspension was homogenized in a tight fitting teflon homogenizer, with two passes, each of 5 seconds at 1400 rpm.

The volume of the homogenate was made up to 180 ml. with sucrose solution, pH was readjusted to 7.8, and the suspension was again centrifuged at 26,000 g for 15 minutes. The resulting pellet had three layers; a small top, a large middle, and small bottom layer. Only the middle dark brown layer was collected as described above. The mitochondrial suspension was homogenized, and the volume was adjusted to 60 ml with the sucrose solution. The pH was adjusted to 7.8. This suspension was centrifuged at 26,000 g for 15 minutes. This time again the dark brown middle layer was collected, suspended in a small volume of sucrose solution, and homogenized. The protein concentration was adjusted to 20-40 mg protein per milliliter. Protein determinations were done by Lowry's method (1951)

and the average yield of heavy beef heart mitochondria
(HBHM) was about 1 mg protein per gram of initial mince.

Preparation of Sub-mitochondrial Particles (SMP)

The sub-mitochondrial particles from beef heart
mitochondria were obtained by the method of Hansen and
Smith (1964). HBHM were isolated as described above, and
were suspended in a solution 0.25M in sucrose, 0.01M in
Tris-HCl, pH 7.8, 1mM in ATP, 1mM in $MgCl_2$, and 1mM in
potassium succinate (this solution is abbreviated as
STAMS), at a protein concentration of 30 mg/ml; pH of the
suspension was adjusted to 7.8 with 1N KOH. This suspen-
sion was centrifuged at 26,000g for 15 minutes. The dark
brown middle layer of HBHM was collected as described
before, and suspended at a protein concentration of 30
mg/ml in a solution 0.25M in sucrose, 0.01M in Tris-HCl
(pH 7.5), 1mM in ATP, 1mM in potassium succinate, 5mM in
$MgCl_2$, and 10mM in $MnCl_2$. The pH of the solution was ad-
justed to 7.5 with either 1N HCl or KOH. Aliquots of about
15 ml. were placed in a tube, kept in ice, and subjected
to 10Kc sonic irradiation for 2 minutes from a sonifer
(Braunsonic 1510) at a power output between 100 to 150
Watts (P.E.P.). The pH was adjusted to 7.5 and the re-
sulting suspension was centrifuged at 14 x 10^4g-min. (20,0(
rpm, 7 minutes, No. 40 rotor in a Beckman L3-40 ultracen-
trifuge). The supernatant was collected by decantation

and was centrifuged at 40,000 rpm in a Beckman Type 40 rotor for 45 minutes in a Beckman L3-40 ultracentrifuge (105,000 × g). As a result of this a tightly packed reddish brown pellet was obtained. The surface of the pellet was rinsed with a few milliliters of a solution 0.25M in sucrose and 0.01M in Tris-Hcl, pH 7.5, and the pellet was suspended to a protein concentration of about 20 mg/ml in sucrose-Tris solution described above. Protein determinations were done by the method of Lowry. (1951).

Negative Staining

For negative staining a drop of the SMP suspension diluted to approximately 1 mg protein/ml, was placed on the formvar and carbon-coated grids and stained with 2% phosphotungstic acid (PTA), pH 7.1. Phosphotungstic acid was used because it is thought to be an effective stain, for the many polar atoms in its periphery impose an ordered structure on adjacent water molecules and leave the structure beyond it undistorted so that water can evaporate with little damage to the surface of the molecule (Blundell and Johnson, 1976).

Freeze-etching of SMP

a. Freeze-etching of glycerolated SMP

For freeze-etching in glycerol, the SMP suspension was centrifuged for 45 min. at 40,000 rpm in a Type 40

rotor in a Beckman L3-40 ultracentrifuge. The resulting
pellet was suspended in 20% glycerol at the protein con-
centrations of 10-15 mg/ml. After 2 hours in glycerol
solution, drops of sample were frozen in liquid Freon 22
and transferred to a Balzers freeze-fracture device and
fractured at $-100^\circ C$ as described by Malhotra and Tewari
(1973). The preparations were etched for 70 seconds
with a liquid N_2 cooled knife right over the specimens at
a vacuum of 1×10^{-6} torr and replicated with platinum-
carbon. The replicas were soaked in chromic acid over-
night, washed for 1 hour in household detergent solution,
followed by three washings in distilled water and eventual-
ly these were picked up on the formvar-coated copper grids.

b. Freeze-etching in water

The SMP suspension was centrifuged for 45 minutes
at 40,000 rpm in a Type 40 rotor in a Beckman L3-40 ultra-
centrifuge. The resulting pellet was suspended in dis-
tilled water at a concentration of approximately 5 mg
protein/ml and centrifuged again at 40,000 rpm for 45
minutes, in a Type 40 rotor in a Beckman L3-40 ultracen-
trifuge. The pellet so obtained was suspended in distilled
water at a protein concentration of 10-15 mg/ml. Drops
of the sample were frozen in liquid Freon 22 and processed
for freeze-etching as described before in the case of the
SMP suspended in 20% glycerol.

Electron Microscopy of Negatively Stained SMP Preparations

In the high resolution electron microscopy of
biological or organic specimens, the radiation damage poses
a very severe limitation over the obtainable resolution.
In spite of the continual developments in the instrumen-
tal resolving power and the applications that have been
made with these improvements to the inorganic materials,
the best resolution of periodic structure obtained with
stained biological specimens is usually 2 to 3 nm. The
resolution, attained with the unstained biological speci-
mens is far worse than the above-mentioned value. How-
ever, a few exceptions to the general limitations on re-
solution in organic materials have been found, but these
exceptions always occur in materials that are unusually
resistant to the radiation damage (Glaeser, 1975). From
a simple calculation it can be shown that the usual ex-
posures needed in high resolution electron microscopy lead
to a radiation dose of 10^{10} to 10^{11} rads being deposited
in the sample (Glaeser, 1971; Glaeser *et al.*, 1971; Grubb
and Keller, 1972). On the other hand, a dose of 10^9 rads
is sufficient to destroy the original properties of most
organic materials and a dose of 10^6 rads will inactivate
most of the enzymes and kill the most resistant of all
living things (Glaeser, 1975). An idea of the severity
of radiation damage can also be obtained from the follow-
ing example (Stenn and Bahr, 1970): In a 0.1μ thick

organic specimen, at 100kV a single electron sustains approximately one inelastic collision in passing through it. Each collision has an energy yield of the order of 32eV (the energy yield of a C-C single bond is of the order of 5eV). At a magnification of 10,000x, a current density of at least 10^{-2} amp/cm^2 is required and from this it has been calculated (Stenn and Bahr, 1970) that energy absorbed by 1g of specimen in a 1s exposure is sufficient to bring 20g of ice to steam.

An estimate of the electron fluxes to "see" the molecular detail has been made by Glaeser (1971). The detectability of an object feature of the characteristic dimension d is determined by the ratio of signal to noise in the measurement. In order to record the image features with low contrast under the conditions of severely restricted exposure, the signal-dependent noise, associated with statistical fluctuations in the intensity from one area to another in the image, must be given careful consideration (Glaeser, 1971; Glaeser, 1975). Image contrast can be defined as the ratio between spatial variations in the intensity and local average of intensity (Glaeser, 1975):

$$C = \frac{\Delta I}{\langle I \rangle} \qquad \ldots (1)$$

In the above equation C denotes inherent image contrast. The incidence of the electrons at the image plane is a

random process (Glaeser, 1971). If the total number of electrons passing into a given image "point" (picture element) is n, then the statistical fluctuation, or "counting error" is $\sqrt{n}$ (Glaeser, 1971). It is beneficial to express this fluctuation in terms of the following parameters:

(1)   the area of picture element, d^2;

(2)   the current density through the object, j;

(3)   the "integration time" or exposure time, t; and

(4)   the fraction of electrons passing through the specimen that actually enter the lens aperture and contribute to the image, f (Glaeser, 1971). And thus,

$$n = fjtd^2 \qquad \qquad \ldots (2)$$

In case of limited exposure, a certain statistical fluctuation occurs in the particle flux from one image point to another, leading to the generation of spurious structure, or spatial noise, with contrast (Glaeser, 1971):

$$C' = \frac{\Delta n}{n} \qquad \qquad \ldots (3)$$

where

$$\frac{\Delta n}{n} = \frac{1}{\sqrt{fjd^2t}} \qquad \qquad \ldots (4)$$

Therefore, in order to "resolve" a low contrast image feature, the inherent contrast must exceed the statistical fluctuations by the minimum acceptable signal to noise-ratio, S/N.   It leads to the following inequality (Glaeser, 1971):

$$Cd \geq \frac{S/N}{\sqrt{fjt}}$$ . . . (5)

According to this equation the product of resolution and contrast must exceed a certain constant, which in turn is determined by the conditions of measurement (f, j, and t) and by the conditions of analysis (minimum ajceptable S/N) (Glaeser, 1971). Therefore, to record some object feature with contrast C at the resolution d, the integrated flux density it must be greater than or equal to $(S/N)^2/C^2d^2$. For a specimen with a value of S/N = 5, the value usually quoted for the visual perception of structure (Rose, 1948), and an inherent image contrast C = 0.1, the minimum resolution attainable with a flux density of 10^{17} electrons/cm^2 (the maximum permissible dose for negatively stained catalase crystals) is of the order of 1.6nm (Glaeser, 1971).   To resolve up to 0.5nm, a current density of 10^{18} electrons/cm^2 is needed which would totally destroy the specimen. Thus the radiation damage leads to a paradoxical situation.   In order to achieve high resolutions, we need greater electron fluxes, but these electron fluxes would destroy the organic specimen's structure.

Disordering effect of the high vacuum in the electron microscope also causes extensive damages to the biological or organic specimens (Stenn and Bahr, 1970).

The suggestion has often been made that the problem of radiation damage would be greatly reduced if the specimens could be examined in a vacuum of 10^{-8} torr or better. This suggestion is based on the observation that the specimens undergo dramatic modifications and it is associated with residual gases in the microscopes at a pressure of 10^{-5} to 10^{-6} torr. Specimen contamination and specimen etching are also one of the common problems encountered in preserving the specimen structure in the electron microscope.

Negative staining alleviates these damaging effects to some extent. Salts used in the negative staining lose water quicker than the protein molecules in the drying process and hence preserve the structure of the hydrated protein (Horne and Whittaker, 1962; Johnson and Horne, 1970), and form a glass around the specimen which is likely to prove more resistant to the beam damage than the biological molecule alone. It was due to this reason that in the present study, the negative staining method was employed to visualize the SMP preparations in the electron microscope. The electron microscope was also suitably modified to provide low-dosage conditions for the electron-microscopy of the negatively stained SMP preparations. The overall objective was to reduce the specimen damage, due to the electron beam, in the electron microscope.

The following method was used for taking the bright field electron micrographs of the negatively stained

6MP preparations. A Philips EM 300 electron microscope, equipped with a liquid N_2 cooled anti-contamination device, operating at 100kV with 50µm (first) and 50µm (secone) condenser apertures was used. No objective aperture was used. A narrow coherent illuminating beam could be obtained by exciting the first condenser lens, strongly. At first the grid was scanned at low magnifications using very low illumination levels, just enough to reveal the outline of the structures. Once a suitable area was found, it was brought on the optical axis of the electron-microscopy (the marked spot on the large viewing screen) and the magnification was increased to the desired level. The specimen was then moved slightly using the translation controls, so that the area of interest was removed far from the center of the screen. Following this a narrow coherent illuminating beam was obtained, as described above, to enable the focusing to be done. The beam was then expanded to give the desired intensity on the screen (this intensity of illumination was kept as low as possible with the photographic exposure times kept at 4-16 sec.) and the area of interest was brought back and placed on the optical axis of the electron microscope and photographed.

Electron Microscopy of Freeze-etch Replicas

Freeze-etch replicas were examined routinely in a Philips EM300 electron microscope operating at 80kV with a liquid N_2 cooled anticontamination device using

300μm (first) and 300μm (second) condenser apertures and a 25μm objective aperture.

Optical Diffraction

Optical diffraction experiments were carried out on a Polaron electron micrograph optical-diffractometer model M802. Optical elements in this optical diffracto-meter are mounted on a 3 metre length of double rail pre-cision optical bench. The optical diffractometer lens system shown in Figure 2 can be easily divided into three parts:

a.   the electron micrograph illuminating system;

b.   the transform imaging system; and

c.   the image reconstitution system.

a. Micrograph illuminating system

A 2 milliwatt helium-neon laser provides a source of coherent light of λ = 632.8nm. It gives a beam of about 2mm cross-sectional diameter, and it is mounted on adjust-able legs. A spatial filter, SP, consisting of microscope objective fitted with the front face of laser and adjus-able iris diaphragm is employed to prevent any extraneous light from laser. The rays become divergent in passing through the objective and are converted by a planoconvex lens B to a collimated beam of light approximately 1 cm. in diameter incident on the electron-micrograph C. The electron micrograph is suitably masked to expose the par-

ticular area of interest only, and mounted on a mechanical stage which provides adjustments for selecting any area on the micrograph.

b.   Transform imaging system

A plano convex lens D is used to image the optical Fourier transform of the micrograph. A two-lens telephotosystem consisting of lenses E and F is employed to magnify the image of the transform. The transform can be viewed by placing a ground glass screen in the mask holder G and recorded with 5 x 4 Polaroid Land camera back provided. The polaroid print of the transform is used for making a metal mask which allows the primary beam and selected diffracted beams to go into the image reconstruction system. The mask is fitted in holder G in image reconstitution experiments.

c.   Image reconstitution system

The rays which pass through the mask are reconstructed using lens H and J. Together these lenses form a telephoto system which is reversed and symmetrical with respect to lenses E and F. Lens K, which is final, is a power projecter lens. Again, the reconstituted image may be viewed using the ground glass screen or can be recorded by the camera.

However, in the present study, where the transform (optical Fourier transform) was of main interest, the lenses H, J and K (image reconstitution lenses) were re-

moved and other lenses were rearranged on the entire
optical bench to increase its magnification.

Both the negatives of the electron micrographs
of negatively stained SMP (at the magnification of x 25,000
--40,000) and their enlarged transparencies (at the magni-
fication of x 60,000--80,000) were used in the optical dif-
fraction experiments. These were suitably masked to ex-
pose only the particular areas of interest and were used as
objects in the optical diffractometer. A total of 20 dif-
ferent specimens were analyzed.

Micrographs of the freeze-etch replicas of SMP
preparations were enlarged to a final magnification of
x 242,000. Holes were punched in the micrograph at the
locations where there seemed to be particles on the con-
vex fracture face of SMP. Then the micrograph was kept on
a black paper with its face down, so that the holes ap-
peared black on white background. This was then photo-
graphed on a Kodak Panatomic X film. The resulting nega-
tive reduced the size of the mask (representing the ar-
rangement of particles on the convex fracture face of a
SMP) 19 times and was finally used as an object in the op-
tical diffractometer. A total of 10 specimens were ana-
lyzed. Photographic negatives of the optical diffraction
patterns (optical Fourier transforms) were used for mea-
surements.

Negative Staining

a. Relationship between the electron microscope image
    and the specimen

In the process of negative staining the specimen
is surrounded by an aqueous solution of an electron dense
salt (the negative stain) which dries out into a thin
film. This thin film, in effect, is an amorphous electron
dense 'glass' (Horne and Whittaker, 1962) in which the speci-
men is buried. The negative stain penetrates into the
hydrophobic regions of the biological specimen and the re-
sulting image in the electron microscope reveals the un-
penetrated regions as relatively electron-transparent
structures against an electron dense background. Thus the
amplitude contrast is provided by the scattering of elec-
trons by the stain (Blundell and Johnson, 1976).

In the electron microscopy contrast is achieved
by both the amplitude contrast and by the phase contrast
effects which are created by contributions from defocus-
ing and spherical aberration of the objective lens. Phase
contrast plays the predominant role at medium and high re-
solutions. Since the Fourier transform of the image dis-
plays these phase contrast effects more simply than the
image itself (Erickson and Klug, 1971), these can be

readily investigated with the help of an optical diffrac-
tometer or computer transforms. According to the wave
theory of image formation, the effects of defocusing and
spherical aberration are attributed to a phase shift
($\chi\{\alpha\}$) which occurs at the diffraction plane of the micro-
scope, in the scattered electron wave. It is a function of
scattering angle α and is given by

$$\chi(\alpha) = 2\pi/\lambda \left(-\tfrac{1}{4}C_s\alpha^4 + \tfrac{1}{2}\Delta f\alpha^2\right) \qquad \ldots \quad (6)$$

where C_s is the coefficient of spherical aberration and
Δf is the defocusing (positive for a weak or underfocused
lens) (Erickson and Klug, 1971). Transform of the phase
contrast image, T_{ph}^i, is related to the transform of the
object, $T^0(\alpha,\phi)$ by the expression:

$$T_{ph}^i(\alpha,\phi) = -T^0(\alpha,\phi)A(\alpha)f(\alpha)\sin\chi(\alpha) \qquad \ldots \quad (7)$$

where $A(\alpha)$ accounts for the effect of an objective aper-
ture and is called aperture function ($A[\alpha] = 1$ if $\alpha \leq$ de-
fining aperture angle; $A(\alpha) = 0$ elsewhere), α is the scat-
tering angle, $f(\alpha)$ is the atomic scattering factor for the
elastic scattering of electrons, and ϕ is the azimuthal
coordinate (Erickson and Klug, 1971). This equation tells
us that the relation of the object to the image formed in
the presence of aberrations is very simple and direct if
expressed in terms of Fourier transforms. According to

this expression the image transform is directly proportional to the true object transform, modulated by three factors. Most important factor among the three factors is $\text{Sin}\chi(\alpha)$, which is also called as the transfer function (Hanzen, 1971).

The transfer function will have the following effects on the terms corresponding to a spatial resolution of λ/α: (a) if $\text{Sin}\chi(\alpha)$ is close to 1, then these terms would provide their full weight to the Fourier transform and details corresponding to this resolution would be faithfully imaged; (b) if $\text{Sin}\chi(\alpha)$ is close to zero, then the terms of spatial resolution will be removed from the transform and will not contribute to the final image; (c) if $\text{Sin}\chi(\alpha)$ changes sign then terms corresponding to this resolution will contribute to the final image with reverse contrast leading to the risk of false image. Therefore it is required that $\text{Sin}\chi(\alpha)$ be relatively invariant over the range of resolutions of interest to us. Curves for $\text{Sin}\chi(\alpha)$ for different degrees of underfocus were computed by Erickson and Klug (1971). It was concluded that 90nm underfocus would be a favourable choice for imaging details between 2 to 0.5nm resolution, while a greater degree of underfocus would be suitable for imaging details between 5 to 2nm range. Erickson and Klug (1971) also examined the effect of transfer function in real images (negatively stained catalase crystals) and concluded that at $\Delta f = 540$nm many of the high resolution components are enhanced and

micrograph contains more detail and contrast. At this
underfocus value, the phase contrast effect is sufficient
to increase the transfer function for periodicities in the
range of 2 to 5nm by about five times to that of the
in-focus image. Since this range of detail is of particu-
lar interest in protein studies, 500nm was thought to be
the optimum underfocus setting for such work. This is a
very important study because it provides insight into one
of the most important aspects of image formation. In the
present work too, most of the electron micrographs are mo-
derately underfocused. In the present work, the electron
micrographs of negatively stained SMP were taken at two
underfocus settings: (a) at ~100nm underfocus (for imaging
details between 2.0 to 0.5 nm resolution); (b) at ~550nm
underfocus (for enhancing the high resolution components
in the range 2.0 to 5.0 nm. It was in accordance with the
conclusion drawn by Erickson and Klug (1971) with respect
to the implication of their work in biological electron
microscopy. In their own words,

> Finally it is comforting to be able to confirm
> quantitatively that the moderately underfocused
> micrographs used in most biological electron
> microscopy are valid images, frequently the
> best possible in terms of resolution and con-
> trast, with no artifacts in the low and medi-
> um resolution range of interest.

b. Negatively stained SMP and cristal membrane fragments

In a typical SMP preparation, negatively stained,
such as one shown in Figure 3, knobs approximately 10nm in

diameter can be easily identified on the inner mitochondrial membrane vesicles (SMP). These inner mitochondrial membrane vesicles have the inner-mitochondrial membrane in the inside-out configuration, and these vesicles (SMP), therefore enable F_1-ATPase knobs to be visualized. These knobs look more like polyhedral structures. It was Racker ('70, '76) who first of all demonstrated that the knobs present on the SMP are F_1-ATPase ("coupling factor"). In most cases these knobs appear to be attached to the membrane by the stalks (Figure 4). These stalks are approximately 6nm long and 3nm wide, and correspond to the oligomycin sensitivity conferring proteins ("OSCP", Racker, 1970). On the edge of the SMP, the F_1-ATPase knobs can be very clearly seen to be attached by the stalks to the SMP, in most of the cases. These stalks are separated from one another by approximately an ~11nm-wide gap, as measured on the edges of the negatively stained SMP.

Frozen beef heart mitochondria, when they are subjected to swelling and alternate freezing and thawing in 10mM Tris chloride (pH 8.0), their cristal membranes undergo "tubularization" (Green, 1976). Results from such a phenomenon are shown in Figure 5. Regular-looking arrays of knobs connected to the membrane by the stalks, on either side of the "tubular" membrane, are discernable. This tubularization may be due to the transition of a single flat continuous membrane sheet to a large number of interconnecting tubules (Green, 1976). According to this

author tubularization of the inner mitochondrial membranes means that the components of this membrane can exist in either of two states--the "two-dimensional" state of the flat membrane sheet and the "three-dimensional" state of tubular evaginations. In such structures, described as "tubular" by Green, the knobs have the diameter of 11.0nm (corresponding to the F_1-ATPase knobs) and the stalks are ~6nm long and ~3nm wide. The distance between the two stalks on the "tubular" membrane is ~11nm.

Freeze-etching

a. Interpretation of freeze-etch replicas

The evidence was first presented by Branton and his coworkers in support of their hypothesis that during, "freeze-etching", the fracture plane passes through the interior of the biological membranes, so that the inner faces become exposed (Branton, 1966). Face views of the fractured membranes show particles of ~10nm diameter. Their number is subject to variation from one membrane type to the other. Branton argued that these particles belonged to the membrane interior. It was observed that upon etching, fractured membranes frequently display a narrow ridge ~3nm thick. Where the face view changes into cross-fracture, the ridge is seen to be continuous with one of the two parallel ridges that form the usual appearance of cross-fractured membranes. Therefore it was concluded

that the membrane was split. Similar argument has also

been applied to the swollen nerve myelin, rod outer seg-

ment membranes, and swollen lecithin lameller phases

(Branton, 1967, Clark and Branton, 1968; Staehlin, 1968).

After considering actual dimensions and spacings of the

ridges, it becomes clear that an ~5.5nm-wide gap must

exist between them. Langmuir-Blodgett technique (Langmuir,

1917; Blodgett, 1935; Langmuir, 1939) has been employed to

assemble radioactively labelled bilayers and multilayers.

These were frozen and split and the radioactivity of the

separated layers was measured. The results thus obtained

are explainable only by assuming that assemblies split

along a plane defined by the methyl end groups of the indi-

vidual monolayers (Deamer and Branton, 1967). After frac-

turing and shadowing under the conditions of freeze-etching

(-100°C), very smooth surfaces were noticed. Only a few

particles could be found on these surfaces. Similarly

membranes which have little or no protein, generally dis-

play smooth fracture faces as in the nerve myelin (Branton,

1967; Malhotra *et al.*, 1975), liposomes (James and Branton,

1971) and lameller lipid phases (Deamer *et al.*, 1970).

However, more information about the nature of the surface

revealed after freeze-fracturing was obtained by experi-

ments employing chemical prefixation. Aldehyde fixation

which cross-links the proteins, did not bring about any

change in the appearance of the membranes after freeze-

fracturing and etching. However, an improvement in their

overall mechanical stability was noted (Jost, 1965; Branton and Park, 1967; Tillack and Marchesi, 1970). When the aldehyde-fixed membranes were subjected to lipid extraction prior to the freeze-fracturing and etching, the splitting of membranes was abolished completely. Although the trilameller structure could still be seen in the thin sections (Park and Branton, 1966; Fleisher et al., 1967; Branton and Park, 1967). These results clearly indicate that the lipids or lipid regions are required to achieve freeze-fracturing of the membranes along an inner plane.

Deamer and Branton (1967) have proposed an hypothesis to explain the splitting behaviour of the biomembranes. They explain their arguments on Kauzman's (1959) thermodynamic considerations of the temperature dependence of the energy of hydrophobic bonding. Freezing is known to weaken the hydrophobic bonds and so the hydrophobic interior of the biomembranes becomes more susceptible to splitting in the frozen state than any other zone. However since the frozen membrane is not in a thermodynamically equilibrium state, their argument should be weighed with caution.

Experiments with the membrane surface markers (covalently bound Ferritin or attached F-Actin) demonstrate that these markers can only be seen after etching; they never appear on cleaved membrane faces (Pinto de Silva and Branton, 1970; Tillack and Marchesi, 1970). This lends strong support to the membrane splitting hypothesis. By

employing the techniques for the production of complimen-
tary replicas from both (complimentary) fracture faces of
a membrane (Sleytr, 1970; Wehrli *et al.*, 1970; Chalfont
and Bullivant, 1970), it was found that both fracture faces
were unetchable. It can be simply explained on the basis
of the membrane splitting hypothesis.

Branton (1967; 1971) has argued that the molecular
correlates of the membrane intercalated particles occurring
within freeze-fractured biomembranes, are proteins. It
has been shown that they are pronase sensitive and their
occurrence is influenced by the "metabolic activity" of a
cell or membrane type.

Wallach and Zahler (1966) were the first to
point out the possibility and likelihood of hydrophobic
protein-lipid interaction in biomembranes. A large amount
of evidence has accumulated in the last few years regarding
the membrane intercalated particles and now the idea is
widely adopted that these membrane intercalated particles
represent proteins (including the polypeptides and oligo-
mers (Bullivant, 1974; Bretscher and Raff, 1975). Recently
(Branton *et al.*, 1975) a nomenclature for the freeze-
etching has been proposed which is simple and uniform to
describe the various fracture faces and surfaces. Accord-
ing to this scheme of nomenclature, for any membrane that
can be split, the half closest to cytoplasm, nucleoplasm,
chloroplast stroma or mitochondrial matrix is designated
as "protoplasmic half" abbreviated as P; the half closest

to the extracellular space, exoplasmic space, or endoplas-.
mic space is designated as extracellular, exoplasmic or
endoplasmic half, abbreviated as E. E is also used to
designate the half membranes closest to the space between
the inner and outer membranes of mitochondria.

After labelling the half membrane as P or E, the
particular aspect seen in the electron microscope can be
recognized as a true surface of the membrane or as a
fracture face. The surface is named either PS or ES and
it is the hydrophilic portion of the membrane usually re-
vealed after etching; the fracture face is designated as
PF or EF.

b.   Freeze-etching of SMP

Sub-mitochondrial particles are inside-out
inner mitochondrial membrane vesicles, obtained from the
mitochondria. Therefore, in SMP, the face of the inner
mitochondrial membrane, which is closest to the mitochon-
drial matrix in intact mitochondria, faces the outside of
the vesicle. And the outer surface of the inner mitochon-
drial membrane in intact mitochondria faces the inside of
the vesicle in SMP.

In a sub-mitochondrial particle, therefore, the
protoplasmic half of the inner mitochondrial membrane faces
towards the exterior of the vesicle while the exoplasmic
half faces towards the interior of the vesicle (Fig. 6a).
For the sake of explanation the sub-mitochondrial particles

can be considered to be the vesicles of varying size. During the fracturing, the fracture plane "hits" those vesicles which are within the reach of this plane. The membrane of these vesicles splits in half between the phospholipid leaflets thus revealing, on a profile in the form of a spherical surface, an internal fracture face of the membrane studded with particles. If the plane hits the vesicle above the equator, a convex fracture surface is formed; if it hits below the equator, the surface is concave (Weibel, Losa and Bolender, 1976). In SMP, the convex fracture face is the exoplasmic fracture face (abbreviated as EF) and the concave fracture face is the protoplasmic fracture face (abbreviated as PF). The corresponding etched true surfaces are called exoplasmic surface (the ES) and protoplasmic surface (the PS) (see Figure 6b).

i. Freeze-etching of glycerolated SMP

The total number of replicas examined was 40 and they were selected from 18 freeze-fracture experiments.

Typical fracture faces revealed after freeze-fracturing and etching of SMP in 20% glycerol are exhibited in Figures 7, 8 and 9. Both convex and concave fracture faces were obtained. No true surface was exposed as a result of the etching of glycerolated SMP preparations. The membrane intercalated particles are present on both the convex and the concave fracture faces. The convex fracture face is the exoplasmic fracture face (EF) and the

concave fracture face is the protoplasmic fracture face
(PF). The intramembranous particles with diameters rang-
ing from about 5.0nm to 16.0nm are present on both the
fracture faces.—It seems that the convex fracture face
(EF) has a lower packing density of particles as compared
with the packing density of particles on the concave frac-
ture face (PF). There are approximately 1300 ± 300 parti-
cles/μm^2 in the convex fracture face and about 2000 ± 200
particles/μm^2 in the concave fracture face. It indicates
the asymmetry in the particle distribution in the two halves
of the membrane.

The size distribution of the particles is given
in Figure 11. Most of the particles fall in the size
range 10-12nm, but there is a significant number of par-
ticles in the 7 to 10nm range too. The significance of
this distribution of intramembranous particles in respect
of the known composition of the inner motichondrial mem-
brane is discussed on p. 51. On the concave fracture face,
a few of the particles are sometimes seen to be arranged
in some kind of irregular rows, and their pattern is un-
even (Figure 10).

ii. Freeze-etching of SMP in water

When the sub-mitochondrial particles are suspend-
ed in water and subjected to freeze-fracturing and etching,
at least two types of membrane faces must be exposed in
principle. One is the fracture face containing the

membrane-intercalated particles of the hydrophobic interior

and the other is the etch face (surface). A typical con-

vex fracture face (EF) exposed in the freeze-fracturing and

etching of SMP suspended in distilled water is shown in

Figure 12. It displays typical intramembranous particles

ranging from about 5.0nm to 16.0nm in diameter. The pack-

ing density of these particles is about 1600 particles/μm

in this fracture face.

However, of the total of 38 replicas (obtained

from 15 freeze-etching experiments) examined, only one

replica showed two SMP with etch surfaces. The main reason

for getting the etch surfaces with such difficulty seems

to lie in the very small surface area of the sub-

mitochondrial particles in the suspension. On fracturing,

most of the surface area of the SMP is fractured, and it

appears that very little area is left for the subsequent

etching. And this, perhaps, renders the probability of

obtaining meaningful etching very low.

The etch surface (PS) is smooth and does not show

the presence of any particles on it (Figure 12). On the

contrary, a negatively stained preparation of the same

SMP preparation displays the typical ~10nm F_1-ATPase par-

ticles (knobs) attached to the membrane. This observation

raises some very interesting questions. Why does the

F_1-ATPase not show up as the particles on the etch surface

(PS) of the inner mitochondrial membrane in the freeze-

etch replica of the SMP preparation? Is it being lost in

the freeze-etching experiment, from the surface of SMP?

Could it be possible that the negative staining is causing

the "lollipop"-like appearances of F_1-ATPase on the SMP?

These questions are discussed on

Optical Diffraction Studies

a. Negatively stained SMP preparations.

A typical optical Fourier transform of the
electron micrograph of a negatively-stained sub-
mitochondrial particle is displayed in Figure 13b. The
corresponding electron micrograph of the negatively stained
sub-mitochondrial particle is shown in Figure 13a. Only
a part of the surface (the image) of the sub-mitochondrial
particle was subjected to optical diffraction. The remain-
der of the area was covered. This was achieved by using a
black mask with a circular hole in it, which allowed the
desired area to remain exposed to the laser beam, in the
optical diffraction experiment. This area is shown under
the circle in Figure 13a. Therefore, the optical Fourier
transform of the electron micrograph (Figure 13b) also
contains the Fraunhofer diffraction pattern of a circular
hole; which displays a central peak called the airy disk,
surrounded by successively weaker rings. In some cases
instead of using a circular mask, a rectangular mask was
used. For example, a rectangular mask was applied to the
electron micrograph of a negatively stained SMP (Figure
14a) to leave open a rectangular area and mask out the

remainder of the area. The corresponding optical Fourier trans-
form is shown in Figure 14b. The transform contains the Fraun-
hofer diffraction pattern of a rectangular opening too,
because of the rectangular mask used. The diffraction pat-
tern of a rectangular opening consists of a central spot,
the shape of which is inversely related to the shape of
the aperture; that is, more extended along the direction
of the shorter side of the rectangle and vice versa. In
the crossed pattern, there are sets of successively weaker
spots (see appendix for details).

As is evident from the optical Fourier transforms
of the electron micrographs of the negatively stained sub-
mitochondrial particles, the scattering object appears to
be a non-periodic one. The optical Fourier transforms lack,
any periodic elements which must have been represented on
a reciprocal lattice, provided there had been any regular
surface lattice or repeating subunit structure in the plane
of inner mitochondrial membrane present in SMP. It seems
that the lattice of F_1-ATPase molecules is a completely
disordered one. From the optical Fourier transforms of the
electron micrographs of negatively stained sub-mitochondrial
particles, it can be reasonably deduced that there is a lack
of any two-dimensional repeating or subunit structure within
the plane of inner mitochondrial membrane (in SMP). The
optical Fourier transforms of the SMP resemble the optical
Fourier transforms of non-periodic objects.

For the sake of comparison the optical Fourier

transform of the electron micrograph (Figure 15b) of a freeze-fractured and etched replica of catalase crystal is shown in Figure 15c. Figure 15a shows a catalase crystal negatively stained with 2% PTA (pH 7.2). For freeze-fracturing and etching the catalase crystals were suspended in water. The optical Fourier transform of the fractured face of catalase shows distinct repetitive details limited to a resolution of 3.2nm. Basic spacings are 16.1nm and 6.5nm. The optical diffraction pattern shows diffraction spots, symmetrically arranged on the reciprocal lattice. In the electron micrograph of the negatively stained catalase crystal, the lattice repeats are 19nm and 7.8nm.

b. Freeze-etch replicas.

The optical Fourier transform of the mask (Figure 16b) representing the arrangement of particles on the convex fracture face (Figure 16a) of SMP, obtained by freeze-fracturing and etching in 20% glycerol, is shown in Figure 16c. It contains information only about the particles of interest and thus the confusion likely to be caused by both the platinum grain and non-uniform appearance of structures as caused by platinum shadow in the electron micrographs of the freeze-fractured (etched) replica has been avoided. The optical Fourier transform of the mask, representing the arrangement of particles on the convex fracture face of SMP, reveals a diffuse ring

extending from about 9-12nm, with an average radius cor-

responding to 10.5nm. The spatial frequencies fall pre-

dominantly between about 9-12nm range. This demonstrates

lack of any regular two-dimensional periodic arrangement

of particles in the hydrophobic interior of the membrane.

Because if there were any regular two-dimensional lattice

of the intramembranous particles in the hydrophobic interi-

or of the membrane, the optical Fourier transform should

have displayed sharp diffraction spots of ring(s).

DISCUSSION

F$_1$-ATPase

The negatively stained preparations of SMP distinct-
ly display the presence of typical ~10.0nm F$_1$-ATPase
("coupling factor F$_1$") knobs attached by the stalks to the
inner mitochondrial membrane (of SMP). These stalks are
the correlates of the oligomycin sensitivity conferring
proteins. It is reasonable to assume that if these
~10.0nm F$_1$-ATPase knobs, as visualized by the negative
staining, are present and attached to the SMP, the freeze-
etching technique would be most suitable for demonstrating
the presence of these particles on the surface of SMP.
However, in the present study, deep-etched submitochondrial
particles were found to be devoid of any membrane-attached
particles. The etch-surface of inner mitochondrial mem-
brane in the SMP appears to be smooth. This observation
leads to a paradoxical situation. It seems that results of
freeze-etch studies are in agreement with Sjöstrand's sug-
gestion that the "knobs" or "lollipop"-like structures are
artifacts produced by the negative staining procedure in
that the ATPase molecules would "pop out" due to osmotic
shock during the specimen drying. But this argument has
two problems. First of all, it has been demonstrated that
the F$_1$-ATPase knobs are present even when the osmotic
shock is reduced to a minimum by negative staining in am-

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monium molybdate (Muscatello and Horne, 1968; Muscatello
and Carafoli, 1969). Though it is known that the negative
stain dries before the specimen by the measurement of
^3H_2O concentrations of negatively stained preparations
during drying (Johnson and Horne, 1970), the shrinkage
artifacts are still a matter of controversy. On the other
hand, the phosphotungstic acid is expected to be a "good"
stain because the large number of polar atoms on its peri-
phery induce an ordered structure on the adjacent water
molecules and leave the structure beyond it undistorted
so that the water can evaporate from the stain with little
damage to the surface of the specimen. Therefore it seems
less likely that the phosphotungstic acid would induce the
artifacts in negative staining. Recently, the stalked knobs
have been claimed to be visualized in the ultrathin sec-
tions of mitochondria and submitochondrial particles, pre-
ferably after glutaraldehyde fixation and staining with
methanolic uranyl acetate (Telford and Racker, 1973).

However, as compared with packing density of
particles in the negatively stained, unfixed preparations
of inner mitochondrial membrane which exhibit approximately
3000 particles/μm^2 of the membrane area (Stoeckenius, 1963),
packing density of particles in ultrathin sections (Tel-
ford and Racker, 1973) is much less (Zingsheim and Plattner,
1976). However when the particle packing density of face
views of negatively stained inner mitochondrial membranes
following glutaraldehyde fixation (Keyhani, 1972; Stiles
and Crane, 1966) is compared with the particle packing

density in the ultrathin sections of the mitochondria and SMP (Telford and Racker, 1973), both are found to be in agreement. Thus the negative staining and thin sectioning data appear to be reconciled. But the freeze-etch data presents the problem in its interpretation. Recently an interesting observation has been made by Beechey and his coworkers (Beechey et al., 1974). These authors have demonstrated that in the submitochondrial particles extracted with the diethyl ether, the typical inner mitochondrial membrane knobs, which are correlated with ATPases activity, fail to show up in negative staining. The vesicles (SMP) appear to be less regular. One would assume that either the inner membrane knobs have been contracted into the membrane or the PTA now cannot penetrate the membrane and reveal the knobs. However, in these diethyl-ether treated SMP, the membrane bound ATPase is still sensitive to aurovertin, an antibiotic that binds to the ATPase molecule itself (that is the inner membrane knobs). Therefore it appears that the diethyl ether extraction has little effect on the ATPase molecule itself. However, diethyl ether extraction causes a loss of the sensitivity of the ATPase activity to oligomycin, venturicidin, DCCD and triethyl tin. It causes relatively little loss of the lipid (14% of the total membrane phosphorus is extracted) and almost no extraction of protein (just 0.05%) from the membrane. It appears that the lipid loss may be responsible for contracting the F_1-ATPase within the lipoid environment

and thereby rendering it inaccessible to the negative stain.

Penefsky and Warner (1965) have proposed a structure for the mitochondrial ATPase which consists of 11 subunits each with a molecular weight of approximately 26000 and the entire complex has a molecular weight of 284,000 (estimated by the sedimentation velocity). Forrest and Edelstein (1970) have confirmed this value. However, subsequent examinations of the molecular weights have led to a revised figure of 340,000-384,000 for the enzyme isolated from rat liver, ox heart and *Saccharomyces cerevisiae* mitochondria (Lambeth *et al*., 1971; Lambeth and Lardy, 1971; Catterall and Pedersen, 1971; Senior and Brooks, 1971; Tzagoloff and Meagher, 1971). Purified preparations of ATPase molecules from these sources show, on negative staining, the subunits arranged in a hexagonal array. The maximum distance across the hexagon is approximately 10nm (Kagawa and Racker, 1966; Schatz *et al*., 1967; Kopaczyk *et al*., 1968; Tzagoloff and Meagher, 1971; Pedersen and Catterall, 1973). If the average molecular weight of the subunits is 55,000 and the subunits are spherical, then based on these assumptions it can be calculated that the diameter of the subunits is approximately 5.5nm and a hexagonal array of these subunits would give rise to a structure with the maximum dimensions of 15nm x 5nm (Beechey, 1974). However, this is not in accordance with the sizes measured in the electron micro-

scope. Therefore it seems more likely that the subunits

are not spherical. If one assumes that the subunits are

ellipsoids with a minor axis of 3.3nm, the length of the

major axis is 11.4nm (Beechey, 1974). A hexagonal array

of six of these subunits with parallel major axes gives

a structure that agrees well with most of the electron

microscope images. But this model does not account for at

least one and possibly three other subunits which are

associated with the six subunits that have been considered

here.

Beechey (1974) has presented a diagrammatic sum-

mary of the information available on the protein subunit

structure of the oligomycin-sensitive ATPase, isolated

from *Saccharomyces cerevisiae* (Figure 17). The structure

appears to be a hexagon made up of subunits 1 and 2

(diameter 5nm). The subunit 3 (diameter 4.4nm) is thought

to be connected to the cylindrical oligomycin-sensitivity-

conferring protein (5nm long, 3.0nm diameter) in the cen-

tre of the hexagon. 8a and 8b (3.0nm diameter) and 9

(2.6nm diameter) are displayed arranged around the oligo-

mycin-sensitivity conferring protein cylinder. Though the

calculated dimensions of this model (15nm x 9.4nm) agree

well with the measurements on the negatively-stained

oligomycin sensitive ATPase (15nm x 10nm), it must be

stressed that this model should be viewed with caution.

The subunits in this model are assumed to be spherical in

shape, which is probably not the case for the ATPase sub-

units (they are most probably ellipsoidal).

Beechey (1974) envisages the above-described complex of subunits to traverse the thickness of the inner mitochondrial membrane and not protrude from the membrane as observed in the negative staining. This model readily explains the absence of any particles on the etched surface (PS) of the submitochondrial particles that correspond with the stalked knobs seen in the negatively-stained preparations of SMP. This model also explains the presence of a membrane bound ATPase in the diethyl ether extracted SMP (Broughall *et al.*, 1973) which in negative staining reveal the absence of typical inner mitochondrial membrane stalked knobs. Perhaps on negative staining the phosphotungstic acid penetrates between the ATPase molecules with the attached oligomycin sensitivity conferring protein and rest of the oligomycin sensitive ATPase complex to extrude the ATPase and oligomycin sensitivity conferring protein molecules (Cunningham *et al.*, 1967; Beechey *et al.*, 1973.

However, the above given explanation has some serious problems. According to a model of the mitochondrial proton pump (the mitochondrial ATPase system) (Figure 18) described by Racker (1976), it contains a minimum of eight polypeptide chains that are requried for function. The biggest component where the energy transformation takes place is the ATPase (F_1). There are two other coupling factors, OSCP, an oligomycin sensitivity confer-

ring protein and another coupling factor, F_6, that are required for the attachment of F_1 to the membrane. F_0 is a hydrophobic protein factor (Kagawa *et al.*, 1973) containing a proteolipid. It is a membranous preparation from mitochondria and confers oligomycin sensitivity to the F_1-ATPase and is responsible for the transmembranous movement of protons. F_2 is a coupling factor that still needs further characterization (Racker *et al.*, 1970). Though the precise function of F_2 is still unknown, it seems to stimulate the energy coupling by decreasing the permeability of mitochondrial membranes to protons (Racker, 1976). The proteolipid is the most intriguing component of the ATPase system. It is known to interact with energy transfer inhibitors such as DCCD (N,N' dicyclohexyl carbodiimide). The precise molecular correlates of the stalk are also unknown. It could be OSCP or F_6 or both or neither (Racker, 1976). A reasonable alternative is also δ subunit of F_1 which is required for the attachment of the water-soluble ATPase to the membrane (Smith *et al.*, 1975). Presumably the stalk is formed by all three as shown in Figure 18. Racker's model clearly displays the extramembranous location of F_1-ATPase and is in agreement with the results obtained by negative staining of SMP in the present study. Results obtained by thin sectioning of SMP (Telford and Racker, 1973) also support the extramembranous location of F_1-ATPase. Moreover, the reactivity of F_1-ATPase in SMP to nonpermeant diazobenzene is

comparable to that of F_1-ATPase in solution (Schneider et al., 1972) quantitatively. These results would be difficult to reconcile with any model which shows the F_1-ATPase to be submerged in the membrane, and strongly favours the extramembranous location of F_1-ATPase.

There is another line of argument which also favours the extramembranous location of F_1-ATPase. It is known that the negative stains are hydrophilic and hence they should interact with the hydrophilic surfaces only. If the F_1-ATPase was submerged in the lipid milieu of the bilayer in the inner mitochondrial membrane, then it should not be "stained" at all by the PTA in the negative staining. But, since in the negatively stained preparations of SMP the typical "stalked knobs" are clearly seen projecting out from the membrane, it is only reasonable to assume that the F_1-ATPase projects out from the inner mitochondrial membrane and it is not an artifact of negative staining. However this model is unable to explain the smooth appearance of the etched surface of SMP in the freeze-etch experiments done with the SMP suspended in water. More work is clearly needed in this area to unravel the secret of this phenomenon. There may be "non-sublimable" water present on the surface of the SMP and it will render a smooth profile to the etched surface.

In the freeze-fracture of the SMP preparations in 20% glycerol, the typical membrane intercalated particles are observed on both the concave and the convex frac-

ture faces. There is an asymmetry in the distribution of these particles between the two membrane halves. The protoplasmic half has more particles (2000 ± 200 particles/μm^2) in comparison with the exoplasmic half (1300 ± 300 particles/μm^2). These particles most likely represent the components of respiratory chain (Wrigglesworth et al., 1970). A theoretical frequency distribution of the components of the respiratory chain (Klingenberg, 1968) has been calculated from a knowledge of the approximate molecular weights and molar composition of the respiratory chain (Wrigglesworth et al., 1970). Cytochromes would comprise the majority of protein components in such a distribution. Cytochromes fall in the particle size range 4-6nm with a peak at 6nm due to cytochromes a and a_3.

The distribution is given a low frequency tail by the ATPase and various dehydrogenases, between 7-13nm. Assuming a Pt deposition thickness of 1-1.5nm, a particle size distribution has been calculated for the intramembranous particles observed on the fracture faces of the SMP (Figure 11). From this distribution it is obvious that the maximum number of particles belong to 10-12nm range, and it is followed by the particles in the 5-7nm range. Therefore, the freeze-fracture particles do not correspond one-to-one with the members of the respiratory chain (the molecular correlates of the members of the respiratory chain). Some types of associations between the molecules of various members of the respiratory chain would give rise to such

a situation, and it is reasonable to assume so as it would facilitate the cooperative interactions in the respiratory chain.

It is not surprising to find that the particles on the fracture faces are not representative of the individual protein molecules. When membranes reconstituted with rhodopsin are freeze-fractured, despite the low molecular weight of the prosthetic group of rhodopsin (26,500-28,500; Bownds and Gaid-Huguenin, 1970), the fracture faces show ~20nm large particles (Hong and Hubbel, 1972). These particles are too large to represent individual rhodopsin molecules, and therefore it is quite likely that they are multimers. MN-glycoprotein, which is obtained from the erythrocyte membranes (glycophorin, molecular weight ~55,000; Steck, 1974), can be incorporated into liposomes. This protein undergoes multimerization above a critical concentration in the liposomes and shows up as ~8nm large particles in fracture faces, after freeze-fracturing (Grant and McConnell, 1974; Sergrest et al., 1974). Recently Fisher and Stoeckenius (1977) have correlated the freeze-fractured replicas of the purple membrane preparations with the electron density projection worked out by U..... and Henderson (1975). These authors have determined that each particle in the fracture face contains 9-12 bacteriorhodopsin molecules--63 to 84 transmembrane α-helices.

Optical Diffraction Studies

The results of the optical diffraction experi-

ments on the electron micrographs of negatively stained SMP clearly demonstrate the absence of any subunit structure in the plane of inner-mitochondrial membrane in SMP. Furthermore, it also suggests that the lattice of F_1-ATPase molecules is highly disordered. The data from the optical diffraction experiments on the masks obtained from the electron micrographs of the fracture faces of the SMP also demonstrate a lack of any subunit structure in the hydrophobic interior of the membrane. The intramembranous particles are not arranged in any regular two-dimensional lattice, and the average particle-to-particle distance is 10.5nm. These results negate the presence of the "much discussed" pronounced subunit structure (Green, 1974; Green, 1972; Fernández-Morán et al., 1964) in the plane of the inner mitochondrial membrane. These conclusions are in agreement with the observations of Finean and his co-workers (Thompson et al., 1968) who have studied the inner mitochondrial membrane organization using the technique of small-angle X-ray diffraction. The diffraction patterns obtained from hydrated pellets of the inner mitochondrial membrane (obtained from rat liver mitochondria) showed two or three orders of a lamellar repeat of 11.8nm at water contents of 10-30%. The lameller phase is thought to re-present the intact membrane which retains water essential to its integrity. Subsequent alterations in the diffrac-tion patterns on further drying probably represented mole-cular reorganization within the membrane. These authors

found no X-ray diffraction evidence in support of a pronounced subunit structure within the plane of the inner mitochondrial membrane.

These results, which negate the presence of the subunit or any two-dimensional periodic structure within the plane of the inner mitochondrial membrane, point against the "tripartite repeating unit" (TRU) concept of the inner mitochondrial membrane organization given by Green and his colleagues (Fernandez-Moran et al., 1964; Green and Ji, 1972; Green, 1974). First these authors had invoked the concept of the lipoprotein subunits, which was refuted later on (Stoeckenius and Engelmen, 1968). Now, Green has proposed a model (Green, 1974) in which the mitochondrial supermolecule is envisaged to contain five complexes: the four complexes of the electron transfer chain and a central tripartite repeating unit, which consists of the systems involved in ATP synthesis (the F_1-ATPase) and active transport (the membrane-forming basepiece). But on the basis of the observations made in the present study it seems that there is no repeating unit in the inner mitochondrial membrane.

The concept of the lipoprotein subunits has been abandoned now. In some membranes the presence of a subunit structure (here the word subunit structure refers to the two-dimensional periodic structure within the plane of the membrane as defined in the introduction and therefore it should not be confused with the lipoprotein sub-

units) has been demonstrated. Subunit X-ray diffraction patterns (arising due to the proteins within the plane of the membrane) have been recorded from the disc membranes obtained from the retina, sarcoplasmic reticulum membranes, hepatic gap junctions, purple membranes of the halophilic bacterium *Halobacterium halobium*, and from the quantasomes of the chloroplasts. The subunit diffraction is diffuse and of weak intensity and is therefore not easily re-corded. The first X-ray evidence for a subunit structure within the plane of the membrane was obtained from the subunit diffraction pattern recorded from fully hydrated, partially hydrated or air-dried isolated disc membrane preparations (Blasie *et al*, 1965). In the normal hydrated state the lattice of rhodopsin molecules is disordered but in the air-dried disc membranes, rhodopsin molecules are arranged in a square lattice of 7.0 x 7.0nm. In the case of the fully hydrated sarcoplasmic reticulum membranes, a subunit pattern consisting of two diffuse reflections has been recorded. In the normal hydrated state the lattice of ATPase molecules is disordered with an average center-to-center nearest-neighbour distance of about 7.3nm (Worthington and Liu, 1973). Dupont and his coworkers (Dupont *et al.*, 1974) have obtained a subunit pattern from the X-ray study on the membrane fragments from *Tor-pedo* electric tissue. In the partially dehydrated elec-troplax membranes, the cholinergic receptor molecules are arranged in a hexagonal array of 9.0nm in size. In the case

of the purple membrane, the unusually regular arrangement of the protein and lipid components was first demonstrated by Blaurock and Stoeckenius (1971). The X-ray diffraction pattern of purple membrane in water consisted of a series of sharp rings, the spacings of which corresponded to a two-dimensional hexagonal crystal with a 6.3nm unit cell. [Recently Unwin and Henderson (Unwin and Henderson, 1975; Henderson and Unwin, 1975) have achieved a notable breakthrough in the analysis of periodic arrays of biological molecules and they have determined the three-dimensional structure of purple membrane at 0.7nm resolution. The unstained purple membranes in 1% glucose were exposed to very low electron doses, low enough to avoid the destruction of their structure, to obtain the "low dose images". The data obtained from low does images was combined with the data from the electron diffraction patterns. The low dose images were processed to provide the phases (the phases were suitably corrected for the contrast transfer function by utilizing the optical Fourier transforms of the normal high-dose images), and the amplitudes of the structure factors were obtained by measuring the intensities in the electron diffraction patterns. By employing the data obtained for various tilt angles, a three-dimensional Fourier synthesis was computed according to the method given by DeRosier and Klug (1968). The map reveals that the bacteriorhodopsin protein in the membrane is globular and it extends to both sides of the membrane.

It is made up 7 α-helices packed 1.0-1.2nm apart and 3.5-4.0nm in length running perpendicularly to the plane of the membrane. The molecules are arranged around a three-fold axis with a space of 2.0nm in diameter at the centre, which is filled with lipid. The membrane displays a surprisingly great degree of order. Purple membrane is the only example of a biomembrane where we know the structure of a membrane-bound protein in situ.] The gap junctions are made up of the units hexagonally arrayed with two-dimensional crystal-like regularity in the plane of the membrane. Goodenough and Stoeckenius (1972) have obtained the subunit reflections from gap junctions corresponding to a hexagonal lattice of 8.6nm.

Very recently Caspar and his group have used X-ray crystallographic methods and electron microscope image analysis to correlate the structure and composition of gap junction plaques isolated from mouse liver (Caspar et al., 1977; Makowski et al., 1977). They have shown that there is a significant short-range disorder in junction lattice, even though the long-range order of array is remarkable. This disorder has provided more information about the nature of forces that hold the array together. In the case of gap junctions and purple membranes, the subunit structure remains unaffected by drying.

It is now known on the basis of the observations of Blasie and Worthington (1969), who had employed the radial distribution functions to analyze the subunit

structure, and that of Poo and Cone (1973), that the rho-
dopsin molecules in the normal hydrated disc membranes have
freedom of motion within the plane of membrane and behave
like a "planar liquid". The subunits in the sarcoplasmic
reticulum have also been demonstrated to behave in the same
way (Worthington, 1976).

What are the probable reasons for the absence of
a subunit structure in the plane of the inner mitochondrial
membrane? It is known in the case of the cytochrome c
that it moves between its oxidase and reductase in the lipid
milieu of the membrane (Chance, 1974). For such trans-
lational and rotational motions that the cytochrome c under-
goes, the membrane lipids must be in a sufficiently fluid
state to allow for such motions. And this more fluid lipid
environment would reduce the possibility of the organization
of membrane proteins in any regular two-dimensional lattice
in the plane of the membrane. Most of the membranes which
have distinctive subunit structure have a relatively rigid
lipid environment. For example in the case of the purple
membrane which has a regular subunit structure, the bac-
teriorhodopsin molecules exist in a very rigid lipid en-
vironment (Blaurock and Stoeckenius, 1971). However, there
is still a possibility of the existence of regions of high
lipid fluidity and relatively rigid lipid environments oc-
curring together in the inner mitochondrial membrane. The
evidence for the presence of relatively rigid environments
in the inner mitochondrial membrane came from the experi-

ments with cytochrome oxidase, which demonstrated that it is probably completely immobilized in the inner mitochondrial membrane or its rotation is confined to a single axis coinciding with the symmetry axis of heme a_3 (Junge and DeVault, 1975).

The absence of subunit or any two-dimensional periodic structure within the plane of inner mitochondrial membrane indicates that there is, perhaps, a considerable freedom of molecular motion within the plane of the membrane.

However, in relation to the implications of the present work it should be borne in mind that the results reported here are based on the studies of SMP and it is possible that their organization may not be a true represenation of the life-like structure of the inner mitochondrial membrane. This constraint is of particular signifi- cance in view of the current concept of the "dynamic" biomembrane organization and function.

PLATES AND FIGURE LEGENDS

Figure 1. A three-dimensional drawing of a triple-layered
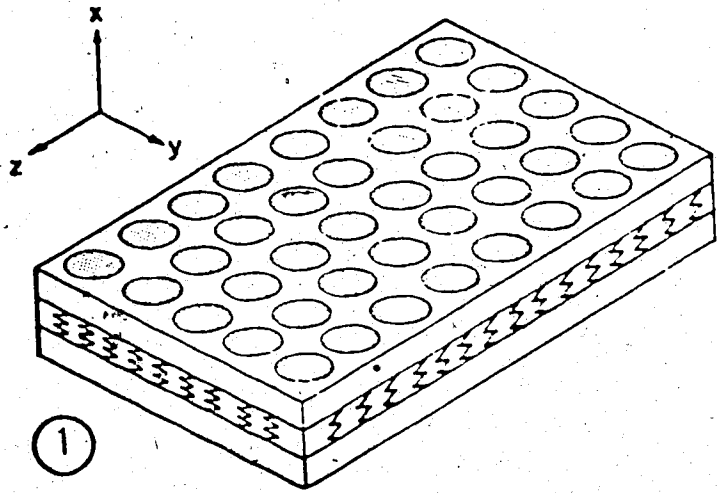   membrane with a regular subunit structure
          (Worthington, 1976).

Figure 2. Optical diffractometer lens system. Ray diagram showing diffraction and reconstitution positions (Polaron Instruments Inc.).
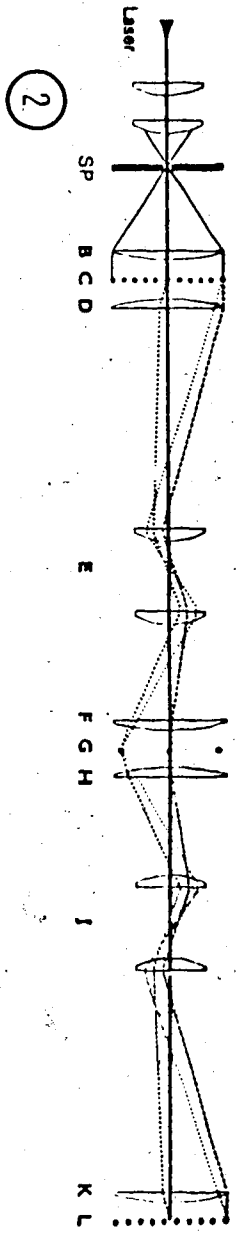
Laser SP BCD E FGH I KL

2

PLATE 1

Figure 3. A typical SMP preparation negatively stained
           with 2% PTA (pH 7.1). F_1-ATPase knobs are
           seen attached to the surface of SMP.

PLATE 2

Figure 4.   An SMP preparation negatively stained with
            2% PTA (pH 7.a) showing F_1-ATPase knobs at-
            tached to the SMP by stalks (arrows point at
            the stalks).
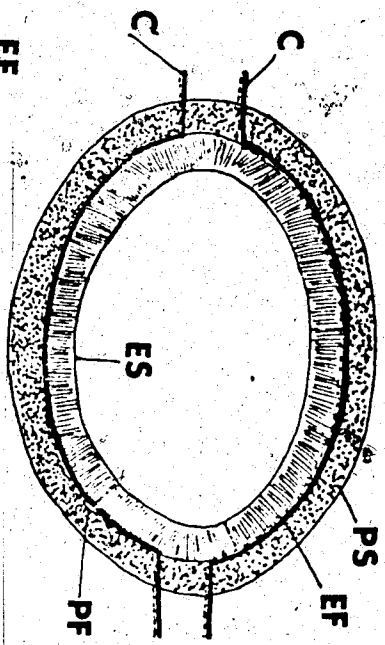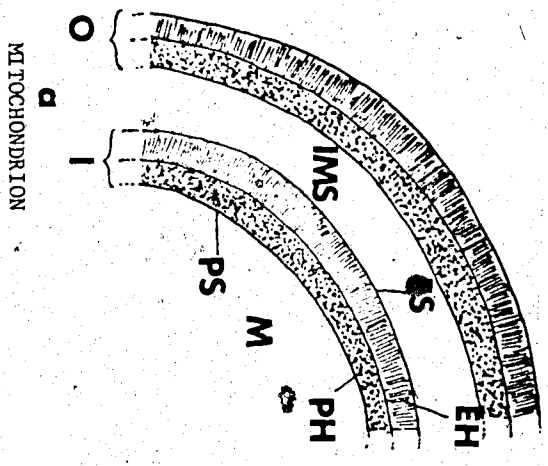
PLATE 3

Figure 5.   A negatively stained (with 2% PTA; pH 7.1)
            "tubularized" preparation of membrane fragments
   of mitochondrial cristae.

Figure 6. Freeze-etch nomenclature for SMP; membranes of mitochondria; freeze-etching of SMP in glycerol; freeze-etching of SMP in water.

C   = convex fracture plane
C' = concave fracture plane
EF = exoplasmic fracture face
EH = exoplasmic half
ES = exoplasmic surface
I   = inner mitochondrial membrane
O   = outer mitochondrial membrane
IMS = the space between the inner and outer mitochondrial membrane
M   = matrix
PH = protoplasmic half
PS = protoplasmic surface
PF = protoplasmic fracture face
SP = surface proteins
W   = water

MITOCHONDRION

O

a

I

IMS

S

PS

M

PH

EH

SMP IN WATER

c

EH

ES

PH

EF

C

SP

V

SMP IN 20% GLYCEROL

b

C

C

ES

PS

EF

PF
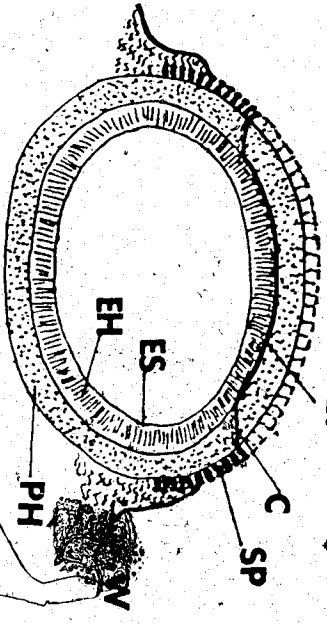
PLATE 4

Figure 7. Fracture faces revealed after freeze-etching
SMP suspended in 20% glycerol. Arrow indicates
the direction of shadowing.

EF = exoplasmic fracture face (convex)
PF = protoplasmic fracture face (concave)

PLATE 5

Figure 8.   Typical fracture faces revealed after freeze-
            etching SMP in 20% glycerol.   Arrow indicates
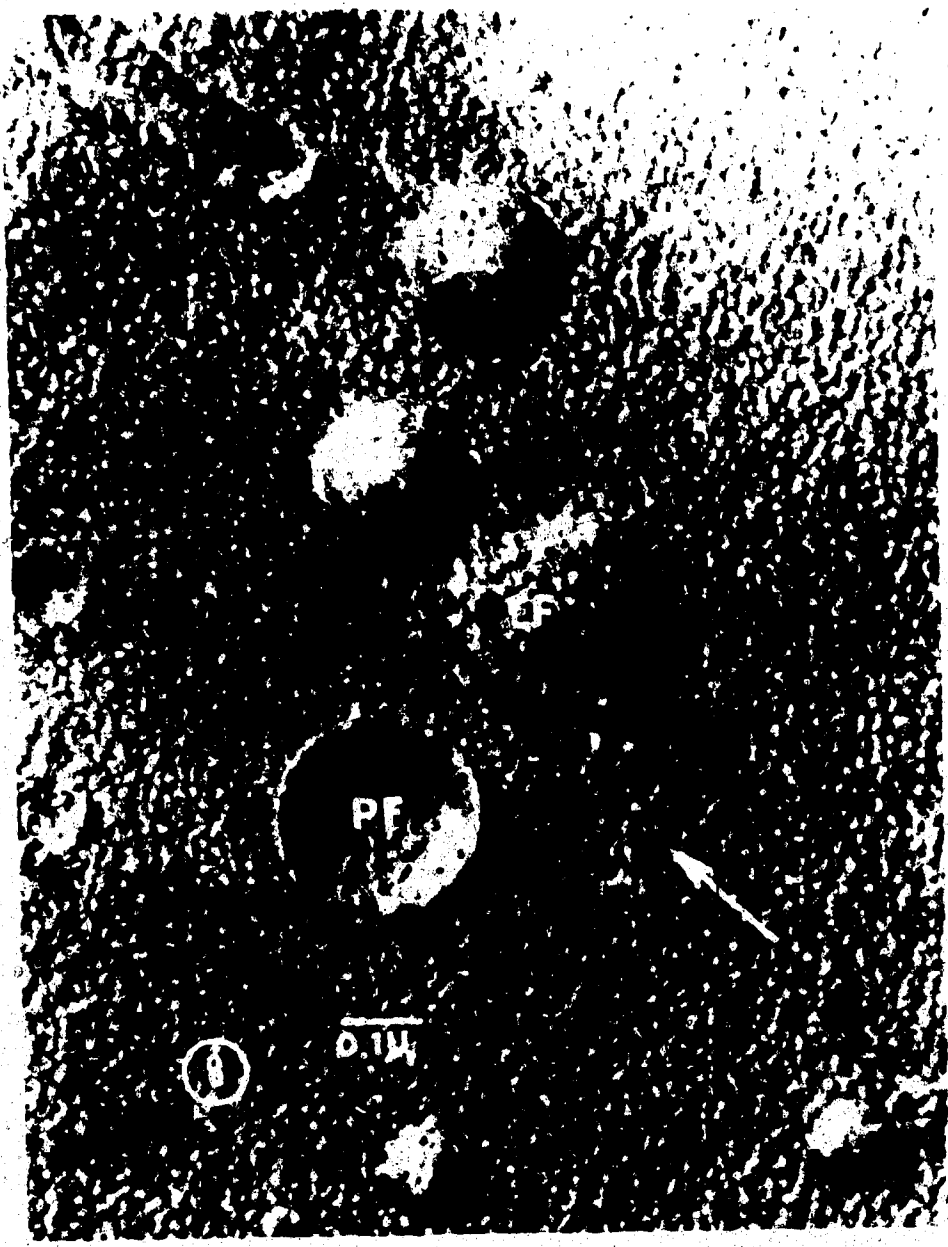   the direction of shadowing.

PLATE 6

Figure 9. Typical fracture faces obtained by freeze-etching
SMP preparation suspended in 20% glycerol.
Arrow indicates the direction of shadowing.

PLATE 7

Figure 10.   A convex fracture face (EF) revealing the
             intramembranous particles arranged in irregu-
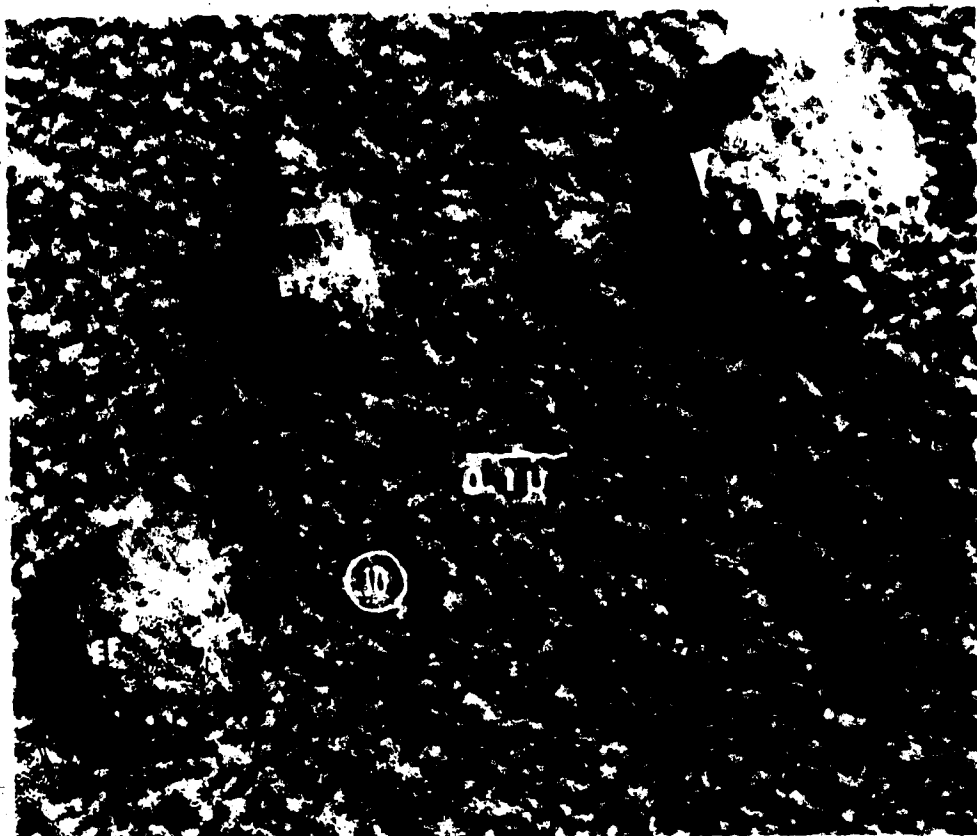   lar rows. Arrows indicate the rows.

Figure 11. Frequency distribution of particle sizes on the SMP fracture faces. Note that most of particles fall in 10-12nm diameter range. Over 200 particles were counted.
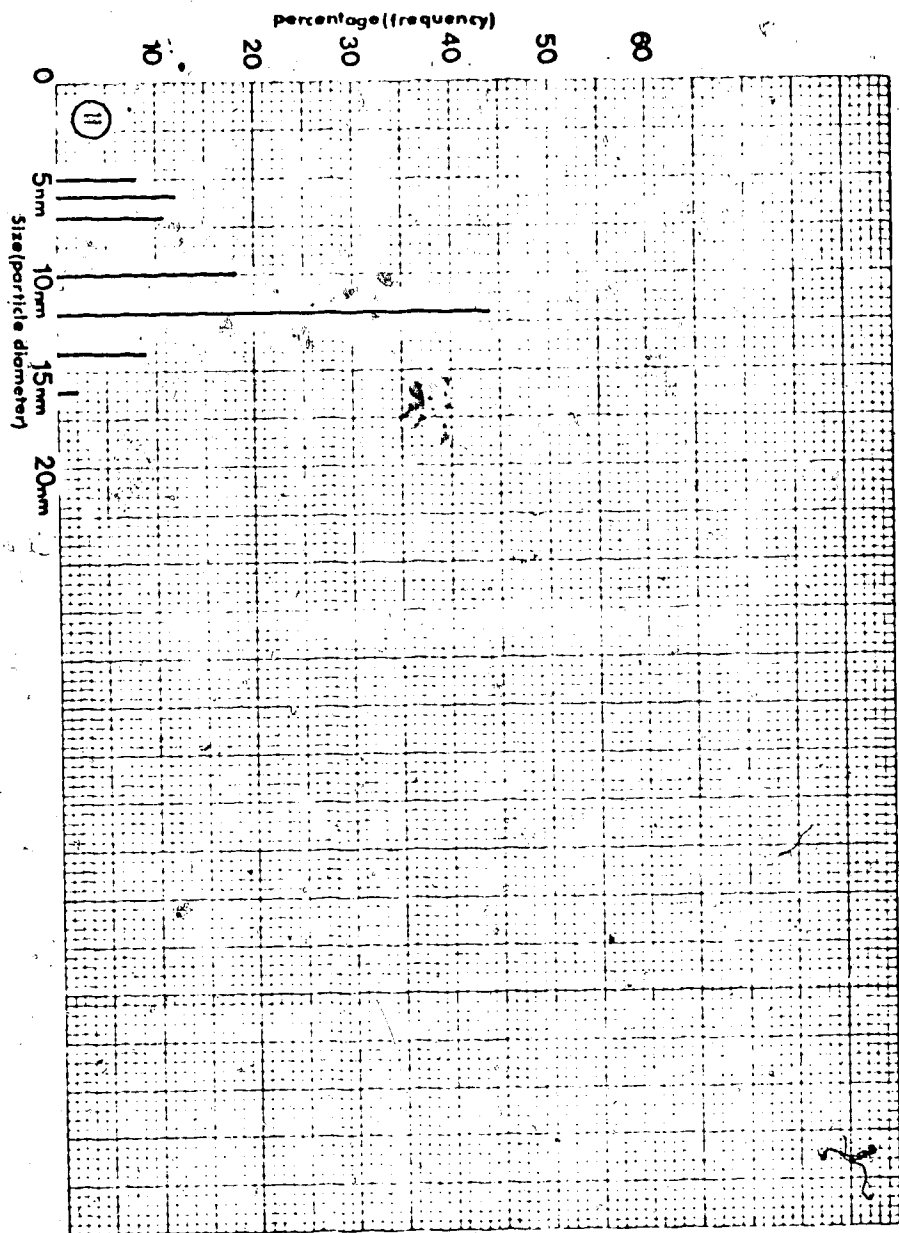
PLATE 8

Figure 12.   A typical SMP, freeze-etched in water. It
             displays a rather smooth etched surface (PF)
             and a typical convex fracture face (EF).
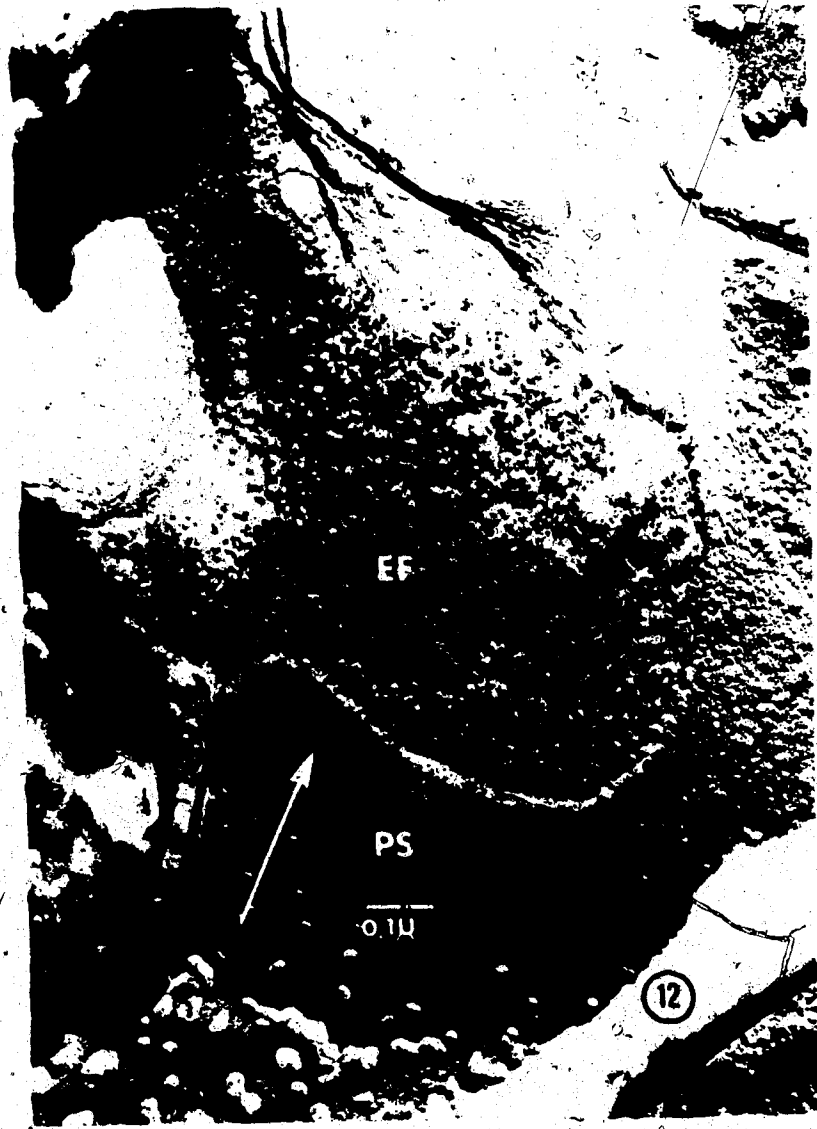   Arrow indicates the direction of shadowing.

EF

PS

0.1μ

⑫

PLATE 9

Figure 13a. A typical negatively stained SMP subjected to
optical diffraction. The area under the circle
was left open and the remainder was masked
out in the optical diffraction experiments.

13b. Optical Fourier transform of the area enclosed
within the circle in Figure 13a. It reveals an
absence of any periodic elements and therefore
it eliminates the possibility of a subunit
structure within the plane of membrane in SMP.

13b' and 13b". Optical Fourier transform of the same
area as shown in Figure 13a, obtained by using
shorter exposure times. These, too, reveal an
absence of periodic elements.

b

⑬

b   b
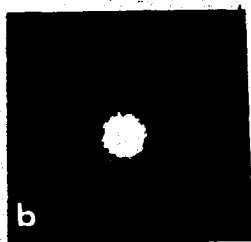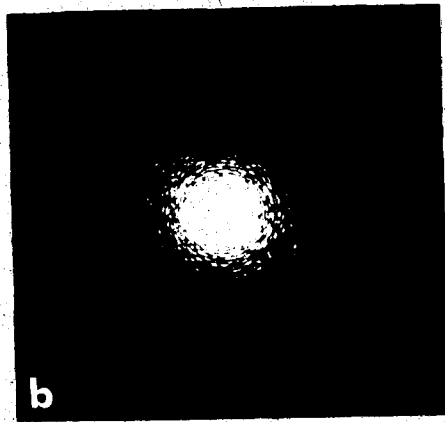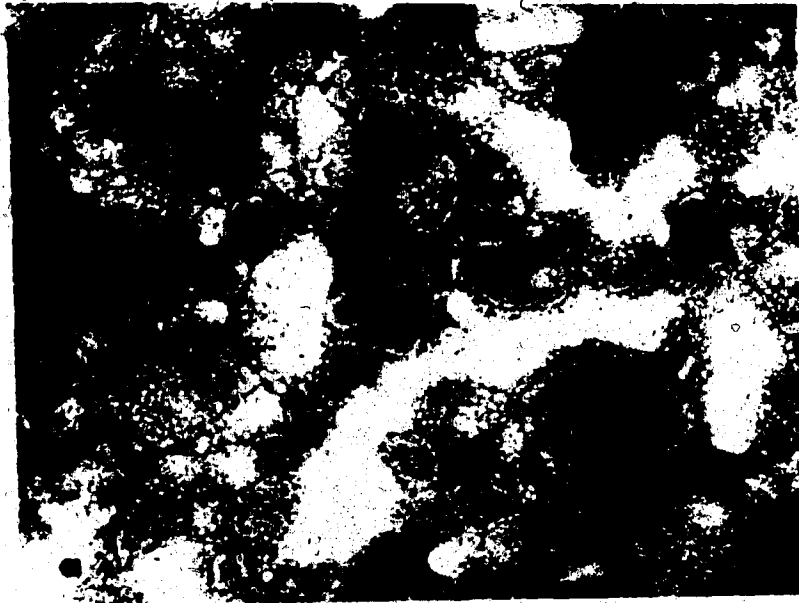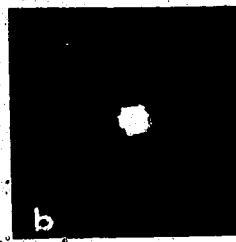
PLATE 10

Figure 14a. An SMP negatively stained with 2% PTA (pH 7.1).
The area within the rectangle was subjected to
optical diffraction.

14b. Optical Fourier transform obtained from area
enclosed within the rectangle in Figure 14a.
It reveals an abstinence of periodic elements.

b

PLATE 11

Figure 15a. A catalase crystal negatively stained with
            2% PTA, pH 7.1. The lattice repeats are 19nm
            and 7.8nm.

    15b.   A catalase crystal freeze-etched in water.
            It reveals a fracture face. The area within
            the rectangle was subjected to optical dif-
            fraction.

    15c.   Optical Fourier transform of the fracture face
            enclosed within rectangle in Figure 15b. It
            shows distinctive repetitive details to a re-
            solution of 3.2nm. Basic spacings are 16.1nm
            and 6.5nm.

b

0.1 μ

0.1 μ

c
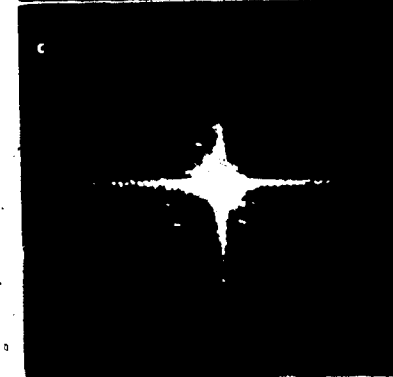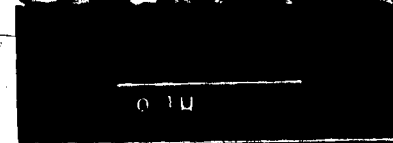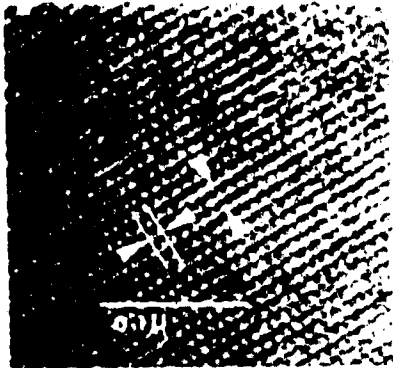
15

PLATE 12

Figure 16a.   Convex fracture face (EF) of a freeze-etched
              SMP. The fracture face displays typical
              intramembranous particles.

       16b.   The mask prepared by punching holes at parti-
              cle positions in the micrograph shown in Fi-
              gure 16a.

       16c.   The optical Fourier transform of the mask
              shown in Figure 16b. The diffraction pattern
              shows a diffuse ring extending from about 9-12nm,
              with a mean radius of 10.5nm.
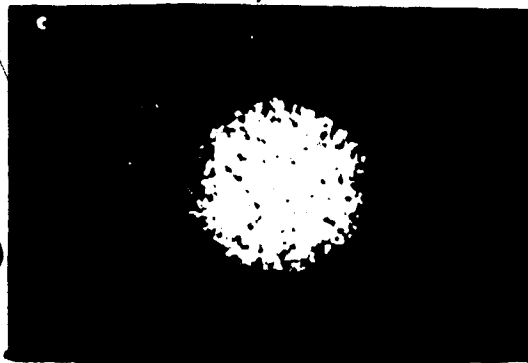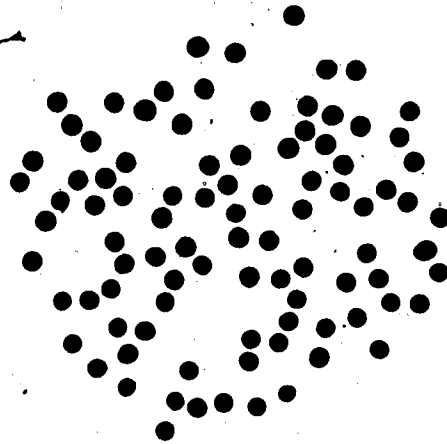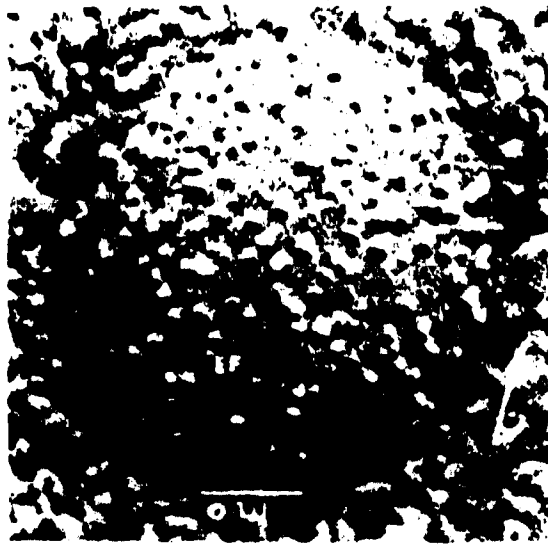
Figure 17. Diagrammatic summary of subunit structure of
            the oligomycin-sensitive ATPase isolated from
            Saccharomyces cerevisiae (Beechey, 1974).

Figure 18. Structure of the mitochondrial proton pump
            (Racker, 1976). It is intentionally incomplete
            with respect to the subunit structure of F_1-
            ATPase, because the topographical positions of
            the γ and ε subunits (which are deleted in the
            drawing) are not clearly known.
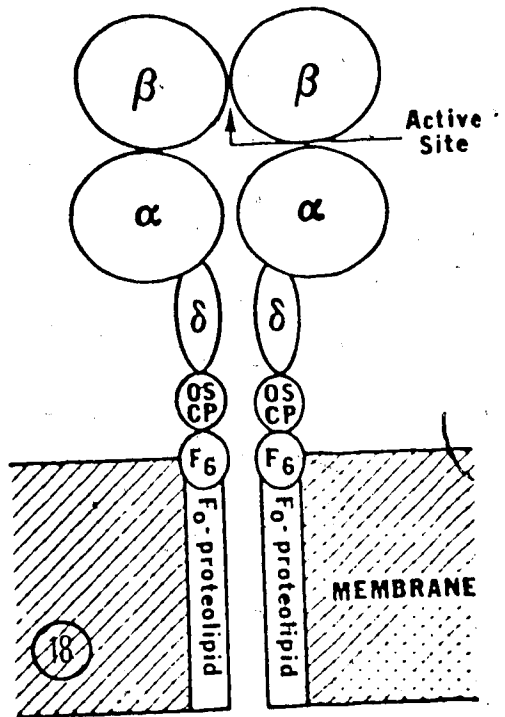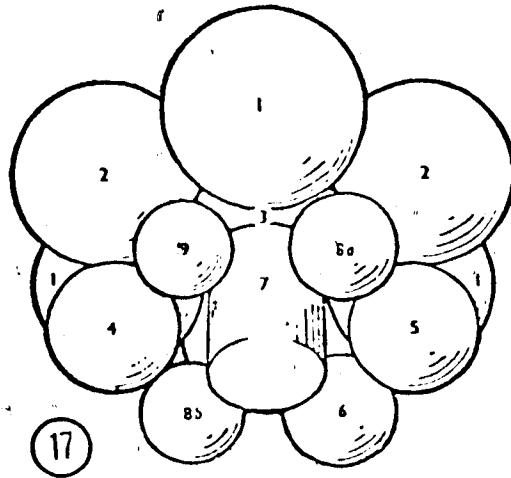
Figure 19.   Path difference for rays scattered at an angle
               α from two points A_1 and A_2 (Lipson and Lip-
               son, 1969).

Figure 20.   Image formation in a single-lense optical
               system illuminated with a coherent beam
               (Johansen, 1975).

Figure 21.   Path differences between waves resulting from
               scattering in a plane object at two object
               points O and P. The angular coordinates of
               the interference pattern at point Q are α
               and β (Johansen, 1975).

Path difference = $x \sin \theta$

OBJECT LENS IMAGE FOCAL PLANE IMAGE
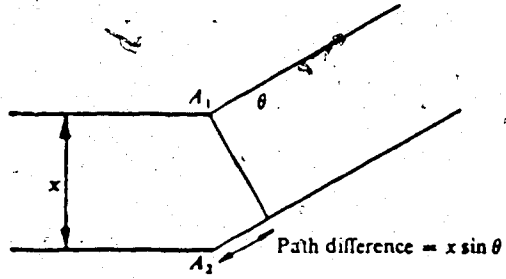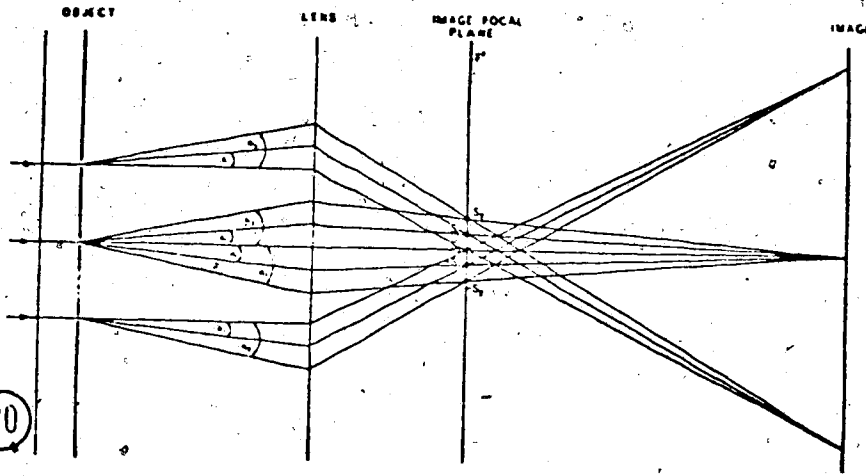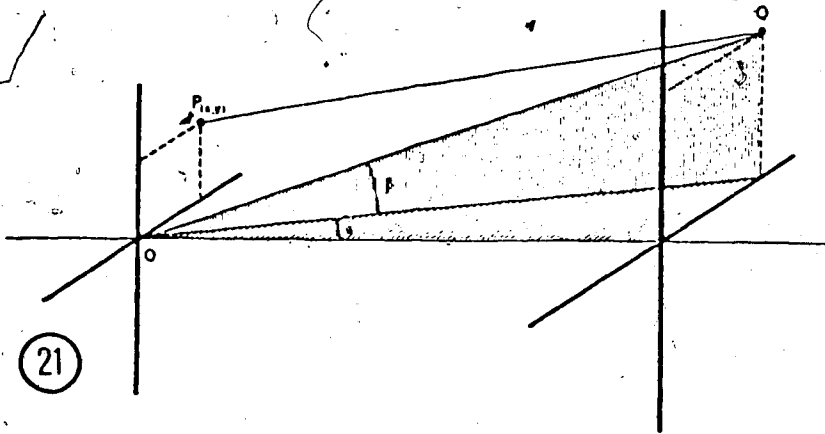
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APPENDIX

A. ABBE'S THEORY OF IMAGE FORMATION

B. CONCEPT OF OPTICAL DIFFRACTOMETER

C. ANALYSIS OF PERIODIC SPACINGS AND
   ORIENTATIONS IN THE ELECTRON
   MICROGRAPH

A. ABBE'S THEORY OF IMAGE FORMATION

According to the Fourier theory of Fraunhofer diffraction, the Fraunhofer diffraction pattern of an object is the Fourier transform of the object (Lipson and Lipson, 1969). Amplitude and the phase of the radiation at any point in the diffraction pattern are the amplitude and the phase at the corresponding point in the Fourier transform. Suppose a radiation of wavelength λ is scattered by two points A_1 and A_2 (Figure 19) placed at a distance x apart. If the radiation is incident normally to a line separating two points, the phase difference for waves diffracted at an angle α is equal to $(2\Pi x/\lambda)\sin\alpha$, which can be written as $kx\sin\alpha$ too (where $k = 2\Pi/\lambda$). If there is a collection of points A, each scattering an amplitude $f(x)$, the complete wave diffracted in the direction α is given by

$$\Psi = \int_{-\infty}^{\infty} f(x)\exp\left(-ikx\sin\alpha\right)dx \qquad \ldots (8)$$

(Lipson and Lipson, 1969). Ψ is the Fourier transform of $f(x)$ in terms of the variable $k\sin\alpha$, which is also called as u. This treatment is one dimensional.

According to Zernike (1946) there are two steps. of diffraction, in the process of image formation, in an

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optical instrument: the incident light wave is first dif-
fracted by the object and the image is formed by a second
diffraction process in which diffraction of the diffraction
pattern, produced in the back focal plane of the objective
lens, takes place.

However in case of the incoherent illumination,
waves will be inclined to the object from a variety of
angles: If only one such wave inclined at an angle β to
the object is considered, the Fourier transform can be
written:

$$\Psi(u,\beta) = \int_{-\infty}^{+\infty} f(x)\exp\left\{-ikx(\sin\alpha + \sin\beta)\right\}dx \qquad \ldots \ (9)$$

(Lipson, 1972). However, unless f(x) is an extraordinarily
simple function, the problem becomes extremely complicated
to deal with; therefore in order to explain the image form-
ation process a simple case of plane parallel illumination
will be used (Johansen, 1975). In order to complete the
requirements of forming an image, it is necessary for an
optical instrument to recombine all the scattered waves from
one point in the object to a single point in the space
because of the fact that the object scatters the plane
parallel wave. The occurrence of an image point will be
determined by whether all the waves arrive in phase. For
the sake of clarity and simplicity, the object is taken to
be a diffraction grating illuminated by a coherent plane
wave as shown in Figure 20. The incident radiation will be

scattered by the object with the maxima of successive

orders at the angles . . . $-\alpha_2$, $-\alpha_1$, 0, α_1, α_2 . . . .

These rays would be refracted by the lens so that all rays

entering a given angle, α_n, will intersect at a point F'

in the image focal plane, also known as the back focal

plane.

The diffraction pattern of the object is in the

back focal plane of the lens, each point $-s_2$, $-s_1$, s_0,

s_1, s_2 being a center of coherently reconstructed rays

scattered from the object at a given angle. In this plane,

intensity of each point would be observed. (The wave Ψ

is a complex quantity and has both amplitude and phase.

However in the diffraction pattern only intensity can be

recorded. Intensity is equal to $|\Psi|^2$ or $\Psi\Psi^*$, where Ψ^*

is the complex conjugate of Ψ.) If the rays are allowed to

go further from the back focal plane F' to the image plane

I, they will recombine to give the image of the object.

To produce an image, the relative phases between

the object points and image points must be the same, and

as is obvious the phase change between the object and the

back focal plane, F' is opposite and equal to the phase

change between F' and the image plane. It therefore im-

plies that the phase relation between the back focal plane

of the lens and the image is the inverse of that between

the image and the focal plane. Therefore, it can be con-

cluded that the image is the inverse Fourier transform of

$\Psi(u)$,

$$f''(x) = \int_{-\infty}^{+\infty} \Psi(u)\exp(iku)du \qquad \ldots (10)$$

(Lipson, 1972), indicating the inverse nature of the trans-
formation by omitting the minus sign in the exponential.

So far only the objects in one dimension have
been considered by the theory, but two-dimensional plane
objects perpendicular to the lens system can also be con-
sidered easily. According to Lipson (1972), each point in
the object is defined by a vector r, direction of the inci-
dent beam is defined by a vector s_0 and that of diffracted
beam by a vector s. If the diffracting object is repre-
sented by the function f(r), the total scattered wave would
be:

$$\Psi = \int_{-\infty}^{+\infty} f(r)\exp\frac{2\Pi i |r|}{\lambda}(\cos\alpha - \cos\beta)dA \qquad \ldots (11)$$

Here dA is the element of area around point r, and α and β
are angles of incidence and scattering respectively. By
giving the vectors s and s_0 moduli of $1/\lambda$, equation (11)
can be written as:

$$\Psi = \int_{-\infty}^{+\infty} f(r)\exp 2\Pi i (r \cdot S - r \cdot s_0)dA \qquad \ldots (12)$$

or

$$\Psi = \int_{-\infty}^{+\infty} f(r)\exp(2\Pi i r \cdot s)dA \qquad \ldots (13)$$

where $S = s - s_0$.

This general expression (Lipson and Taylor, 1958; Lipson, 1972) is valid for all angles of incidence. Equation (13) represents the Fourier transform of the object f(r). To define a complete Fourier transform, two quantities, the phase angle and intensity are needed at each image point. Since the diffraction pattern in the back focal plane of the objective lens·gives only intensity $|\Psi|^2$, it is referred to as the optical transform.

B.   CONCEPT OF OPTICAL DIFFRACTOMETER

It was assumed in the one-dimensional diffraction theory that the beam illuminating the diffracting object was a plane coherent wave, and that the lens was located close to the object. By mounting a collimating lens in front of the diffracting object, this illumination condition can be achieved to a certain approximation in the optical diffractometer. Under these conditions Fraunhofer diffraction is observed, and its theory has been treated in detail by many authors (Sears, 1949; Goodman, 1968; Lipson and Lipson, 1969). Lipson (1972) has provided a simplified view of the theory which considers a plane diffracting object (Figure 2) illuminated by a plane wave travelling perpendicular to it. It is to be determined, then, how the waves from all points (P), having the coordinates (x,y), add together in a parallel plane at a larger distance (ℓ). If it is supposed that point Q is located at such a plane, the coordinates are $\ell\alpha$ and $\ell\beta$, where α and β are the angular coordinates of the line OQ. The path differences OQ-PQ are equal to $2\ell(\alpha x + \beta y)$, where α and β are very small. The diffraction function $\Psi(\alpha,\beta)$, then, can be obtained by using equation (13). In order to determine the diffraction patterns of various objects, this equation can be used. Of special importance is the object of

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an opening where f(x) is constant, it is needed to inte-
grate the expression

$$\Psi(\alpha, \beta) = \int \exp\{-ik(\alpha x + \beta y)\}dxdy \qquad \ldots \quad (14)$$

over the area of the opening.

When the periodicities present in an electron micrograph are to be analyzed in an optical diffractometer, usually a rectangular mask is applied leaving out the area of the micrograph which is of no interest. Upon illumination this single opening would produce Fraunhofer diffraction pattern with a central spot. Shape of the central spot is inversely related to the shape of the aperture and in the crossed pattern there are sets of successively faint spots. It can also be treated purely analytically, since the two dimensions can be dealt with separately. Assuming that the two sides of the rectangle are a and b, equation (14) will become

$$\Psi(\alpha, \beta) = \int_{-a/2}^{+a/2} \exp(-ik\alpha x)dx \int_{-b/2}^{+b/2} \exp(-ik\beta y)dy \quad \ldots \quad (15)$$

After integration,

$$\Psi(\alpha, \beta) = ab\frac{Sin\frac{1}{2}k\alpha a}{\frac{1}{2}k\alpha a} \cdot \frac{Sin\frac{1}{2}k\beta b}{\frac{1}{2}k\beta b} \qquad \ldots \quad (16)$$

which represents the intensity distribution and shape of the Fraunhofer diffraction pattern (Lipson and Lipson,

1969). A circular mask can also be used instead of rec-
tangular opening to leave out the parts of the micrograph
that are of no interest. Goodman (1968) and Lipson and
Lipson (1969) have presented the theory, explaining the
Fraunhofer diffraction pattern of a circular opening, ex-
pressing it in terms of Bessel functions. The diffraction
pattern of a circular hole consists of a central peak,
called the Airy disk, surrounded by the successive weaker
rings.

C. ANALYSIS OF PERIODIC SPACINGS AND ORIENTATIONS IN THE ELECTRON MICROGRAPH

The periodic spacings and orientations in the electron micrograph are determined by the analysis of the optical diffraction pattern in a way that is precisely similar to the determination of interplanar spacings and orientations in the specimen by the analysis of the electron diffraction pattern (Beeston, 1973; Horne and Markham, 1973). The distance on the optical diffraction pattern between any particular first order diffraction spot and th central spot is inversely related to the periodic spacings between the repeat features of the micrograph, which has been subjected to optical diffraction. The periodic spacing, d, in the electron micrograph and the distance between the associated first order spot in the optical diffraction pattern and the central spot, R, are related by the following equation:

$$Rd = \lambda L \qquad \qquad \ldots (17$$

where λ is the wavelength of the laser light (632.8nm for the helium neon laser) and L is the diffraction camera length. Camera length of the optical diffractometer can be calibrated by a number of suitable test objects (Horn

and Markham, 1973,) for example, negatively stained micro-
crystals of catalase, platinum pthalocyanine crystals, etc.