

1 High-Frequency Dynamic Nuclear
2 Polarization NMR for Solids: Part 1 - An
3 Introduction

4

5 Michelle Ha and Vladimir K. Michaelis*

6

7 Department of Chemistry, University of Alberta, Edmonton, Alberta, Canada T6G 2G2

8

9

10

11

12 *Correspondence: vladimir.michaelis@ualberta.ca

13

14

15

16

17

18

19 **Abstract**

20 Dynamic nuclear polarization (DNP) NMR spectroscopy, a high-polarization method, is rapidly
21 changing the landscape of what is possible in solid-state nuclear magnetic resonance (NMR)
22 spectroscopy. To date, there have been over 200 publications discussing high-frequency DNP
23 NMR of solids with more than half being released within the past few years. Below we provide
24 for researchers that may be interested in this high-sensitivity technique an introduction to high-
25 frequency DNP NMR spectroscopy, including instrumentation, mechanisms, polarizing agents
26 and sample preparation. While there are many applications utilizing high-frequency DNP NMR,
27 Part II will deal with recent advances in method development and applications to biomolecular
28 solids and materials science.

29 **Introduction**

30 Solid-state nuclear magnetic resonance (NMR) spectroscopy is a mature field and arguably
31 one of the most robust analytical techniques for characterizing atomic- and molecular-level
32 structure in solids. It can be found in nearly every scientific discipline such as biomolecular,
33 chemical, materials and earth science due to the unique ability to probe sub-nanometer short- and
34 medium-range structure of ordered and disordered solids. A particular strength of NMR
35 spectroscopy is its ability to elucidate various isotropic and anisotropic interactions that are rich
36 in atomic- or molecular-level structural and dynamic information. For example, dipolar coupling
37 (a through space interaction) is readily used to address medium-range order within solids.¹⁻⁵ The
38 isotropic chemical shift is vital in identifying functional groups within organic molecules,
39 polymorphs in pharmaceutical compounds, or coordination environments and bonding
40 arrangements important in materials science and geoscience.^{6, 7} The quadrupolar interaction,
41 affecting over 70% of the NMR-active nuclei on the periodic table, is highly sensitive to the

42 overall molecular and atomic environment. The magnitude and shape of the interaction has aided
43 in studying many chemical systems.^{8,9}

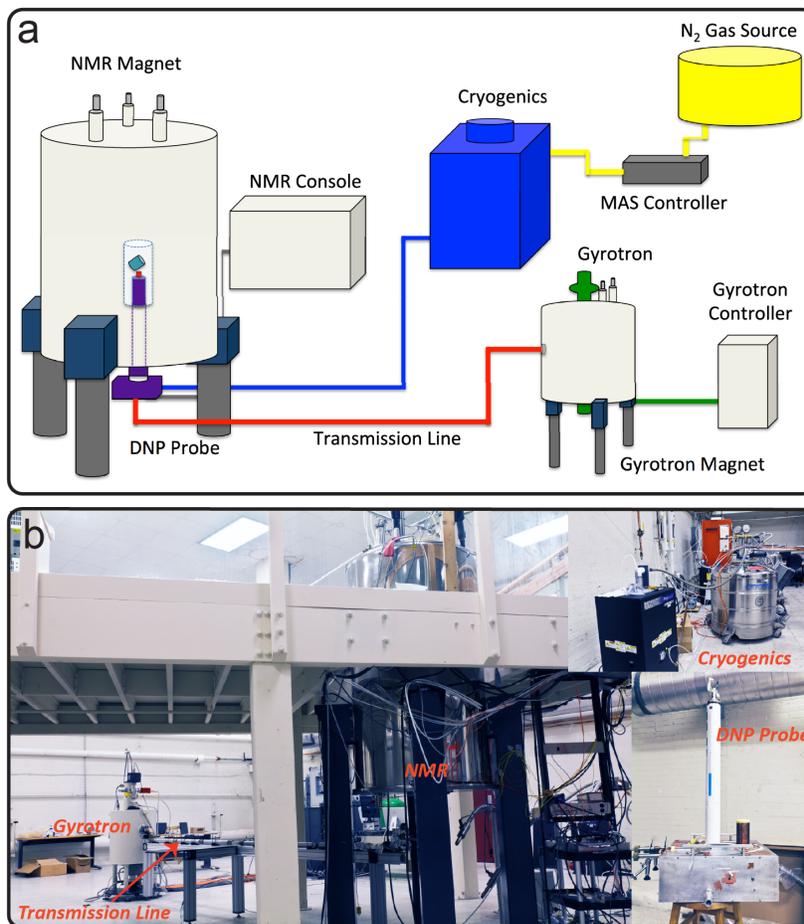
44 The large versatility of NMR spectroscopy can be overshadowed by the small nuclear
45 Zeeman polarization resulting in poor overall sensitivity. Solids are particularly disadvantaged as
46 many suffer from broad NMR resonances ranging between 10's of kHz (dipolar coupling,
47 magnetic shielding) to MHz (quadrupolar coupling) and a range in relaxation properties (*e.g.*, T_1
48 and T_2). Over the past 50 years, several innovations have advanced the field, providing practical
49 gains in sensitivity. The first breakthrough was made by Andrew and Lowe^{10,11} who introduced
50 magic-angle spinning; another important advance was the introduction of cross-polarization¹²
51 (CP). In the early 1990's progress in high-field NMR magnets surged, with the introduction of
52 ultrahigh-field magnets from a host of commercial and home-built systems; today commercial
53 systems exist as high as 1 GHz, with plans for 1.2 and 1.3 GHz in the very near future.^{13,14} At
54 each stage these developments have left a lasting impact in the field of NMR spectroscopy.

55 With the recent commercialization of dynamic nuclear polarization (DNP), a high-
56 polarization technique in NMR spectroscopy, the field is undergoing a rapid change due to the
57 technique's unprecedented sensitivity.^{15,16} DNP NMR involves transferring the large electron
58 polarization of unpaired electron spins to nearby insensitive nuclei via microwave irradiation of
59 the electron-nuclear transitions, resulting in sensitivity gains in varying orders of magnitude (*i.e.*,
60 10 to 10³) over traditional NMR spectroscopy (*e.g.*, up to 658-fold (γ_e/γ_n) increase in the case of
61 ¹H). This makes otherwise impossibly long experiments practical to complete. For example, an
62 enhancement factor (ϵ) of 20 reduces the required experimental time by a factor of 400 (20²),
63 reducing experimental times from months to hours and opening new frontiers for scientific
64 exploration.

65 To gain a better understanding of the various components necessary for high-frequency
66 DNP NMR, this article contains two parts, below (Part I) will provide an overview of the
67 instrumentation, mechanisms, polarizing agents, and sample preparation that are commonly used
68 within the field; Part II highlights the new areas DNP NMR is making available to the research
69 community including DNP method development, biomolecular solids and materials science.
70 Many excellent reviews have emerged over the years and the following articles are recommended
71 for interested readers.¹⁷⁻²⁸

72 **High-Frequency DNP NMR Instrumentation**

73 Robust and reliable instrumentation is essential to performing experimental high-frequency DNP
74 research and is still a key barrier in implementing DNP NMR within international research
75 groups. A DNP NMR spectrometer (Figure 1) is comprised of four major components: (i) a solid-
76 state NMR spectrometer, (ii) a microwave device and accessories, (iii) cryogenics and (iv) a DNP
77 NMR probe.



78
 79 **Figure 1: (a) Schematic of a high-frequency DNP NMR Spectrometer and (b) photo of a 699 MHz / 460 GHz**
 80 **DNP NMR Spectrometer (Francis Bitter Magnetic Laboratory, Massachusetts Institute of Technology,**
 81 **Cambridge, MA, USA) including (i) solid-state NMR magnet, (ii) gyrotron and transmission line, (iii)**
 82 **cryogenics and (iv) DNP NMR probe.**

83 *i. Solid-state NMR Spectrometer*

84 A conventional NMR spectrometer (*i.e.*, superconducting magnet, spectrometer,
 85 pneumatic control system, user interface, *etc.*) is required to perform NMR experiments. For
 86 DNP NMR, the magnet bore must be 89 mm (*i.e.*, wide-bore) rather than a standard bore of 51
 87 mm to accommodate the probe electronics, dewar, microwave waveguide, sample eject and
 88 insulation requirements for performing low-temperature experiments. Another consideration is
 89 whether the main magnetic field is coupled to an outer coil that can be adjusted to varying
 90 degrees (*i.e.*, a sweep coil) to increase or decrease the field strength in fine steps. This is

91 particularly important for radical development and for other targeted applications. The sweep coil
92 enables one to optimize the strength of the main magnetic field for advanced applications.

93 *ii. Microwave Devices*

94 One of the most challenging pieces of instrumentation to develop is a source that
95 generates high-frequency microwaves. With high-frequency DNP NMR now ranging from 5 to
96 19 T (200 to 800 MHz), microwave devices must generate microwaves in the 140 to 527 GHz
97 frequency range. The most commonly applied device is known as a gyrotron. This is in part due
98 to the significant development in gyrotron technology for the application in DNP NMR carried
99 out by Temkin and Griffin at MIT,²⁹⁻³³ by Osaka University and the University of Fukui³⁴⁻³⁶ and
100 commercially by Bruker Biospin, Communications and Power Industries (CPI) and by Bridge12.

101 The basis of a gyrotron is its use of stimulated cyclotron radiation generated when present
102 in a superconducting magnet. A gyrotron is capable of generating high-power microwaves at high
103 frequency (> 100 GHz). The fast-wave device is often operated in a higher mode which improves
104 the robustness and cooling abilities providing a microwave device that is extremely stable, with
105 high output power and continuous operation for days to months. The design of these devices is
106 also attractive for their longevity with lifetimes expected to be 10+ years. These features make
107 them ideal for NMR studies where experiments often require days to weeks of continuous
108 acquisition. As discussed above, the NMR spectrometer may be equipped with a sweep coil to
109 allow fine adjustments of the field strength. If this is not possible, an alternative is having a
110 tunable microwave device such as a tunable gyrotron, as one can fluctuate the output of the
111 microwave frequency while keeping the main magnetic field of the NMR instrument constant.^{33,}
112 ³⁷⁻⁴⁰ However, an issue to be aware of is maintaining a stable and constant microwave output over
113 a wide tuning range.

114 Alternative microwave sources, optimal for applications below 5 T, including extended
115 interaction Klystrons (EIK), oscillators (EIO) and amplifiers (EIA), have also been successfully
116 applied to higher field strengths; for example the Tycko group has an operational system at 9.4 T
117 (400 MHz, ^1H and 264 GHz, e^-).⁴¹ Unfortunately, these alternatives suffer from limited output
118 power at high frequencies, ~ 5 W at 265 GHz, which affects enhancements and strains the
119 devices, limiting their longevity (lifetimes of $\sim 10,000$ hrs, ~ 1.2 yrs at 265 GHz). Furthermore,
120 commercial units are unavailable for applications above 265 GHz. As advancements and demand
121 of microwave technology continue, these sources may be attractive for certain DNP applications
122 as they become available at higher magnetic fields and/or microwave output powers.

123 *iii. Cryogenics*

124 Conventional DNP NMR relies on the ability to cool the sample to cryogenic
125 temperatures to improve the electron and nuclear relaxation behaviour, aiding the effective
126 transfer of bulk polarization within the sample. This can be accomplished by using a cryogenic
127 heat exchanger whereby a sealed can within a larger container of liquid nitrogen is pressurized to
128 provide a stream of gas at cryogenic temperatures.⁴² By adjusting the pressure within the heat
129 exchanger one can control the output temperature. Finer control can easily be achieved using a
130 cryogenic temperature control (*e.g.*, Lakeshore unit) that can be equipped with a heater to
131 regulate the output gas to within 1 K or better. Similar types of devices have been successfully
132 implemented using liquid He although the increasing cost of He is a prominent concern.
133 Breakthroughs in DNP have occurred with recirculated He; in particular, the groups of De
134 Paëpe⁴³ and Matsuki³⁵ have successfully designed a closed-loop system for their DNP NMR
135 instrument that can operate at He temperatures. Tycko *et al.*^{44, 45} and Levitt *et al.*⁴⁶ have also
136 recently contributed to the He-cooled MAS NMR area. The major expense of DNP beyond the

137 initial investment in infrastructure is its thirst for cryogenics (both N₂ and He). Hence, the authors
138 believe a worthwhile goal of this field as it matures should be to strive to reduce the operating
139 costs associated with the cryogenics. For example, cryogenic pre-chillers and nitrogen gas
140 generators have been successfully used in Osaka University³⁶ and MIT.⁴⁷

141 *iv. DNP NMR Probe*

142 The NMR probe is responsible for sending and detecting the signal during an NMR
143 experiment, and if necessary, cooling the sample, rotating the sample and providing the conduit
144 for microwave irradiation prompting the transfer of polarization between electron and nuclear
145 spins. A cryogenic DNP NMR probe is based on conventional home-built or commercial MAS
146 NMR probes, but a few key modifications are required. The lines that provide a stream of
147 compressed gas (*i.e.*, drive and bearing) to levitate and rotate the rotor must be vacuum jacketed.
148 The insulation enables the transfer of chilled gas for the drive and bearing to cool the sample and
149 probe. Typical spinning frequencies are between 4 and 20 kHz, but newly designed commercial
150 probes are available capable of spinning samples up to 40 kHz (depending on rotor size &
151 experimental temperature). In the foreseeable future, frequencies >40 kHz will surely be attained
152 as room temperature probes are capable of spinning frequencies beyond 100 kHz.⁴⁸ The stator
153 housing requires a modification near the coil to accommodate the microwave transmission line,
154 responsible for guiding and projecting the microwave beam onto the rotor. The probe is often
155 encased within a vacuum jacketed dewar to aid in localized cooling of the probe, and more
156 importantly to protect the bore of the NMR magnet (T = ~290 K) from the cryogenic
157 temperatures (< 120 K) located within the probe. Samples can be placed into zirconia or sapphire
158 NMR rotors, the latter appear to provide larger enhancements, although thin-wall zirconia is
159 proving quite successful commercially and is slightly more robust. The drive caps can be Torlon[®]

160 or Vespel[®] that are glued using cryo-epoxy or machined zirconia in combination with a small
161 polymer plug.

162 It should be noted that several variations from this general design model do exist. A few
163 noteworthy differences include: (i) an added variable temperature line (3rd vacuum jacketed line)
164 that can accommodate liquid N₂ or He to assist in cooling;^{44, 47} (ii) the microwave waveguide
165 may be directed at the top of the stator or pointed to the head of the NMR rotor;³⁶ (iii) sample
166 ejection^{47, 49, 50} for ease of changing samples (above or below models exist); and (iv) the gyrotron
167 may be placed above the probe using the same superconducting magnet to produce microwaves
168 and record the NMR experiment. As the probe body is cooled this does affect the overall
169 behaviour of the electronics which can have positive benefits in generating high RF fields (*i.e.*,
170 $\gamma B_1/2\pi$) but it can also affect the tuning circuit. In regards to the RF design, the most effective
171 approach has been a topic of discussion for many years; issues include whether the probe design
172 should be transmission line *vs* locally-tuned and weighing the benefits of implementing a
173 balanced RF circuit design.^{44, 45, 51-58}

174 As the field has rapidly advanced with the introduction of commercial units in 2010,
175 systems have been successfully implemented in several research groups worldwide; a few are
176 summarized in Table 1.

177

178 **Table 1: Examples of homebuilt and commercial DNP NMR instruments**

Type	Location/ Manufacturer	B ₀ (MHz / GHz)	Microwave Source	Completed
Home-Built	MIT	211 / 140 ^{30, 59}	Gyrotron	<i>ca.</i> 1991-1993
		380 / 250 ^{29, 38, 60}	Gyrotron	<i>ca.</i> 2002-2003
		700 / 460 ^{23, 33, 47}	Gyrotron	<i>ca.</i> 2011-2013
		500 / 330 ^{32, 39}	Gyrotron	<i>ca.</i> 2014-2016
		800 / 527 ³¹	Gyrotron	<i>in-prep.</i>
Home-Built	NIH	400 / 263 ⁴¹	Diode	<i>ca.</i> 2009-2010
Home-Built	Osaka	600 / 395 ^{36, 40}	Gyrotron	<i>ca.</i> 2010-2012
		700 / 460 ^{34, 35}	Gyrotron	<i>ca.</i> 2015-2016
Home-Built	Warwick	284 / 187 - 600 / 395 ⁶¹	Gyrotron	<i>ca.</i> 2012
Home-Built	Washington U. St. Louis	300 / 198 ⁶²	Gyrotron	<i>in-prep.</i>
Commercial	Bruker Biospin ^{48, 50}	400 / 263	Gyrotron	<i>ca.</i> 2009-2010
		600 / 395	Gyrotron	<i>ca.</i> 2011-2012
		800 / 527	Gyrotron	<i>ca.</i> 2012-2013

179

180 **DNP Mechanisms**

181 For solids using a continuous microwave source there are four DNP mechanisms that can
182 be considered in order to achieve bulk polarization transfer between an unpaired electron source
183 (polarizing agent) and a nucleus: (i) thermal mixing (TM), (ii) Overhauser (OE), (iii) solid effect
184 (SE), and (iv) cross effect (CE). The latter mechanism is by far the most targeted area in high-
185 frequency DNP NMR applications due to the wide array of wide-line nitroxide radicals that
186 favour the polarization of high-gamma nuclei (*i.e.*, ¹H). SE has been effective at lower field
187 strengths⁶³ although a range of developments and results have recently appeared in the
188 literature⁶⁴⁻⁶⁶ so that it is still an active field of study in high-frequency DNP development.
189 Likewise, Overhauser effect has recently emerged as a contender for higher field DNP NMR.^{67, 68}
190 Further developments with radicals could be fruitful as the enhancements scale linearly with

191 magnetic field strength. Below is a brief overview of the CE and SE DNP mechanisms; the
192 following references are provided for a more comprehensive review.^{15, 16, 21, 24, 27, 65, 67-83}
193 The dominant DNP mechanism depends on the targeted NMR-active nucleus and on the EPR
194 characteristics of the selected polarizing agent. For example, the most common approach in DNP
195 NMR applications of solids is through indirect polarization transfer using a CP step ($e^- \rightarrow {}^1\text{H} \rightarrow$
196 X , where X is a lower gamma nucleus). Using this approach one can select for SE by using a
197 narrow-line radical such as Trityl or for CE with TOTAPOL, a wide-line biradical. In other
198 words, it is the relative magnitudes of the electron homogeneous (δ) and inhomogeneous (Δ)
199 linewidths, and the nuclear Larmor frequency (ω_{0I}) that guide the DNP mechanism.

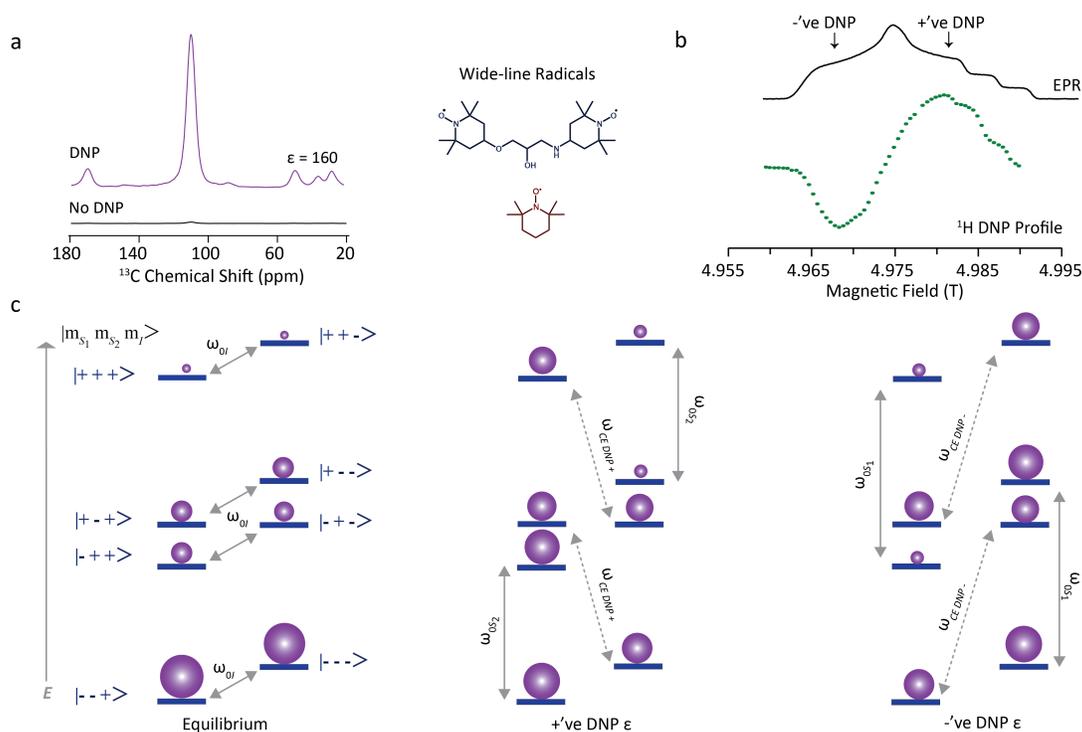
200 *i. Cross Effect DNP mechanism*

201 The CE mechanism can be described as a three-spin flip-flop-flip process between two
202 electrons and a nucleus, which is dominant when $\Delta > \omega_{0I} > \delta$. The difference between the two
203 electron Larmor frequencies should be near the nuclear Larmor frequency to achieve maximum
204 polarization transfer between electron and nuclear spins.^{73, 75, 79, 84}

$$\omega_{0I} = \omega_{0S_2} - \omega_{0S_1} \quad (1)$$

205 To satisfy equation 1 for high-gamma nuclei like ${}^1\text{H}$, the polarizing agent needs to have EPR
206 characteristics (EPR spectrum) of a broadline; this is readily seen in nitroxide-based radicals
207 including monoradicals such as TEMPO and TEMPONE, and biradicals such as SPIROPOL,
208 AMUPOL and TEKPOL. The need to have two electrons in close proximity (*i.e.*, dipolar
209 coupled) while minimizing the paramagnetic bleaching of nuclear spins has pushed the field into
210 biradicals.^{85, 86} Tethered radicals provide the chemical design to reasonably direct orientation and
211 electron-electron distance so that the dipolar coupling is on the order of 20 to 35 MHz, while
212 enabling the concentration of unpaired electrons to be minimized, typically < 15 mM solution

213 (*i.e.*, < 30 mM electrons). In contrast to biradicals, monoradicals with a 40 mM electron
 214 concentration have significantly reduced dipolar couplings of < 2 MHz when present in a
 215 homogenous glassy sample.⁸⁵⁻⁸⁸ The CE mechanism is often the choice for high-frequency DNP
 216 NMR experiments as the mechanism is based on allowable transitions (Figure 2) and loosely
 217 scales with the inverse of magnetic field strength. In the past few years, descriptions of the CE
 218 mechanism taking into account the level crossing that occurs under magic-angle spinning has
 219 shed further light into the spinning rate dependence of the overall enhancement. The reader is
 220 referred to the works by the Tycko⁸⁹ and Vega⁸¹ groups where they discuss modulations of the
 221 energy levels within DNP mechanisms when using magic-angle spinning and the impact it has on
 222 polarization transfer.



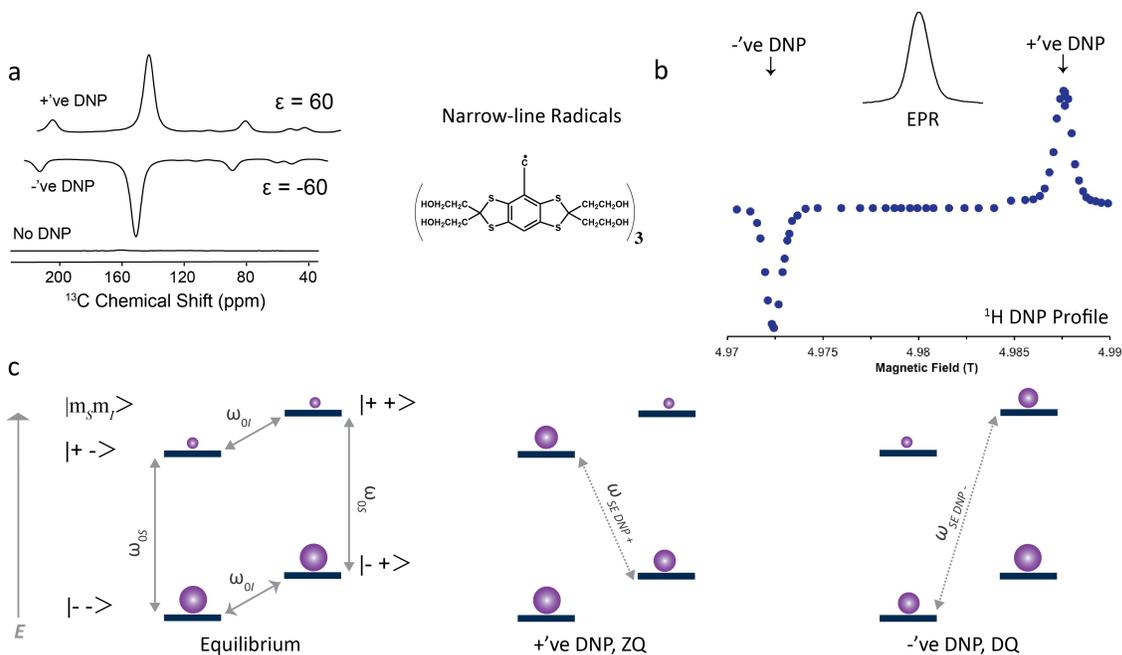
223
 224 **Figure 2: Cross effect DNP: (a) on (DNP) and off (No DNP) ¹³C[¹H] CP MAS NMR experiment on urea using**
 225 **a wide-line nitroxide radical, (b) EPR and ¹H DNP NMR field profile for a nitroxide polarizing agent and (c)**
 226 **energy level diagram for CE DNP displaying spin population distribution for a three-spin (two electrons and**
 227 **one nucleus) system at thermal equilibrium, positive and negative CE DNP conditions. Microwave saturation**
 228 **of the electron transition (ω_{0S1} or ω_{0S2}) leads to a three-spin flip-flop process that distributes the**
 229 **population (ω_{CE}), thus increasing the net nuclear polarization.**

230 *ii. Solid Effect DNP mechanism*

231 The SE mechanism can be described as a two-spin process involving an electron and a
 232 nucleus. The SE mechanism is dominant when the nuclear Larmor frequency is larger than the
 233 electron homogeneous and inhomogeneous EPR linewidths ($\omega_{0I} > \delta, \Delta$) and microwave
 234 irradiation is applied at the electron-nuclear zero- or double-quantum transition as shown in
 235 Figure 3.^{65, 66, 69, 90} The SE matching condition is satisfied when:

$$\omega_{mw} = \omega_{0S} \pm \omega_{0I} \quad (2)$$

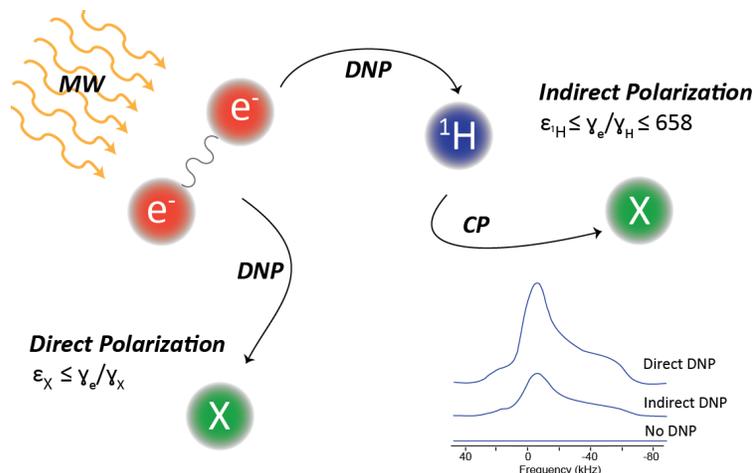
236 where ω_{0S} and ω_{mw} are the electron Larmor and microwave frequencies, respectively. The
 237 mechanism may be observed when a narrow-line radical (e.g., BDPA, trityl, etc.) is used as the
 238 polarizing agent (narrow EPR spectrum) and has an electron spin-lattice relaxation time (T_{1S}) that
 239 is optimized to allow for efficient polarization transfer to nearby NMR-active nuclei.



240
 241 **Figure 3: Solid effect DNP: (a) on (DNP) and off (No DNP) experiment showing the positive and negative DNP**
 242 **enhancement (¹³C[¹H] CP MAS NMR experiment on urea using a narrow-line radical), (b) EPR and ¹H DNP**
 243 **NMR field profile for a narrow-line polarizing agent and (c) energy level diagram for SE displaying spin-**
 244 **population distribution for a two-spin (one electron and one nucleus) system at thermal equilibrium, positive**
 245 **($\omega_{0S} - \omega_{0I}$, ZQ) and negative, ($\omega_{0S} + \omega_{0I}$, DQ) DNP enhancements.**

246 **Polarizing Agents**

247 Dynamic nuclear polarization requires a source of unpaired electron spins, which is
248 typically achieved using exogenous organic-based radicals also known as polarizing agents.
249 Radicals are typically divided into two categories as either narrow-line or wide-line radicals
250 based on their EPR characteristics, since it is the EPR characteristics of the polarizing agent that
251 give insight into what type of DNP mechanism governs the $e^- - n^0$ polarization transfer. For
252 several compelling reasons, including its reduced spin-lattice relaxation, improved sensitivity
253 possible through CP, and a large database of nitroxide radicals, ^1H is often the nucleus of choice
254 for initial polarization at cryogenic temperatures using a wide-line biradical polarizing agent.
255 Subsequent to polarization of ^1H , a CP step is used to observe other low-gamma nuclei. This
256 indirect polarization transfer method ($e^- \rightarrow ^1\text{H} \rightarrow \text{X}$, Figure 4) has been successfully applied to a
257 wide range of solids including biomolecular, materials and surfaces.^{18-20, 91} An alternative to
258 indirect polarization is polarizing an NMR active nucleus (X) directly from a source of unpaired
259 electrons, $e^- \rightarrow \text{X}$ (*i.e.*, direct polarization, Figure 4).^{23, 87, 88, 92-95} This approach is of interest for
260 many chemical systems that do not cross-polarize efficiently by high- γ nuclei (*e.g.*, ^1H or ^{19}F), or
261 those where the high- γ nuclei are absent, and the approach may be of assistance in spectral
262 editing to distinguish between protonated and non-protonated chemical environments.



263

264 **Figure 4: Schematic of indirect (through ^1H 's) and direct e^- - n^0 polarization transfer pathways in DNP NMR of**
 265 **solids. Indirect polarization (most common) enhances ^1H ($\epsilon_{1\text{H}}$), with a theoretical gain in sensitivity of 658,**
 266 **which can then be transferred to some NMR-active nuclei of interest such as ^{13}C , ^{15}N , ^{17}O , etc. Direct**
 267 **polarization (less common) enhances NMR active low-gamma nuclei directly (not through a cross polarization**
 268 **step from ^1H or ^{19}F). The theoretical gain depends on the NMR active nucleus, such as 2,618 for ^{13}C ($\epsilon_{13\text{C}}$),**
 269 **3,311 for ^{29}Si ($\epsilon_{29\text{Si}}$), 4,855 for ^{17}O ($\epsilon_{17\text{O}}$), 6,493 for ^{15}N ($\epsilon_{15\text{N}}$), etc.**

270 The choices can be vast as many organic radicals exist, particularly with the recent
 271 explosive developments in high-frequency DNP NMR and the push for improved biradicals.
 272 Below, we highlight a few of the key polarizing agents that are often used for various
 273 applications, organized by their characteristic EPR spectra of narrow-line (i) and wide-line (ii)
 274 radicals.

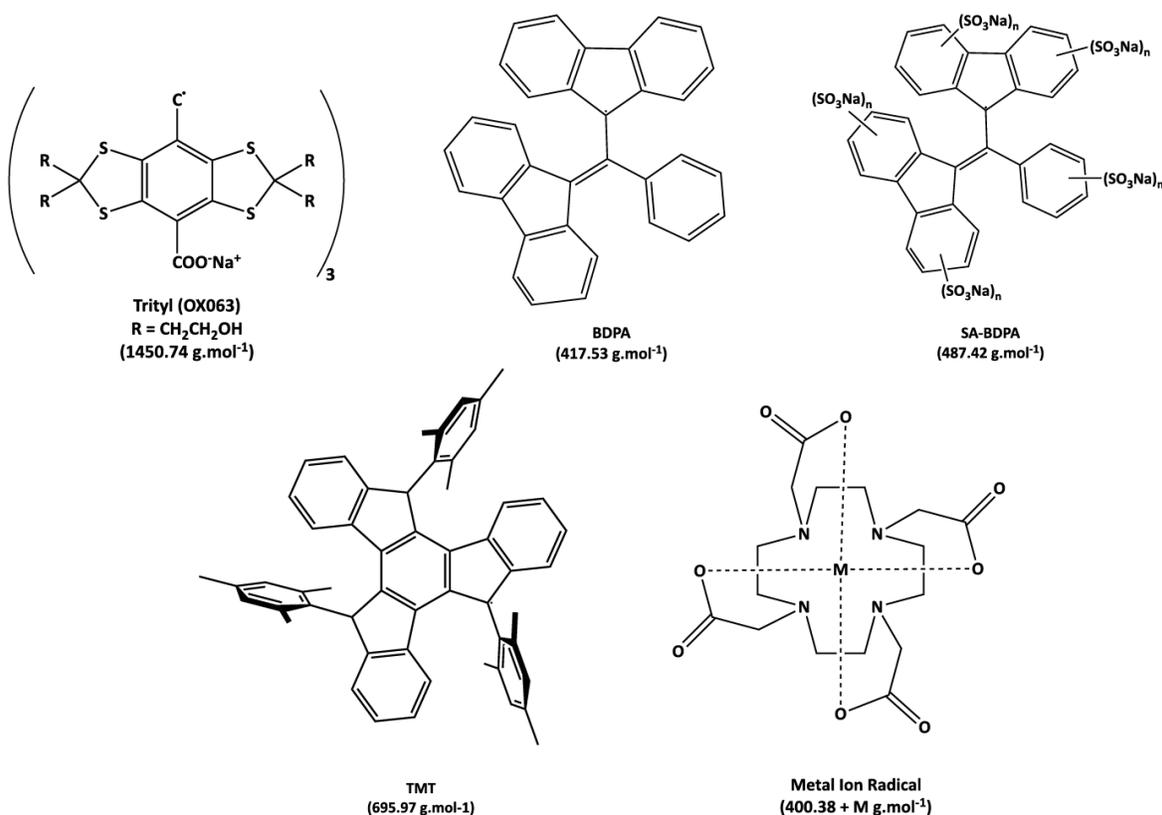
275 *i. Narrow-line polarizing agents*

276 Within the DNP NMR field only a handful of narrow-line radicals are currently viable.
 277 Although limited in options, these chemically designed radicals provide an interesting array of
 278 tunability allowing the selection of different DNP mechanisms and permitting an exploration into
 279 various direct and indirect polarization methods. Generally, these narrow-line radicals have been
 280 used within applied fields (*vide infra*), and can display the CE or SE DNP mechanism depending
 281 on the NMR-active nucleus being probed. In the case of ^1H , these radicals exhibit solid-effect
 282 characteristics which scale unfavourably with field strength dependence ($\epsilon \propto B_0^{-2}$).⁹⁶ This may

283 change as microwave technology emerges offering higher-output power able to circumvent the
284 losses from increased B_0 .

285 Figure 5 illustrates a series of narrow-line radicals that have been successfully applied to
286 various chemical problems. They are promising candidates for low-gamma polarization including
287 ^{13}C ,^{88, 94, 97} ^2H ,⁹³ ^{17}O ,^{23, 87} and ^{29}Si ,⁹⁸ as these begin to satisfy the CE DNP mechanism of low-
288 gamma nuclei via direct polarization. This may be attractive for materials that do not contain
289 high-gamma nuclei or solids that suffer from extremely long ^1H T_1 's. Recently, research has
290 begun to emerge adopting the Overhauser DNP mechanism for ^1H at high magnetic fields using
291 narrow-line radicals.^{67, 68, 99} As the Overhauser effect is the only DNP mechanism that improves
292 with magnetic field strength and requires little microwave power to saturate the electrons,⁹⁹ it
293 may offer significant advantages including permitting the use of low-power microwave sources
294 (*e.g.*, EIK). As the field turns to higher and higher magnetic fields (>800 MHz), alternative
295 radicals will surely be developed.

296



297

298 **Figure 5: Common narrow-line radicals used as polarizing agents in high-frequency DNP NMR.**

299 *ii. Wide-line polarizing agents*

300 The most prominent radicals used in DNP NMR applications are wide-line polarizing

301 agents, typically comprised of nitroxide moieties. These are chosen due to the range of offerings

302 within the literature and/or commercial sources, synthetic tunability and ease in which they

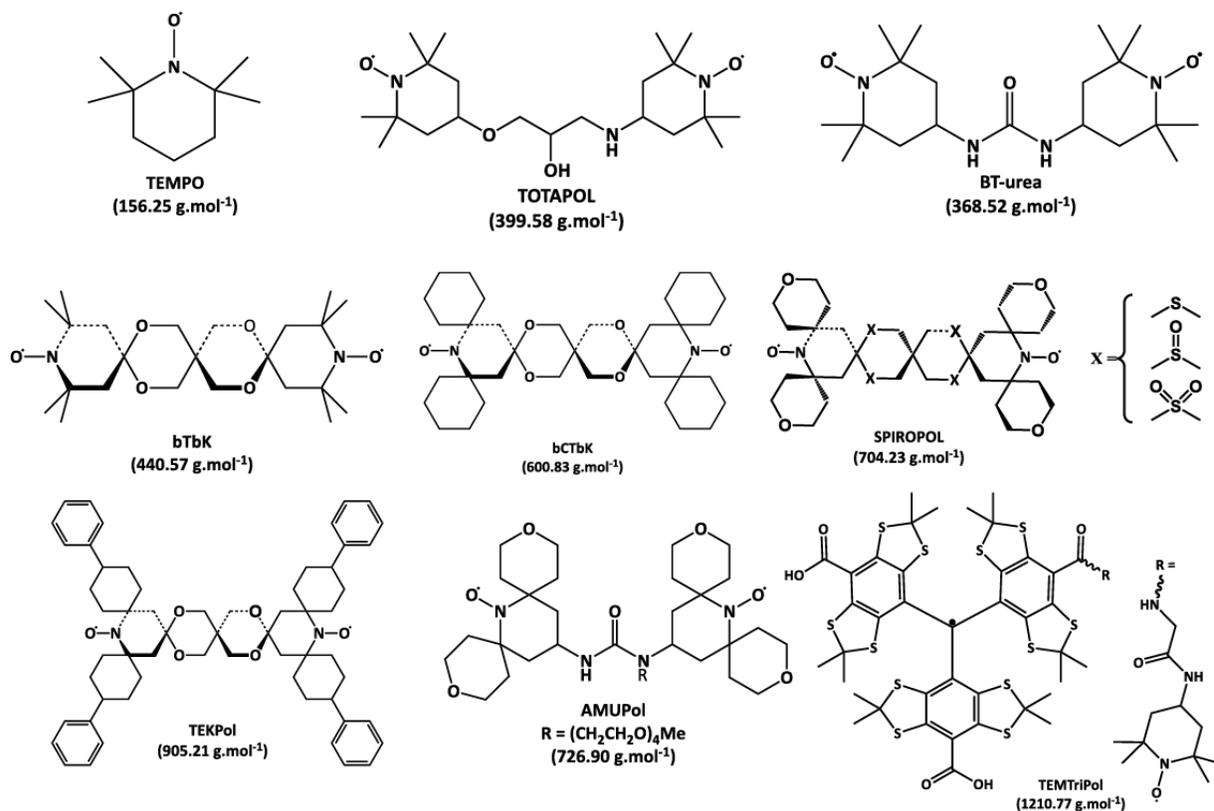
303 satisfy the CE DNP mechanism for ^1H . In most applications of DNP NMR, polarization transfer

304 occurs indirectly through high-polarized protons and a CP step to lower gamma nuclei such as

305 ^{13}C , ^{15}N or ^{29}Si . As outlined above, targeting ^1H is advantageous; in addition, in some cases this

306 approach allows one to tune the ^1H spin-lattice relaxation and ^1H - ^1H spin-diffusion behavior

307 through ^2H exchange (*i.e.*, adjusting the ^1H to ^2H ratios within the solvent and/or solid of interest).



308

309 **Figure 6: Common wide-line radicals used as polarizing agents in high-frequency DNP NMR.**

310 Early work focused on monoradicals, typically involving TEMPO-based derivatives
 311 which are easily sourced. To reach maximum CE DNP enhancements, quantities of 40+ mM
 312 electrons were required as at this level favourable $e^- - e^-$ dipole interactions on the order of a few
 313 MHz were obtained, but this caused significant paramagnetic relaxation, resulting in a loss of
 314 signal as well as reduction in T_2 , affecting resolution. In 2006, Song and Hu, graduate students
 315 within the Swager and Griffin groups (MIT), respectively, synthesized a highly effective and
 316 water soluble biradical known today as TOTAPOL.⁸⁶ This began the movement to utilize
 317 biradicals which permit a reduction of the concentration of the polarizing source (5 to 15 mM
 318 biradical or 10 to 30 mM electrons) and consequently reduce signal quenching¹⁰⁰ while yielding
 319 larger enhancements as the two electrons required within the three-spin CE DNP mechanism are
 320 highly coupled due to the chemical tether. Current research suggests that a dipolar coupling of

321 between 25 and 35 MHz, the relative orientation between the two organic radical moieties and
322 the exchange interaction appear to be important factors.^{96, 101-103}

323 For years two biradicals, bTbK¹⁰⁴ and TOTAPOL⁸⁶ (Figure 6) were the benchmarks for
324 high-frequency DNP, but with the introduction of commercial instrumentation, advances in both
325 organic and water soluble radicals has exploded since 2013. The nature of biradicals have varied
326 using a host of species; some of the most efficient radicals that have emerged include AMUPOL
327 (hydrophilic biradical)¹⁰⁵ and TEKPOL derivatives (hydrophobic biradical),^{103, 106} among many
328 others.^{86, 102, 104-111}

329 Emerging areas in radical development include a report highlighting the benefit of using a
330 mixed biradical that tethers a tempo and trityl moiety. The report demonstrates the excellent high-
331 field enhancements, surpassing conventional biradical nitroxides as well as reducing the signal
332 quenching of the sample, adding another dimension worth exploring in this exciting field.¹⁰¹
333 There have also been advances in the use of paramagnetic metals as DNP polarizing agents
334 including homogeneous solids, biomolecular and inorganic chemical systems.^{64, 112-115} There is no
335 doubt that radical development will continue to advance along with high-frequency DNP.

336 **Sample Preparation**

337 The vast majority of high-frequency DNP NMR experiments applied to chemical systems
338 utilize an *ex situ* polarizing agent with a cryoprotectant. A DNP sample will consist of a
339 polarizing agent, typically a biradical in 10 ± 5 mM concentration, a solvent mixture and the
340 sample of interest. The cryoprotectant (*i.e.*, solvent mixture) is important for several reasons
341 including allowing the homogeneous dispersion of the polarizing agent within the sample,
342 assisting in providing uniform polarization across the sample and protecting the sample from
343 cryogenic temperatures (particularly important in biological specimens). When choosing a

344 solvent mixture one should consider whether the solvent is more likely to have an amorphous-
345 like consistency or to crystalize, as this has drastic effects on the enhancement efficiency as well
346 as on radical homogeneity.¹⁰⁹ It is also important to ensure the solvent does not react with the
347 sample of interest. Typically, a glycerol-water (60:40, v/v) mixture is the most effective solvent,
348 forming an excellent glass at cryogenic temperatures. Glycerol has been used for decades as a
349 cryoprotecting medium in a host of research projects requiring low-temperature experiments.
350 Other solvent mixtures that appear in the DNP literature include DMSO/water (60:40, v/v),
351 dichloroethane/methanol (95:5, v/v), o-terphenyl, *etc.*^{23, 50, 109, 116-120} Typically, hydrophilic
352 solvents are preferred for biomolecular solids while hydrophobic solvents are preferred for
353 inorganic solids.

354 One further step in choosing the appropriate solvent mixture is the ability to exchange
355 isotopes. In general, a ¹H concentration of approximately 10% ¹H within a solvent has been
356 found to be an effective ¹H spin-bath reservoir. Thus, for example, the ¹H concentration in the
357 glycerol/water mixture (colloquially referred to as *DNP Juice*) is diluted to approximately 10 %
358 using a combination of glycerol-*d*₈, D₂O and H₂O (60:30:10, v/v/v).

359 A homogenous sample is one where the radical and chemical sample is readily dissolved
360 in the solvent-matrix and upon quenching a homogeneous glass is formed. This approach often
361 leads to the most effective polarization transfer as the radical is effectively distributed throughout
362 the sample. DNP NMR of a homogeneous, amorphous chemical system can be limited in
363 resolution due to line-broadening stemming from a distribution of chemical shifts, a commonly
364 observed occurrence for many organic and inorganic amorphous materials, as well as from
365 slower side-chain dynamics at cryogenic temperatures. A heterogeneous sample is one where the
366 solvent matrix (radical and solvent) are in contact with the chemical sample, although the sample
367 of interest does not readily dissolve in the matrix. Large enhancements are achievable for these

368 systems although they are typically reduced due to inhomogeneous radical distribution, solvent
369 selection, glassing ability, domain size, pore size, radical-sample interaction and the limitations in
370 effective $e^- - ^1\text{H}$ and $^1\text{H} - ^1\text{H}$ spin-diffusion across the solid.^{23, 66, 83, 121-126} Dynamics and hydrogen
371 concentrations within the sample can also affect the recorded enhancement. An added benefit of
372 heterogeneous DNP within nano- and microcrystalline solids is that resolution is typically
373 unhampered at cryogenic temperatures enabling both a savings in acquisition time as well as
374 high-resolution spectra that provide detailed structural information. It has also been our
375 experience that using low radical concentrations (< 10 mM biradical) tends to diminish the
376 quenching effect on spin-spin relaxation (T_2), providing higher resolution spectra, although the
377 enhancement is reduced (a trade-off between sensitivity gain and spectral resolution). DNP NMR
378 spectroscopy has been successfully applied to a diverse range of homogeneous and
379 heterogeneous biomolecular^{18, 29, 123, 127-140} and inorganic^{18, 20, 23, 121, 123, 125, 126, 141, 142} solids.

380 Other sample preparations have also proven effective for high-frequency DNP NMR
381 including ones that are cryoprotectant-free. An approach that does not need a solvent, or instead
382 one that disperses the radical with a solvent that is subsequently removed. For example, a radical
383 that is introduced onto or into a chemical system such as cellulose or a porous material then
384 followed by evaporation has recently shown promise for natural-abundance systems.¹⁴³⁻¹⁴⁵ A self-
385 cryoprotecting solvent-free approach using *in situ* or *ex situ* sedimented (SED) DNP within an
386 apoferritin complex (480 kDa) and BSA has recently been described.^{23, 146, 147}

387 Although these methods lead to a more heterogeneous distribution of radicals and hence
388 polarization is not uniform within the samples, they maintain excellent sensitivity and produce
389 excellent spectral resolution from an overall smaller effect from paramagnetic broadening. More
390 recently a series of *in situ* methods have appeared using either paramagnetic metals within
391 inorganic solids¹¹⁵ or tagging organic radicals¹⁴⁸ or metals^{112, 114} onto biomolecular solids. These

392 unique approaches to chemically engineering the polarizing agent within the system are
393 interesting avenues that could improve homogenous radical dispersion within the host material
394 such as crystalline solids (for example, alleviate domain size issues in solids) or incorporating
395 radicals as synthetic tags with proteins (for example, incorporating the radical in the lipid within
396 membrane proteins, or using alternative labelling procedures to tag the protein itself).

397 **Summary and Future Outlook**

398 High-frequency DNP NMR will continue to revolutionize our ability to explore the most
399 challenging spectroscopic questions. The balance between radical concentration and resolution is
400 a critical concept that must be carefully evaluated while sample preparation is critical to one's
401 ability to obtain the highest quality spectra. Due to the access of biradicals, glass forming
402 solvents and ease in implementing CE DNP NMR, this area will surely lead in future DNP NMR
403 applications and development for solids. To circumvent some of the enhancement issues at high
404 fields and/or access to high power microwave sources, mixed and narrow-line radical
405 development will surely continue, with the latter turning the attention to Overhauser DNP NMR.

406 As for the next technological challenges we face, DNP NMR growth will continue with
407 development beyond 800 MHz / 527 GHz on the horizon, faster magic-angle spinning (*i.e.*, > 40
408 kHz), reducing the cryogen-usage footprint and method development (*e.g.*, electron decoupling,
409 pulsed vs. CW DNP NMR, frequency tunable microwave devices, *etc.*) to improve the sensitivity
410 gains and resolution at higher magnetic fields and/or temperatures. One can foresee the day
411 where a DNP NMR spectrometer will be as routine in research facilities as liquids and solid-state
412 NMR spectrometers are today.

413 Acknowledgements

414 VKM acknowledges the Natural Sciences and Engineering Research Council of Canada
415 (NSERC) Discovery Grants program and the University of Alberta for funding. MH is partially
416 supported by the Government of Alberta Queen Elizabeth II Graduate Scholarship.

417 References

- 418 1. T. Gullion and J. Schaefer, *J Magn Reson*, 1989, **81**, 196.
- 419 2. J. C. Chan and H. Eckert, *J Magn Reson*, 2000, **147**, 170-178.
- 420 3. E. Daviso, M. T. Eddy, L. B. Andreas, R. G. Griffin and J. Herzfeld, *Journal of Biomolecular*
421 *NMR*, 2013, **55**, 257-265.
- 422 4. M. Weingarth and M. Baldus, in *Advances in Biological Solid-state NMR: Proteins and*
423 *Membrane-Active Peptides*, RSC, 2014, ch. 1, pp. 1-17.
- 424 5. H. Eckert, S. Elbers, J. D. Epping, M. Janssen, M. Kalwei, W. Strojek and U. Voight, in *Topics in*
425 *Current Chemistry*, Springer - Verlag Berlin Heidelberg, 2004, vol. 246, pp. 195-233.
- 426 6. K. J. D. MacKenzie and M. E. Smith, *Multinuclear Solid-state NMR of Inorganic Materials*,
427 Pergamon, 2002.
- 428 7. E. Pretsch, P. Bühlmann and M. Badertscher, *Structure Determination of Organic Compounds*,
429 Springer-Verlag Berlin Heidelberg, 2009.
- 430 8. R. E. Wasylshen, S. E. Askbrook and S. Wimperis, *NMR of Quadrupolar Nuclei in Solid*
431 *Materials*, Wiley, 2012.
- 432 9. A. P. M. Kentgens, *Geoderma*, 1997, **80**, 271-306.
- 433 10. E. R. Andrew, A. Bradbury and R. G. Eades, *Nature*, 1958, **182**, 1659.
- 434 11. I. J. Lowe, *Phys Rev Lett*, 1959, **2**, 285-287.
- 435 12. A. Pines, M. G. Gibby and J. S. Waugh, *Chemical Physics Letters*, 1972, **15**, 373.
- 436 13. J. Bascunan, S. Hahn, D. K. Park and Y. Iwasa, *IEEE Transactions on Applied Superconductivity*,
437 2011, **21**, 2092-2095.
- 438 14. B. Biospin, Bruker Announces Five Ultra-High Field NMR Orders from Europe and Brazil,
439 [http://ir.bruker.com/investors/press-releases/press-release-details/2015/Bruker-](http://ir.bruker.com/investors/press-releases/press-release-details/2015/Bruker-Announces-Five-Ultra-High-Field-NMR-Orders-from-Europe-and-Brazil/default.aspx)
440 [Announces-Five-Ultra-High-Field-NMR-Orders-from-Europe-and-Brazil/default.aspx](http://ir.bruker.com/investors/press-releases/press-release-details/2015/Bruker-Announces-Five-Ultra-High-Field-NMR-Orders-from-Europe-and-Brazil/default.aspx),
441 (accessed 1).
- 442 15. M. Goldman, *Spin temperature and nuclear magnetic resonance in solids*, Clarendon Press,
443 Oxford,, 1970.

- 444 16. A. Abragam and M. Goldman, *Reports on Progress in Physics*, 1976, **41**, 395-467.
- 445 17. A. B. Barnes, G. D. Paëpe, P. C. A. v. d. Wel, K.-N. Hu, C.-G. Joo, V. S. Bajaj, M. L. Mak-
446 Jurkauskas, J. R. Sirigiri, J. Herzfeld, R. J. Temkin and R. G. Griffin, *Applied Magnetic*
447 *Resonance*, 2008, **34**, 237-263.
- 448 18. Q. Z. Ni, E. Daviso, T. V. Cana, E. Markhasin, S. K. Jawla, R. J. Temkin, J. Herzfeld and R. G.
449 Griffin, *Accounts of Chemical Research*, 2013, **46**, 1933-1941.
- 450 19. T. Maly, G. T. Debelouchina, V. S. Bajaj, K. N. Hu, C. G. Joo, M. L. Mak-Jurkauskas, J. R.
451 Sirigiri, P. C. A. van der Wel, J. Herzfeld, R. J. Temkin and R. G. Griffin, *J Chem Phys*, 2008,
452 **128**, 052211.
- 453 20. A. J. Rossini, A. Zagdoun, M. Lelli, A. Lesage, C. Copéret and L. Emsley, *Accounts of Chemical*
454 *Research*, 2013, **46**, 1942-1951.
- 455 21. R. A. Wind, M. J. Duijvestijn, C. Vanderlugt, A. Manenschijn and J. Vriend, *Progress in Nuclear*
456 *Magnetic Resonance Spectroscopy*, 1985, **17**, 33-67.
- 457 22. Y. Su, L. Andreas and R. G. Griffin, *Annual Review of Biochemistry*, 2015, **84**, 465-497.
- 458 23. V. K. Michaelis, T.-C. Ong, M. K. Kiesewetter, D. K. Frantz, J. J. Walish, E. Ravera, C. Luchinat,
459 T. M. Swager and R. G. Griffin, *Israel Journal of Chemistry*, 2014, **54**, 207-221.
- 460 24. V. A. Atsarkin, *Sov. Phys. Usp.*, 1978, **21**, 725.
- 461 25. A. Abragam and M. Goldman, *Nuclear Magnetism: Order and Disorder*, Clarendon Press,
462 Oxford, 1982.
- 463 26. M. L. Mak-Jurkauskas and R. G. Griffin, *eMagRes*, 2007.
- 464 27. C. D. Jeffries, *Dynamic Nuclear Orientation*, Interscience Publishers, 1963.
- 465 28. T. C. Ong, R. Verel and C. Copéret, in *Encyclopedia of Spectroscopy and Spectrometry (Third*
466 *Edition)*, eds. G. E. Tranter and D. W. Koppenaal, Academic Press, Oxford, 2017, pp. 121-127.
- 467 29. V. S. Bajaj, M. K. Hornstein, K. E. Kreischer, J. R. Sirigiri, P. P. Woskov, M. L. Mak-Jurkauskas,
468 J. Herzfeld, R. J. Temkin and R. G. Griffin, *J Magn Reson*, 2007, **189**, 251-279.
- 469 30. G. J. Gerfen, L. R. Becerra, D. A. Hall, R. G. Griffin, R. J. Temkin and D. J. Singel, *J Chem Phys*,
470 1995, **102**, 9494-9497.
- 471 31. S. Jawla, E. Nanni, M. Shapiro, I. Mastovsky, W. Guss, R. Temkin and R. Griffin, 2011,
472 presented at the 36th International Conference on Infrared, Millimeter, and Terahertz Waves,
473 IEEE, Huston, TX, USA.
- 474 32. S. Jawla, M. Reese, C. George, C. Yang, M. Shapiro, R. Griffin and R. Temkin, 2016, presented
475 at the International Vacuum Electronics Conference, IEEE, Monterey, CA, USA.
- 476 33. A. C. Torrezan, S.-T. Han, I. Mastovsky, M. A. Shapiro, J. R. Sirigiri, R. J. Temkin, A. B. Barnes
477 and R. G. Griffin, *IEEE Transactions on Plasma Science*, 2010, **38**, 1150-1159.

- 478 34. T. Idehara, Y. Tatematsu, Y. Yamaguchi, E. M. Khutoryan, A. N. Kuleshov, K. Ueda, Y. Matsuki
479 and T. Fujiwara, *Journal of Infrared, Millimeter, and Terahertz Waves*, 2015, **36**, 613-627.
- 480 35. Y. Matsuki, T. Idehara, J. Fukazawa and T. Fujiwara, *J Magn Reson*, 2016, **264**, 107-115.
- 481 36. Y. Matsuki, H. Takahashi, K. Ueda, T. Idehara, I. Ogawa, M. Toda, H. Akutsu and T. Fujiwara,
482 *Physical Chemistry Chemical Physics*, 2010, **12**, 5799-5803.
- 483 37. A. B. Barnes, E. A. Nanni, J. Herzfeld, R. G. Griffin and R. J. Temkin, *J Magn Reson*, 2012, **221**,
484 147-153.
- 485 38. S. Jawla, Q. Z. Ni, A. Barnes, W. Guss, E. Daviso, J. Herzfeld, R. Griffin and R. Temkin, *Journal*
486 *of Infrared, Millimeter, and Terahertz Waves*, 2013, **34**, 42-52.
- 487 39. A. C. Torrezan, M. A. Shapiro, J. R. Sirigiri, R. J. Temkin and R. G. Griffin, *IEEE Transactions*
488 *on Electron Devices*, 2011, **58**, 2777-2783.
- 489 40. R. Ikeda, T. Idehara, I. Ogawa, Y. Tatematsu, T. H. Chang, N. C. Chen, Y. Matsuki, K. Ueda and
490 T. Fujiwara, 2012.
- 491 41. K. R. Thurber, W.-M. Yau and R. Tycko, *J Magn Reson*, 2010, **204**, 303-313.
- 492 42. P. J. Allen, F. Creuzet, H. J. M. De Groot and R. G. Griffin, *Journal of Magnetic Resonance*
493 *(1969)*, 1991, **92**, 614-617.
- 494 43. E. Bouleau, P. Saint-Bonnet, F. Mentink-Vigier, H. Takahashi, J. F. Jacquot, M. Bardet, F.
495 Aussenac, A. Pura, F. Engelke, S. Hediger, D. Lee and G. De Paepe, *Chemical Science*, 2015, **6**,
496 6806-6812.
- 497 44. K. Thurber and R. Tycko, *J Magn Reson*, 2016, **264**, 99-106.
- 498 45. R. Tycko, *Accounts of Chemical Research*, 2013, **46**, 1923-1932.
- 499 46. M. Concistre, O. G. Johannessen, E. Carignani, M. Geppi and M. H. Levitt, *Accounts of Chemical*
500 *Research*, 2013, **46**, 1914-1922.
- 501 47. A. B. Barnes, E. Markhasin, E. Daviso, V. K. Michaelis, E. A. Nanni, S. K. Jawla, E. L. Mena, R.
502 DeRocher, A. Thakkar, P. P. Woskov, J. Herzfeld, R. J. Temkin and R. G. Griffin, *J Magn Reson*,
503 2012, **224**, 1-7.
- 504 48. M. Rosay, M. Blank and F. Engelke, *J Magn Reson*, 2016, **264**, 88-98.
- 505 49. A. B. Barnes, M. L. Mak-Jurkauskas, Y. Matsuki, V. S. Bajaj, P. C. A. van der Wel, R. DeRocher,
506 J. Bryant, J. R. Sirigiri, R. J. Temkin, J. Lugtenburg, J. Herzfeld and R. G. Griffin, *J Magn Reson*,
507 2009, **198**, 261-270.
- 508 50. M. Rosay, L. Tometich, S. Pawsey, R. Bader, R. Schauwecker, M. Blank, P. M. Borchard, S. R.
509 Cauffman, K. L. Felch, R. T. Weber, R. J. Temkin, R. G. Griffin and W. E. Maas, *Physical*
510 *Chemistry Chemical Physics*, 2010, **12**, 5850-5860.
- 511 51. E. Markhasin, J. Hu, Y. Su, J. Herzfeld and R. G. Griffin, *J Magn Reson*, 2013, **231**, 32-38.

- 512 52. P. K. Gor'kov, W. W. Brey and J. R. Long, *eMagRes*, 2007.
- 513 53. J. Hu and J. Herzfeld, 2011, Pat. # 7936171B2.
- 514 54. R. A. McKay, *eMagRes*, 2007.
- 515 55. J. Schaefer and R.A. McKay, 1999, Pat.# 5861748.
- 516 56. C. V. Grant, C. H. Wu and S. J. Opella, *J Magn Reson*, 2010, **204**, 180-188.
- 517 57. F. D. Doty, *eMagRes*, 2007.
- 518 58. E. K. Paulson, R. W. Martin and K. W. Zilm, *J Magn Reson*, 2004, **171**, 314-323.
- 519 59. L. R. Becerra, G. J. Gerfen, B. F. Bellew, J. A. Bryant, D. A. Hall, S. J. Inati, R. T. Weber, S. Un,
520 T. F. Prisner, A. E. Mcdermott, K. W. Fishbein, K. E. Kreisler, R. J. Temkin, D. J. Singel and R.
521 G. Griffin, *J Magn Reson Ser A*, 1995, **117**, 28-40.
- 522 60. V. S. Bajaj, C. T. Farrar, M. K. Hornstein, I. Mastovsky, J. Vieregge, J. Bryant, B. Eléna, K. E.
523 Kreisler, R. J. Temkin and R. G. Griffin, *J Magn Reson*, 2003, **160**, 85-90.
- 524 61. K. J. Pike, T. F. Kemp, H. Takahashi, R. Day, A. P. Howes, E. V. Kryukov, J. F. MacDonald, A.
525 E. Collis, D. R. Bolton, R. J. Wylde, M. Orwick, K. Kosuga, A. J. Clark, T. Idehara, A. Watts, G.
526 M. Smith, M. E. Newton, R. Dupree and M. E. Smith, *J Magn Reson*, 2012, **215**, 1-9.
- 527 62. E. Saliba, N. Alaniva, F. Scott, B. Albert, C. Gao, M. Mardini, E. Choi, S. Ho Pahng and A. B.
528 Barnes, Breckenridge, 2016.
- 529 63. K. N. Hu, V. S. Bajaj, M. Rosay and R. G. Griffin, *Journal of Chemical Physics*, 2007, **126**.
- 530 64. B. Corzilius, A. A. Smith, A. B. Barnes, C. Luchinat, I. Bertini and R. G. Griffin, *J Am Chem Soc*,
531 2011, **133**, 5648-5651.
- 532 65. B. Corzilius, A. A. Smith and R. G. Griffin, *J Chem Phys*, 2012, **137**.
- 533 66. A. A. Smith, B. Corzilius, A. B. Barnes, T. Maly and R. G. Griffin, *J Chem Phys*, 2012, **136**.
- 534 67. T. V. Can, Q. Z. Ni and R. G. Griffin, *J Magn Reson*, 2015, **253**, 23-35.
- 535 68. M. Lelli, S. R. Chaudhari, D. Gajan, G. Casano, A. J. Rossini, O. Ouari, P. Tordo, A. Lesage and
536 L. Emsley, *J Am Chem Soc*, 2015, **137**, 14558-14561.
- 537 69. A. Abragam and W. G. Proctor, *Cr Hebd Acad Sci*, 1958, **246**, 2253-2256.
- 538 70. M. Afeworki and J. Schaefer, *Macromolecules*, 1992, **25**, 4092-4096.
- 539 71. E. Erb, J. L. Motchane and C. R. Ubersfeld, *Acad. Sci.*, 1958, **246**, 2253.
- 540 72. C. F. Hwang and D. A. Hill, *Phys Rev Lett*, 1967, **19**, 1011.
- 541 73. C. F. Hwang and D. A. Hill, *Phys Rev Lett*, 1967, **18**, 110.
- 542 74. C. D. Jeffries, *Phys Rev*, 1957, **106**, 164-165.
- 543 75. A. V. Kessenikh, V. I. Lushchikov, A. A. Manenkov and Y. V. Taran, *Sov Phys-Sol State*, 1963, **5**,
544 321-329.
- 545 76. A. V. Kessenikh, A. A. Manenkov and G. I. Pyatnitskii, *Sov Phys-Sol State*, 1964, **6**, 641-643.

- 546 77. A. W. Overhauser, *Phys Rev*, 1953, **92**, 411-415.
- 547 78. D. S. Wollan, *Phys Rev B*, 1976, **13**, 3671-3685.
- 548 79. Y. Hovav, A. Feintuch and S. Vega, *J Magn Reson*, 2012, **214**, 29-41.
- 549 80. Y. Hovav, O. Levinkron, A. Feintuch and S. Vega, *Applied Magnetic Resonance*, 2012, **43**, 21-41.
- 550 81. F. Mentink-Vigier, Ü. Akbey, Y. Hovav, S. Vega, H. Oschkinat and A. Feintuch, *J Magn Reson*,
551 2012, **224**, 13-21.
- 552 82. D. Shimon, Y. Hovav, A. Feintuch, D. Goldfarb and S. Vega, *Physical Chemistry Chemical*
553 *Physics*, 2012, **14**, 5729-5743.
- 554 83. K.-N. Hu, *Solid State Nuclear Magnetic Resonance*, 2011, **40**, 31-41.
- 555 84. K. N. Hu, G. T. Debelouchina, A. A. Smith and R. G. Griffin, *Journal of Chemical Physics*, 2011,
556 **134**, 125105.
- 557 85. K.-N. Hu, V. S. Bajaj, M. Rosay and R. G. Griffin, *The Journal of Chemical Physics*, 2007, **126**,
558 044512.
- 559 86. C. S. Song, K. N. Hu, C. G. Joo, T. M. Swager and R. G. Griffin, *J Am Chem Soc*, 2006, **128**,
560 11385-11390.
- 561 87. V. K. Michaelis, B. Corzilius, A. A. Smith and R. G. Griffin, *J Phys Chem B*, 2013, **117**, 14894-
562 14906.
- 563 88. V. K. Michaelis, A. A. Smith, B. Corzilius, O. Haze, T. M. Swager and R. G. Griffin, *J. Am.*
564 *Chem. Soc.*, 2013, **135**, 2935-2938.
- 565 89. K. R. Thurber and R. Tycko, *The Journal of Chemical Physics*, 2012, **137**, -.
- 566 90. C. D. Jeffries, *Physical Review*, 1960, **117**, 1056-1069.
- 567 91. A. N. Smith and J. R. Long, *Analytical Chemistry*, 2016, **88**, 122-132.
- 568 92. O. Lafon, A. S. L. Thankamony, M. Rosay, F. Aussenac, X. Lu, J. Trebosc, V. Bout-Roumazeilles,
569 H. Vezin and J.-P. Amoureux, *Chemical Communications*, 2013, **49**, 2864-2866.
- 570 93. T. Maly, L. B. Andreas, A. A. Smith and R. G. Griffin, *Physical Chemistry Chemical Physics*,
571 2010, **12**, 5872-5878.
- 572 94. J. H. Ardenkjær-Larsen, B. Fridlund, A. Gram, G. Hansson, L. Hansson, M. H. Lerche, R. Servin,
573 M. Thaning and K. Golman, *Proceedings of the National Academy of Sciences*, 2003, **100**, 10158-
574 10163.
- 575 95. A. E. Dementyev, D. G. Cory and C. Ramanathan, *The Journal of Chemical Physics*, 2011, **134**,
576 154511.
- 577 96. K.-N. Hu, C. Song, H.-h. Yu, T. M. Swager and R. G. Griffin, *The Journal of Chemical Physics*,
578 2008, **128**, 052302.

- 579 97. J. H. Ardenkjaer-Larsen, S. Macholl and H. Johannesson, *Applied Magnetic Resonance*, 2008, **34**,
580 509-522.
- 581 98. S. Reynolds and H. Patel, *Applied Magnetic Resonance*, 2008, **34**, 495-508.
- 582 99. T. V. Can, M. A. Caporini, F. Mentink-Vigier, B. Corzilius, J. J. Walish, M. Rosay, W. E. Maas,
583 M. Baldus, S. Vega, T. M. Swager and R. G. Griffin, *The Journal of Chemical Physics*, 2014, **141**,
584 064202.
- 585 100. B. Corzilius, L. B. Andreas, A. A. Smith, Q. Z. Ni and R. G. Griffin, *J Magn Reson*, 2014, **240**,
586 113-123.
- 587 101. G. Mathies, M. A. Caporini, V. K. Michaelis, Y. Liu, K.-N. Hu, D. Mance, J. L. Zweier, M. Rosay,
588 M. Baldus and R. G. Griffin, *Angewandte Chemie International Edition*, 2015, **54**, 11770-11774.
- 589 102. C. Sauvée, G. Casano, S. Abel, A. Rockenbauer, D. Akhmetzyanov, H. Karoui, D. Siri, F.
590 Aussenac, W. Maas, R. T. Weber, T. Prisner, M. Rosay, P. Tordo and O. Ouari, *Chemistry – A*
591 *European Journal*, 2016, **22**, 5598-5606.
- 592 103. D. J. Kubicki, G. Casano, M. Schwarzwaldler, S. Abel, C. Sauvee, K. Ganesan, M. Yulikov, A. J.
593 Rossini, G. Jeschke, C. Coperet, A. Lesage, P. Tordo, O. Ouari and L. Emsley, *Chemical Science*,
594 2016, **7**, 550-558.
- 595 104. Y. Matsuki, T. Maly, O. Ouari, H. Karoui, F. Le Moigne, E. Rizzato, S. Lyubenova, J. Herzfeld, T.
596 Prisner, P. Tordo and R. G. Griffin, *Angew Chem Int Edit*, 2009, **48**, 4996-5000.
- 597 105. C. Sauvée, M. Rosay, G. Casano, F. Aussenac, R. T. Weber, O. Ouari and P. Tordo, *Angewandte*
598 *Chemie International Edition*, 2013, **52**, 10858-10861.
- 599 106. A. Zagdoun, G. Casano, O. Ouari, M. Schwarzwälder, A. J. Rossini, F. Aussenac, M. Yulikov, G.
600 Jeschke, C. Copéret, A. Lesage, P. Tordo and L. Emsley, *J Am Chem Soc*, 2013, **135**, 12790-
601 12797.
- 602 107. A. Zagdoun, G. Casano, O. Ouari, G. Lapadula, A. J. Rossini, M. Lelli, M. Baffert, D. Gajan, L.
603 Veyre, W. E. Maas, M. Rosay, R. T. Weber, C. Thieuleux, C. Coperet, A. Lesage, P. Tordo and L.
604 Emsley, *J Am Chem Soc*, 2012, **134**, 2284-2291.
- 605 108. M. K. Kiesewetter, B. Corzilius, A. A. Smith, R. G. Griffin and T. M. Swager, *J Am Chem Soc*,
606 2012, **134**, 4537-4540.
- 607 109. T. C. Ong, M. L. Mak-Jurkauskas, J. J. Walish, V. K. Michaelis, B. Corzilius, A. A. Smith, A. M.
608 Clausen, J. C. Cheetham, T. M. Swager and R. G. Griffin, *J Phys Chem B*, 2013, **117**, 3040-3046.
- 609 110. M. K. Kiesewetter, V. K. Michaelis, J. J. Walish, R. G. Griffin and T. M. Swager, *The Journal of*
610 *Physical Chemistry B*, 2014, **118**, 1825-1830.
- 611 111. E. L. Dane, B. Corzilius, E. Rizzato, P. Stocker, O. Ouari, T. Maly, A. A. Smith, R. G. Griffin, O.
612 Ouari, P. Tordo and T. M. Swager, *J. Organic Chem.*, 2012, **77**, 1789-1797.

- 613 112. M. Kaushik, T. Bahrenberg, T. V. Can, M. A. Caporini, R. Silvers, J. Heiliger, A. A. Smith, H.
614 Schwalbe, R. G. Griffin and B. Corzilius, *Physical Chemistry Chemical Physics*, 2016, **18**, 27205-
615 27218.
- 616 113. B. Corzilius, *Physical Chemistry Chemical Physics*, 2016, **18**, 27190-27204.
- 617 114. P. Wenk, M. Kaushik, D. Richter, M. Vogel, B. Suess and B. Corzilius, *Journal of Biomolecular*
618 *NMR*, 2015, **63**, 97-109.
- 619 115. B. Corzilius, V. K. Michaelis, S. Penzel, E. Ravera, A. A. Smith, C. Luchinat and R. G. Griffin, *J*
620 *Am Chem Soc*, 2014, **136**, 11716-11727.
- 621 116. D. A. Hall, D. C. Maus, G. J. Gerfen, S. J. Inati, L. R. Becerra, F. W. Dahlquist and R. G. Griffin,
622 *Science*, 1997, **276**, 930-932.
- 623 117. G. R. Eaton, S. S. Eaton, D. P. Barr and R. T. Weber, *Quantitative EPR*, SpringerWien, New
624 York, 2010.
- 625 118. T. Wada, M. Yamanaka, T. Fujihara, Y. Miyazato and K. Tanaka, *Inorganic Chemistry*, 2006, **45**,
626 8887-8894.
- 627 119. A. Zagdoun, A. J. Rossini, D. Gajan, A. Bourdolle, O. Ouari, M. Rosay, W. E. Maas, P. Tordo, M.
628 Lelli, L. Emsley, A. Lesage and C. Copéret, *Chemical Communications*, 2012, **48**, 654-656.
- 629 120. S. Y. Liao, M. Lee, T. Wang, I. V. Sergeyev and M. Hong, *Journal of Biomolecular NMR*, 2016,
630 **64**, 223-237.
- 631 121. G. T. Debelouchina, M. J. Bayro, P. C. A. v. d. Wel, M. A. Caporini, A. B. Barnes, M. Rosay, W.
632 E. Maas and R. G. Griffin, *Phys. Chem. Chem. Phys.*, 2010, **12**, 5911-5919.
- 633 122. W. R. Gunther, V. K. Michaelis, M. A. Caporini, R. G. Griffin and Y. Román-Leshkov, *J Am*
634 *Chem Soc*, 2014, **136**, 6219-6222.
- 635 123. P. C. A. van der Wel, K. N. Hu, J. Lewandowski and R. G. Griffin, *J Am Chem Soc*, 2006, **128**,
636 10840-10846.
- 637 124. S. Lange, A. H. Linden, Ü. Akbey, W. Trent Franks, N. M. Loening, B.-J. v. Rossum and H.
638 Oschkinat, *J Magn Reson*, 2012, **216**, 209-212.
- 639 125. A. J. Rossini, A. Zagdoun, F. Hegner, M. Schwarzwälder, D. Gajan, C. Copéret, A. Lesage and L.
640 Emsley, *J Am Chem Soc*, 2012, **134**, 16899-16908.
- 641 126. A. J. Rossini, A. Zagdoun, M. Lelli, J. Canivet, S. Aguado, O. Ouari, P. Tordo, M. Rosay, W. E.
642 Maas, C. Copéret, D. Farrusseng, L. Emsley and A. Lesage, *Angewandte Chemie International*
643 *Edition*, 2012, **51**, 123-127.
- 644 127. M. L. Mak-Jurkauskas, V. S. Bajaj, M. K. Hornstein, M. Belenky, R. G. Griffin and J. Herzfeld,
645 *Proceedings of the National Academy of Sciences of the United States of America*, 2008, **105**,
646 883-888.

- 647 128. V. S. Bajaj, M. L. Mak-Jurkauskas, M. Belenky, J. Herzfeld and R. G. Griffin, *Proceedings of the*
648 *National Academy of Sciences of the United States of America*, 2009, **106**, 9244-9249.
- 649 129. A. B. Barnes, B. Corzilius, M. L. Mak-Jurkauskas, L. B. Andreas, V. S. Bajaj, Y. Matsuki, M. L.
650 Belenky, J. Lugtenburg, J. R. Sirigiri, R. J. Temkin, J. Herzfeld and R. G. Griffin, *Phys. Chem.*
651 *Chem. Phys.*, 2010, **12**, 5861-5867.
- 652 130. M. A. Voinov, D. B. Good, M. E. Ward, S. Milikisiyants, A. Marek, M. A. Caporini, M. Rosay, R.
653 A. Munro, M. Ljumovic, L. S. Brown, V. Ladizhansky and A. I. Smirnov, *The Journal of Physical*
654 *Chemistry B*, 2015, **119**, 10180-10190.
- 655 131. B. J. Wylie, B. G. Dzikovski, S. Pawsey, M. Caporini, M. Rosay, J. H. Freed and A. E.
656 McDermott, *Journal of Biomolecular NMR*, 2015, **61**, 361-367.
- 657 132. L. B. Andreas, A. B. Barnes, B. Corzilius, J. J. Chou, E. A. Miller, M. Caporini, M. Rosay and R.
658 G. Griffin, *Biochemistry*, 2013, **52**, 2774-2782.
- 659 133. M. J. Bayro, G. T. Debelouchina, M. T. Eddy, N. R. Birkett, C. E. MacPhee, M. Rosay, W. E.
660 Maas, C. M. Dobson and R. G. Griffin, *J. Am. Chem. Soc.*, 2011, **133**, 13967-13974.
- 661 134. M. J. Bayro, T. Maly, N. Birkett, C. MacPhee, C. M. Dobson and R. G. Griffin, *Biochemistry*,
662 2010, **49**, 7474-7488.
- 663 135. G. T. Debelouchina, G. W. Platt, M. J. Bayro, S. E. Radford and R. G. Griffin, *J. Am. Chem. Soc.*,
664 2010, **132**, 17077-17079.
- 665 136. A. J. Rossini, A. Zagdoun, F. Hegner, M. Schwarzwaelder, D. Gajan, C. Copéret, A. Lesage and
666 L. Emsley, *J. Amer. Chem. Soc.*, 2012, **134**, 16899-16908.
- 667 137. A. Lesage, M. Lelli, D. Gajan, M. A. Caporini, V. Vitzthum, P. Mieville, J. Alauzun, A. Roussey,
668 C. Thieuleux, A. Mehdi, G. Bodenhausen, C. Coperet and L. Emsley, *J Am Chem Soc*, 2010, **132**,
669 15459-15461.
- 670 138. G. T. Debelouchina, M. J. Bayro, P. C. A. van der Wel, M. A. Caporini, A. B. Barnes, M. Rosay,
671 W. E. Maas and R. G. Griffin, *Physical Chemistry Chemical Physics*, 2010, **12**, 5911-5919.
- 672 139. M. Lelli, D. Gajan, A. Lesage, M. A. Caporini, V. Vitzthum, P. Mieville, F. Heroguel, F. Rascon,
673 A. Roussey, C. Thieuleux, M. Boualleg, L. Veyre, G. Bodenhausen, C. Coperet and L. Emsley, *J.*
674 *Am. Chem. Soc.*, 2011, **133**, 2104-2107.
- 675 140. Kendra K. Frederick, Vladimir K. Michaelis, B. Corzilius, T.-C. Ong, Angela C. Jacavone,
676 Robert G. Griffin and S. Lindquist, *Cell*, 2015, **163**, 620-628.
- 677 141. O. Lafon, A. S. L. Thankamony, T. Kobayashi, D. Carnevale, V. Vitzthum, I. I. Slowing, K.
678 Kandel, H. Vezin, J.-P. Amoureux, G. Bodenhausen and M. Pruski, *The Journal of Physical*
679 *Chemistry C*, 2013, **117**, 1375-1382.

- 680 142. D. Lee, N. T. Duong, O. Lafon and G. De Paëpe, *The Journal of Physical Chemistry C*, 2014, **118**,
681 25065-25076.
- 682 143. H. Takahashi, I. Ayala, M. Bardet, G. De Paëpe, J.-P. Simorre and S. Hediger, *J Am Chem Soc*,
683 2013, **135**, 5105-5110.
- 684 144. H. Takahashi, S. Hediger and G. De Paepe, *Chemical Communications*, 2013, **49**, 9479-9481.
- 685 145. A. S. L. Thankamony, O. Lafon, X. Lu, F. Aussenac, M. Rosay, J. Trébosc, H. Vezin and J.-P.
686 Amoureux, *Applied Magnetic Resonance*, 2012, **43**, 237-250.
- 687 146. E. Ravera, B. Corzilius, V. K. Michaelis, C. Rosa, R. G. Griffin, C. Luchinat and I. Bertini, *J. Am.*
688 *Chem. Soc.*, 2013, **134**, 1641-1644.
- 689 147. E. Ravera, B. Corzilius, V. K. Michaelis, C. Luchinat, R. G. Griffin and I. Bertini, *Journal of*
690 *Physical Chemistry B*, 2014, **118**, 1825-1830.
- 691 148. A. N. Smith, M. A. Caporini, G. E. Fanucci and J. R. Long, *Angewandte Chemie International*
692 *Edition*, 2015, **54**, 1542-1546.
- 693