| Technique             |            | Single Rock<br>Chlorophyll |          | Brush<br>a m-2 | Scrape    |           |         | •   | •<br>•                   |
|-----------------------|------------|----------------------------|----------|----------------|-----------|-----------|---------|-----|--------------------------|
|                       | Mean<br>CV | Mean                       | Range    | <del>.</del>   | Mea<br>cv | n<br>Mean | Range   | _   | nificance<br>n vs Scrape |
| Shore                 | 63         | 26.6                       | (7.5-65. | .8)            | 61        | 22.7      | (5.8-36 | .5) | (p < .05)                |
| Midstream             | 58         | 31.1                       | (4.7-84  | .9)            | 65        | 24.1      | (3.7-94 | .2) | N.S.                     |
| Midstream<br>(shaded) | 75         | 16.8                       | (5.8-55  | .9)            | 70        | 16.9      | (0.5-58 | .1) | N.S.                     |

cv = coefficient of variation

(% deviation from the mean <u>Standard Deviation</u> X 100)

Mean

Table 17. Microhabitat mean and range of standing crops, deviation from the mean (cv) and probability of significant differences between sampling techniques (students t-test) (1978).

Although the single rock brush and scrape techniques yielded similar mean standing crop estimates among each of the microhabitats examined, specific differences were evident when seasonal patterns were compared (Figures 28, 29). The rock brush technique yielded higher values during May, July, August and after November (Figure 28). The scrape technique yielded higher estimates of mean standing crop primarily during the fall (Sept-Nov).

Standing crop estimates were lower in the midstream shaded than the unshaded river locations (Table 17). The values which exhibited the highest percent deviation from the mean were found in the shaded microhabitat. Analysis of variance (ANOVA) failed to show significant differences in algal standing crop among the various microhabitats. Paired student t-tests did however indicate a significant difference between the two techniques when applied to the shallow shore microhabitat only.

## 4.1.3 Single Rock Brush vs Scrape Technique (1979)

During 1979, the single rock brush technique was again employed to estimate epilithic algal standing crops; however, microhabitats were not differentiated; instead, rocks were collected at random along transects from shore to shore in the Muskeg-River. The brushing technique provided a higher estimate of epilithic algal standing crop (Figure 28). Differences were most pronounced during June and July. The scrape technique yielded a mean 24.8 mg chl a  $m^{-2}$ , range (5.7-76.3 mg chl a  $m^{-2}$ ), the brush technique a mean of 26.2 mg chl a  $m^{-2}$  (range 1.3-101.1 mg chl  $m^{-2}$ ). Analysis of variance (ANOVA) showed no significant difference (p <0.05) between the two techniques for estimating epilithic algal standing crop.

## 4.2 Substratum Comparison: Oilsand/Rock

During 1977, visual observations indicated that naturally occurring bitumous substratum (oilsand) appeared to support more Nostoc commune than rock substratum. Closer examination using chlorophyll a as a measure of epilithic algal standing crop indicated that the oilsand supported an average 27.7 mg chlorophyll a m-2 while the rock supported an average 60.1 mg chlorophyll a m-2, during the period July 13 through December 9, 1977. A similar trend was again found during 1978 with oilsand averaging 10.8 and rock averaging 48.5 mg chlorophyll a m-2. The rock supported significantly higher (p .05) standing crops. The apparent abundance of Cyanophycean algae associated with oilsand and rock substratum was further examined with respect blue-green algal numbers and nitrogen fixation. The findings will be discussed later.

# 4.3 Seasonal Batterns or Temporal Changes

## 4.3.1 Physical Changes

The following is a presentation of the seasonal dynamics of physical and chemical parameters examined during investigation of the Muskeg, Steepbank, Hangingstone, Ells and MacKay rivers.

## 4.3.1.1 Temperature

The seasonal dynamics of water temperature were similar for the five rivers investigated. Maximum temperatures occurred each July and thereafter declined (Figure 30). Only during 1977 in the Steepbank and Muskeg Rivers did any marked fluctuation occur. The MacKay River exhibited the greatest overall mean temperature (9.6°C), and the widest temperature range (0-23°C). During the winter period and the resultant low temperatures (November - April), ice depths of up to 76.3 cm were encountered on the Ells River.

## 4.3.1.2 Discharge

Discharge patterns were similar in each river (Figure 31). In the MacKay, Ells and Steepbank Rivers maxima occurred in September, 1978 and May, 1979, with the latter being greater. In contrast, these maxima were of comparable size in the Hangingstone and Muskeg Rivers. Discharge during 1977 in both the Muskeg and Steepbank Rivers were much lower than during 1978 and 1979. Discharge in the MacKay River was greater than any of the remaining four rivers.

# 4.3.1.3 Specific Conductance

Mean specific conductance in the five rivers investigated ranged from 10.9 to 23.4 mS/m (Figure 36). It was greatest in the Muskeg River but remained lowest and essentially constant in the Ells River, (i.e. 10.9 + 0.2 mS/m) throughout the study period.

Figure 30.
Water temperatures in the Ells, MacKay, Hangingstone,
Steepbank and Muskeg rivers.

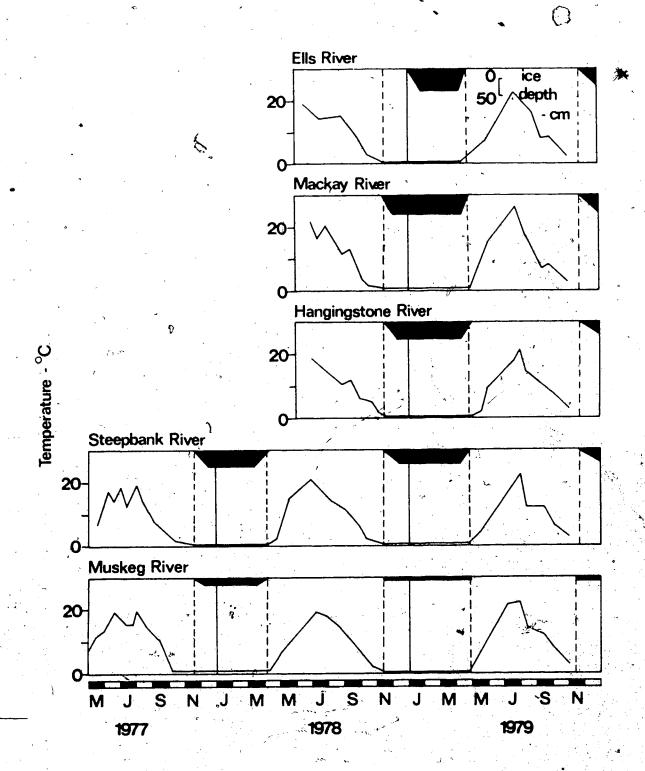


Figure 31.

Discharge (m<sup>3</sup> s<sup>-1</sup>) in Ells, MacKay, Hangingstone, Steepbank, and Muskeg Rivers.

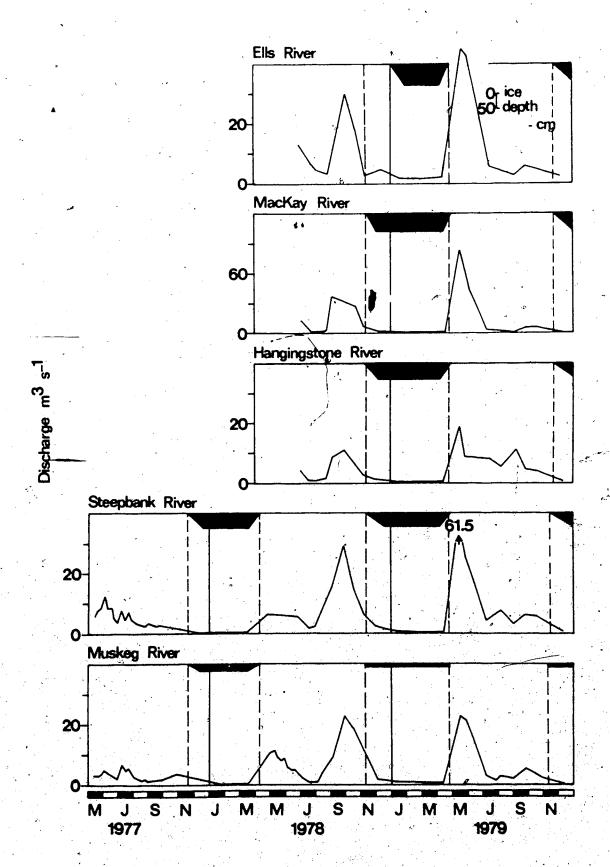
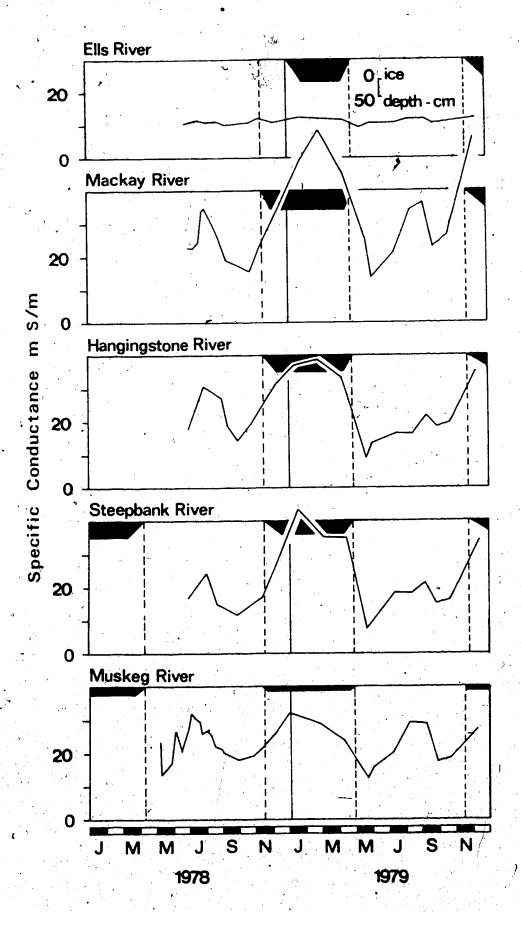


Figure 32.

Specific conductance (mS/m) in the Ells, MacKay Hangingstone, Steepbank and Muskeg Rivers.



In contrast to the Ells River, a distinct seasonal pattern of conductance occurred in the remaining Rivers (Figure 32). All rivers investigated displayed a summer peak which decreased to a minimum during the autumn, then under ice cover conductance increased and the overall maxima occurred.

## 4.3.1.4 <u>Calcium</u>

Calcium concentrations fluctuated least in the Ells River with only a small increase occurring during the winter (Figure 33). Again, in , marked constrast, large winter peaks occurred in all the other rivers 49.5, 44.8, 43.0, 33.1 mg L<sup>+1</sup> in the MacKay, Muskeg, Steepbank and Hangingstone Rivers respectively. During the summer of 1978 calcium levels in the Muskeg River fluctuated, displaying several distinct peaks (Figure 33) which coincided with periods of minimum stream discharge. In the MacKay, Steepbank, and Hangingstone, summer peaks were evident but always smaller than winter ones.

## 4.3.1.5 Sodium

Sodium concentrations in the Ells River remained low  $(3.9 \text{ mg L}^{-1})$  and fluctuated less than in any other river (Figure 34).

As ice formed during the fall, sodium decreased and remained uniform until after ice break-up during early May. The seasonal pattern, however, was more variable in the remaining rivers. In the Mackay, a series of distinct peaks were evident; 52.4 mg  $L^{-1}$  during August of 1978, 57.5 mg  $L^{-1}$  during March of 1979, 51.8 mg  $L^{-1}$  during September, 1979 and 55.1 mg  $L^{-1}$  during December, 1979. These peaks originated and disappeared quickly. In between values fell to minima, ranging from 7.9 to 19.5 mg  $L^{-1}$  sodium. A decreasing stepwise series of peaks from early August, 1978 until a minimum in May, 1979 were noted for the Hangingstone River.

Figure 33.

Calcium (mg 1<sup>-1</sup>) in the Ells, MacKay, Hangingstone, Steepbank, and Muskeg rivers.

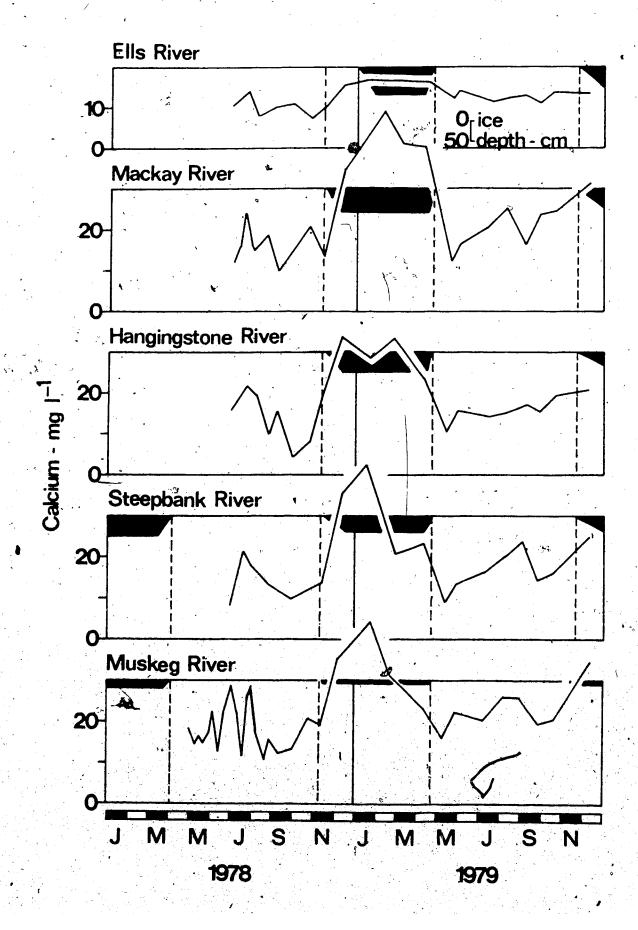
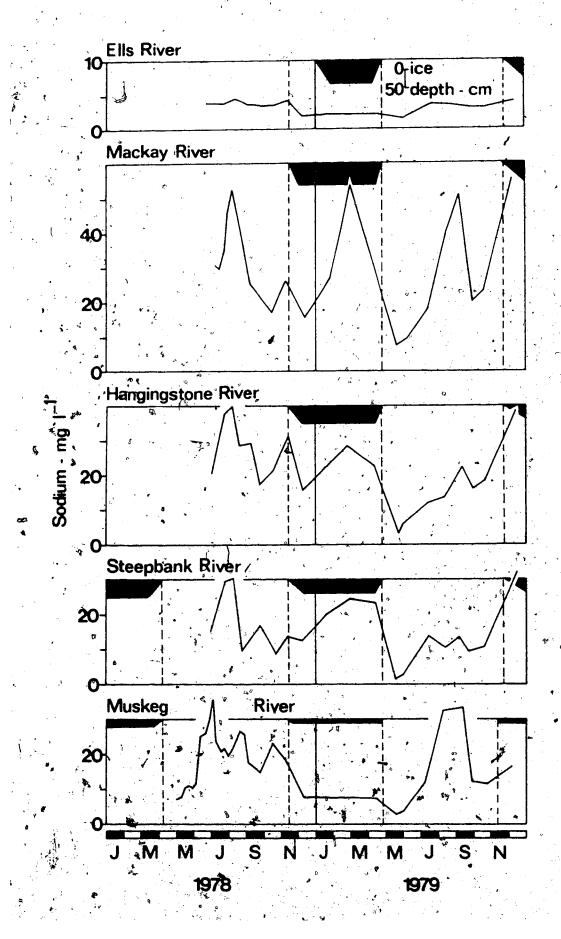


Figure 34.

Sodium (mg 1<sup>-1</sup>) in the Ells, Hnagingstone,
Steepbank, and Muskeg rivers.



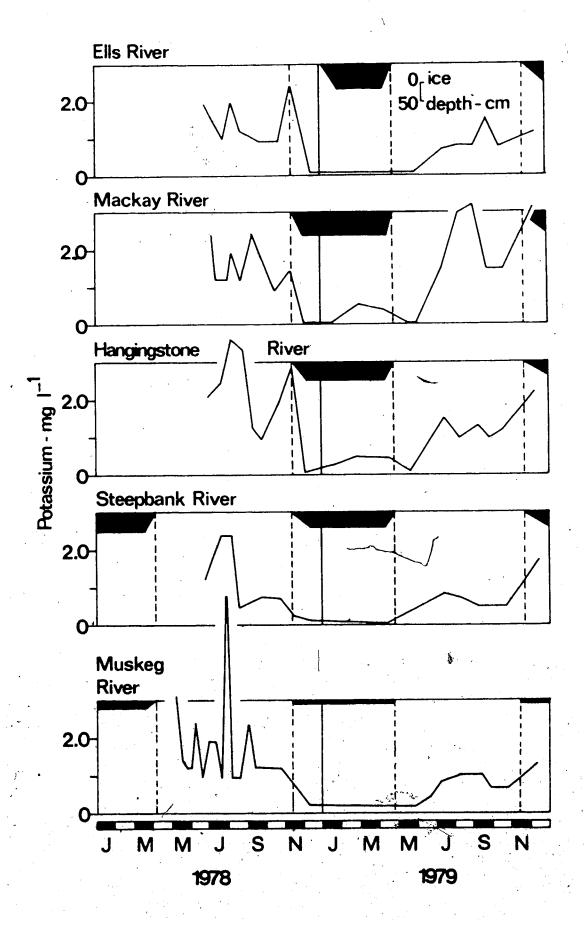
After May, sodium concentrations increased steadily, peaking again during early December at 31.2 mg L $^{-1}$ . Sodium concentrations in the Steepbank River fluctuated similarly, but the winter peak (48.7 mg L $^{-1}$ ) was larger than the autumn one of 16.4 mg L $^{-1}$  (Figure 34). Spring minima in the Steepbank River were also less than the spring minimum in the Hangingstone River, reaching 3.20 mg L $^{-1}$ .

A decreasing stepwise pattern of sodium similar to the Hangingstone was also evident in the Muskeg River during July (35.6 mg  $L^{-1}$ ). August (26.50 mg  $L^{-1}$ ) and during October (18.70 mg  $L^{-1}$ ). Unlike the MacKay, Hangingstone and Steepbank rivers, but like the Ells River, sodium levels fell after mid-October 1978 and remained low all winter with a May, 1979 minimum of 5.7 mg  $L^{-1}$ . Sodium concentrations increased to a maximum of 33.2 mg  $L^{-1}$  during September, 1979, then decreased but continued to show an upward trend during the onset of ice formation and termination of the study.

#### 4.3.1.6 Potassium

Potassium concentrations were quite similar seasonally among the five rivers (Figure 35). Varying sized peaks occurred during the summer growth period and late autumn/early winter season. Two distinct peaks occurred in the Ells and Hangingstone Rivers during August and November (1.94 and 2.40 mg L<sup>-1</sup>; 3.60 and 2.88 mg L<sup>-1</sup>) respectively. One main peak occurred during July/August in the Steepbank River, whereas the potassium levels fluctuated more irregularly in the MacKay and Muskey Rivers (Figure 35). Four peaks were discernable in the MacKay River in July, August, September, and November. Five peaks occurred in the Muskey River during May, June/July, mid July and late August. Potassium levels were low in each river during the winter, but increased from early May minima to summer peaks, and fell to a minima.

Figure 35. Potassium (mg  $1^{-1}$ ) in the Ells, MacKay, Hangingstone, Steepbank and Muskeg rivers.



#### 4.3.1.7 Magnesium

Magnesium fluctuated little in the Ells, ranging from 1.6 to 4.9 mg  $L^{-1}$  (Figure 36). Small minima occurred just prior to ice formation and immediately after ice break-up. Seasonal patterns, however, in the other five rivers were more pronounced, with small summer peaks and very large winter maxima (17.9, 15.8, 14.9 and 14.1 mg  $L^{-1}$ ) in the MacKay, Steepbank, Muskeg and Hangingstone Rivers respectively. Magnesium concentrations decreased rapidly after ice break-up in early May, perhaps due to dilution associated with increased discharge attributed to the spring melting period. Nevertheless, spring minima were followed by increasing magnesium concentrations, and again, a fall minimum associated with peak discharge periods.

The concentrations of magnesium were similar seasonally for all five rivers, ranging from 6.40 mg  $L^{-1}$  in the MacKay River during 1978 to 10.3 mg  $L^{-1}$  in the Muskeg during 1979. Most irregularity occurred during 1978 in the Muskeg River (Figure 36) coincidentally with unusually high discharges during early September.

#### 4.3.1.8 Iron

An investigation of iron concentrations indicated that the MacKay, Hangingstone, and Steepbank Rivers were similar seasonally (Figure 37). Large early winter, late winter/early spring, and autumn (1979) maxima occurred. Iron ranged from 0.18 mg L $^{-1}$  in the Steepbank to 0.36 mg L $^{-1}$  in the MacKay. Two large peaks occurred in the Muskeg River in early May and early December, 1979 (0.73 and 0.99 mg L $^{-1}$  respectively), otherwise values fluctuated little. Iron concentrations remained relatively constant in the Ells River throughout the study, with only slight deviation similar to the seasonal patterns indicated for the MacKay, Hangingstone and Steepbank Rivers.

Figure 36.

Mangnesium (mg 1<sup>-1</sup>) in the Ælls, MacKav, Hnagingstone,

Steepbank and Muskeg rivers.

.

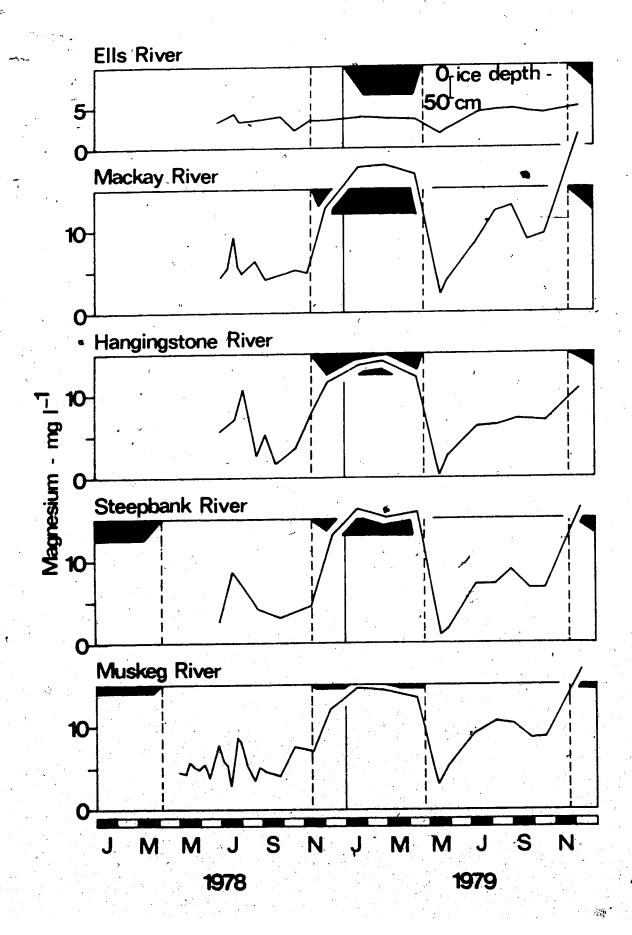
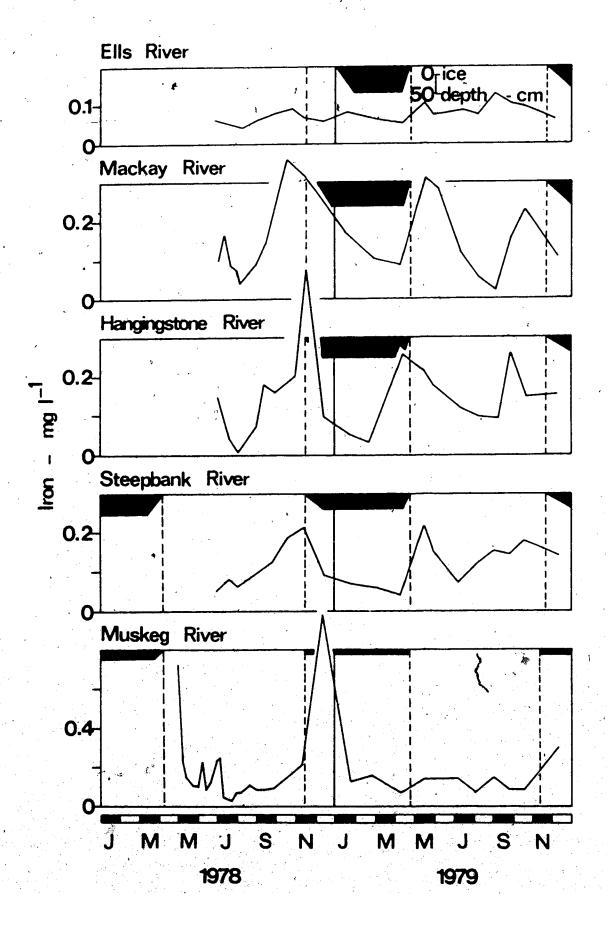


Figure 37. Iron (mg  $1^{-1}$ ) in the Ells, MacKay, Hangingstone, Steepbank and Muskeg rivers.



#### 4.3.1.9 Manganese

Manganese levels fluctuated irregularly in the Muskeg River, particularly during 1978 (Figure 38). In this river two winter maxima of 0.04 and 0.05 mg  $L^{-1}$  declined during ice-out to fluctuate throughout the open water period. One winter peak occurred in the Steepbank River (0.05 mg  $L^{-1}$ ), while a minimum manganese concentration appeared during late winter, and in contrast to the Muskeg River, the levels increased quickly after ice-out and again in early August.

Manganese levels fluctuated irregularly in the Hangingstone River with peaks of 0.075 and 0.095 mg  $L^{-1}$  during the winter season. Fewer rapid and irregular fluctuations occurred in the MacKay River (Figure 38). Here the seasonal pattern was dominated by an autumn and a smaller winter maxima (0.051 and 0.013 mg  $L^{-1}$  respectively). Ells River manganese levels decreased from June (0.01 mg  $L^{-1}$ ) to an overall November minimum of 0.007 mg  $L^{-1}$  during 1978. During 1979, however, manganese concentrations increased irregularly to peak during September, 0.101 mg  $L^{-1}$  (Figure 38).

# 4.3.1.10 Sulphate

Sulphate seasonal patterns were similar for the Mackay, Hangingstone and Steepbank Rivers (Figure 39). The concentrations per river, however, were very different. Sulphate ranged from a minimum of 0.0 mg  $L^{-1}$  in the Muskeg River to a maximum of 53 mg  $L^{-1}$  in the Mackay River.

Small summer peaks of sulphate occurred each year in the MacKay, Hangingstone and Steepbank Rivers (i.e. 53.0, 23.5, 13.4 mg  $L^{-1}$  respectively). Peaks were far less pronounced in the Ells River, fluctuating between 4.4 and 9.0 mg  $L^{-1}$  (Figure 39). However, in the

Figure 38.

Manganese (mg 1-1) in the Ells, MacKay, Hnagingstone, Steepbank and Muskeg rivers.

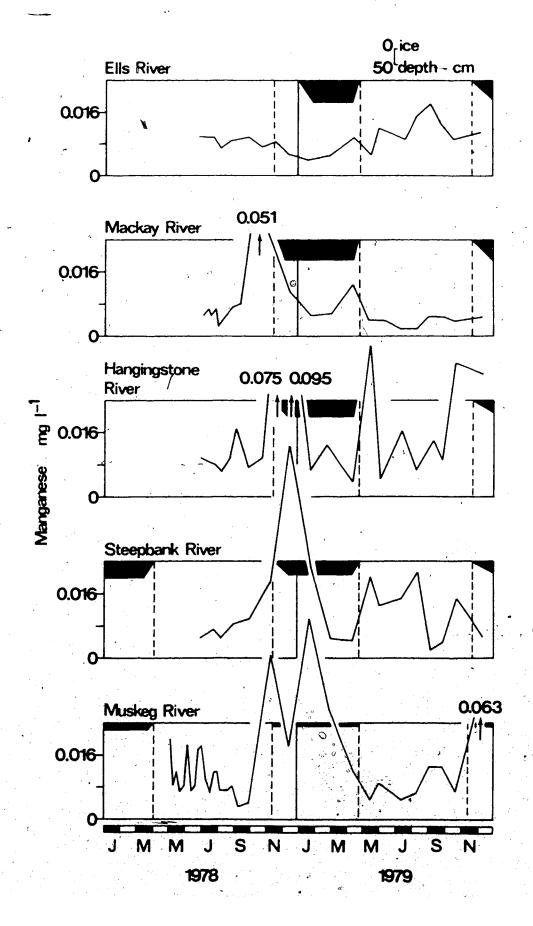
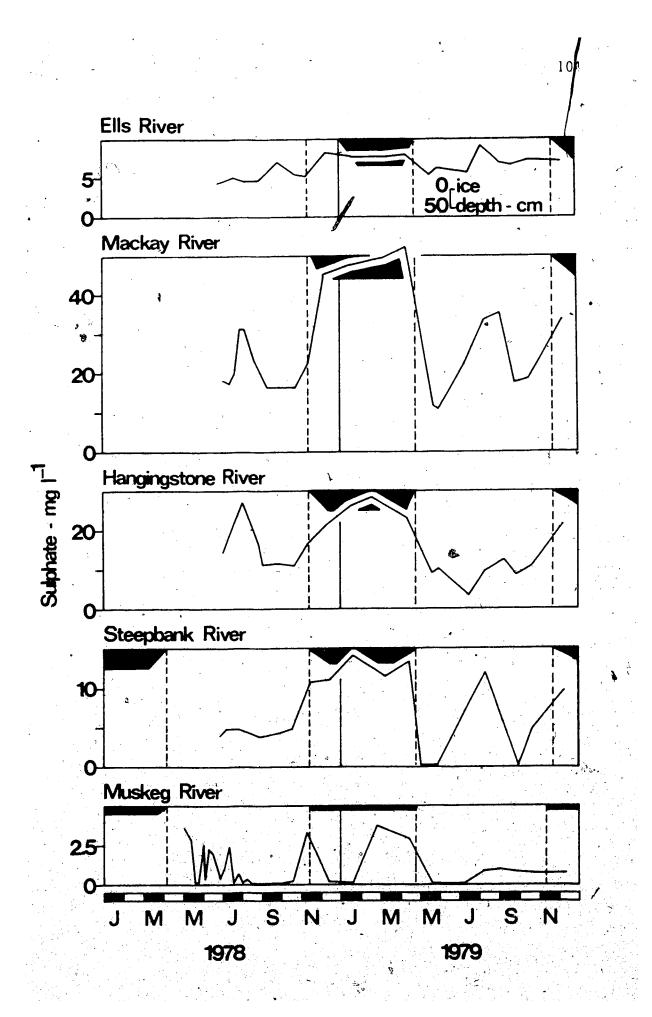


Figure 39.
Sulphate (mg 1<sup>-1</sup>) in the Ells, MacKay, Hangingstone,
Steepbank and Muskeg rivers.



Muskeg River, sulphate levels fluctuated widely during 1978,  $L^{-1}$  and the large of the Muskeg River; one in November (3.3 mg  $L^{-1}$ ) and the other in February (3.75 mg  $L^{-1}$ ) Figure 39.

#### 4.3.1.11 aChloride

Chloride was undetectable in the Ells River. Seasonal patterns were, however, similar in the MacKay, Hangingstone and Steepbank Rivers with large winter maximums and a smaller summer peak (Figure 40). Peak values of 20.0, 30.0 and 8.0 mg  $L^{-1}$  were found in these three rivers respectively. Seasonal patterns were similar in the Muskeg River; however, summer father than winter maximum occurred, and fluctuations were more frequent during 1978 than during 1979. Two peaks of 35.6 and 25.0 mg  $E^{-1}$  occurred during June and July of 1978, and of 27.5 and 42.5 mg  $E^{-1}$  during August and September of 1979.

## 4.3.1.12 Nitrate-nitrogen

Distinct seasonal nitrate-nitrogen fluctuations did not occur in the Ells River (Figure 41). More distinctive patterns occurred in the other four rivers (Figure 41), where the concentrations of nitrate-nitrogen were low during mid-summer, rising to a numn/early winter peaks. Then, as ice formed it declined in all but the Muskeg Rivers. The greatest variability occurred in the Muskeg River during 1978 (Figure 41).

# 4.3.1.13 Phosphate-phosphorus

Phosphate-phosphorus concentrations fluctuated widely and irregularly in the Ells River. Four major peaks occurred in late August (0.15 mg  $L^{-1}$ ), early December (0.24 mg  $L^{-1}$ ), and mid-January, 1979 (0.26 mg  $L^{-1}$ ) and mid-May (0.29 mg  $L^{-1}$ ), (Figure 42). Low summer phosphate-

Figure 40.
Chloride (mg l<sup>-1</sup>) in the Ells, MacKay, Hangingstone,
Steepbank and Muskeg rivers.

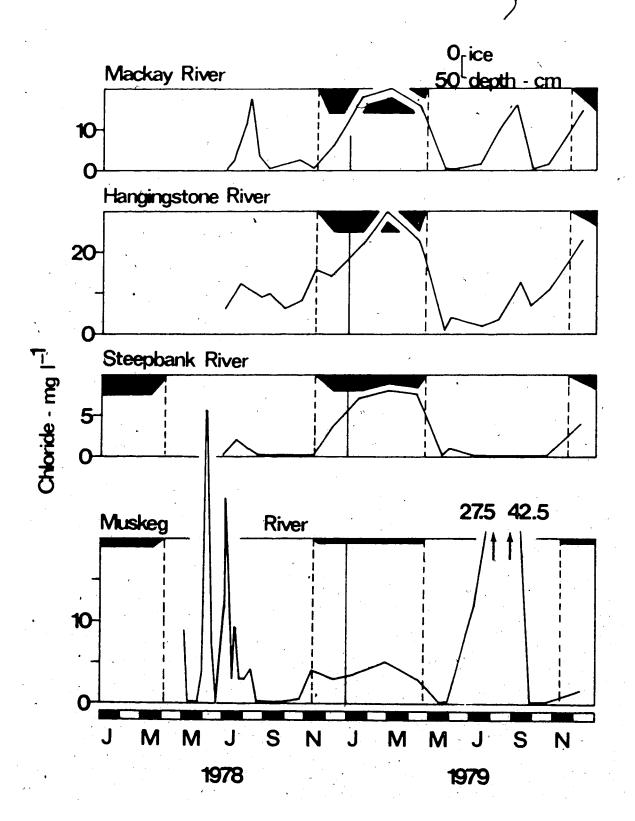


Figure 41. Nitrate-Nitrogen (mg  $1^{-1}$ ) in the Ells, MacKay, Hangingstone, Steepbank, and Muskeg rivers.

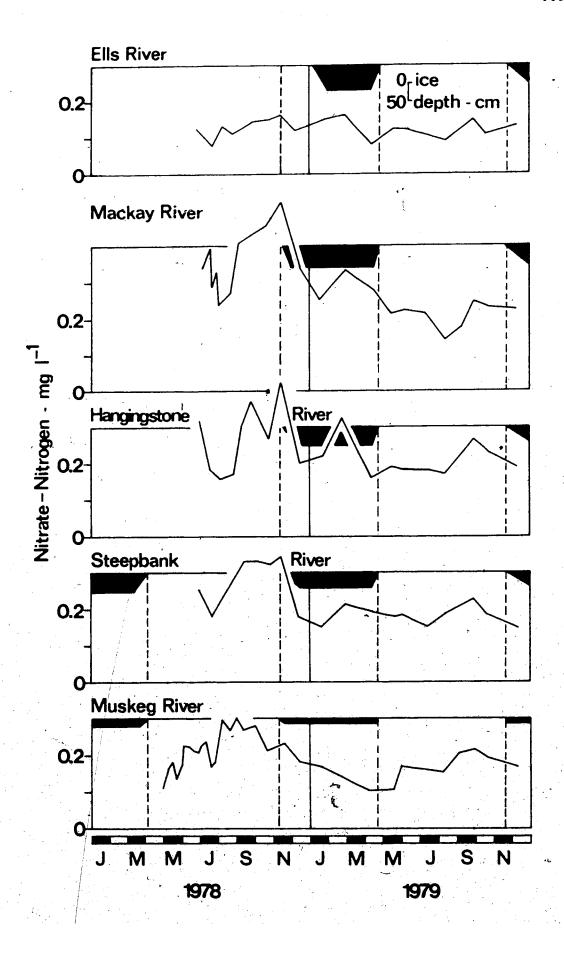
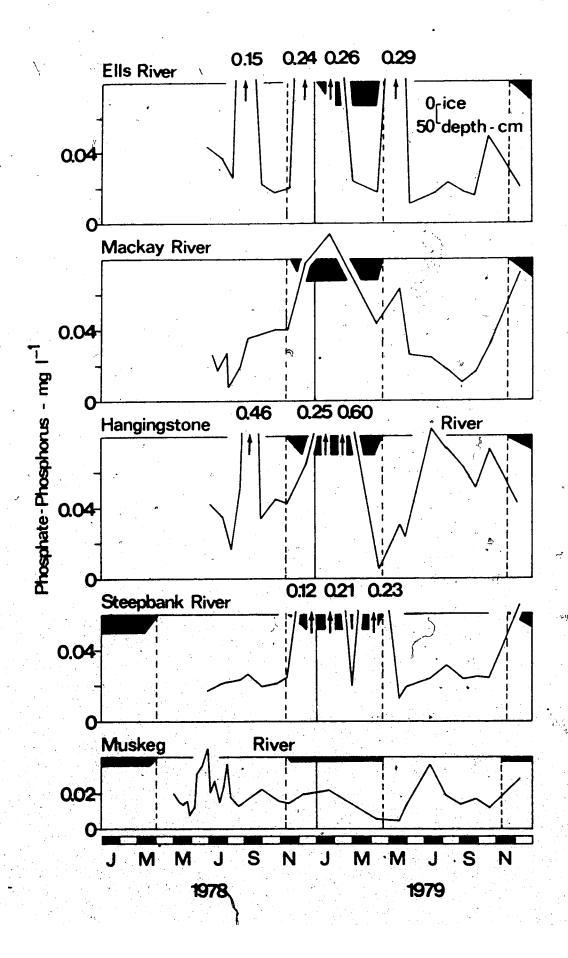


Figure 42.

Phosphate-Phosphorus (mg 1-1) in the Ells, MacKay,

Hangingstone, Steepbank and Muskeg rivers.



phosphorus values increasing to a mid-winter maximum of 0.094 mg  $L^{-1}$  occurred in the MacKay River. Large winter maxima occurred in the Hangingstone and Steepbank Rivers. In both a minimum occurred in May, however, this was followed by a dramatic increase to 0.084 mg  $L^{-1}$  during mid-July in the Hangingstone River only. The overall Hangingstone maximum was 0.60 mg  $L^{-1}$  and the Steepbank maximum was 0.23 mg  $L^{-1}$  also during the winter of 1979.

## 4.3.1.14 Dissolved silica

Dissolved silica fluctuations were quite similar in all but the Ells River (Figure 43). In the Ells, dissolved silica fluctuated the least with only slight fall peaks occurring. In the other rivers it increased to a winter maximum then declined with the onset of spring. (Figure 43). While mean dissolved silica levels were lower in the MacKay River than the Muskeg, Steepbank and Hangingstone rivers, the dissolved silica content of the Ells River was three times lower again. Despite the numerous differences concerning dissolved silica concentration the trend toward higher winter concentration was evident.

# 4.3.1.15 pH and alkalinity

In all rivers, pH varied from being acid to basic with maximum and minimum values occurring during the summer and winter months respectively (Figure 44). In the Ells River, pH ranged from 6.0 to 9.0 and this range was the widest among the five rivers.

Alkalinity was maximum in all but the Ells River during the winter period. Small peaks of alkalinity occurred during the early summer, in the five rivers (Figure 45). Alkalinity ranged from .80 meq  $L^{-1}$  in the Ells to a maximum of 5.3 meg  $L^{-1}$  in the Steepbank River.

Figure 43.

Dissolved Silica (mg l<sup>-1</sup>) in the Ells, MacKay, Hangingstone, Steepbank and Muskeg rivers.

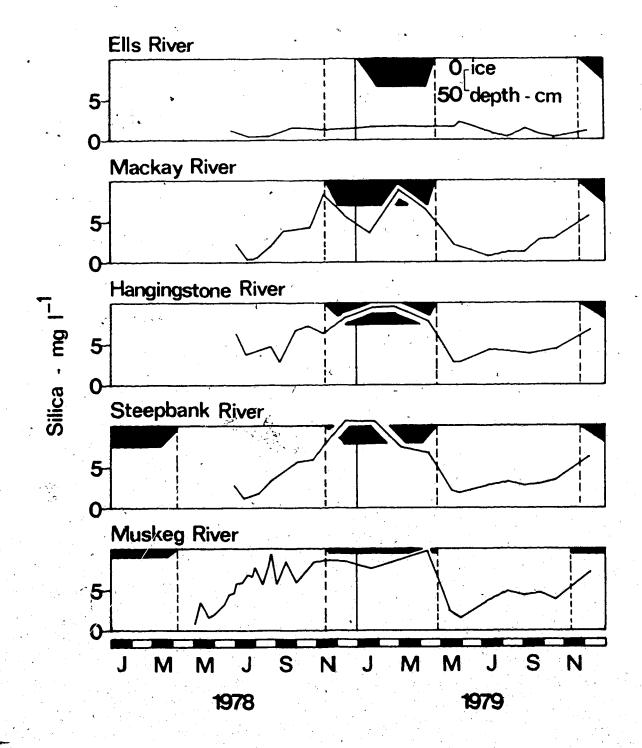


Figure 44.
pH in the Ells, MacKay, Hangingstone,
Steepbank and Muskeg rivers.

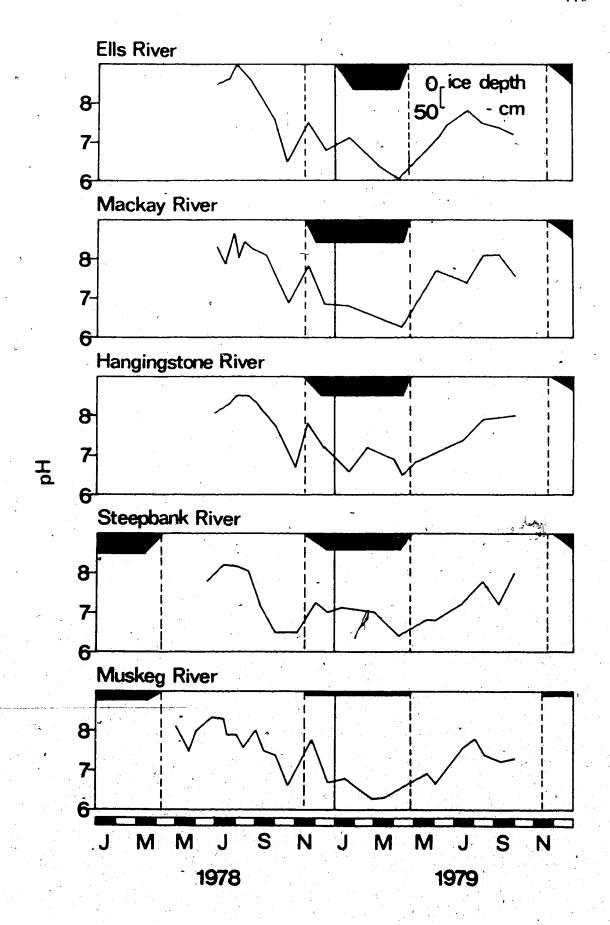
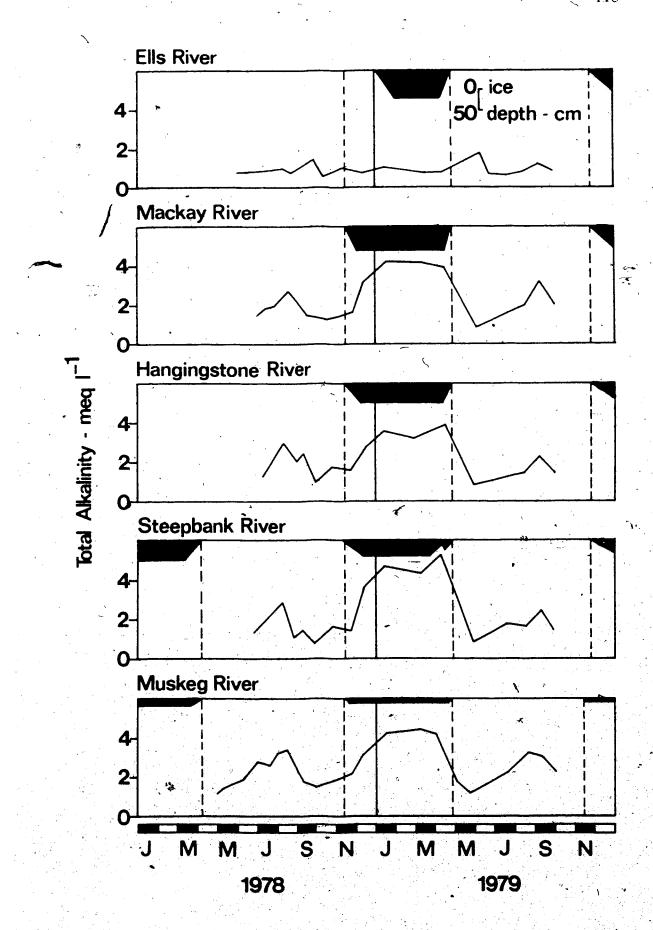


Figure 45.
Total Alkalinity (meg 1<sup>-1</sup>) in the Ells, MacKay, Hangingstone, Steepbank and Muskeg rivers.



## 4.3.2 Temporal Epilithic Algal Changes

## 4.3.2.1 Species Composition and Cell Numbers

## 4.3.2.1.1 Species Composition

Algae from four divisions dominated the epilithon in all five rivers. These were the Cyanophyta (blue-green algae), Chlorophyta (green algae), Bacillariophyta (diatoms), and Rhodophyta (red algae). A list of all the species encountered is presented in Table 18. Also indicated is whether they were found freely suspended in the river water (phytoplankton).

In the Muskeg River, Cyanophycean algae dominated with two species, Lyngbya aerugineo-caerulea and Phormidium sp., being the most important (Figure 46 and 47). During July 1978 this division accounted for more than 90% of the total epilithic community, decreasing to 53% by late August. Two other Cyanophycean algae, Calothrix breviarticulata and C. braunii, also contributed significantly at times (Figure 46). However, these species appeared to be more specific with respect to their ecological requirements. Diatoms were more prevalent during May and August through October 1978. Synedra ulna and Nitzschia fonticola were important during May and these, together with Gomphonema of vaceum and Synedra rumpens, during the fall period. During 1978, Chlorophycean algae were less important than the two divisions mentioned earlier, however, during September they accounted for 15.3% of the total community when Draparnaldia glomerata was present. Rhodophycean algae, Batra hospermum vagum and Audouinella violaceae, were also present, contributing significantly only during May, July and November of 1978. The Trmer species was most prevalent in May and November, and the latter during July.

Table 18. The complete list of algal species encountered in the five rivers (M = Muskeg River; SB = Steepbank River; HS = Hangingstone River; MK = MacKay River; E = Ells River). (+ = present; - = absent; = phytoplankton).

|  |                |                | <del></del>  |                |                  |
|--|----------------|----------------|--------------|----------------|------------------|
|  | M              | SB             | HS           | MK             | Ε                |
| CYANOPHYTA                                 | ,              |                |              | •              |                  |
| Anabaena affinis Lemm.                     | +              | +              | +            | +              | +                |
| A. inaequalis Borge                        | - '            | +              | -            | -              | -                |
| <u>A variabilis</u> Kütz.                  | +              | +              | +            | +              | 9                |
| A. wisconsinense Prescott                  | +              | -              | -            | -              | -                |
| Aphanizomenon flos-aquae (L.) Ralfs.       | +              | -              | - }          | <b>-</b>       |                  |
| Calothrix braunii Bornet & Flahault        | +              | · <b>+</b>     | + 4          | +              | +                |
| C. breviarticulata West & West             | +              | +              | -            | <b>-</b> .     | <u>-</u> `       |
| C. fusca (Kütz.) Bornet & Flahault.        | +              | <del>-</del> . | -            | -              | -                |
| Chamaesiphon incrustans Grunn.             | +              | +              | -            | -              | -                |
| Chroococcus limneticus Lemm.               | +17            | +              |              | +              | -                |
| Fischerella muscicola (Borzi) Gomont       | - ′            | +              |              | -              | -                |
| Gomphosphaeria aponina Kütz                | - '            | -              | <b>-</b> . ' | +              |                  |
| G. lacustris v. compacta Lemm.             | + ,            | -              | -            | -              | -                |
| Lyngbya aerugineo-caerulea (Kutz.) Gomont  | +              | + .            | +            | +              | +                |
| L. aestuarii (Mert.) Lieb. *               | -              | .+             | -            |                | -                |
| L. epiphytica Hieronymus                   | , <del>+</del> | +              | -            | -              | -                |
| L. nordgaardii Wille                       | +              | +              | -<br>-       | -              | -                |
| L. taylorii Drouet & Strickland            | +              | +              | <del>-</del> | -              | · · -            |
| L. versicolor (Watt.) Gomont               | <b>`</b> +     | +              | -            | ′ -            | . <del>-</del> . |
| Merismopedia elegans A. Braun              | +              | -              | -            |                | -                |
| M. glauca (Ehr.) Naegeli                   | +              | -              |              | +              | •                |
| Microcoleus vaginatus (Vauch.) Gomont      | -              | + .            | -            |                | -                |
| Microcystis aeruginosa Kütz. emend Elenkin | +              | +              | -            | -              | -                |
| Nostoc commune Vaucher                     | .+             | · +            | +            | +              | . + ,            |
| N. microscopicum Carmichael                | +              | +              | +            | • +            | + 1              |
| N. verrusosum Vaucher                      | +              | +              | +            | +              | +                |
| Nostoc sp.                                 | +              | +              | +            | +              | +                |
| Oscillatoria amphibia C.A. Agardh.         | -              | -              |              | +              |                  |
| O. lacustris (Kleb.) Geitler               | +              | +              | _            | . <del>-</del> |                  |
|  |                |                |              | 100            |                  |

| Table to. Continued. | Table | 18. | Continued. |
|----------------------|-------|-----|------------|
|----------------------|-------|-----|------------|

|  | M              | · SB         | HS ,       | MK         | Ε              |
|--|----------------|--------------|------------|------------|----------------|
| O. tenuis C.A. Agardh.                               | + .            | +            | _          | _          | _              |
| Oscillatoria sp.                                     | +'             | +            | _          |            | · +            |
| Phormidium favosum (Bory) Gomont                     | +              | +            | • .<br>-   | _          | _              |
| P. tenue (Menegh.) Gomont                            | +              | +            | _          | _          | _              |
| Rhaphidiopsis sp.                                    | _              | _            | -          | _          | +              |
| Rivularia haematites (D.C.) C.A. Agardh.             | , <del>t</del> | +            |            | _          | -              |
| Schizothrix tinctoria Gomont                         | +              | +            | -          | ,<br>,     | _              |
| Tolypothrix distorta Kütz.                           | _              | +            | _          |            | <b>-</b> ,     |
|  |                |              |            |            | ,              |
| CHLOROPHYTA  |                |              |            |            |                |
| Ankistrodesmus falcatus (Corda) Ralfs.               | +              | +            | + ;        | +          | +              |
| A. spiralis (Turner) Lemm.                           | +              | . <b>-</b>   | _          | <u>.</u>   | -              |
| C. globosa Snow                                      | -              | • •          |            | +          | -              |
| Chaetophora incrassata (Hud.) Hazen                  | +              | <b>+</b> ,   | _          |            | -              |
| Chlamydomonas sp.                                    | +              | +            | +          | +          | +              |
| Cladophora glomerata (L.) Kütz.                      | + .            | +            | +          | +          | +              |
| Chlorella ellipsoidea Gerneck                        | _              | -            | ·<br>      | + .        | -              |
| C. vulgaris Beyer                                    | + 1            | +            | +          | +          | + .            |
| Closterium sp.                                       | +              | <b>*</b>     | +          | , <b>+</b> | + ,            |
| Coelastrum scabrum Reinsch                           | +              |              | <u>.</u>   | 4          | -              |
| Coleochaete divergens Pringscheim                    | +              | -            |            | -          | <b>-</b> '.' . |
| Cosmarium sp.  | +              | +            | <b>-</b>   | +          | -              |
| Crucigenia quadrata Morren                           | +              |              | -          |            | <b>-</b> ,     |
| C. tetrapedia (Kirch.) West & West                   | -              | +            | -          | -          | <b>-</b> ;     |
| <u>Dictyosphaerium</u> <u>ehrenbergianum</u> Naegeli | +              | +_           |            | -          | -              |
| D. pulchellum Wood                                   | +              | +            | -          | -          | + 10           |
| Draparnaldia acuta (C.A.Ag.) Kütz.                   | +              | •            | _          | -          |                |
| D. glomerata (Vauch.) C.A. Ag.                       | +              | +            | -          | 1 · · · ·  | <b>-</b> ,     |
| Elaktothrix sp.                                      | · <b>-</b>     | +            | . <b>.</b> | -          | <u>.</u>       |
| Gloeocystis gigas (Kutz.) Lager                      | -              | -            | _          | . <b>+</b> | _              |
| Hyalotheca sp.                                       | _              | , <b>+</b> , | _          | • .        | -              |
| Mougeotia sp.  | +              | +            | •          | . ,        | ÷ .            |
|  | <i>Y</i> .     | ,            |            |            |                |

| TOUR TOUR CONTENTIACE | Table | 18. | Continued. |
|-----------------------|-------|-----|------------|
|-----------------------|-------|-----|------------|

| Table 18. Continued.                        | M            | SB         | HS           | MK          | E              |
|---|--------------|------------|--------------|-------------|----------------|
|   |              |            | 113          | TIK         |                |
| ,   | <del></del>  |            | r            |             | <del></del>    |
| Microspora loefgrenii (Nordst.) Lager       | -            | +          |              | ٠ + ،       | -              |
| M. pachyderma (Wille) Lager                 | -            | -          | -            | -           | +              |
| Oedogonium sp.                              | +            | . <b>+</b> | +            | -           | +              |
| Pediastrum biradiatum Meyer                 |              | + _        | -            | -           | -              |
| P. biradiatum v. emarginatum                |              |            |              |             | •              |
| f. convexum Prescott                        | · -          | -          |              | + ,         | -              |
| P. boryanum (Turp.) Meneghini               |              | -          | -            | -           | +              |
| <u>Pithophora</u> varia Wile                | +            | +          |              |             | -              |
| Pleurotaenium spp.                          | -            | -          | +            | -           | -              |
| Rhizoclonium hieroglyphicum (C.A. Ag.) Kütz | +            | +          | -            | -           | · - 🐃          |
| Scenedesmus acutiformis Schroeder           | -            | -          | · -          | + _         | <del>-</del> . |
| S. bijuga (Turp.) Lager                     | + .          | _ '        |              | + _         | - 1            |
| S. dimorphus (Turp.) Kütz.                  | -            |            | -            | -           | +              |
| S. obliguus (Ţurp.) Kütz.                   | .+           | <b>_+</b>  | -            | +           | • -            |
| S. quadricauda (Turp.)de Breb.              |              | - ,        | <del>-</del> | +           | . –            |
| Sorastrum spinulosum Naegeli                | . =          | -          | -            | +           | -              |
| Sphaerocystis schroeteri Chodat             |              | •          | -            | +           | -              |
| Sphaeroplea annulina (Roth.) C.A. Agardh.   | -            | <b>±</b>   | -            | +           | <u>-</u>       |
| Spirogyra sp.                               | +            | +          |              | +           |                |
| Stigeoclonium sp.                           | +            | +          | 5. <b>+</b>  | +           | <u>.</u> + .   |
| S. pachyderm Prescott                       | *** <b>+</b> | +          | -            | . <b>-</b>  | _              |
| Staurastrum sp.                             | . <b>-</b>   | +          | +            | · <u>-</u>  | - 4            |
| Tetraedron *asymmetricum \Prescott          | -            | + -        | -            | •           | -              |
| <u>Ulothrix</u> sp.                         | _+           | .+         | +            | * * *       | . + .          |
| U. subconstricta G.S. West                  | +            | +          | -            | -           | -              |
| U. subtilissima Rabenhorst                  | <b>-</b> , " | <b>,</b> + | -            | <u> -</u> . |                |
| U. zonata (Weber & Mohr) Kutz.              | + .          | + '        | , <b>-</b>   | -           | -              |
| Zygnema sp.                                 | +            |            | -            | •           |                |
| •   |              |            |              |             |                |
| RHODOPHYTA                                  |              |            |              |             |                |
| Batrachospermum vagum (Roth.) C.A. Agardh.  | +            | + .        |              | -           | -              |
| Audouinella violacea (Kütz.) Hamel          | +            | , +        | -            | -           | _              |

| Table | 18. | Continued. |
|-------|-----|------------|
|       | ٠   | •          |
|       |     |            |

|                                      | М        | SB            | нs         | MK             | E      |
|--------------------------------------|----------|---------------|------------|----------------|--------|
|                                      |          |               |            |                |        |
| A. <u>Pygmaea</u> Kütz.              | +        | +             | -          | -              | -      |
| EUGLENOPHYTA                         |          |               | a*         |                | ,      |
| Phàcus sp.                           | - `      | . +           | <u>-</u>   | <sup>.</sup> . |        |
| CHRYSOPHYTA                          | •        |               |            |                |        |
| Dinobryon sertularia Ehr.            | -        | -             | -          | + _            | -      |
| Mallomonas caudata Iwanoff           | +        | +             | +          | +              | +      |
| СКУРТОРНУТА                          |          |               |            |                |        |
| Cryptomonas erosa Ehr.               | +        | + :           | -          |                | -      |
| C. ovata Ehr.                        | +        | +             | _          | -              | -      |
| •                                    |          |               |            |                |        |
| BACILLARIOPHYTA                      |          |               |            | ٠.             | •      |
| Achnanthes lanceolata Breb.          | +        | <b>;</b> +    | +          | +              | +      |
| A. lanceolata v. rostrata Hust.      | +        | +             | +          | +              | + .    |
| A. minutissima Kütz.                 | + ,      | $\mathcal{A}$ | +          | +              | + ,    |
| A. peragallii Brun & Herbaud         |          | •             | -          | . +            | +      |
| Amphipleura lindheimeri Grun         | -        | - ·           | . <b>.</b> | +              | +      |
| A. pellucida Kütz.                   | +-       | +             | + •        | +              | +      |
| Amphora ovalis Kütz.                 | +        | +             | . 🕳        | _              | + .    |
| A. perpusilla Grun.                  | -        | -             | <b>-</b> , | •              | +      |
| Asterionella formosa Hass.           | <b>}</b> | +_            | · <u>-</u> | -              | +_     |
| Caloneis alpestris (Grun.) Cl.       | + •      | _             |            | -              | -<br>- |
| Cocconeis pediculus Ehr.             | + .      | +             | +          | +              | + ,    |
| C. placentula Ehr.                   | +        | +             | +          | +              | +      |
| C. placentula v. euglypta (Ehr.) Cl. | +        | + 1           |            | <b>-</b> .     | -      |
| Cyclotella catenata Brun.            | •        | <b></b> .     | +          |                | -      |
| C. comta (Ehr.) Kütz.                | -        | . +           |            | +              | +      |
| C. kutzingiana Thwaites              | -        | -             | +          | -              | · -    |
| C. meneghiniana Kütz.                | +        | +             | ° +        | +              | +      |
| Cymatopleura solea (Breb.) W.Sm.     | +        | -             | +          | -              | +.     |

Table 18. Continued.

| ,                                    | M              | SB   | .HS              | . MK                                  | E ·            |
|--------------------------------------|----------------|--|------------------|---------------------------------------|----------------|
|                                      |                | <del></del>                                    | <del></del>      | <del></del>                           |                |
| Cymbella amphioxys (Kütz.) Grun.     | -              | -  | •                | ·<br>-                                | <b>,</b> +     |
| C. cistula (Hemprich) Grun.          | +              | +  | +                | -                                     | <b>*</b> +*    |
| C. lanceolata (Ehr.) V.H.            |                | -  | -                | +                                     |                |
| C. naviculiformis Auerswald          | +              | <b>-</b>                                       | -                | <u>-</u> ,                            | +              |
| C. prostrata (Berkeley) Cl.          | +              | , <b>+</b>                                     | · +              | -                                     | +              |
| C. sinuata Greg.                     | +              | +  | +                |                                       | +              |
| C. tumida (Breb.) V.H.               |                | -  | · +              | -                                     | +              |
| C. turgida (Greg.) Cl.               | +              | -  |                  | -                                     | <b>+</b>       |
| C. ventricosa Kütz.                  | +              | +  | +                | +                                     | +              |
| Diatoma elongatum Agardh.            | +              | +  | +                | +                                     | +              |
| D. anceps (Ehr.) Grunn.              | +              | -  | . <b>-</b>       | -                                     | - 🐧            |
| D. vulgare Bory                      | +              | +  | +                | +                                     | + 1            |
| D. vulgare v. grandis (Smith) Grun.  | +              | +  | +                | +                                     | +              |
| D. vulgare v. ovalis (Fricke) Hust.  | ·<br>•, ·      | -  |                  | <u>.</u> .                            | +.             |
| D. vulgare v. producta Grun.         | " <b>-</b>     | -  | -                | +                                     | +              |
| Epithemia argus Kūtz.                | +              |  | -                | -                                     | -              |
| E. sorex Kütz.                       | +              | ., +   | +                | + ,                                   | +              |
| E. turgida, (Ehr.) Kūtz.             | +              | +  |                  | -                                     | -              |
| E. turgida v. granulata (Ehr.) Grun. | + .            | <b>.</b> • • • • • • • • • • • • • • • • • • • | , + <sup>1</sup> | +                                     | + 1 .          |
| E. zebra (Ehr.) Kütz.                | -              | +  |                  | •                                     | •              |
| Eunotia Zunaris (Ehr.) Grun.         | + .            |  | -                | •                                     | - *            |
| E. pectinalis v. minor (Kütz.) Rabh. | - '            | · <b>-</b>                                     | , <b>-</b>       | + .                                   | * · · <u>·</u> |
| E. valida Hust.                      | , <del>-</del> | _  | -                | +                                     | <b>-</b>       |
| Fragilaria capucina Desm.            | +-             | +  | +                | <del></del>                           | +              |
| F. capucina v. acuta Grun.           | _              |  | -                | •                                     | <b>+</b>       |
| F. capucina v. Lanceolata Grun.      | . <del>-</del> | -  | -                | , • .                                 | +*             |
| F. construens (Ehr.) Grun.           | +              | +  | +                | • • • • • • • • • • • • • • • • • • • | +              |
| F. construens v. venter (Ehr.) Grun. | +              | + ,  | + '              | +                                     | +              |
| F. crotonensis Kitton                | -              | +_   | +_               | •                                     | -              |
| F. <u>l'eptostauron</u> (Ehr.) Hust. | -<br>          | •  | <del>-</del> -   | -                                     | +              |
| F. pinnata' Ehr.                     | -              | -  | +                | •                                     | +              |
| F. vaucheriae (Kütz.) Boye Pet.      | •              | +  | +                |                                       | +              |
|                                      |                | ,  |                  |                                       |                |

| Table | 1Ω  | Continued. |
|-------|-----|------------|
| lable | 18. | Continued. |

| ,   |                | ,  |                                     |                    | _            |
|---|----------------|--|-------------------------------------|--------------------|--------------|
|   | M              | SB                                       | HS .                                | MK                 | Ε            |
|   | <del></del>    | <del></del>                              |                                     | <del></del>        |              |
|   |                | •  |                                     |                    |              |
| F. virescens v. capitata Krasske          | -              | +  |                                     | +                  | -            |
| Frustulia rhomboides v. amphipleuroides   |                |  |                                     | e e                |              |
| Grun.                                     |                | +,                                       | • -                                 | -                  | + '          |
| Gomphonema acuminatun Ehr.                | +              | +  | -                                   | ₹ ,                | <b>~</b>     |
| G. acuminatum v. coronata (Ehr.) W.Sm.    | -              | -  | +                                   |                    | - 8          |
| G. abbreviatum (Agardh.) Kütz.            | +              |  | + \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ | . <del></del>      | +            |
| G. angustatum v. producta Grun.           | 7              | -  | -                                   | -                  | · +          |
| G. bohemicum Reichelt & Fricke            | +              | +  | + '                                 | +                  | +            |
| G. constrictum Ehr.                       | + '            | -  | - ,                                 | -                  | -            |
| G. gracile Ehr.                           | -              | <b>+</b> . ,                             | -                                   | -                  | . •          |
| G. lanceolatum Ehr.                       | + .            | +  | +                                   | +                  | +            |
| G. longipes v. subclavata Grun.           | -              | +  | +                                   | 7                  | +            |
| G. olivacium (Lyngb.) Kütz.               | +              | +  | +                                   | +                  | , <b>+</b> , |
| G. parvulum Kütz.                         | +              | +  | +                                   | +                  | +            |
| G. parvulum v. exilis Grun.               | +              | +  | +                                   | +                  | +            |
| G. ventricosum Greg.                      | , <b>+</b>     | -  |                                     | <b>-</b> ,         | -            |
| Gyrosigma acuminatum Kūtz.                | + `            | +  | +                                   | -                  | ;· +         |
| Hantzschia amphioxys f. capitata 0. Mull. | -              | -  | <b>-</b> ·                          | +                  | +            |
| Melosira granulata (Ehr.) Ralfs.          | - ,            | <b>-</b> ,                               |                                     | -                  | +            |
| M. islandica O. Mull.                     | + .            | +  | +                                   | 1+                 | + .          |
| M. varians C.A. Agardh.                   | +              | +  | +                                   | +                  | -            |
| Meridion circulare Agardh.                | +              | +  | +                                   | •                  | +            |
| Navicula bacilliformis Grun.              | -              | <u>-</u>                                 | _                                   | _                  | +            |
| N. cryptocephala Kütz.                    | +              | +  | +                                   | +                  | +            |
| N. cuspidata Kütz.                        | +              | +  | _                                   |                    | + 1          |
| N. dicephala (Ehr.) W.Sm.                 | , <del>-</del> | _  |                                     | + ,                |              |
| N. gracilis Ehr.                          | +              | +  | •                                   | +                  | +            |
| N. graciloides A. Mayer                   | +              | +  | . <b>+</b>                          | -                  | +            |
| N. hungarica v. capitata (Ehr.) Cl.       | _              |  | -                                   | <br>. <del>.</del> | +            |
| N. lapidosa Krasske                       |                | in i | +                                   | _                  |              |
| N. minima v. atomoides (Grun.) Cl.        | _              | 1_                                       |                                     |                    | +            |
| N. placentula (Ehr.) Grun.                |                | +  | _                                   | +                  | +            |
| Fragation of Chit of City                 |                |  | _                                   | •                  | •            |

| Table | 18. | Continued. |
|-------|-----|------------|
|       |     | CONCINUCA. |

|  |       |              |                |        | <del></del>                           |
|--|-------|--------------|----------------|--------|---------------------------------------|
| N. placentula v. rostrata A. Meyer       | -     | _            | + .            | +      | +                                     |
| N. pupula Grun.                          | +     | +            | -              | -      | -                                     |
| N, pupula v. rectangularis (Greg.) Grun. | -     | _            | +              | -      | _                                     |
| N. radiosa Kütz.                         | +     | +            | +              | ·<br>+ | +                                     |
| N. rhychocephala Kütz.                   | +     | +            | -              | +,     | -                                     |
| N. scoliopleuroides Quint                | -     | <b>-</b> .   |                | •      | +                                     |
| Neidium affine (Ehr.) Cl.                | -     | _            | -              | +      | -                                     |
| N. affine v. amphirhynchus (Ehr.) Cl.    | -     | -            | -              | +      |                                       |
| Nitzschia acicularis W.Sm.               | +     | -            | -              | _      | -                                     |
| N. acuta Hantzsch.                       | _     | <u> </u>     | +              | · _    | -                                     |
| N. amphibia Grun.                        | +     | +1           | ·<br>•         | · ·    | -                                     |
| N. clausii Hantzsch.                     | _     | - }          | -              | +      | _                                     |
| N. commutata Grun.                       |       | +            | -              | -      | <b>-</b>                              |
| N. fonticola Grun.                       | + '   | +            | <b>-</b> ,     | _      | +                                     |
| N. gracilis Hantzsch.                    | + ,   | +            | <del>.t</del>  | +      | +                                     |
| N. hantzschiana Rabh.                    | -     | +            | -              | -      | <b>.</b>                              |
| N. heurfleriana Grun.                    |       |              | -<br>-         | ·<br>• | +                                     |
| N. ignorata                              |       | •            |                |        | +                                     |
| N. palea (Kütz.) W.Sm.                   | - '   | +            | ·              | +      | +                                     |
| N. paleacea Grun.                        |       |              | -              | -      | +                                     |
| N. recta Hantzsch.                       | _     | _            | +              |        |                                       |
| N. romana Grun.                          | -     | -            | <del>-</del> , |        | 4                                     |
| N. sublinearis Hust.                     | +     |              | -              | +      | •                                     |
| Opephora martyi Heribaud                 | - · · |              | -              | _      | +                                     |
| Pinnularia gibba Ehr.                    | +     |              | +              | +      |                                       |
| P. mesolepta (Ehr.) W.Sm.                | +     | +            | , <b>+</b>     | -      |                                       |
| P. molaris Grun.                         |       | <del>,</del> | +              | _      | • • • • • • • • • • • • • • • • • • • |
| P. nodosa v. constricta Mayer            | -     | •            | ·<br>• ·       | -      | +                                     |
| P. viridis v°. sudetica (Hilse) Hust.    |       | -            | -              | +      | _                                     |
| Rhoicosphenia curvata (Kütz.) Grun.      | +     | + 3          | +              | +      | +                                     |
| Rhopalodia gibba (Ehr.) O. Mull.         | +     | +            | +              | +      |                                       |
| R. gibberula (Ehr.) O. Mull.             | +     | +            | +              | +      |                                       |
|  |       |              |                |        |                                       |

Table 18. Concluded.

| М                                       | SB           | HS                                      | MK         | E                                     |                |
|---|--------------|---|------------|---------------------------------------|----------------|
|   | · · · · · ·  |   |            |                                       |                |
| R. parallela (Grun.) O. Mull.           | -            | +                                       | +          | +                                     | -              |
| Stauroneis anceps Ehr.                  | -            | · ·                                     | -          | <b>+</b> .                            | -              |
| S. phoenicenteron Ehr.                  | -            | -                                       | -          | -                                     | +              |
| S. legumen Ehr.                         | +            | · -                                     | 7          | -                                     | -              |
| Stephanodiscus astraea (Ehr.) Grun.     | -            | -                                       | £ +        | <b>+</b>                              | + _            |
| S. hantzschii Grun.                     | +            | <u>.</u> +                              | +          | + _                                   |                |
| <u>Surirella</u> <u>angustata</u> Kütz. | -            | -                                       | +          | +)                                    | +              |
| S. didyma Kutz.                         | -            | -                                       | , <b>-</b> | -'                                    | +              |
| S. delicatissima Lewis                  | -            | -                                       | `-         | . +                                   | -              |
| Surirella linearis v. helvetica         |              |   |            |                                       |                |
| (Brun.) Meister                         | +            | -                                       |            | <u>-</u>                              | -              |
| S. ovalis Breb.                         | -            |   | -          | +                                     | · <del>=</del> |
| S. robusta v. splendida (Ehr.) V.H.     | -            | ** -                                    | -          | +                                     | +              |
| S. tenera Greg.                         | , , <b>-</b> | -                                       | -          | -                                     | +              |
| Synedra cyclopum Brutschii              | -            | -                                       | -          | -                                     | +              |
| S. capitata Ehr.                        | +            | •                                       | ·          |                                       | <u></u>        |
| S. pulchella Kütz.                      | +            | +^                                      | الم        |                                       | <del>-</del> , |
| S. rumpens Kutz.                        | +            | Ý                                       | -          | -                                     | +              |
| S. rumpens v. familiaris (Kutz.) Grun.  | +            | +                                       | -          | • • • • • • • • • • • • • • • • • • • | +              |
| S. ulna (Nitzsch.) Ehr.                 | +            | +                                       | +          | + .                                   | +              |
| Tabellaria fenestrata (Lyngby.) Kütz.   | +            | , · · · · · · · · · · · · · · · · · · · | +          | +                                     | <b>.</b>       |
| T. flocculosa (Roth.) Kütz.             | + ]          | _                                       | -          | -                                     | <b></b> ****** |
|   |              | •                                       |            |                                       |                |

Figure 46.
Seasonal succession of the dominant epilithic algae in the Muskeg River.

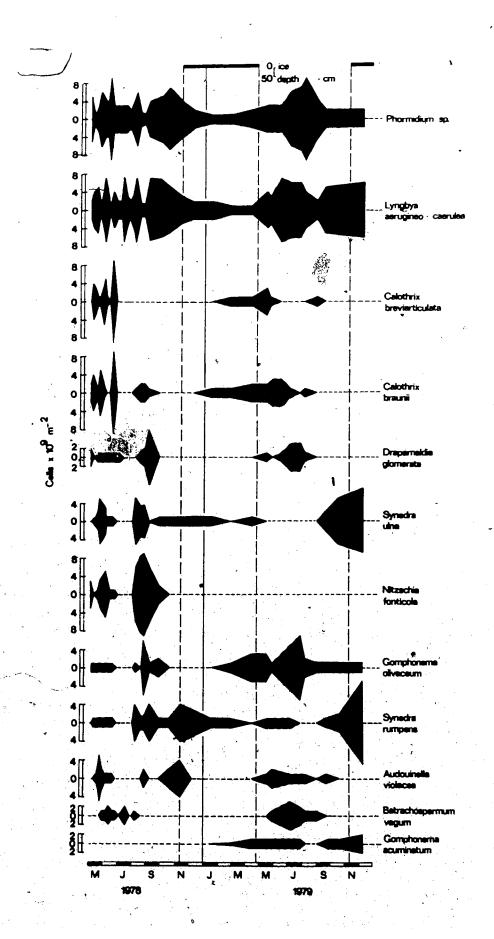
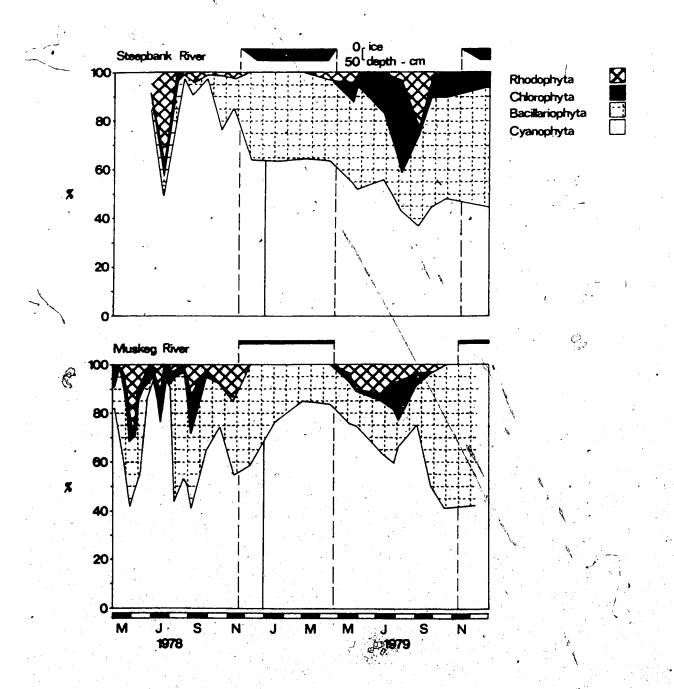


Figure 47.
Seasonal changes in the percentage composition of the epilithon in the Steepbank and Muskeg rivers.

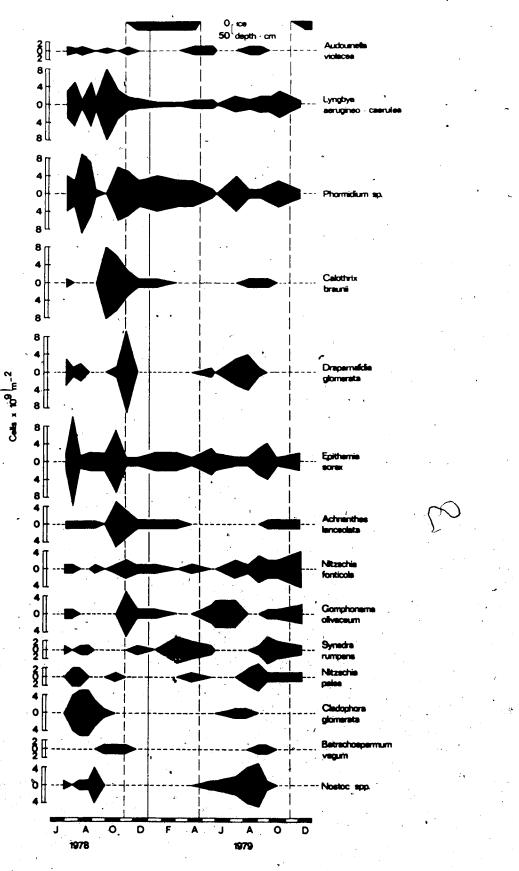


From December 1978 until April 1979 only Cyanophycean and Bacillariophycean algae were found. Two species of Calothrix began growth in early December and slowly increased in numbers during the winter to peak during May and early June 1979. Also, both Lyngbya aerugineo-caerulea and Phormidium sp. were present and these too began increasing in numbers, under ice cover, during March. The winter diatom community comprised mainly Synedra ulna, Gomphonema oliaceum (after January, 1979), Synedra rumpens and again after January, as well as for the first time, Gomphonema acuminatum (Figure 46).

From May 1979 Cyanophycean algae decreased in importance from 75% to 59.7% by late July. After an increase during August and early September, corresponding to large Phormidium and Lyngbya populations, further decreases to 41% by mid-October occurred. Diatoms continued to constitute the second major algal group from May to late July 1979 and again from early September until early December when they accounted for 56% of the benthic population. Gomphonema olivaceum, particularly, was important from May to late July, while Synedra ulna and Synedra rumpens dominated from early September onwards. Rhodophycean algae were again present during 1979. Audouinella violaceae peaked in late May and Batrachospermum vagum in mid-July. From May to early July, Rhodophycean algae were less conspicuous accounting for 10 to 12% of the total populations. Chlorophycean algae were less conspicuous with Draparnaldia glomerata the most important species during August. (Figure 46 and 47).

Eyanophycean algae were also dominant in the Steepbank River (Figures 47 and 48). Their importance, however, declined from a contribution of 85% of the total population during November 1978 and

Figure 48.
Seasonal succession of the dominant epilithic algae in the Steepbank River:



Phoritidium sp. were again the most important species (Figure 48). A large population of Calothrix braunii developed during the autumn and early winter of 1978, but during 1979 the populations of these three algae never reached the 1978 levels. Increasing from a minimum of 49.5% in mid-July of 1978 the Cyanophycean algae attained a peak contribution of 98% by mid-August. In contrast, levels never exceeded 64.7% during 1979 (March) and by early December they had fallen to 44.5%.

During 1978 in the Steepbank River, Audouinella violaceae and Batrachospermum vagum were important, also peaking in mid-July (39.3%). Both species were present during 1979, however, they contributed slightly less and they appeared later in the year (August - September). Diatoms were important and steadily increased as the Cyanophycean algae" decreased. Epithemia sorex was most prevalent during July and late autumn/early winter 1978. Achnanthes lanceolata peaked along with Epithemia during this latter period. Then, as ice formed Nitzschia fonticola and Gomphonema olivaceum peaked. All these populations however, decreased before March when numbers of Synedra rumpens increased. Peak diatom contributions occurred during 1978 October through December, averaging 24.3% of the total epilithic population. Levels were much higher during 1979, reaching 49.8% by early December. Spring 1979 diatom populations were dominated by Epithemia sorex and Gomphonema olivaceum. Then, from a low of 15.3% in August, diatom contributions increased as Epithemia sorex, Nitzschia fonticola, Gomphonema olivaceum, Synedra rumpens and Nitzschia palea became dominant. Chlorophycean algae were also more prevalent during 1979, particularly from July to August when large populations of Draparnaldia

glomerata and Cladophora glomerata developed (figures 47 and 48) contributing 28 to 38% of the benthic community.

Cyanophycean algae were the most important algal group in the MacKay River (Figure 49), particularly during 1978. However, during the following year diatoms assumed a far greater importance, accounting for 100% of the community on June 2 and December 9 (Figure 49). The epilithon of the MacKay River was also dominated by far fewer algal species than the other four rivers (Figure 50). Lyngbya aerugineo caerulea, Calothrix braunii and Anabaeana affinis were the dominant Cyanophycean algae during 1978 (Figure 50), Cladophora glomerata and Chlamydomonas spp., the dominant Chlorophycean algae; and Epithemia sorex the dominant diatom. During 1978, Lyngbya and Calothrix maxima coincided during mid-July when Cyanophycean algae accounted for 99.3% of the total epilithic algal community, whereas, during 1979 only the spring peaks coincided. The summer peak of Calothrix occurred during mid July while that of Lyngbya did not occur until early September. At each time, Cyanophycean algae accounted for 92.9% and 55.6% of the total -community. Chlorophycean algae constituted 47.3% of the total community in early August 1978, corresponding to populations of Chlamydomonas spp. and Cladophora glomerata.

The only other major contribution occurred under ice-cover in April (47.4% Cladophora glomerata). From here, no one particular Chlorophycean alga dominated and as a group occurred only in small numbers. Diatoms accounted for a far greater proportion of the epilithon during 1979, particularly because of Epithemia sorex which had a restricted distribution during 1978. In contrast, during 1979 it was present on all but one sampling date in far greater numbers.

Figure 49.
Seasonal changes in percentage composition of the epilithon in the MacKay and Hangingstone rivers.

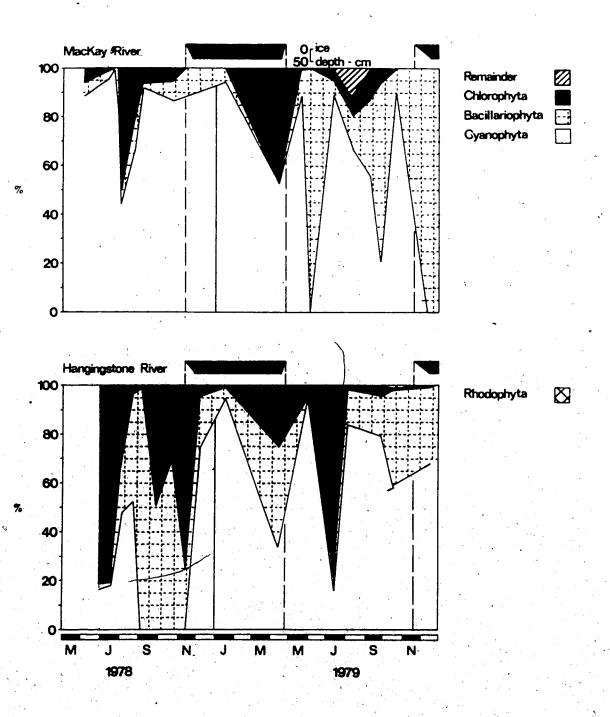
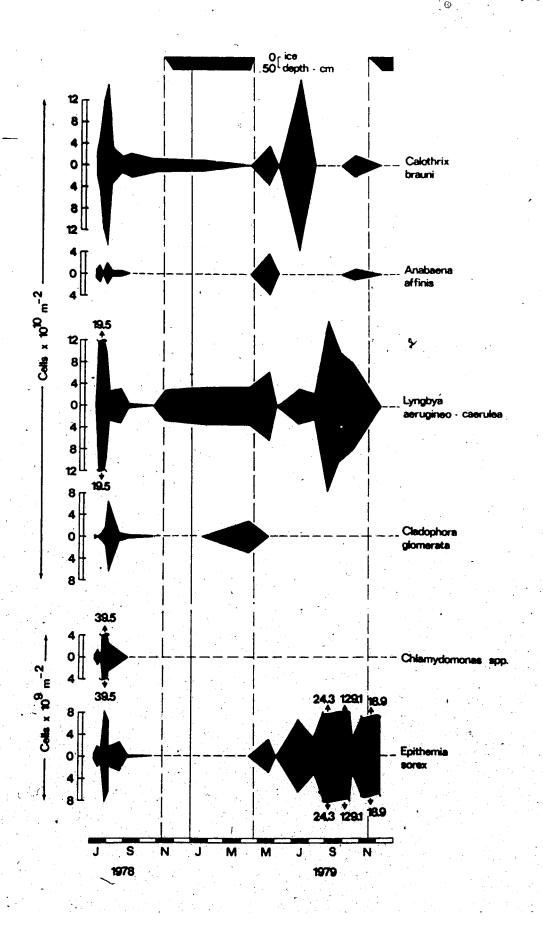


Figure 50.
Seasonal succession of the dominant epilithic algae in the MacKay River.

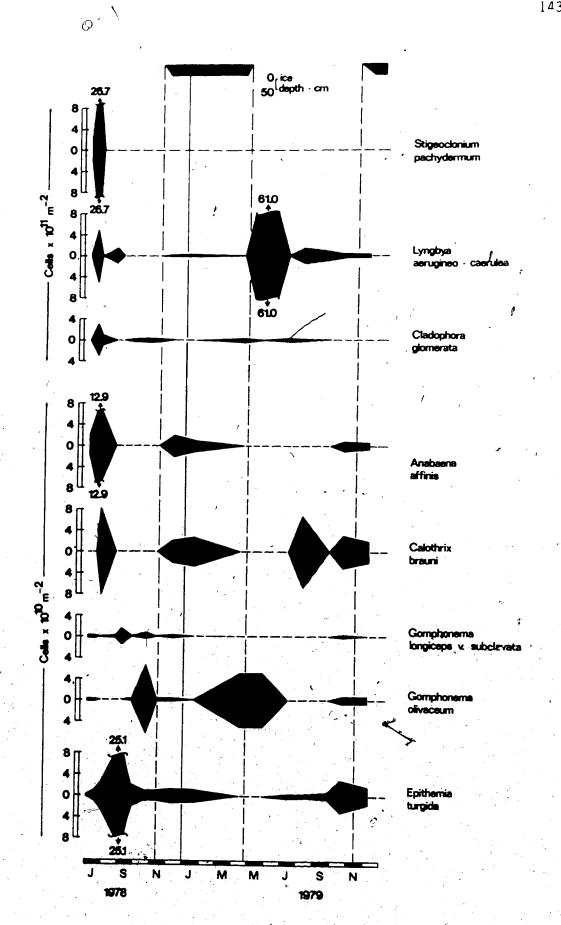


In the Hangingstone River Cyanophycean algae were not as abundant as in the other four rivers during 1978 (Figure 49). Initially, Chlorophycean algae accounted for 81% of the total population, with Stigeoclonium pachydermum being most important (Figure 51). However, after mid-July this alga occurred rarely. By early August, Cyanophycean algae had increased to 47.9%, reaching the 1978 maximum of 52.3% at the end of this month. Anabaena affinis, Calothrix braunii, and Lyngbya aerugineo-caerulea were the dominant Cyanophycean algae. From late August until November no Cyanophycean algae were found. Instead, by early September diatoms acounted for 98.8% of the total community (mainly Epithemia turgida). Dietem contribution thereafter decreased, reaching a low of 4.3% in mid-January. Concomitant with this decrease, Chlorophycean algae (mainly <u>Cladophora</u> glomerata) increased in importance, and also Cyanophycean algae (from November 1978) when Anabaena affinis and Calothrix Braunii reappeared. By mid-April diatoms were the most important (40.3% - mainly Gomphonema olivaceum) followed by Cyanophycean and Chlorophycean algae; 33.2% and 26.5% respectively. However, by the end of May the Cyanophycean algae accounted for 93.9% of the total community when Lyngbya aerugineo-caerulea dominated. Chlorophycean algae dominated during July (77.3% - mainly Chlorella Thereafter they were quite insignificant with Cyanophycean vulgaris). algae (mainly <u>Calothrix</u>) and diatoms (mainly <u>Epithemia</u> <u>turgida</u>) dominating.

Cyanophycean algae in the Ells River dominated on all but two occasions (Figure 52 and 53). The dominant species were Lyngbya aerugineo-caerulea, Calothrix braunii and Anabaena affinis. Lyngbya was always present producing its largest populations during autumn 1978 and

Figure 51. Seasonal succession of the dominant epilithic algae in the Hangingstone River.

1



7 /

Figure 52.
Seasonal changes in the percentage composition of the epilithon in the Ells River.

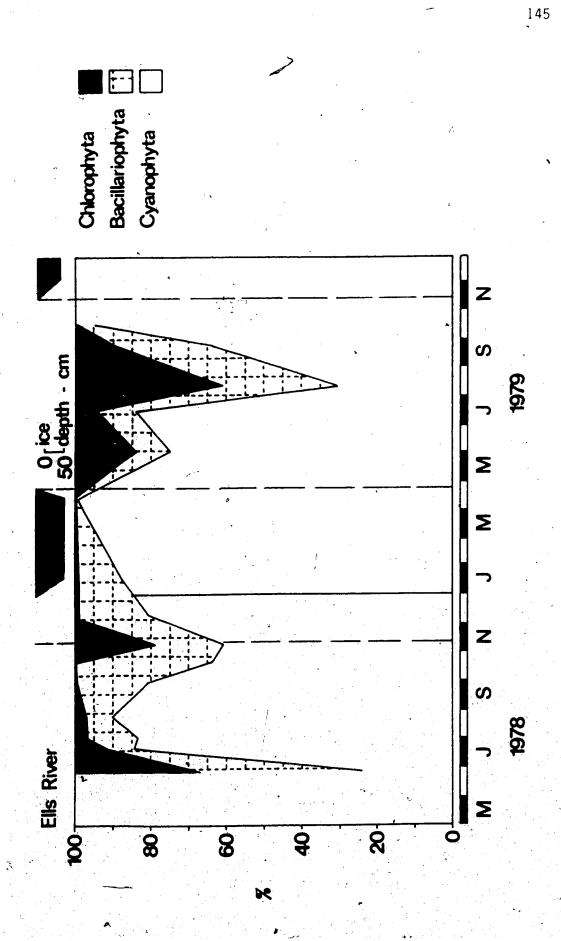
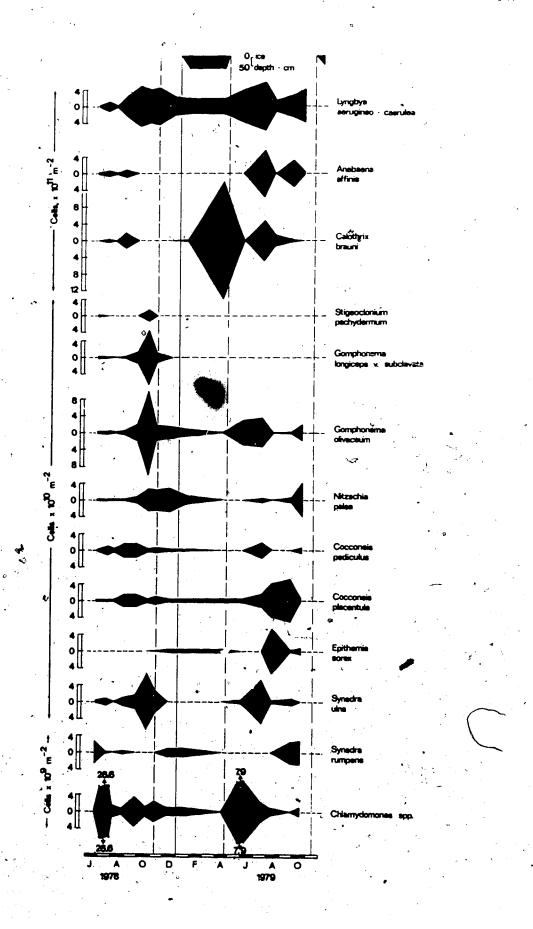


Figure 53.
Seasonal succession of the dominant epilithic algae in the Ells Riyer.



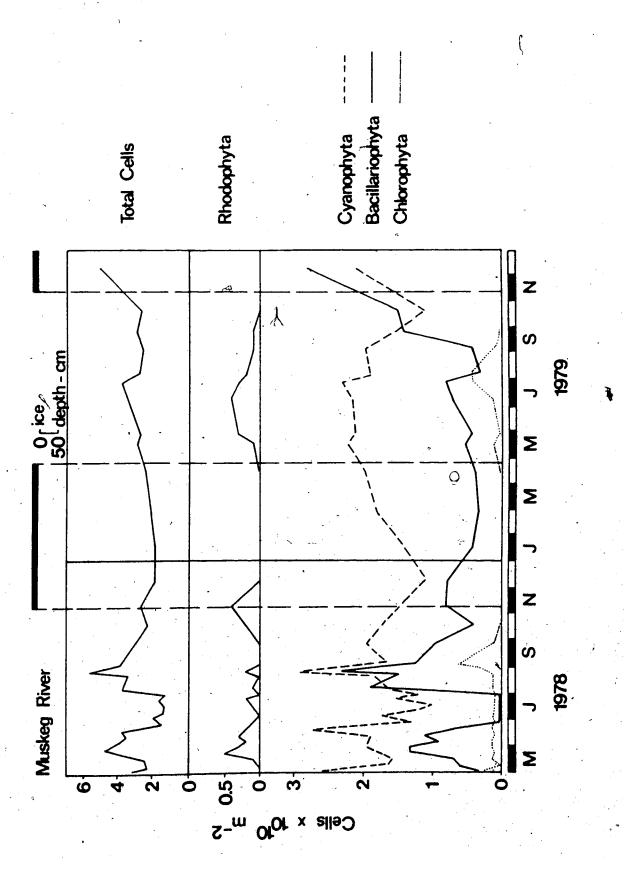
July 1979; <u>Calothrix braunii</u> peaked in April and July 1979, although like <u>Anabaena affinis</u>, smaller populations were present during 1978 with a small peak occurring in September; and <u>Anabaena</u> peaked in July and late September 1979.

On those two occasions that Cyanophycean algae did not dominate, late June 1978 and early August 1979 respectively, diatoms and chlorophycean algae were numerically more important. In late June 1978, Synedra ulna, Synedra rumpens, Cocconeis pediculus, Gomphonema olivaceum and Gomphonema longiceps var. subclavata were all present (Figure 53); in August 1979, Epithemia sorex and Cocconeis placentula were the dominant diatoms. In both instances the dominant Chlorophycean algae were Chlamydomonas spp.

## 4.3.2.1.2 Cell Numbers

In the Muskeg River total cell numbers peaked in the spring and autumn of 1978, but in 1979 a summer peak in July occurred between a much smaller spring and a larger autumn peak (Figure 54). The dominant species during these peaks are shown in Table 19. The species composition was very similar in both years in the spring and autumn maxima. However, Nitzschia fonticola and Synedra ulna were not present in the 1979 spring peak, and Batrachospermum vagum and Gomphonema olivaceum present in spring 1979 were absent in 1978. Similarly, Draparnaldia glomerata and Gomphonema olivaceum were present during the summer maximum of 1979. Gomphonema acuminatum only contributed significantly in autumn 1979. Cyanophycean algae were almost always the dominant algae numerically (Figure 54). Exceptions occurred during the autumn each year. Diatoms comprised the next most numerous division followed by Chlorophycean and Rhodophycean algae. The range of total

Figure 54.
Seasonal changes in total cells, Cyanophyta, Bacillariophyta, Chlorophyta and Rhondophyta in the Muskeg River.



cells, diatoms, Chlorophycean, Cyanophycean and Rhodophycean algae for the Muskeg river is presented in the following Table 20.

Table 19: The algae dominant during the spring, summer, and autumn cell number peaks in the Muskeg River.

| Year  | Ma                                      | ximum                         |                               |
|-------|---|-------------------------------|-------------------------------|
|       | Spring                                  | Summer                        | Autumn                        |
| 1978. | Phormidium sp.                          | no maximum                    | Phormidium sp.                |
| •     | Lyngbya aerugineo-caerulea              | -                             | Lyngbya<br>aerugineo-caerulea |
|       | Calothrix breviarticulata               |                               | Calothrix braunii             |
|       | <u>Calothrix</u> <u>braunii</u>         |                               | Draparnaldia<br>glomerata     |
| ~     | Synedra ulna                            |                               | Synedra ûlna                  |
|       | Nitzschia fonticola                     |                               | Nitzschia<br>fonticola        |
|       | Audouinella violacea                    |                               | Gomphonema olivacium          |
| 1979. | Phormidium sp.                          | <u>Phormidium</u> sp.         | Lyngbya<br>aerugineo-caerulea |
| · .   | Lyngbya aerugineo-caerulea              | Lyngbya aerugineo<br>caerulea | - Synedra ulna                |
|       | <u>Calothrix</u> <u>breviarticulata</u> | Draparnaldia<br>glomerata     | Synedra rumpens               |
|       | <u>Calothrix</u> <u>braunii</u>         | Gomphonema<br>olivacium       | Gomphonema<br>acuminatum      |
|       | Gomphonema olivaceum                    | Batrachospermum<br>vagum      | <u>Phormidium</u> sp.         |
|       | Audouinella violacea                    |                               |                               |
|       | Batrachospermum vagum                   |                               |                               |

Table 20

Mean and Range of total cells numbers, Diatoms, Cyanophycean, Chlorophycean and Rhodophycean algae in the Muskeg River for the study period

| -<br>           | Range<br>cells 10 <sup>10</sup> m <sup>-2</sup> | Mean (arithmetic) $\frac{\text{cells } 10^{10}\text{m}^{-2}}{\text{cells } 10^{10}\text{m}^{-2}}$ |
|-----------------|---|---|
| Total Cells     | 1.30 - 5.63                                     | 2.90  |
| Cyanophyta      | 1.02 - 2.90                                     | 1.78  |
| Bacillariophyta | 0.01 - 2.30                                     | 0.79  |
| Chlorophyta     | 0 - 0.61  | 0.12  |
| Rhodophyta      | 0 - 0.50  | 0.13  |

During 1978 in the Steepbank River, two algal maxima occurred, one during late July and a second, the largest, during October (Figure 55). Thereafter the populations remained high, decreasing only slightly during March of 1979. The winter population was dominated by Phormidium sp., Epithemia sorex, and Synedra rumpens (table 21). A small spring peak was apparent in 1979 followed by larger summer and winter peaks. Again, species composition was very similar at the maxima each year, although in spring 1979 Phormidium sp. was not as prevalent as in 1978 and the two diatoms Gomphonema olivaceum and Synedra rumpens were important only in 1979. Also, four species not found in 1978, Draparnaldia glomerata, Nitzschia palea, Nostoc spp. and Gomphonema olivaceum were dominant during summer 1979. Draparnaldia glomerata also occurred during the autumn peak in 1978. Only this species and Calothrix braunii were absent from the 1979 autumn peak.

Figure 55.
Seasonal changes in total cells, Cyanophyta, Bacillariophyta, Chlorophyta and Rhondophyta in the Steepbank River.

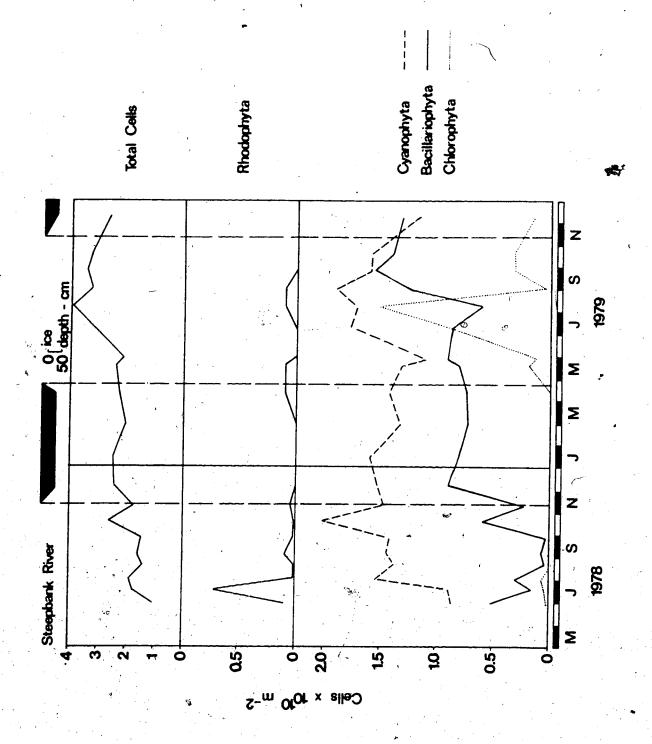


Table 21: The algae dominant during the spring, summer, and autumn cell number peaks in the Steepbank River.

|                                       | ,  |                                |                                   |
|---------------------------------------|--|--------------------------------|-----------------------------------|
| Year                                  | Max                                      | imum                           | •                                 |
|                                       | Spring                                   | Summer                         | Autumn                            |
| 1978.                                 | Lyngbya aerugineo-caerulea               | Lyngbya aerugineo-<br>caerulea | Lyngbya<br>aerugineo-<br>caerulea |
|                                       | Epithemia sorex                          | <u>Phormidium</u> sp.          | <u>Phormidium</u> sp.             |
|                                       | Phormidium sp.                           | Cladophora<br>glomerata        | Calothrix<br>braunii              |
|                                       |  | Epithemia sorex                | Draparnaldia<br>glomerata         |
| CZ .                                  | 4  | Draparnaldia<br>glomerata      | Epithemia sorex                   |
|                                       |  |                                | Achnanthes<br>lanceolata          |
| , , , , , , , , , , , , , , , , , , , | <b>◆</b>                                 |                                | Gomphonema<br>olivaceum           |
| <del>.</del>                          |  |                                | Nitzschia<br>fonticola            |
| 1979.                                 | Epithemia sorex                          | Lyngbya aerugineo-<br>caerulea | Lyngbya<br>aerugineo-<br>caerulea |
|                                       | <u>Lyngbya</u> <u>aerugineo-caerulea</u> | Draparnaldia<br>glomerata      | <u>Phormidium</u> sp.             |
|                                       | Gomphonema olivaceum                     | Epithemia sorex                | Epithemia sorex                   |
|                                       | Synedra rumpens                          | Nitzschia palea                | Nitzschia<br>fonticola            |
|                                       |  | Nosto spp.                     | Synedra rumpens                   |
| •                                     |  | Gomphonema<br>olivaceum        | Achnanthes<br>Tanceolata          |
| 4                                     |  | Cladophora<br>glomerata        | Gomphonema<br>olivacium           |

Cyanophycean algae were always most numerous except in early December 1979 (Figure 55, Table 21, 22), followed by diatoms. Only during mid-summer 1979 did a very large Chlorophycean community develop. The largest Rhodophycean population occurred in July 1978 when Audouinella violaceae was present.

Table 22

Mean and Range of total cell numbers, (Diatoms, Cyanophycean, Chlorophycean, and Rhodophycean algae) in the Steepbank River for the study period

|                 | Range<br>Cells 10 <sup>10</sup> m <sup>-2</sup> | Mean<br>Cells 10 <sup>10</sup> m <sup>-2</sup> |
|-----------------|---|--|
| Total Cells     | 1.02 - 3.93                                     | 2.36   |
| Cyanophyta      | 0.86 - 2.00                                     | 1.45   |
| Bacillariophyta | 0.03 - 1.55                                     | 0.66   |
| Chlorophyta     | 0 - 1.51  | 0.18   |
| Rhodophyta      | 0 - 0.70  | 0.07   |

Sampling on the Mackay river did not commence until early July of 1978. Thereafter a large summer maximum of epilithic algae occurred (Figure 56). Cyanophycean algae were invariably the most numerous being followed by diatoms and Chlorophycean algae (Table 24). No autumn maxima occurred during 1978, but in 1979 spring, summer and autumn peaks appeared, each slightly larger than the previous season. However, the 1979 population numbers never approached the size of the 1978 epilithic numbers. Little change occurred in the dominant of these peaks (table 23) and diatoms along with Cyanophycean algae became very important during autumn 1979 with massive populations of Epithema sorex developing.

Figure 56.
Seasonal changes in total cells, Cyanophyta, Bacillariophyta and Chlorophyta in the MacKay River.

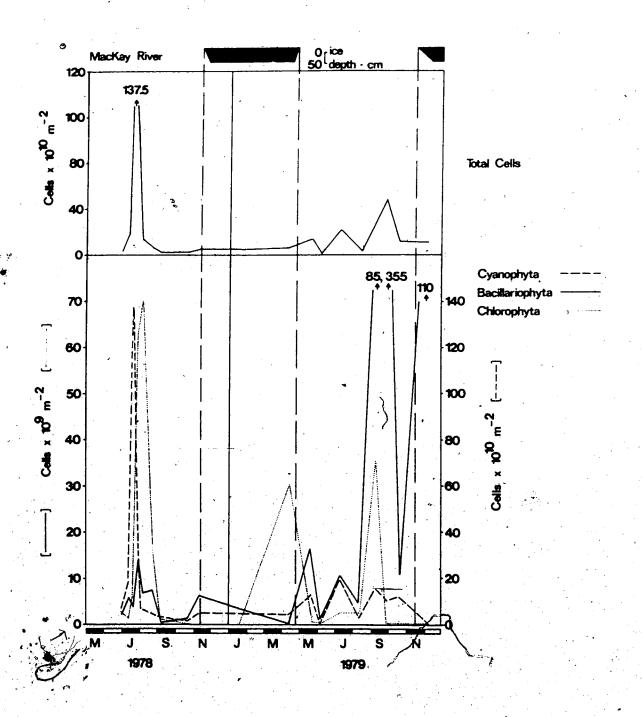


Table 23. The algae dominant during the spring, summer, and autumn cell, number peaks in the MacKay River.

| Year  |                                | Maximum                    |                                   |
|-------|--------------------------------|----------------------------|-----------------------------------|
|       | Spring                         | Summer                     | Autumn                            |
| 1978. | Not sampled                    | Calothrix braunii          | No maximum                        |
|       |                                | Lyngbya aerugineo-caerulea | ^1                                |
| •     |                                | Cladophora glomerata       |                                   |
|       |                                | <u>Chlamydomonas</u> spp.  |                                   |
|       | •                              | Epithemia sorex            | <b></b>                           |
| 1979. | Calothrix braunii              | Lyngbya aerugineo-caerulea | Lyngbya<br>aerugineo-<br>caerulea |
|       | Anabaena affinis               | Calothrix braunii          | Epithemia sorex                   |
|       | Lyngbya aerugineo-<br>caerulea | Epithemia sorex            |                                   |
|       | Epithemia sorex                |                            |                                   |

TABLE 24

Mean and Range of total cell numbers (Diatoms Cyanophycean and Chlorophycean algae) in the Mackay River

| •               |     | R     | ange       | Mea          | n        |
|-----------------|-----|-------|------------|--------------|----------|
| <b>n</b>        | , A | Cells | 1010 m-2   | <u>Cells</u> | 1010 m-2 |
| Total Cells     |     | 0.1   | 0 - 137.50 | · /          | 20.30    |
| Cyanophyta      | •   | 0-    | - Î36.50   | -            | 15.40    |
| Bacillariophyta | •   | 0     | - 35.50    | e<br>e       | 3.39     |
| Chlorophyta ;   |     | 0     | - 7.05     |              | 1.38     |

Sampling on the Hangingstone River also commenced during early July of 1978. During that year a large summer epilithic community developed, followed by a much smaller autumn one. The spring and summer peaks of 1979 merged and only a slight fall peak, was evident (Figure 57).

Table 25: The algae dominant during the spring, summer, and autumn cell number peaks in the Hangingstone River.

| Year     |             | Maximum                         |                         |
|----------|-------------|---------------------------------|-------------------------|
|          | Spring      | Summer                          | Autumn                  |
| 1978.    | not sampled | Stigeoclonium pachydermum       | Epithemia<br>turgida    |
|          |             | Lyngbya aerugineo-caerulea      | Gomphonema<br>olivacium |
|          | v           | Cladophora glomerata            | • . •                   |
| •        |             | Anabaena affinis                | •                       |
|          |             | <u>Calothrix</u> <u>braunii</u> | in the second second    |
| <b>.</b> |             |                                 | •                       |

1979. Spring and Summer

Lyngbya aerugineo-caerulea

Calothrix braunii

Gomphonema olivaceum

Calothrix braunii

Epithemia turgida

Stigeoclonium pachydermum was found only during July 1978.

Gomphonema olivaceum was dominant during autumn 1978 as well as during the spring/ summer maximum of 1979. These algae formed only a small population during autumn 1979 (Table 25, Figure 57). Cyanophycean algae were again numerically dominant but, unlike the other four rivers, the second most numerous division was Chlorophycean algae followed by diatoms (Table 26).

TABLE 26

Mean and Range of total cell numbers (Diatoms, Cyanophycean, Chlorophycean algae) in the Hangingstone River

|                 | A          | Range                                  | Mean                             |
|-----------------|------------|--|----------------------------------|
| v               | 1 <u>1</u> | cells 10 <sup>10</sup> m <sup>-2</sup> | <u>cells 10<sup>10</sup> m-2</u> |
| Total Cells     |            | 29.00 - 653.16                         | 75.80                            |
| Cyanophyta      |            | 0 - 613.57                             | 40.70                            |
| Bacillariophyta | `y`.       | 0.30 - 31.50                           | 5.64                             |
| Chlorophyta     | đ          | 0 - 299.70                             | 23.80                            |

Unlike the MacKay and Hangingstone Rivers, the Ells river did not have a very large summer peak of epilithic algae during 1978. Instead, 1979 was the year of maximum epilithic abundance. Autumn maxima were, however, comparable for both 1978 and 1979 (Figure 58). Cyanophycean algae were dominant, with <u>Lyngbya aerugineo-caerulea</u> dominant all the time (Table 27). In this river more diatom species contributed significantly than in the other four rivers (Table 27).

Figure 57.
Seasonal changes in total cells, Cyanophyta, Bacillariophyta and Chlorophyta in the Hangingstone River.

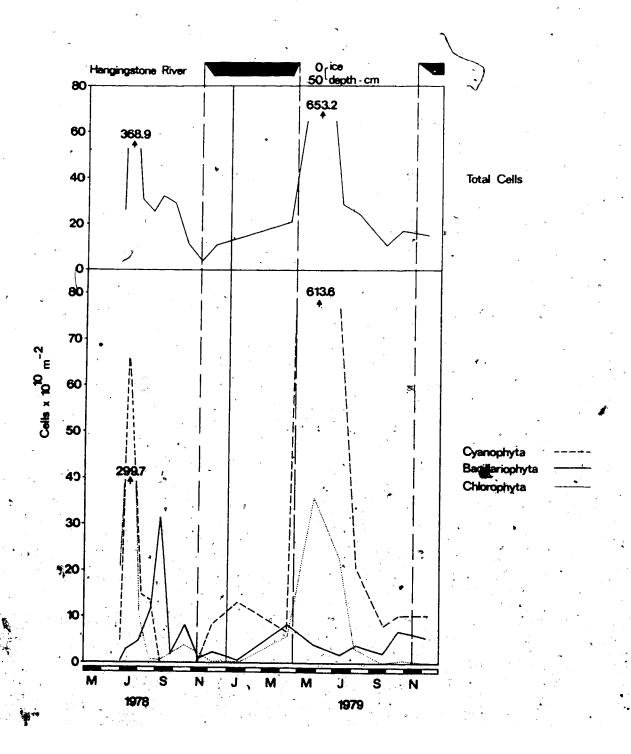


Table 27: The algae dominant during the spring, summer, and autumn cell number peaks in the Ells River

| Year                                 | Maximum                                  |                                    |
|--------------------------------------|--|------------------------------------|
| Spring                               | Summer                                   | Autumn                             |
| 1978. not sampled                    | Lyngbya aerugineo-caerulea               | Lyngbya aerugineo-<br>caerulea     |
|                                      | Anabaena affinis                         | Stigeoclonium<br>pachydermum       |
|                                      | Cocconeis pediculus                      | Gomphonema longiceps v. subclavata |
| •                                    | Synedra rumpens                          | Gomphonema olivaceum               |
| •                                    | Chlamydomonas spp.                       | Cocconeis pedicultus               |
|                                      |  | <u>Cocconeis</u> placentula        |
| •                                    |  | Synedra ulna                       |
|                                      | *  | <u>Chlamydomonas</u> spp.          |
| 1979. <u>Lyngbya</u><br>aerugineo-ca | Lyngbya <u>aerugineo-caerulea</u> erulea | Lyngbya aerugineo-<br>caerulea     |
| Calothrix braun                      | ii Anabaena affinis                      | Gomphonema olivaceum               |
| Gomphonema                           | Calothrix braunii                        | Nitzschia palea                    |
| <u>'olivaceum</u>                    | Gomphonema olivaceum                     | Cocconeis placentula               |
|                                      | Synedra ulna                             | Epithemia sorex                    |
|                                      |  | Synedra rumpens                    |

Lyngbya was the dominant Cyanophycean algae in the summer of 1978, whereas it, along with Anabaena affinis and Calothrix sp., were codominant in 1979. Two diatoms, Gomphonema olivaceum and Synedra ulna were also present in 1979, whereas Synedra rumpens had been in 1978. The autumn peaks comprised many diatom species, some of which occurred both years (Table 27). However, only in 1978 was Stigeoclonium pachydermum found.

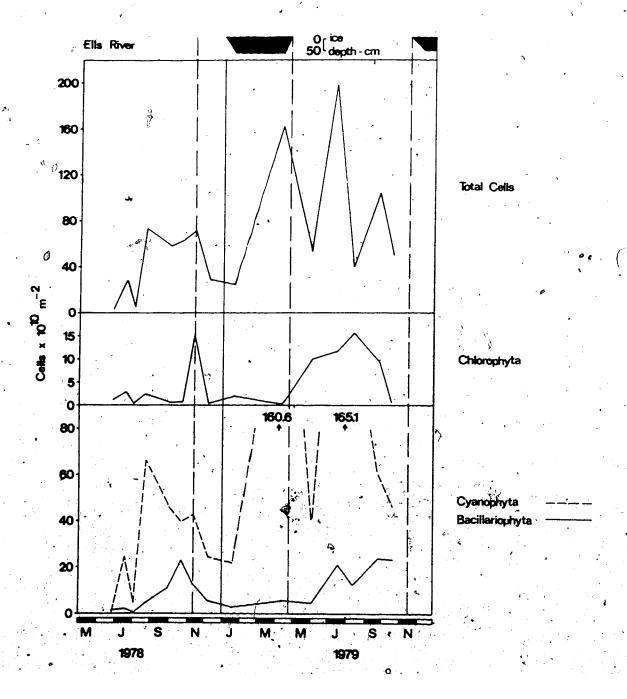
In summary, therefore Cyanophycean algae were generally dominant followed by diatoms and Chlorophycean algae in the Ells River (Table 28).

Table 28

Mean and Range of the Total Cell Numbers (Diatoms, Cyanophycean and Chlorophycean algae in the Ells River

|   |                 | Range cells 10 <sup>10</sup> m <sup>-2</sup> | Mean (arithmetic) cells 10 <sup>10</sup> m <sup>-2</sup> |
|---|-----------------|--|--|
| ì | Total Cells     | 3.70 - 197.70                                | 63.80  |
|   | Cyanophyta      | C.90 - 165.12                                | 50.70  |
|   | Bacillariophyta | 0.51 - 23.98                                 | 8.50   |
|   | Chlorophyta     | 0 - 15.49                                    | 4.60   |

Figure 58.
Seasonal changes in the total cell numbers of Cyanophyta, Bacillariophyta and Chlorophyta in the Ells River.



### 4.3.3 Temporal Standing Crop Changes

### 4.3.1 Epilithon

Seasonal fluctuations of the epilithic standing crops in the Muskeg, Steepbank, Hangingstone, MacKay and Ells Rivers were compared using the singular rock brushing technique and transect methods in conjunction with primary productivity studies. The results are presented in figure 62. In the Muskeg River during 1978, chlorophyll a concentrations fluctuated more than during 1979, with spring, late summer and early winter peaks evident. The spring peak, like cell counts in 1979 was small, with the standing crop rising to a maximum in early December (Figure 59).

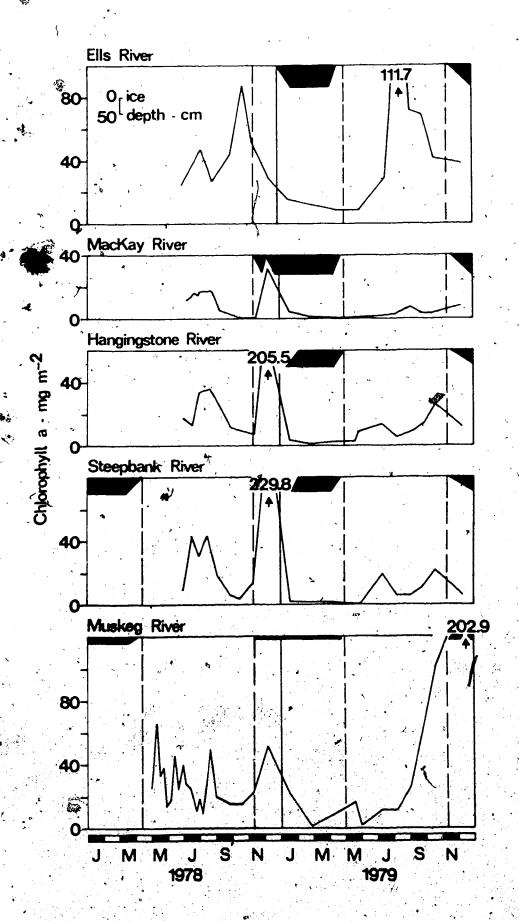
A summer/autumn chlorophyll peak was evident in the Steepbank River while a massive peak (229.8 mg chlorophyll  $\underline{a}$  m<sup>-2</sup>) occurred in early December 1978. During spring of 1979 a spring maximum was not detected; however, a small summer and a distinct autumn maxima occurred (Figure 59). Thus, a bimodal peak of epilithic standing crop was found to occur in the Steepbank River as well.

Standing crops in the Hangingstone River were very similar to those in the Steepbank River; the algal standing crop peaks occurred at the same time and achieved nearly the same size (205.5 mg chlorophyll  $\underline{a}$  m<sup>-2</sup>) and 229.8 mg chlorophyll a m<sup>-2</sup>.

Chlorophyll a, like cell counts in the MacKay River, were lower than the other four rivers. Although this river supported a smaller epilithic biomass, a summer peak was evident as well as a late winter maximum following ice formation during 1978. Standing crops remained lower during 1979 with only a small peak during the autumn.

Epilithic standing crops fluctuated with similar seasonal patterns in the Ells River; however, during both 1978 and 1979 the autumn maximum

Figure 59. Seasonal fluctuations in epilithic algal standing crop, as measured by chlorophyll a content in the Ells, MacKay, Hangingstone, Steepbank and Muskeg rivers.



preceded that of the other four rivers. The development of late summer maxima was also followed by declining autumn standing crops despite the fact that species composition changes as noted earlier were similar to the other rivers.

A comparison of the standing 2rop means for the five rivers indicated that for the duration of the study, the Ells River supported the highest standing crop, followed by the Muskeg, Steepbank, Hangingstone and MacKay Table 29).

Table 29

Mean and Range of Standing Crop for Epilithon
As Measured by Chlorophyll a content

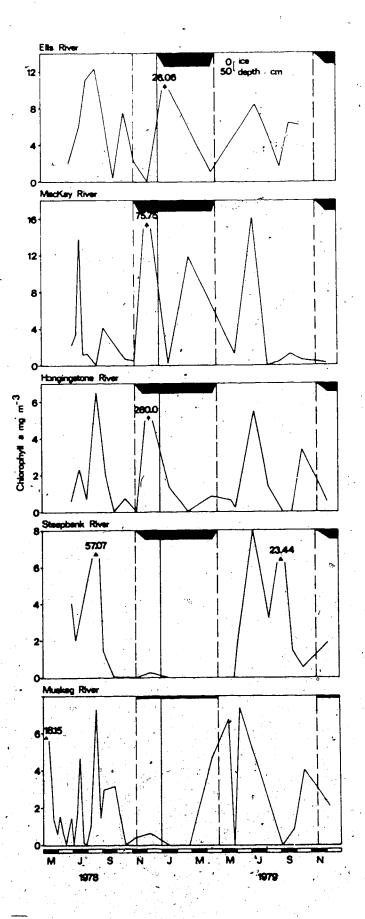
| River          | Range<br>Chlorophyll <u>a</u><br>mg m <sup>-2</sup> | #Mean<br>Chlorophyll a<br>mg m-2 |
|----------------|---|----------------------------------|
| Muskeg         | 1.33 - 202.95                                       | 30.46                            |
| Steepbank      | 40 - 229.84   | 22.99                            |
| MacKay         | 0.11 - 30.66  | 7.94                             |
| Hangingstone , | 1.42 - 205.46                                       | 22.35                            |
| E11s           | 8.38 - 111.72                                       | 43.23                            |

# 4.3.3.1 Phytoplankton

Microscopic examination of planktonic lotic algae collected from the five rivers indicated that both senescing epilithic and lentic species were present. It was noted that desmids probably derived from muskeg runoff were also present. The standing planktonic crop fluctuations were very irregular (Figure 60).

Figure 60.

Standing crop, as measured by chlorophyll a content, of the planktonic algae in the Ells, MacKay, Hangingstone, Steepbank and Muskeg rivers.



P

Despite the irregular nature of standing crop fluctuations, spring, summer and autumn peaks as well as some winter ones (eg. Ells, MacKay, Hangingstone) did occur. Peak planktonic standing crops frequently occurred during periods of peak epilithic standing crops and thereafter, suggesting that detached epilithic algae are the major component of the planktonic algae in these rather small brown water rivers. Planktonic standing crops were highest in the Hangingstone and lowest in the Muskeg River (Table 30).

Table 30

Range of Standing Crop for Plankton as Measured by Chlorophyll a content

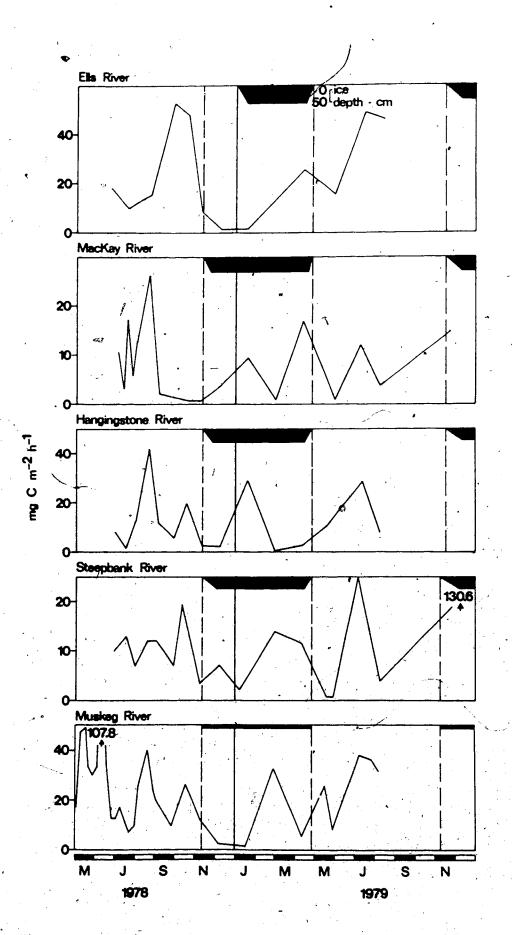
|              | Range                          | " Mean |
|--------------|--------------------------------|--------|
| River .      | Chlorophyll <u>a</u><br>mg m-3 | mg m-3 |
|              | •                              |        |
| Muskeg       | 0 - 18.15                      | 2.93   |
| Steepbank    | 0 - 57.07                      | 5.42   |
| MacKay       | 0 - 75.75                      | 7.12   |
| Hangingstone | 0 - 280.00                     | 15.11  |
| Ells         | 0.01 - 26.06                   | 6.44   |

# 4.3.4 Temporal Productivity Changes

# 4.3.4.1 Epilithon

Seasonal primary productivity fluctuations for each of the five rivers are illustrated in Figure 61. They were more variable than those for standing crops. Summer maxima occurred in all rivers, but spring and autumn maxima did not occur consistently except in the Muskeg River. Winter peaks occurred under ice and snow cover during January 1979 in the Hangingstone and MacKay rivers and during March in the Muskeg and

Figure 61.
Epilithic algal primary productivity in the Ells, MacKay,
Hangingstone, Steepbank and Muskeg rivers.



Steepbank rivers, and late April in the Ells and MacKay Rivers. The range of productivity values for the five rivers is presented in table 31.

Mean and range of epilithic <sup>14</sup>C uptake for each of the five rivers

| River '      | • | Range                                 | Mean          |
|--------------|---|---------------------------------------|---------------|
| -            |   | mg C m <sup>-2</sup> hr <sup>-1</sup> | mg C m-2 hr-1 |
| Muskeg       | • | 1.1 - 107.8                           | 25.3          |
| Steepbank    | • | 0.6 - 130.6                           | 16.3          |
| Hangingstone | • | 0.1 - 41.9                            | 12.4          |
| MacKay       |   | 0.5 - 26.0                            | 8.3           |
| Ells         | j | 1.1 - 52.5                            | 23.2          |

Mean productivity among the five rivers ranged from 8.3 mg C m $^{-2}$  hr $^{-1}$  (MacKay) to 25.3 mg C m $^{-2}$  hr $^{-1}$  (Muskeg). Although epilithic productivity was on average higher in the Muskeg River, the highest value was found in the Steepbank River.

Multiple linear regression of open water period primary production (epilithic) versus algal numbers, temperature, silica, pH, phosphorus, velocity, discharge, nitrogen, and total alkalinity revealed that diatom numbers were primarily related to productivity in the Muskeg River (r=.64) (Appendix 6 and 7). The remaining variables increased the multiple r value to 0.87. In the Steepbank River nitrate-nitrogen (r=.75) discharge and pH were primarily related to productivity. Phosphorus was the more important variable (of those examined) related to productivity in the Hangingstone River (r=.41), and the Ells River (r=.64). In the MacKay River Chlorophycean numbers (r=.51) were most

nighly correlated with epilithic productivity. Multiple linear regression analysis of these sample variables versus productivity (open water) for all the rivers combined indicated that Cyanophycean algal numbers are most highly correlated with productivity (r=.28). The remaining variable yielded a multiple r of 0.54 indicating that other factors influence primary productivity among these brown water alkaline streams. Mean epilithic chlorophyll a and productivity were positively correlated (r=0.869, p< 0.10), suggesting the presence of a possible biotic factor which influences epilithic primary productivity (Appendix 8).

### 4.3.4.2 Phytoplankton

Phytoplankton productivity was highly variable among the five rivers. Table 32 below presents the 1979 findings.

Table 32

Mean and range for phytoplankton <sup>14</sup>C uptake during 1979

| Location     |            | Range         |   | Mean       |  |
|--------------|------------|---------------|---|------------|--|
|              |            | mg C m-3 hr-1 | · mg                                    | C m-3 hr-1 |  |
| Muskeg       |            | 7.4 - 90.5    | •                                       | 46.5       |  |
| Steepbank    | , <u>,</u> | 0.8 - 61.2    | • · · · · · · · · · · · · · · · · · · · | 26.8       |  |
| Hangingstone |            | 14.2 - 51.2   |   | 34.3       |  |
| Ells -       |            | 26.4 - 77.5   |   | 49.4       |  |
| MacKay       |            | 0.2 - 68.9    |   | 34.4       |  |

The highest mean <sup>14</sup>C uptake rate of 49.4 mg C m<sup>-3</sup> hr<sup>-1</sup> was found to occur in the Ells River, which is located downstream of the Gardiner Lakes. Peak phytoplankton productivity occurred in the winter (January) under ice cover in Muskeg, during April (ice out) in the Steepbank and

Hangingstone and July in the MacKay and Ells. Phytoplankton productivity and standing crops were not related statistically.

### 4.3.4.3 Microhabitat Differences

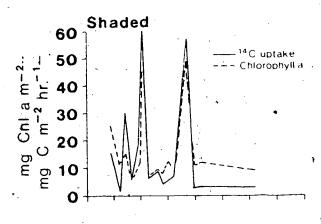
Since epilithic algal standing crops varied depending upon the microhabitat it followed that productivity would vary too. To confirm this, primary productivity was compared among the shore, midstream and midstream artificially shaded sites in the Muskeg River during 1978. The results are presented in figure 62.

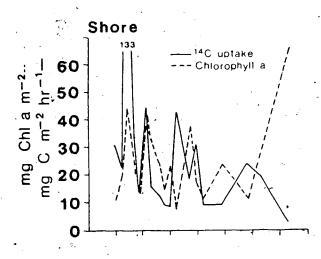
At the onset of this study on 29 May, productivity was higher at the shore site than midstream. It reached maximum at both sites in early June (133.0 and 83.0 mg C m<sup>-2</sup> hr<sup>-1</sup> respectively). Second peaks occurred at both sites in August but that at the shore site occurred earlier than the peak at the midstream site (Figure 62). Higher epilithic algal productivity continued at this midstream site until the late fall. It declined at both sites as winter approached and ice formed in early November. At these two sites there was little correlation between standing crop size and productivity (r=0.608 and r=0.237, p=0.05) whereas there was at the artificially shaded site (r=0.961, p=0.05). Shading also appeared to retard and delay standing crop and productivity peaks (Figure 62).

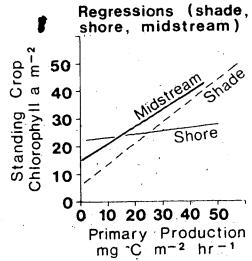
Expressing the productivity as a function of standing crop or as a photosynthetic index (mg C (mg chlorophyll  $\underline{a}$ )<sup>-1</sup> m<sup>-2</sup> hr<sup>-1</sup>) found the shore epilithic algae with the highest mean value while the lowest was occurred at the midstream site (Table 33).

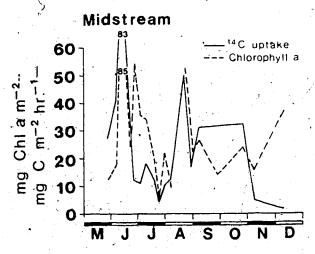
Figure 62.

Epilithic primary productivity, standing crops and linear regressions for shore, midstream and shaded midstream microhabitats in the Muskeg River during 1978.









Production, photosynthetic index and standing crops for the Muskeg River microhabitats during 1978

| Microhabitat      | Mean<br>Primary<br>Productivity       | Mean<br>Photosynthetic<br>Index   | Mean * Standing Crop |              |
|-------------------|---------------------------------------|---|----------------------|--------------|
| /                 | mg C m <sup>-2</sup> hr <sup>-1</sup> | $\begin{array}{c} \text{mg C (mg Chl }\underline{a})^{-1} \\ \text{m-2 hr-1} \end{array}$ | mg Chl               | <u>a</u> m-2 |
| Shore             | 22.                                   | 9 0.86  |                      | 26.6         |
| Midstream         | 21.                                   | 0.69  | . •                  | 31.1         |
| Midstream (shaded | ) 13.                                 | . 0.82  |                      | 16.8         |

At the midstream shaded site epilithic standing crops were 56% smaller and productivity 60% smaller than at the unshaded site.

## 4.3.4.4 Substratum Comparison

As mentioned earlier naturally occurring bitumen in the Steepbank River supported an algal community, consisting primarily of Cyanophycean algae. Therefore standing crops and productivity were also compared between the epi-bituminous algae and the epilithic algae (Figure 63). Primary productivity associated with both limestone rock and bitumen remained similar June through early September of 1978; however, it dramatically increased on the rock substratum during late October when algal numbers (particularly diatoms) were also higher. Productivity declined on both substrata during ice formation in early November. Primary productivity ranged from 3.2 to 27.5 mg C m<sup>-2</sup> hr<sup>-1</sup> for limestone attached epithilic algae and 0.18 to 11.7 mg C m<sup>-2</sup> hr<sup>-1</sup> for bitumen. Cyanophycean algae were numerically dominant for both substrata accounting for 86 to 99% of the limestone epilithon and 73 to 99% of the oilsand benthos.

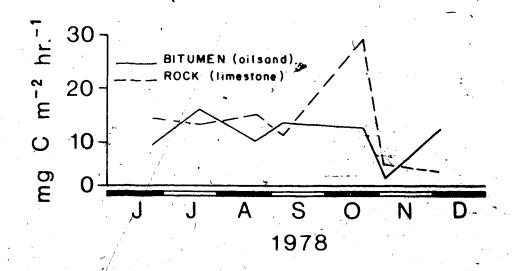


Figure 63 (a) Epilithic productivity on oilsand and rock substratum.



Figure 63 (b) Photograph of the substratum found in the Steepbank River.

Microalgae were more abundant on the limestone while macroalgae particularly Nostoc commune were more abundant upon the oilsand substrate. This investigation provided evidence that primary productivity during 1978 was 22% higher for epilithic algae or limestone than those or the bitumen, and the difference was statistically significant for p=0.05.

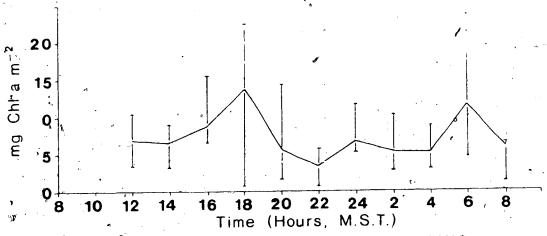
## 4.3.4.5 Diurnal Experiment

There have been few studies of diurnal productivity for northern latitudes (Murphy, 1984; Ilmavita, 1977). Therefore, one such experiment was conducted over the period July 28/29, 1979. The results are presented in figure 64.

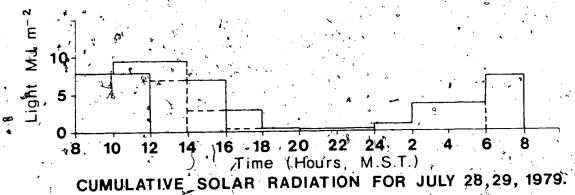
Epilithic algal productivity was highest during the midday period (1200-1600 hr), and minimal during 0200-0600 hrs. Moreover, during the mid-day productivity and bicarbonate alkalinity were inversely related; alkalinity ranged from 3.4 to 3.7 meq L<sup>-1</sup> with the minimum extending until 2000 hrs despite  $^{14}$ C uptake declines from 42.2 mg C m<sup>-2</sup> hr<sup>-1</sup> (midday) to 1.3 mg C m<sup>-2</sup> hr<sup>-1</sup> at 2000 hrs. pH ranged from 6.6 to a maximum of 7.2 by 1400 hours. Diurnal fluctuations of pH and bicarbonate attributed to physical and biotic factors were in fact minor compared to those reported for productive systems (Wetzel, 1975).

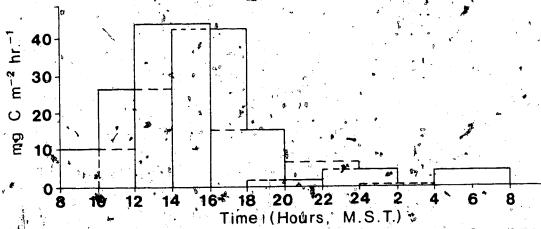
Temperatures in the Muskeg River were lowest at 0400 hrs  $(18^{\circ}\text{C})$  just prior to sunrise and maximum during early evening 18-2000 hrs  $(19^{\circ}\text{C})$ .

Benthic algal standing crop estimates were also variable throughout the experiment. The range of standing crop estimates were higher during the early evening and morning. Simple linear regression indicated that Figure 64.
Diurnal primary productivity, and standing crops for the Muskeg River during 1979.



MAXIMUM, MINIMUM AND MEAN EPILITHIC STANDING CROPS FOR EACH INCUBATION.





EPILITHIC 14C UPTAKE DURING JULY 28, 29, 1979, FOR THE MUSKEG RIVER.

14C uptake and chlorophyll <u>a</u> were highly correlated during this short term study (r=.745) (p = <0.05). Photosynthetic efficiency reached a maximum of 5.01 mg C <sup>m-2</sup> hr<sup>-1</sup> during 1200 - 1600 hours, the period of peak insolation (Figure 64). However <sup>14</sup>C uptake and insolation were in part, negatively correlated (r-.397, p = 0.05).

## 4.3.5 Temporal Nitrogen Fixation Changes

### 4.3.5.1 Epilithon

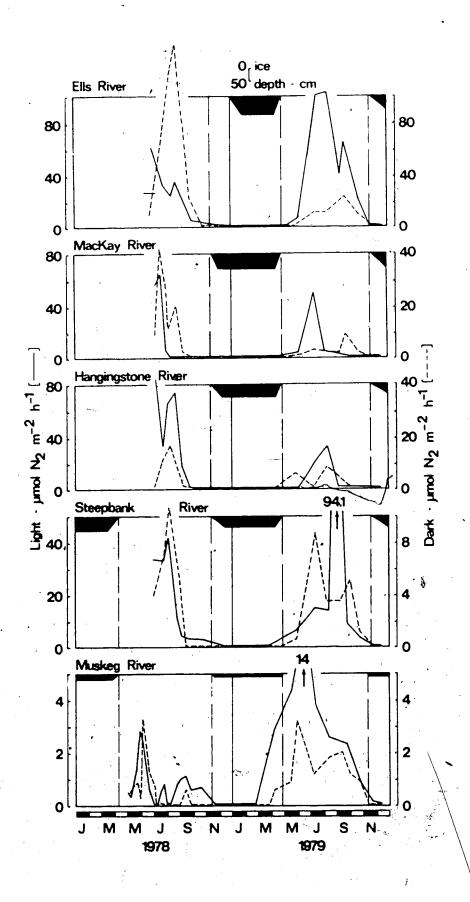
Temporal nitrogen fixation patterns were similar in all five rivers (Figure 65). Maxima occurred during the spring/summer period, but not always corresponding to the largest Cyanophycean populations. The dominant Cyanophycean species or potential algae capable of elemental nitrogen fixation indicated (Table 34) below were similar in all rivers.

Table 34,

The dominant Cyanophycean algae coinciding with the nitrogen fixation maxima in the five rivers

| <u>Species</u>                | 0 ,            | ` <b>.</b> ' | *        |               | Riv            | er             | ,   |            | 1   | •           |
|-------------------------------|----------------|--------------|----------|---------------|----------------|----------------|-----|------------|-----|-------------|
|                               | Mus<br>1978    |              |          | pbank<br>1979 | Hangin<br>1978 | gstone<br>1979 |     | 1979       | 7   | 11s<br>1979 |
| Anabaena affinis              | <b>-</b> ·     | ·            | <b>-</b> | -             | +              | -              | +   | -          | + . | +           |
| Calothrix brauni              | <u>i</u> .+,   | + "          | -        | +             | +              | +              | . + | +          | + \ | +           |
| C. breviarticulat             | <u>a</u> + `,  | + .          | -        | -             |                | -              |     | <b>-</b> ' | -   | <b>-</b> ′  |
| Lyngbya aerugined<br>caerulea | <u>)-</u><br>+ | · +          | +        | +             | +              | +              | +   | +          | +   | <b>.</b>    |
| Nostoc Spp                    | -<br>0         | • .          | · +      | +             | + ,            | <b>-</b> .     |     | +          | +   | <b>+</b>    |
| <u>Oscillatoria</u> sp.       | ° - a          | . <b>-</b>   | - :      | -             | -              | •              | . o | <b>-</b>   | +   | -           |
| Phormidium sp.                | . +            | +            | +        | + .           | -              | · -            | -   | <b>.</b>   | · - | -           |

Figure 65.
Epilithic nitrogen fixation in the Ells, MacKay, Hangingstone, Steepbank and Muskeg rivers.



Nostoc spp. were particularly abundant in the Ells, MacKay, Steepbank and Hangingstone (only during 1978). The timing of peak nitrogen fixation varied among the rivers; moreover differences between the two years, and among the rivers also occurred with respect to both light and dark fixation rates. Dark nitrogen fixation rates although displaying similar temporal fluctuations as light fixation rates were sometimes greater especially during 1978 in the Ells River (Figure 65). At that time (fall) epilithic algal populations, which were dominated by Nostoc commune, had accummulated to a depth of 1 cm. The reverse occurred the following year when fall discharge in the Ells was significantly lower than observed during 1978. Light and dark fixation maxima also did not always coincide, for example during 1978 and 1979 in the Muskeg and Steepbank rivers (Figure 65).

¢

Epilithic nitrogen fixation was consistently minimal during the winter (cold water) periods, therefore the five rivers were compared for both the entire study period and the open water period of each year (April - November, Table 35).

#### Table 35

The mean number of epilithic Cyanophycean algae over the entire study period, and the range of nitrogen fixation for the open water periods of 1978 and 1979.

| River                              | Cyanoph  | yta                             | Nitrogen F                      | ixation |                                   |              |
|------------------------------------|----------|---------------------------------|---------------------------------|---------|-----------------------------------|--------------|
| $\overline{x}$ cells $10^{10}$ m-2 |          | 010 <sub>m</sub> -2             | u moles N <sub>2</sub> hrā m²²² |         |                                   |              |
|                                    | e<br>Eps | Over Entir<br>Light             | e Period<br>Dar                 | •       | en Water<br>Light                 | 1978<br>Dark |
| Muskeg                             | 1.78     | $\frac{0-14.30}{x} = 1.52$      | $\frac{0-3.18}{x} = 0.65$       |         |                                   |              |
| Steepbank                          | 1.45     | $\frac{0-31-94.10}{x} = 12.26$  | $\frac{0.01-16.80}{x} = 2.26$   |         |                                   | )<br>'h.     |
| MacKay                             | 15.40    | $\frac{0.09-64.00}{x} = 8.72$   | $\frac{0.86-42.00}{x} = 5.99$   |         |                                   |              |
| Hanging-<br>stone                  | 46.70    | $\frac{0-84.00}{x} = 17.2$      | $\frac{0}{x} = 2.20$            |         |                                   | )            |
| Ells                               | 50.7     | $\frac{0.01-110.00}{x} = 27.14$ |                                 |         | $00 \ \frac{0.39-143}{x} = 49.90$ |              |
|                                    |          |                                 |                                 |         |                                   |              |

| ***          | Open Water 1<br>Light             | 1979<br>Dark                    |
|--------------|-----------------------------------|---------------------------------|
| Muskeg       | 0.02-14.30                        | <u>0</u> .04-3.18               |
| Steepbank    | $\bar{x} = 3.62$ 0.31-94.10       | $\bar{x} = 1.33$<br>0.33-8.5    |
| Mackay       | x = 17.53 $1.02-49.00$            | x = 3.41 $0.44-2.5$             |
| Hangingstone | x = 8.68 $0.59-33.40$             | $\bar{x} = 1.53$<br>0.11-8.8    |
| Ells         | $\overline{x} = 11.38$ 0.71-110.0 | $\bar{x} = 2.49$<br>- 0.53-21.9 |
|              | $\bar{x} = 56.19$                 | $\overline{x} = 12.4$           |

The Ells and Muskeg Rivers possessed the largest and smallest Cyanophycean populations and similarly the highest and lowest rates of nitrogen fixation respectively.

The data acquired through these investigations were employed to determine the relationship between epilithic nitrogen fixation and factors previously shown to affect fixation rates for each river and for all rivers combined. With the exception of temperature all other parameters were log transformed in order to yield a normally distributed data base.

Epilithic algal nitrogen fixation was consistently negatively correlated with ambient nitrate-nitrogen concentrations. Negative relationships also occurred for Cyanophyta numbers and nitrogen fixation in the Steepbank, Hangingstone and Ells Rivers where Nostoc commune were always observed; temperature was negatively correlated with nitrogen fixation in the Muskeg River. Interestingly, Nostoc spp. were very fare in the Muskeg River, while filamentous and colonial Cyanophytes were abundant.

Multiple linear regression was also performed to determine which of the parameters already outlined affected nitrogen fixation appreciably (Appendix 9). Among each of the rivers the primary variable affecting nitrogen fixation was ambient nitrate-nitrite nitrogen concentration. For all rivers combined however, the primary variable affecting nitrogen fixation was chlorophyll <u>a</u> followed closely by temperature and nitrate-nitrite nitrogen.

## 4.3.5.2 Microhabitat Differences

Nitrogen fixation by the epilithic algae was also measured at shore, midstream and artificially shaded midstream sites in the Muskeg

River (1978). Species composition, was similar at the three sites but the dominant Cyanophycean algae differed (Table 36).

Table 36.

| De                      | ominant species, nur<br>three Muskeg Rive |                              |                      |  |
|-------------------------|---|------------------------------|----------------------|--|
| Site<br>Muskeg<br>River | Dominant<br>cyanophycean<br>algae         | x<br>cyanophycean<br>Numbers | Nitrogen<br>Light    | Dark                                     |
| Shore                   |   | Cells 10 <sup>10</sup> m     | u mol N <sub>2</sub> | m-2 hr-1                                 |
| ) م                     | Lyngbya aerugineo-                        | 1.48                         | 0.40                 | 0.13                                     |
| (15 cm dep              | th) <u>caerula</u>                        |                              | •                    |  |
|                         | Phormidium sp (s)                         | ·.                           |                      |  |
|                         | Calothrix braunii                         |                              |                      | ·  |
| Midstream               |   | 1.70                         | 0.63                 | 3 0.38                                   |
|                         | Lyngbya-aerugineo-                        | _                            |                      |  |
| •                       | caerula                                   |                              |                      | •  |
|                         | Phormidium sp (s)                         | •                            |                      |  |
| · · · · ·               | Calothrix braunii                         |                              |                      | ia.                                      |
| Artificial              | ly <u>Aphanothece</u>                     | 1.27                         | 0.15                 | 0.07                                     |
| Shaded                  | <u>nidulans</u>                           |                              | **                   | •  |
|                         | Calothrix braun                           | <u>ii</u>                    | · ·                  | en e |
|                         | Microcystis inc                           | erta                         |                      |  |

Heterocystous and non-heterocystous filamentous species were similar at the shore and midstream sites. (ie. Lyngbya, Phormidium, Calothrix). Colonial (Aphanothece, Microcystis) and filamentous Calothrix forms were dominant under the artificial shade. At the shore and midstream sites Cyanophycean numbers fluctuated similarly but the June and September peaks were more pronounced at the latter site (figure 66). Under shade, numbers fluctuated far least with the maximum occurring in October. Mean numbers were highest at midstream and least under shade (Table 36). The differences in algal numbers were statistically different for shore vs midstream (p=.025), midstream vs shaded midstream (p=.005), and shore vs midstream (p=0.05).

Nitrogen fixation (both light and dark) was highest at the midstream (unshaded) site (Table 36) with pronounced June, July, and September peaks (Figure 66). Significant differences occurred between the midstream shaded and unshaded sites (p=0.005), and between the shore and unshaded midstream (p=0.05). The difference between the shore and shaded midstream epilithic nitrogen fixation was not statistically significant.

### 4.3.5.3 Substrata Comparison

As indicated earlier both limestone rocks and naturally occurring oilsand or bitumen supported an abundance of Cyanophycean algae in the Steepbank River, therefore nitrogen fixation was assayed for these substrata. Numbers peaked in July on both substrata (excluding Nostoc Commune (Figure 67). Afterwards numbers decreased rapidly upon the bitumen, and thereafter remained low. In contrast upon the limestone they remained high and even peaked again in October. The temporal fluctuations of Nostoc commune were identical upon both substrata.

Figure 66.
Epilithic nitrogen fixation and Cyanophycean numbers in shore (less than 15 cm depth) midstream and artificially shaded midstream communities of the Muskeg River during 1978.

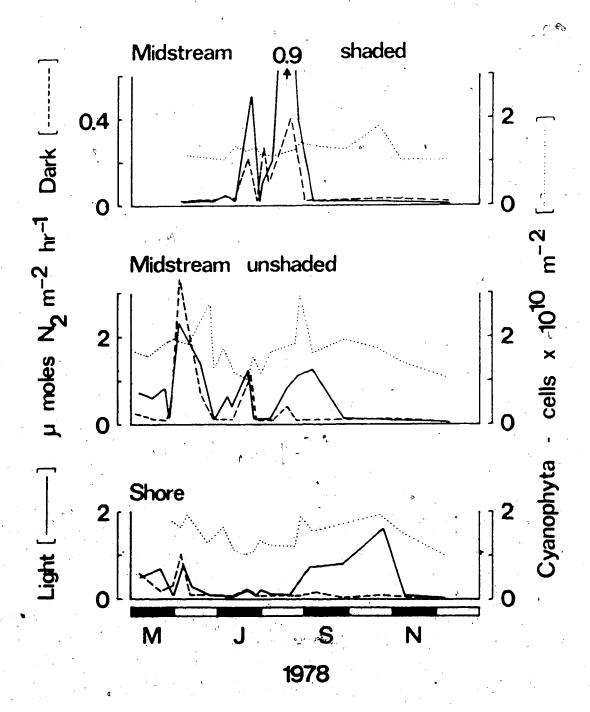
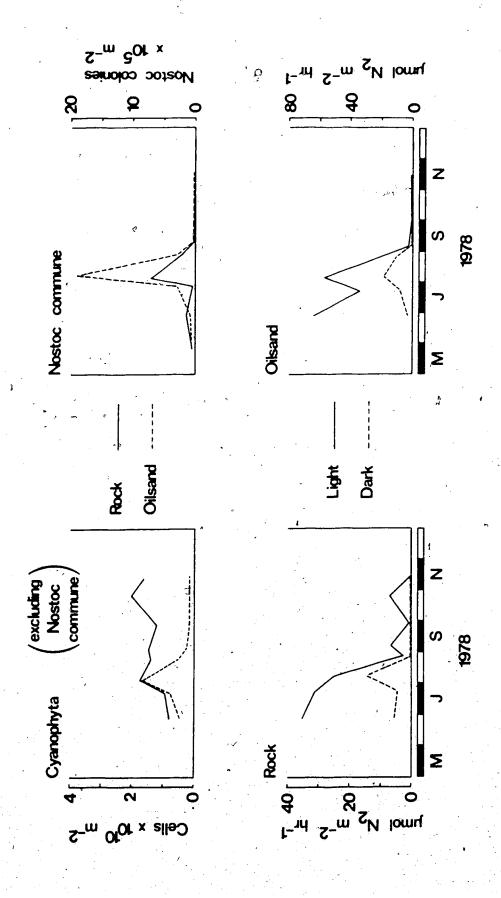


Figure 67.

Nitrogen fixation and Cyanophycean algal numbers for epilithic algao attached to limestone rock and oilsand substratum in the Steepbank River.



However, Nostoc numbers were greatest upon the bitumen during the period of peak standing crop (Figure 67). The differences did not prove to be statistically significant. Statistically significant differences were however found for total Cyanophycean numbers (p = 0.05) between the two substrata. The following Table 37 indicates mean algal numbers and mean nitrogen fixation rates for the Steepbank River open water period of 1978 are presented in Table 35. Identical sampling dates and assay times were compared.

Table 37

Mean algal numbers and nitrogen fixation for oilsand and rock substrata in the Steepbank River during 1978

| Nostoc Commune Colonies 104m-2 |      | Total<br>Cyanophycean No.<br>cells 10 <sup>8</sup> m <sup>-2</sup> | Nitrogen Fi<br>umol N <sub>2</sub> m <sup>-2</sup> h | Nitrogen Fixation umol N <sub>2</sub> m <sup>-2</sup> hr <sup>-1</sup> |  |  |
|--------------------------------|------|--|--|--|--|--|
| Substrate                      | a    |  | &<br>Light   | Dark   |  |  |
| Rock                           | 13.6 | 156.1  | 13.4   | 3.3  |  |  |
| Oilsand                        | 32.0 | 60.0   | 20.1 .   | 5.1  |  |  |

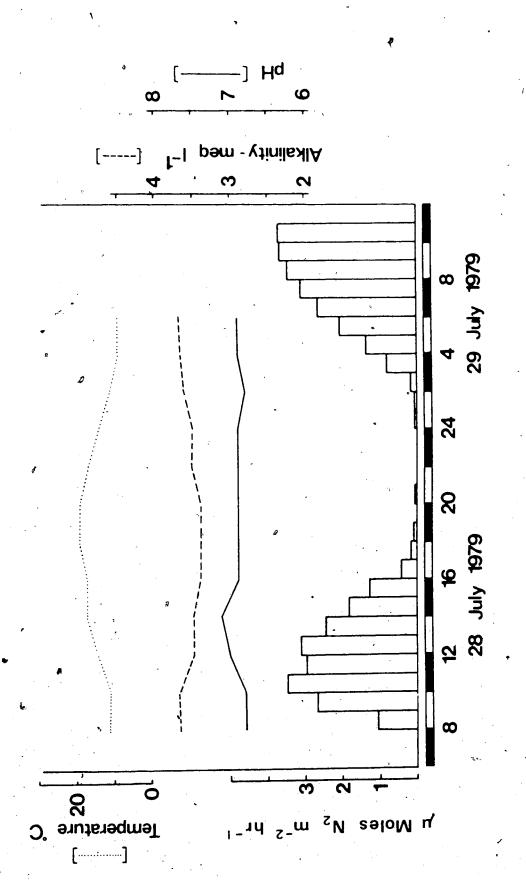
Nitrogen fixation although higher on average for epilithon attached to oilsand underwent similar temporal trends in the Steepbank River characterized by summer (July, August) peaks (Figure 67).

## 4.3.5.4 Diurnal Experiment

Diurnal epilithic nitrogen fixation was also assayed during July 28, 29, 1979, in the Muskeg River. On that date Lyngbya aerugineo-caerula, Phormidium sp., Chaemosiphon incrustans and Calothrix braunii dominated almost exclusively.

Water temperature when the experiment began at 08:00 hours was  $18^{\circ}$ C increasing between 10:00 and 12:00 hours (Figure 68) to a maximum of  $19^{\circ}$ C between 18:00 and 20:00 hours, whereupon it declined to  $10^{\circ}$ C by

Figure 68.
The diurnal pattern of epilithic algal nitrogen fixation, pH, alkalinity and temperature for the Muskeg River during 1979.



04:00 hours during predawn of July 29. Bicarbonate alkalinity varied little while pH increased from 6.8 at 08:00 hours to a maximum of 7.2 at 14:00 hours before decreasing to, and remaining at 6.9 (except for a decline at 02:00 hours (Figure 68). Incident irradiance was greatest between 10:00 and 11:00 hours on both July 28 and 29.

During the first incubation period (08:00 to 13:00 hrs.) light fixation was high (2.85 umol of  $N_2$  m<sup>-2</sup>hr<sup>-1</sup>) (Figure 68). It decreased slightly between 13:00 and 18:00 hours and more sharply between 18:00 and 23:00 hours to 1.15 umol  $N_2$  m<sup>-2</sup>hr<sup>-1</sup>. This followed by an abrupt increase corresponding to an increase in irradiance levels as dawn drew into early morning. In fact the highest fixation rate of 3.15 umol.  $N_2$  m<sup>-2</sup>hr<sup>-1</sup> occurred during the morning period. Afterwards rates decreased even though irradiance levels continued increasing. Less variability was observed for dark fixation rates (Figure 68).

Nitrogen fixation efficiency (umol  $N_2$  mg<sup>-1</sup> Chl a m<sup>-2</sup>hr<sup>-1</sup>) in clear glass bottles was maximum between 18:00 and 23:00 hours whereas dark bottle efficiency was highest between 23:00 and 04:00 hours (Figure 68). 4.3:6 Cluster and Principal Components Analysis

Cluster and principal components analyses was applied to a set of fourteen physical, chemical and biological attributes of the five rivers under investigation. A total of 92 cases were included in the program analysis for the Muskeg, Steepbank, Hangingstone, Ells and MacKay, 1978 and 1979 data pool. The first three component eigenvalues greater than 1.5 were utilized for ecological interpretation of the similarities and differences among the rivers. Ibaney (1983) and Lengendre and Legendre (1979) recommended the use of eigenvalues greater than 1. Since they usually explain the majority of the data variance within the first two

or three vectors (de Emiliani and Depestris, 1982, Charlton, Hamilton, Cross, 1985).

Table 38

Eigenvector coefficients, eigenvalues, total variance and

| cummulative variance accounted for by the first five components |        |        |        |        |        |  |  |
|---|--------|--------|--------|--------|--------|--|--|
| Eigenvector   | I      | II     | III    | ΙV     | ٧.     |  |  |
| Coefficients  |        |        |        | ,      |        |  |  |
| Prod. ( <sup>14</sup> C uptake)                                 | .104   | 0.169  | 0.164  | -0.490 | -0.102 |  |  |
| Std. Crp (Chl a)  | .023   | -0.241 | -0.211 | -0.129 | .620   |  |  |
| Temperature   | .332   | -0.173 | .426   | -0.035 | -0.082 |  |  |
| Discharge   | .232   | .528   | -0.138 | -0.025 | .132   |  |  |
| Velocity  | .273   | .513   | -0.160 | .007   | .161   |  |  |
| Nitrogen  | -0.136 | .260   | .297   | .402   | .164   |  |  |
| Phosphorus  | -0.188 | -0.097 | -0.257 | .583   | 0.078  |  |  |
| Silica  | -0.463 | .008   | -0.011 | -0.006 | .244   |  |  |
| pH  | .292   | -0.209 | .426   | .252   | .070   |  |  |
| T. Alkalinity   | -0.454 | -0.188 | .052   | -0.077 | -0.047 |  |  |
| N-Fixation  | .288   | -0.291 | .087   | .183   | .212   |  |  |
| Nmbs (Chlorophyta)  | .191   | -0.231 | -0.216 | -0.022 | .398   |  |  |
| Nmbs (Cyanophyta)   | .210   | -0.135 | -0.306 | .062   | -0.502 |  |  |
| Nmbs<br>(Bacillariophyta)                                       | .167   | -0.173 | -0.466 | .364   | -0.028 |  |  |
| Eigenvalue  | 3.21   | 2.32   | 1.74   | 1.28   | 1.11   |  |  |
| Total Variance  | 22.93  | 16.58  | 12.39  | 9.16   | 7.96   |  |  |
| Total Variance (cummulative %)                                  | 22.93  | 39.51  | 51.90  | 61.06  | 69.02  |  |  |

The first three principal components accounted for nearly 52% of the total data pool variance (Table 38) and can be interpreted as representing the lotic seasonal influence. The first component with 22.93% of the variance can be interpreted as representing the spate (spring/fall) influence. This finding emerges from the highly positive weight exerted by the contributions of river temperature, pH, nitrogen fixation, velocity and discharge. During the spring and fall periods, North Eastern Alberta river temperatures and nitrogen fixation are low while velocity, discharge and pH are high, indicating that high discharge, velocity and low temperature may stimulate biological related pH increases by non-nitrogen fixers such as diatoms. Moreover, high negative scores for vector I were attributed to the variable silica, alkalinity, nitrogen and phosphorous.

The second component with 16.58% of the variance can be interpreted as representing the lotic influence. This finding emerges from the highly positive weight exerted by the contributions of lotic discharge, velocity, nitrogen and the negative scores (Table 38) for nitrogen fixation, algal standing crop and numbers of Chlorophyta. The climatological factors which increase discharge and velocity also increase lotic nitrogen and silica concentrations. Conversely high discharge and velocity disrupt the benthic algal communities reducing nitrogen fixation, chlorophyll a and particularily the numbers of Chlorophycean algae.

The third component explained 12.39% of the variance and can be interpreted again as the nutrient influence. Temperature, pH, and nitrogen variables provided a significant contribution to the eigenvector. Diatoms, blue green algal numbers and phosphorus provided

negative scores (Table 43).

Four major clusters were defined using the same data subjected to principal components analysis. Cluster analysis placed the data from all the rivers (1978/79) into groups (clusters) based upon similarities among the fourteen physical, chemical and biological values included in the program. This technique consisted of calculating a similarily matrix between samples using the squared Euclidean Distance Method (Sneath and Sokal, 1973). Clustering was then performed by using Ward's Hierarchical Fusion Method (Ward 1963).

The nature of the three vectors is illustrated in Figure 69 and the distribution of clusters along vectors I, II and III is presented in figure 70.

Cluster 3 consisted of 15 cases where the Muskeg, Steepbank, MacKay and 1979 Hangingstone winter data (16 Nov. - 16 April) grouped together. Silica, total alkalinity, phosphorous and nitrogen were the principal components explaining this cluster. Within this cluster the mean values for the principal components were 8.1 mg  $L^{-1}$ , 4.3 meq  $L^{-1}$ , 0.08 mg  $L^{-1}$ , and 0.21 mg  $L^{-1}$  respectively.

Cluster 2 consisted of 19 cases which were skewed to the positive side of vector II. Discharge, velocity, nitrogen and silica were the most important components defining this cluster. The mean values were 22.67 m<sup>-3</sup>s<sup>-1</sup>, 0.98 cms<sup>-1</sup>, .30 mg L<sup>-1</sup>, and 4.6 mg L<sup>-1</sup> respectively. Cluster 2 consisted of spring (17 April - 30 June) and fall (20 Aug. - 15 Nov) data (Figure 69 and 70) from the Muskeg, Steepbank, fall/78 Hangingstone, MacKay and spring/79 Ells. Hence cluster 2 emphasizes the similarity among the Muskeg, Steepbank, and MacKay Rivers during the spring and fall when spates occur.

Cluster 4 was widely divided among the negative sides of all except vector I. Consisting of 17 cases (Figure 70) cluster 4 contained Ells and Hangingstone data from all seasons and MacKay River (fall/winter). Four of the 17 cases represented summer data. Cluster 4 provides evidence that although the Ells, Hangingstone and MacKay (fall/winter) Rivers are dissimilar with respect to biotic communities, temperature, pH and nitrogen as well as nitrogen fixation are similar. Cluster four contains data having the greatest numbers of benthic algae an average temperature of 7.5°C; pH of 7.4 and nitrogen .18 mg  $L^{-1}$ . Indeed this cluster provides evidence of the physico-chemical factors which are particularly beneficial to benthic algal community development.

Cluster 1 consisted of 41 cases (Figure 69, 70) which were skewed from median position to positive along vectors I and III. This grouping includes data from all rivers particularily for the spring/summer seasons. Cluster 1 contains data having the highest average temperature (15.1°C), the highest pH (7.9), an average discharge of 47 m<sup>3</sup>s<sup>-1</sup>, and velocity of .37 cms<sup>-1</sup>. Cluster one provides evidence that the five rivers examined during this study are similar with respect to a few physical and chemical characteristics during the summer, and these factors are the primary components explaining their particular limnology.

Figure 69.
The relative location of clusters I, II, and III and their principal components.

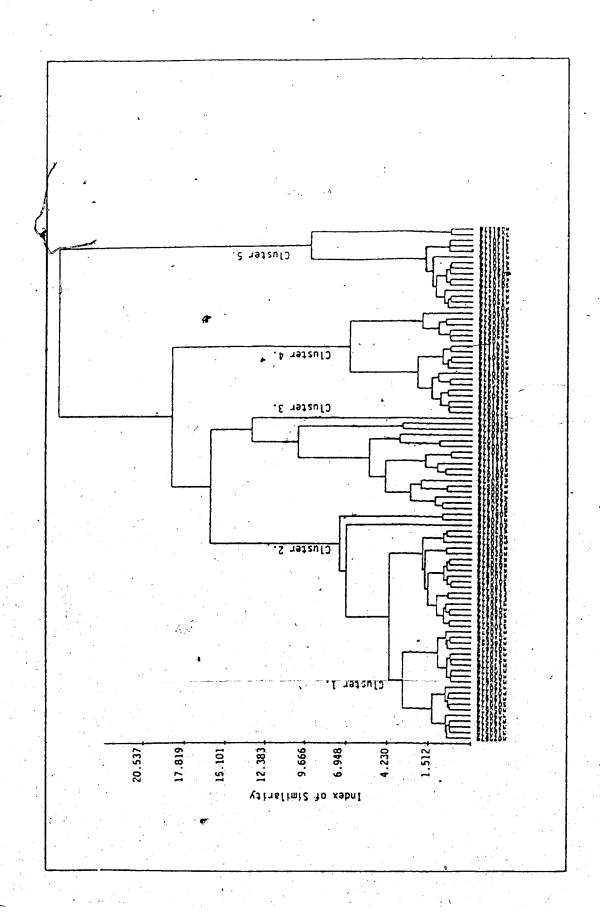
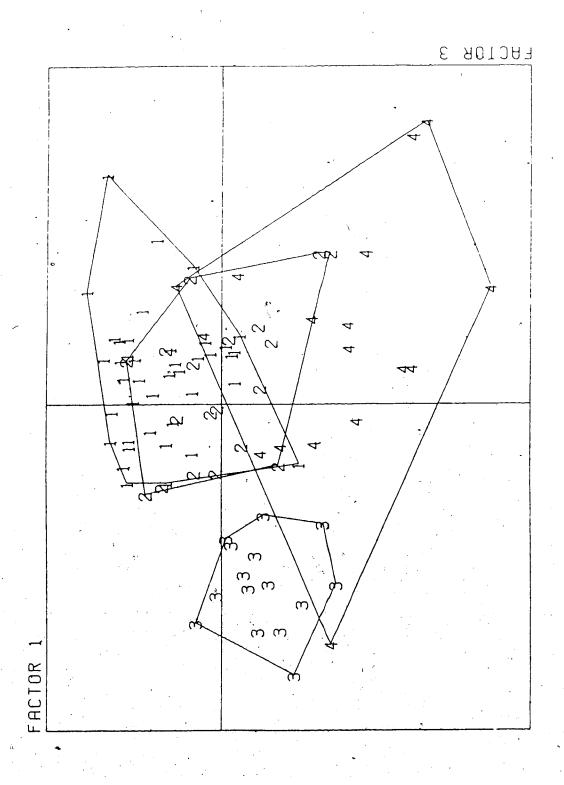


Figure 70.
The relative location of the first four clusters derived for the Ells, MacKay, Hangingstone, Steepbank and Muskeg rivers



#### 5. Discussion

### 5.1 Longitudinal Characteristics

Rivers occurring in the oil sands region of northeastern Alberta are atypical at their origin compared to the numerous descriptions quoted in Whitton (1975). The five rivers are not characterized by swift currents, steep gradients and pronounced erosion at their source. They originate in Muskeg, except for the Ells River which emerges from the Gardiner Lakes. The Muskeg, Steepbank, Hangingstone, and MacKay are slow flowing, meandering creeks with predominantly an organic substratum at their origin. Evidence of any erosion did not appear until the middle to lower reaches of all rivers. Haaslam, (1978) reported that channel size, width and depth generally increase in a downstream direction, in this respect these rivers are typical except for the Ells which was considerably wider in the upper reach.

Calcium was the major cation in all but the MacKay River where sodium was the most abundant. A consistent pattern emerged with respect to major anions whether the results were expressed as mg or meq  $1^{-1}$  with  $HCO_3^- > SO_4^= > Cl^-$  in all but the Muskeg River. Here chloride replaced sulphate mainly due to the high concentrations originating from the catchment area sediments or ground waters. Schwartz (1979) reported that 12 to 40% of the Muskeg River stream flow during late spring, summer and autumn comprises ground waters derived from a watertable situated at a depth of 0.5 to 3.5 meters. This factor contributes to increasing ionic concentration in a downstream direction, particularly chloride. Nothing is known about the ground water regimes of the other four rivers. However, least effect would undoubtedly be found in the Ells River.

Several algae were cosmopolitan, being found in all rivers (e.g. Lyngbya aerugieo-caerulea, Chamydomonas sp., Chrorella vulgaris, Achnanthes lanceolata, Cocconeis pediculus, Cocconeis placentula, Cyclotella meneghiniana, Cymbella ventricosa, Epithemia argus, Fragilaria capucina, Gomphonema lanceolatum, Gomphonema olivaceum, Navicula cryptocephala, Nitzschia palea, and Synedra ulna). All the cosmopolitan algae formed a dominant population at, at least one site in each river, except Epithemia argus. Only Lyngbya aerugineo-caerula and Cocconeis placentula did so in all rivers. The former species is tolerant of both hard water and dystrophic conditions, (Prescott, 1962); the latter prefers moderately alkaline waters (pH 7-9), unpolluted waters, (Schoeman, 1973). Some species had very restricted distributions being found in only one river. For example, Gomphosphaeria lacustris v. compacta, Crucigenia quadrata, Cryptomonas erosa, Achnanthes sp., Cymbella turgida, Eunotia lunaris, Fragilaria leptostauron, Gomphonema acuminatum v. coronata, Gomphonema ventricosum, Navicula gracilis, · Pinnularia mesolepta, and Tabellaria fenestrata were found only in the Muskeg River; Hyalotheca sp., Pediastrum biradiatum, Gomphonema gracile, and Nitzschia hantzschiana were confined to the Steepbank River; Microspora pachyderma, Asterionella formosa, Cyclotella comta, Cyclotella kutzingianum, Diatoma elongatum, Diatoma vulgare v. ovalis, Fragilaria construens, Fragilaria construens v. binodis, Navicula minima v. atomoides, and Stephanodiscus astraea were confined to the Ells River; Pediastrum boryanum was confined to the Hangingstone River; and to the MacKay River were confined Chroococcus limneticus, Gomphosphaeria aponina, Oscillatoria amphibia, Chlorella ellipsoidea, Gloecystis gigas, Oedogonium sp., Pediastrum biradiatum v. emarginatum f. convexum,

Scenedesmus acutiformis, Scenedesmus quadricauda, Sphaerocystis schroeteri, Sphaeroplea annulina, Ulothrix sp., Dinobryon sertularia, Mallomonas sp., Cymbella lanceolata, Eunotia pectinalis v. minor, Eunotia valida, Hantzschia amphioxys, Neidium affine, Neidium affine v. amphirhynchus, Stauroneis anceps, and Surirella ovalis.

In general, these rivers were dominated by Cyanophycean algae except for the MacKay River. In fact, this group accounted for 97.3% of the total epilithic community in the Steepbank River. The dominant species were Lyngbya aerugineo-caerula, Anabaena affinis, Calothrix braunii, and Nostoc spp. In the Muskeg River blue green algae accounted for 87.4% with Lyngbya aerugineo-caerulea most important compared to Anabaena affinis and Calothrix braunii. On average, Chlorophycean algae only accounted for 8.1% although at some sites some accounted for as much as 41.1% (e.g. site 4 - Cladophora glomerata). Diatoms accounted for only 4.3%. Cyanophycean algae were less important in the Ells River (75.4%). Here diatoms were important (21.7%). Again Lyngbya aerugineocaerulea was the important Cyanophyte. A variety of diatoms were important depending upon the site but included Achnanthes lanceolata, Cymbella sinuata, Cymbella ventricosa, Diatoma vulgare, Diatoma vulgare v. grandis, Gomphonema olivaceum, Cocconeis placentula, Cyclotella kutzingianum, Fragilaria pinnata, Nitzschia palea, Nitzschia recta, Navicula minima v. atomoides, and Cyclotella meneghiniana. even more important compared to blue green algae in the Hangingstone River (37.0 and 47.0% respectively). Lyngbya lerugineo-caerulea, Anabaena affinis and Oscillatoria sp. were the important blue green algae and Navicula graciloides, Achnanthes lanceolata, Epithemia sorex, Cocconeis placentula and Synedra ulna comprised the important diatoms.

Overall, Chlorophycean and Rhodophycean algae were minor components (11.9 and 3.3% respectively) even though, for example Batrachospermum vagum accounted for 23.4% at site 3 of the Hangingstone River. The MacKay River was the most diverse and on average all algae groups contributed at least 2.0%. Chlorophycean algae were the most important (58.1%). Several species contributed with none being absolutely dominant (e.g. Chlorella vulgaris, Chlamydomonas spp.), Cryptophycean algae (Cryptomonas ovata and Rhodomonas minutum) comprised 10.2%, Cyanophycean algae (mainly Lyngbya aerugineo-caerulea, Oscillatoria sp., and Anabaena affinis), 8.3%; diatoms (Pinnularia gibba, Navicula cryptocephala, Fragilaria vaucheriae, Cocconeis pediculus, Epithemia sorex) 6.3%; Chrysophyta and Euglenophyta both 5.1%; and Pyrrophyta and Rhodophyta 2.5% and 2.0% respectively.

A.

Chlorophyll a analyses provided evidence that the five rivers supported higher standing crops in the upper and middle reaches. Crowther, (1980) reported similar findings for benthic invertebrates. Hence the rivers investigated during this study are more productive in the upper and/or middle reaches where the relative contribution of detritus is also highest. High middle reach algal standing crops were associated with an opening of the forest canopy (which allowed more light to reach the substratum) and a predominance of rock substratum. Rivers in North-Eastern Alberta therefore exhibit a high degree of physical, chemical and biotic longitudinal variation which probably functions to maintain lotic system stability.

# 5.1.1 Epilithic Algal Sampling Technique Comparison

An investigation of three different sampling techniques applied to four microhabitats throughout this study of the Muskeg River provided further evidence of the extremely heterogeneous nature of epilithic algal communities. Well defined differences among standing crop estimates were found as shore, midstream, riffle and artificially shaded microhabitats were compared. The differences were more evident on a daily or weekly basis than for long term studies. The various sampling techniques provided evidence of the complex variability which occurs in rivers both temporally and spatially. No particular technique was significantly different as a method for estimating epilithic algal standing crop; however, the differences with respect to time and equipment were profound. The quadrat technique was extremely monotonous and very time-consuming. The single rock brush technique was less demanding than the quadrat; however, the need to transport rocks from remote areas for area determination is bardly economical. The scrape technique on the other hand was efficient, rapid and required a minimum of sampling equipment. It is particularly useful for remote areas and specifically when a limited space is available in which to transport supplies (i.e. a helicopter). All of the sampling techniques are potentially biased by the collector during substrate collection and selection of the scraping site on the rock surface. Moreover, the presence of macro-algae i.e. Cladophora spp. and Nostoc spp. clog pipettes during the subsampling further introducing potential error. Thus it would appear that any of these techniques is useful for long term studies. Care, must however be taken to survey all microhabitats and particularly during the periods of abundant epilithic community development.

### 5.2 Seasonal Patterns or Temporal Changes

#### 5.2.1 Physical and Chemical Changes

#### 5.2.1.1 Temperature

Throughout the five rivers water temperatures were lowest during the period of ice cover (generally November through April) and the highest during late July or early August. Boon and Shires (1976) indicated that flow rate, however small, maintains river temperatures in northeast England, and also temporary flow pools may contribute to increased river water temperature. Temporary pools were more abundant throughout the upper reaches of both the Muskeg and MacKay Rivers (Charlton, Hickman, Jenkerson 1981) and may account for the greater mean temperatures in these rivers. Water temperatures annually declined with increasing September discharge and increased at the onset of spring breakup and increased discharge, illustrating the role of runoff and radiation in the maintenance of water temperatures. There was no evidence suggesting that increasing discharge during late winterresulted in marked temperature increases prior to the breakup; which agrees with the findings of Langford (1970). Thus, direct radiation and re-radiation from the streambed are the two principle factors contributing to the observed temperatures.

## 5.2.1.2 Discharge

Discharge (current speed) exerts a profound mechanical force in lotic systems which is both destructive (McIntire, 1966) and beneficial (Ruttner 1960) to organisms. It affects both species composition and morphology (McIntire, 1966) as well as biomass accumulation (Reisen et al (1970). In addition to the physical forces exerted by discharge, its increase causes dilution of ions and nutrients thus the nutrient status of water.

Discharge peaks occurred bi-annually in April/May and September/
October in all five rivers. High spring discharge is attributed to
runoff and snowmelt while high fall discharge is attributed to runoff
increase resulting from a reduction of the deciduous plant water uptake
throughout each basin. The fall and spring spates in the Hangingstone
and Muskeg Rivers in fact were of comparable size. Spring spates were
larger than fall ones in the remaining rivers. The MacKay which drains
the largest area also possessed on average the greatest discharge rates.

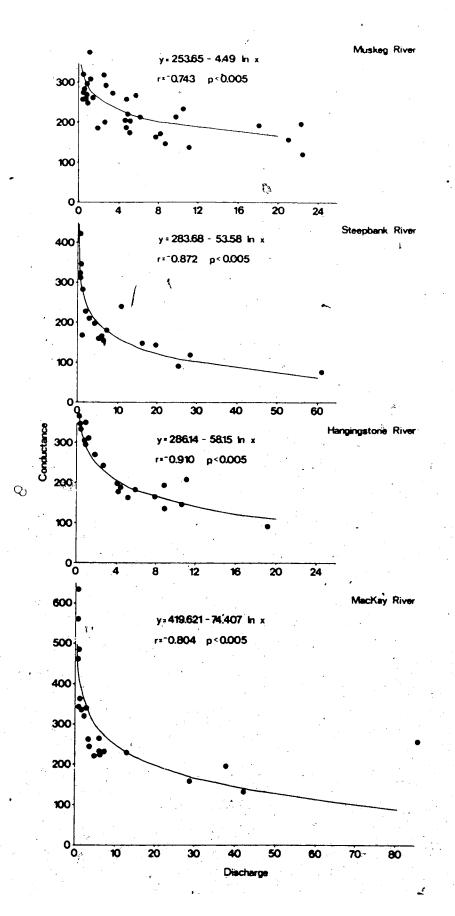
5.2.1.3 Specific Conductance

High winter specific conductance was characteristic of all rivers, except the lake-fed Ells River. Schwartz (1979) attributed high winter conductivity to the increasing role played by groundwater inputs in relation to river discharge. Conversely during periods of ascending discharge, conductance decreased due to dilution effects. Specific conductance, was negatively correlated in a logarithmic manner with discharge rates in all but the Ells River (Figure 71, Appendix 10). The headwater lakes of the Ells had a buffering effect upon discharge rates which maintained specific conductance within a very narrow range and while conductance among all the rivers increased in a downstream direction the amplitude and values remained lower throughout the Ells. Thus the origin of a river plays a major role with respect to specific conductance.

# 5.2.1.4 Calcium

Calcium concentrations in all of the rivers investigated were highest during the winter period. Schwartz (1979) indicated that calcium is one of the dominant cations found in groundwater examined throughout the Muskeg River catchment area. Calcium, like conductance

Figure 71.
Relationships between conductance and discharge.



was negatively correlated in a logarithmic manner with discharge rates in all but the Ells River (Appendix 10). Thus it would appear that groundwater again plays a major role with respect to maintaining winter discharge among the Muskeg, Steeplebank, Hangingstone and MacKay Rivers. Calcium among all the rivers averaged  $18.9~\text{mg}~\text{L}^{-1}$ . This represents a value less than that reported for European rivers (i.e.  $31~\text{mg}~\text{L}^{-1}$ ) as well as North American rivers (i.e.  $21~\text{mg}~\text{L}^{-1}$ ) (Whitton, 1975). Calcium concentrations excluding the Ells River were however similar to those reported for North America (i.e.  $20.5~\text{mg}~\text{L}^{-1}$ ).

#### 5.2.1.5 Sodium

Sodium was also significantly correlated with discharge among all five rivers. Temporal fluctuations were however irregular in all but the Ells River. Neill and Evans (1979) indicated that sodium increases associated with spring thaw are derived from surficial runoff and lake contributions. High winter values are undoubtedly derived from groundwater sources. Wetzel (1975), indicated that sodium concentrations vary little seasonally in lakes due to the conservative nature of this ion, hence sodium concentrations fluctuate little in the Ells River which originates as a lake.

On a worldwide basis lotic sodium is highly variable. European rivers average 5.4 mg  $L^{-1}$ , North American rivers average 9 mg  $L^{-1}$  Whitton (1975). With the exception of the Ells River the average sodium concentration among these lotic systems is 24.6 mg  $L^{-1}$ . High values for this region are possibly related to considerable subsurface groundwater flow (Neill and Evans, 1979) which is extremely high in chloride (i.e. 10,000 mg  $L^{-1}$ ) and total dissolved solids (19,000 mg  $L^{-1}$ ) (Smith, 1979).

#### 5.2.1.6. Potassium

With the exception of the Hangingstone River potassium concentrations were unrelated to discharge. In the Hangingstone River the relationship was linear (Appendix 10). Potassium undoubtedly enters these rivers in surface runoff being mobilized in particulate form or leached from the soils during the melting of the snow as well as periods of high rainfall. This can easily occur during the summer months because 50 to 75% of the annual rainfall occurs during this period, (Smith, 1979). Potassium concentrations also fluctuated most widely during 1978 (spring) coincident with above average rainfall further evidencing surface runoff mobilization.

Potassium concentrations among the five rivers averaged 1.2 mg  $L^{-1}$  with the highest occurring in the Steepbank. The average among the North Eastern Alberta rivers was lower than average values indicated by (Whitton, 1975) for European rivers (1.7 mg  $L^{-1}$ ) and North American rivers (1.4 mg  $L^{-1}$ ).

### 5.2.1.7. Magnesium

Calcium, magnesium and sodium were the dominant cations found in Muskeg River basin groundwater respectively (Schwartz, 1979). Magnesium concentrations in particular were highly correlated with discharge for all the rivers. Lotic concentrations were also particularly high during the winter months in all rivers except the Ells. Values fluctuated least in this river, averaging 3.6 mg L $^{-1}$ . In the remaining rivers the concentration of magnesium averaged 8.0 mg L $^{-1}$ , hence exceeding the mean values reported for European rivers (i.e. 5.6 mg L $^{-1}$ ) and North American rivers (i.e. 5.0 mg L $^{-1}$ ) (Whitton, 1975). Magnesium compounds are more soluble than their calcium counterparts and precipitate only at pH values greater than 10.

Since pH values were never this high in any of the rivers, summer fluctuations can only be attributed to algal uptake and allochthonous inputs. Moreover, magnesium adsorbs silicate at low  $\rm SiO_2$  concentrations (3.0 mg L<sup>-1</sup>) thus affecting its availability to diatoms.  $\rm SiO_2$  values less than 3.0 mg L<sup>-1</sup> were detected in all of the rivers studied.

#### 5.2.1.8. Iron

Iron was not limiting in any of the rivers. Mean concentrations among the five rivers ranged from 0.08 mg L<sup>-1</sup> (Ells) to .18 mg L<sup>-1</sup> (Muskeg). The highest values were detected during the winter except in the Ells River. Due to the presence of high dissolved organic matter content originating in the bogs which constitute a greater portion of each river basin, biologically available iron is abundant. Sapiro (1957) in Wetzel (1975) indicated that high iron concentrations are associated with high levels of humic acids, tannic acids, Hem (1960) in Wetzel (1975) and other lignin derivatives. The fron complexes in fact impart an intense yellow-brown color of each river. Moreover, Wetzel (1975) indicated that the typical range of total iron found in oxygenated surface waters is about 0.05 to 0.2 mg L<sup>-1</sup>. All of the rivers exceed this range due to the presence of bog waters within the river basins.

### 5.2.1.9. Manganese

Hutchinson (1957) indicated that manganese concentrations in aquatic systems are variable in relation to lithology and drainage patterns. Wetzel (1975) reported that values generally range between 0.1 and 0.85 mg  $L^{-1}$  and average 0.035 mg  $L^{-1}$ . Compared to these values manganese concentration in North Eastern Alberta rivers is higher than average (i.e. 0.14 mg  $L^{-1}$ ). Sources of manganese accounting for elevated values include local alkaline soils Wetzel (1975), groundwater sources

(4)

(Schwartz, 1979) and drainage from the coniferous forests which occur within every drainage basin in the area (Wetzel, 1975).

Although manganese concentrations were high on average, values as low as 0.002 mg  $L^{-1}$  were detected in both the MacKay and Steepbank Rivers. Wetzel (1975) indicated that concentrations less than 0.05 mg  $L^{-1}$  are inhibitory to the development of green and blue-green algae in streams while strongly favoring diatom growth. Minimum concentrations were found in these lotic systems during the spring spate coincidentally with high relative percent contribution by diatoms in all rivers. Although values less than 0.05 mg  $L^{-1}$  were detected in all rivers during the study it did not entirely inhibit the blue-green algae perhaps due to the short duration of times in which concentrations were inhibitory.

### 5.2.1.10 Sulphate

The sources of sulphate ions in relation to lotic ecosystems include weathering of local rocks, soils, decomposition of organic matter, atmospheric precipitation and fall out from local industries. Whitton (1975) reported that North American Rivers average 20 mg L<sup>-1</sup> and European Rivers 24 mg L<sup>-1</sup> sulphate. In the MacKay River yalues were higher than the North American or European average. (ie 27.3 mg L<sup>-1</sup>). Moreover sulphate was negatively correlated with discharge (r = -0.806, p < 0.005). Values were lower in the remaining rivers and similarly the relationship between discharge and sulphate was negative in the Steepbank (r = -0.780, p < 0.005) and Hangingstone (r = -0.919, p < 0.005). Sulphate and discharge were not related statistically in the Muskeg and Ells Rivers. Sulphate concentrations generally increased during the winter period in all rivers. Wetzel (1975) attributed this pattern to bacterial metabolism. It is likely however that groundwater inputs also

influence winter concentrations. Spring and fall inputs attributed to runoff and precipitation are masked by the dilution effects of elevated discharge.

#### 5.2.1.11 Chloride

While chloride influences general osmotic salinity, balance and ion exchange, aquatic biota do not result in it having significant spatial or temporal variation in systems Wetzel (1975). Chloride originates from the weathering and leaching of calcareous deposits as well as atmospheric precipitation. In the AOSERP study area, large winter chloride maximums were detected for the Steepbank, Hangingstone and MacKay Rivers thereby indicating potential influences attributed to ground water inputs wither than weathering of local calcareous deposits. Although chloride increased during the winter in the Muskeg River, summer concentrations were greater than winter ones, indicating the effects of weathering and leaching of local calcareous deposits in this basin. All potential sources of chloride had little influence on the Ells River as chloride was undetectable. Since a great range of mean values ie. O to  $11.5~\mathrm{mg}~\mathrm{L}^{-1}$  were found for this area it would appear that an additional factor influences lotic chloride concentration. Smith (1979) indicated that marine rock formations occur widely and pronounced during the winter. Values however remain close to those reported by Wetzel (1975) ie 8.3 mg  $L^{-1}$  for fresh water systems.

## 5.2.1.12 Nitrate Nitrogen

One of the most common sources of nitrogen used for growth by algae is nitrate (Stewart, 1974). The sources of nitrate-nitrogen to aquatic systems include precipitation, nitrogen fixation and runoff. Nitrate

nitrogen is readily leached from soils (Whitton, 1975) hence at the source of the Ells River (the Birch Mountains) limited precipitation on the eastern slopes may account for the lack of distinct seasonal patterns (Neil and Evans, 1979). In the remaining rivers distinct fall peaks were evident coincidentally with the period of increased rain and runoff as well as leaf drop throughout the basins. In the Muskeg River nitrate-nitrogen fluctuated most dramatically during the 1978 fall. spate. Moreover nitrate-nitrogen was significantly correlated with discharge in the Steepbank River, and in a positive manner. Similar trends were also evident in the Muskeg, Hangingstone and MacKay rivers but only at the p 0.25 level. Such trends provide further evidence that the major nitrogen source stems from allochthonous material reaching the rivers in surface runoff. Further sources of nitrogen originate within the rivers themselves from the nitrogen-fixing blue-green algae and associated bacteria. The former represent the major algal group of these rivers.

## 5,2.1.13 Phosphate-phosphorus

The five northeastern Alberta Rivers discussed herein represent lotic systems unaffected by phosphorus additions originating from human activity. The rivers do however originate in areas rich in organic matter and can therefore be expected to exhibit higher than average total phosphorus concentrations (Wetzel, 1975). The range of values for most uncontaminated surface waters is between 0.10 and 0.50 mg L<sup>-1</sup> phosphate phosphorus. This value however is not directly comparable to Wetzels (1975) total phosphorus ranges, as it excludes phosphorus associated with colloidal particulate material removed by filtration.

Akena (1979) reported that total phosphorus ranges from 0.005 mg  $L^{-1}$  to 0.56 mg  $L^{-1}$  in the Muskeg River. Hence dissolved phosphorus in the Muskeg River (max value 0.352 mg  $L^{-1}$  Appendix 10a) represents 63% of the total phosphorus reported by Akena (1979). By addition, our values fall within those reported by Wetzel (1975) for uncontaminated surface waters.

The sources of phosphate phosphorus to lotic systems include decomposition of organic matter, sediment release, runoff and groundwater inputs. These factors with the exception of surface runoff contributed significantly to winter maximum concentrations among all rivers. Low summer values are related to adsorption of phosphate ions on sediments (Akena, 1979) and biological uptake by algae as well as other aquatic vegetation. Values were particularly high in the Hangingstone River (ie 0.60 mg  $L^{-1}$ ) during the winter period and as low as the Muskeg River during the summer period  $(0.006 \text{ mg L}^{-1})$ . This finding may suggest a human influence in the Hang Angstone River basin over the winter period, an impact possibly related to the town of Fort McMurray. In the Muskeg and Steepbank Rivers phosphate-phosphorus was negatively related to discharge (r = -0.295, p < 0.05; r = -0.625, p<.005 respectively). Similar effects of dilution on phosphate-phosphorus by increased water discharge have also been reported by Wang and Evans (1970) for the Illinois River, USA and Charlton et al (1985) for the Bow River located in Southern Alberta.

## 5.2.1.14 Dissolved Silica

Dissolved silica, the major element essential for the formation of diatom frustules, originates from the degradation of alumino-silicate minerals. The amount of silica in solution is chemically modified by

surface adsorption (on inorganic particulates) of silicic acid which reduces solubility (Akena, 1979), algal uptake and dilution from spring runoff. Dissolved silica and discharge were negatively correlated (p.005) in all but the MacKay and Ells Rivers (Appendix 10), hence disputing Akena (1979) where he indicated that silica is only slightly affected by streamflow and primarily affected by algal uptake in the spring. Winter high values are primarily derived from shallow groundwater entering streamflow during leaseflow periods.

### 5.2.1.15 pH and Total Alkalinity

All of the rivers considered during the study were yellowish brown to dark brown in color. Except for the Ells the uppermost regions which meander through muskeg were darker in color. Despite the acid nature of their origins, the water at the routine sampling sites was basic due to the presence of limestone substratum and local geology. The pH which ranged from winter lows of 6.0 in the Ells also increased to 9.0 during the summer months. Similar seasonal patterns were detected in the other Total alkalinity on the other hand peaked during the winter rivers. months when biological activity was reduced and decreased during the spring and fall seasons as biological activity accelerated. pH and total alkalinity therefore mirror each other, i.e., the primary producers reduce ambient free CO2 and calcium bicarbonate (through the precipitation of CaCO3) while increasing the pH. Moreover the continual addition of free carbon dioxide from the atmosphere allows aquatic mosses (Fontinalis) and the red alga Batrachospermum to survive. Ruttner, (1960) indicated that these species specifically require free CO2 in order to photosynthesize.

### -5.2.2 Temporal Epilithic Algal Changes

#### 5.2.2.1 Species Composition and Cell Numbers

#### 5.2.2.1.1 Species Composition

Numerically, Cyanophycean algae were the most important epilithic algae. Lyngbya aerugineo-caerula, Phormidium sp., Calothrix braunii and Anabaena affinis and Nostoc spp., frequently dominated. The epilithic communities were often several millimeters thick. Diatoms followed the blue-greens namely, Synedra ulna, Synedra rumpens, Gomphonema olinaceum, G. acuminatum, G. longiceps v. subclavata, Nitzschia fonticola, N. palea, Achnanthes lanceolata, Epithemia sorex, E. turgida, Cocconeis placentula and C. pediculus. Fjerdingstad (1950) found that Phormidium spp. and Nitzschia palea are particularly tolerant of organic enrichment. Marker, (1976a) noted that Phormidium grew best in the shade of riparian vegetation, and deep pools where the substrata was relatively stable. Hence it would appear that Phormidium is particularly well adapted to development in these rivers which are not only rich in organic matter but also have reduced light penetration due to their yellowish-brown to dark brown color. Schoeman (1973) noted That Nitzschia palea and Nitzschia fonticola both inhabit oxygen-rich, alkaline freshwaters and are good indicators of eutrophic conditions. Both Synedra ulna and S. rumpens favour weakly alkaline water and S. ulna a's well as <u>Achnantes lanceolat</u>a and Cocconeis pediculus are particularly intolerant of strong oxygen deficiencies (Schoeman, 1973). The remaining diatom species range from pH-indifferent to weakly alkaline, indicators.

In the Hangingstone River Chlorophycean algae rather than diatoms were the second most abundant. Stigeoclonium pachydermum and

cladophora glomerata were dominant. These species are morphologically similar i.e. branched, firmly attached thalli (Prescott, 1961), which were tolerant of the rapid abrasive nature of the Hangingstone River. Rhodophycean algae were found throughout all rivers except at the Ells. Batrachospermum vagum found in the Muskeg and Steepbank Rivers has been shown to prefer shaded cool (less than 20°C) flowing waters (Parker et al, 1973). Since this species was also found at unshaded sites it would appear that the characteristically brown water again favours the development of epilithic communities adapted to lower light intensities. Billard (1966) found that both the red alga Batrachospermum and Audouipella require low light. The critical factor however which influenced either genus was temperature. Light and temperature also play critical roles with respect to diatom populations however it is difficult to separate these variables due to their interrelations in photosynthesis (Whitton, 1975).

## 5.2.2.1.2 Cell Numbers

Moore (1974), after examining nine rivers of southern Baffin Island found that benthic algal numbers exhibit a bimodal peak, one in June-July the other in September. A similar pattern was found for the rivers investigated during this project. Cell numbers however found for north eastern Alberta Rivers were higher than those reported by Moore (1974). Mean cell-numbers ranged from 2.36 10<sup>10</sup> m<sup>-2</sup> in the Steepbank to 63.80 10<sup>10</sup> m<sup>-2</sup> in the Ells. Spectacular development of diatom populations were observed during the fall season in most rivers and during the summer as well in the Muskeg and Ells Rivers. Marker (1976a) similarly reported summer peaks in Bere Stream, England. During the summer months however, it appeared that diatoms were more abundant in the swiftly

flowing shallow river regions. Marker (1976a) attributed this pattern to diffusion of gases, nutrients and light attenuation. Reisen and Spencer (1970), ascribed the seasonal patterns to high flow. McIntire (1968) showed that biomass increased in artificial streams as a result of increasing irradiance and current velocity. Gruendling (1971) found that a myriad of factors, chémical and physical, as well as biotic cause cell numbers to fluctuate. Among the five rivers no one over-riding nutrient or physical factor was responsible for controlling fluctuations and population sizes of the major algal groups. Dissolved silica appeared to be potentially limiting to diatom growth in two rivers, the Hangingstone (e.g., entire study period r = 0.359, p < 0.10; open water period 1978 r = -0.614, p <0.10) and the MacKay (e.g., open water period 1978 r = -0.558, p< 0.10) rivers. Therefore, even this relationship was inconsistent from year to year in these two rivers. Wang and Evans (1969), and Edwards (1974) also found similar relationships in their studies, whereas Marker (1976) did not in shallow chalk streams in southern England. \( \) No other nutrient correlated with diatom growth in any river investigated here. Irradiance was correlated with diatom growth in the MacKay and Ells rivers (e.g., MacKay River (a) entire study period r = -0.305, p < 0.25; open water period 1978 r = -0.558, p< 0.10 and Ells River (a) open water period 1978 r = -0.472, p < 0.10, (b) open water period 1978 r = -0.870, p<0.05). Temperature was also correlated with diatom growth in the Ells River (open water period 1978 r = -0.836, p< 0.05). Current velocity was correlated with diatom growth in only the Muskeg (open water period 1978 r = 0.256, p < 0.25), MacKay (open water period 1978 r = -0.645, p<0.10) and Ells (a) open water period 1978 r = 0.626, p< 0.25, (b) open

water period 1979 r = -0.417, p< 0.25) rivers and discharge only in the latter two rivers (e.g., MacKay River open water period 1978 r = 0.739, p< 0.05, and Ells River (a) open water period 1978 r = 0.599, p< 0.25 and (b) open water period 1979 r = -0.472, p<0.25). Therefore, physical factors appeared more important than nutrients, a phenomenon not uncommon with diatoms in flowing systems (c.f. Moore 1977).

Nutrient correlations were less clear with the Chlorophyta. Calcium was implicated in the Muskeg River (open water period 1978 r = -0.523, p < 0.10), silica in the Steepbank River (entire study period 1979 r = -0.264, p < 0.25, open water period 1978 r = -0.753, p < 0.05; open water period 1979 r = 0.541, p < 0.25) and iron in the MacKay River (open water period 1978 r = -0.609, p < 0.10); none was limiting in either the Hangingstone or Ells rivers. Neither irradiance nor temperature was correlated with Chlorophycean growth or demise. However, current velocity and discharge were negatively correlated with Chlorophycean growth in the Steepbank river (open water period 1978discharge r = -0.664, p< 0.10, current velocity r = -0.786, p < 0.05), Hangingstone (open water period 1978-discharge r = -0.452, p < 0.25, current velocity r = -0.454, p < 0.25; open water period 1979-discharge r= -0.976, p < 0.005, current velocity r = 0.938, p < 0.005), MacKay (entire study period-discharge r = -0.259, p < 0.10, current velocity r = -0.259-0.466, p < 0.05; open water period 1978-discharge r = -0.557, p < 0.10, current velocity r = -0.722, p < 0.05; open water period 1979-discharge r = -0.440, p < 0.25, current velocity r = -0.764, p < 0.05) and Ells (open water period 1978-discharge r = -0.521, p < 0.25, current velocity r =-0.514, p < 0.25) rivers. Thus in spite of the variability, physically disruptivé forces were most important in potentially affecting

Chlorophycean algal populations particularly during the open water period of 1978.

Only nitrate-nitrogen correlated with the Cyanophyta and that was in the Hangingstone River (entire study period r = -0.222, p < 0.25; open water period 1978 r = -0.581, p < 0.10). Of the physical factors only current velocity and discharge were important particularly in the Hangingstone (entire study period-discharge r = -0.325, p< 0.25, current velocity r = -0.301, p <0.25; open water period 1978-discharge r =-0.627, p < 0.10, current velocity r = -0.639, p < 0.10; open water period 1979-discharge r = -0.696, p < 0.10, current velocity r = -0.647, p <0.10) and MacKay (open water period 1978-discharge r = -0.371, p < 0.25, current velocity-no significant relationship). Again physical forces appeared most important. Members of the Rhodophyta correlated with irradiance in the Muskeg (open water period 1978 - r = -0.421, p< 0.10) and Steepbank (entire study period r = 0.412, p <0.05; open water period 1979 - r = 0.494, p < 0.25) rivers as well as a number of other parameters in the Steepbank River (e.g., nitrate-nitrogen, current velocity and discharge).

## 5.2.3 Temporal Standing Crop Changes

## 5.2.3.1 Epilithon

Chlorophyll <u>a</u>, the primary photosynthetic pigment in all oxygen evolving photosynthetic organisms has been used extensively as a measure of algal standing crop. (Marker and Gunn (1977), Waters (1961), Moss (1976b), Round (1981)).

Marker and Gunn (1977) found that due to the presence of various intermediate degradation products chlorophyll  $\underline{a}$  does not necessarily correspond to live-cell counts. A similar lack of continuity was noted

during this study. Standing crop estimates were highest during the fall season of 1978 in all rivers except the Muskeg and Ells which had higher standing crop estimates during 1979. Interestingly peak standing crops occurred earlier in the fall of both years at the Ells River site.

Mean epilithic chlorophyll a values for the five rivers (Table 30) → ranged from 7.94 to 43.2 mg chlorophyll a m $^{-2}$ . The Ells river which supported the highest mean chlorophyll a is similar to the calcareous lowland streams of England during the summer  $(40 - 100 \text{ mg m}^{-2})$  (Marker, 1976a). Peak fall values in Northeastern Alberta are similar to spring peak chlorophyll a values reported by Marker (1976a) i.e. 200 - 300 mg chlorophyll  $\underline{a}$  m<sup>-1</sup>. The range of values reported for other epilithic communities i.e. hot springs and calqareous mountain rivers however exceed standing crop estimates for the rivers investigated during this study (Round, 1981). Due to the extreme variability among lotic systems not only temporally but also annually, it would appear that changes in chlorophyll a are connected to the changes in population structure. Specific dominance by diatoms i.e. spring or fall may yield particularly large standing crop estimates using chlorophyll a as an indicator. The presence of large numbers of blue green algae on the other hand may yield an underestimate of chlorophyll a due to assessory pigments which remain undetected when only chlorophyll a corrected for phaeophytin is used to measure epilithic algal standing crop.

Correlations between standing crop and various other physical and chemical factors over the entire study period and the open water periods of 1978 and 1979 were examined to provide possible information about any potential controlling factors. The results are present in Appendix 11. Over the entire study period, no one nutrient was limiting to standing.

crop size. This was undoubtedly due to physical limitations during the winter months coupled with nutrient increases under ice-cover through decreased flow. No nutrient correlated in the Muskeg and MacKay Rivers (Appendix 11) while those that did in the Steepbank River were positively correlated. Only in the Ells River did significant negative correlations occur (e.g. with  $\rm SiO_2$  and  $\rm PO_4^{-1}$ ). Thus, in general nutrient levels are more than adequate to support algal populations. Of the physical factors, current velocity and discharge were consistently the most important except in the Ells where irradiance was the major. limiting factor. Thus, physically disruptive forces appear to be more important during the entire study period than nutrient levels in all but the Ells River.

Physical factors in particular can change substantially and rapidly, sweeping away not only the algae but also the rocks to which they attach. Moreover, accumulation of epilithon increases resistance to flow particularly if the organic matrix becomes impregnated with sediment and detritus. Both Hynes (1970) and Whitton (1975) emphasized the effects of current. McIntire (1966) demonstrated its devastating impact in laboratory streams. Blum (1963) concluded that it is a primary factor which governs not only organic accumulation but also the distribution of species.

## 5.2.3.2 Phytoplankton

The temporal patterns of northern lotic phytoplankton wax and wane have been related to factors such as temperature and nutrient availability (Moore 1979). Egborge (1973, 1974) indicated that Oshun River phytoplankton were predominantly benthic detached and epilithic species. Whitton (1975) however stated that true phytoplankton populations develop in large slow flowing rivers, and Hynes (1970) noted

that backwater phytoplanktonic communities develop in lotic systems.

Although planktonic chlorophyll <u>a</u> appeared to be related to benthic chlorophyll <u>a</u> the correlation was not statistically significant for p = <0.05. Planktonic chlorophyll <u>a</u> fluctuated widely and highest values often coincided with or followed periods of high benthic chlorophyll <u>a</u>. Diatoms were the dominant algal division found in the plankton, mainly senescent epilithic species. Hooper (1947) similarly reported that diatoms were more abundant in the plankton of five rivers located in the Yukon and Mackenzie River systems and he related their origin to local lakes and ponds. Antoine (1983) reported finding direct correlations between water temperature and transparency with the phytoplankton whereas water level and currents manifested inverse correlations for the Tigris River, Iraq. Unfortunately however he disregarded the epilithic community which may in fact be similarly responding to physical factors. Moreover most of the species which he identified were also detected in the epilithic communities of Northeastern Alberta Rivers.

## 5.2.4 Temporal Productivity Changes

## 5.2.4.1 Epilithon

The difficulties associated with the highly dynamic and heterogenous nature of lotic systems has given rise to several incomparable alternative approaches to assaying primary production (Odum 1956, Wright and Milles 1967, Duffer and Dorris 1966). Their methods ranged from measurement of diurnal dissolved oxygen most commonly to the use of pH and carbon dioxide fluctuations. Whitton (1975) however pointed out that only approximate estimates of gross photosynthesis could be gained through these methods.

Although the closed circulating chambers developed during this study were smaller and less regulated with respect to internal circulation of water than those employed by Marker (1976c) they were dependable and portable. Moreover, the use of the same technique yielded comparisons among five rivers.

The temporal variation of primary productivity fluctuated more than standing crop estimates. Both standing crop and primary production were on average highest in the Ells River followed by the Muskeg River and Steepbank respectively. The Steepbank and Hangingstone supported closely similar epilithic standing crops and were similarly productive. The MacKay River which had a particularly diverse eplithon had the lowest standing crop and production. The same assay techniques were applied to larger rivers located in Southern Alberta (Charlton, Hamilton, Cross, unpublished). They reported much higher mean standing crop values as well as higher rates of <sup>14</sup>C uptake. Southern Alberta Rivers

however in addition to having a more temperate climate are dramatically influenced by man-induced eutrophication. The Bow River for example during a two year period averaged 141 mg chlorophyll a m<sup>-2</sup> and 77 mg C m<sup>-2</sup> hr<sup>-1</sup> (Charlton, Cross, Hamilton unpublished). Northeastern Alberta Rivers averaged 25 mg Chlorophyll a m<sup>-2</sup>, and 17 mg C m<sup>2</sup> hr<sup>-1</sup>.

Epilithic algal standing crops were correlated with primary productivity in all but the MacKay River during the open water period of 1979 (Appendix &). The factors which controlled epilithic primary productivity were more variable. Actually, no factor examined correlated with epilithic primary productivity during the open water period of 1978 other than standing crops in the Muskeg, MacKay and Hangingstone Rivers. In the Ells and MacKay Rivers however many factors

correlated with productivity. A summary of these factors is given in Appendix 8.

The mean specific rates of photosynthesis were also compared among the five rivers. The highest rate was calculated for the Muskeg River  $(8.64 \text{ mg C (mg Chlorophyll a})^{-1}.\text{hr}^{-1})$  followed by the Ells  $(3.6 \text{ mg C (mg Chlorophyll a})^{-1}.\text{hr}^{-1})$ , the Steepbank  $(3.01 \text{ mg C (mg Chlorophyll a})^{-1}.\text{hr}^{-1})$  and the Hangingstone  $(0.82 \text{ mg C (mg Chlorophyll a})^{-1}.\text{hr}^{-1})$ . These values are greater than found by McConnell and Sigler (1959), Bowbowna (1972), and Marker (1976b) for benthic algae in rivers. The mean value in the MacKay River was similar to that reported by Hickman (1971a) for epiphytic algae in a small pond. Values for the other rivers were similar to values found by Hickman and Klarer (1975) for epiphytes in Lake Wabamum, Alberta (range 5% to  $20.8 \text{ mg C (mg chlorophyll a})^{-1}.\text{h}^{-1})$ .

Although phytoplankton standing crops were on average higher in the Hangingstone River the highest mean rate of  $^{14}\text{C}$  uptake was found in the Ells River. Higher production values could be expected in the Ells due to the sampling sites proximity to a series of lakes which give rise to the Ells. Similar values found in the Muskeg probably reflect the presence of actively growing cells originating from the upstream pools and backwaters. Values found for these rivers exceed those found for southern Alberta Rivers (Charlton et al 1985) possibly due to the slow flowing nature at their origin as well as frequent backwaters arising from pools formed by beaver dams and organic obstruction of the flow.

#### 5.2.4.3 Microhabitat Differences

Lotic primary productivity has been shown to vary not only seasonally but spatially. Differences in microcolonization patterns particularly in relation to current velocity have been reported by Round (1981) Gessner (1955) and Gumtow (1955), Korte and Blinn (1983). Patrick (1970) described a stream ecosystem as consisting of mafty small,, compact communities, in which the algae vary in habitat and Gumtow (1955) showed that even within local habitats, distribution. micro-distributional variations are evident. Microhabitat differences have been attributed to a variety of factors including current velocity (Jones 1978, Gumtow 1955), grazing (Hynes, 1970), detrital microcosms (McIntire and Phinney, 1965), light or lack thereof (shading) (Whitton, 1975), (Hynes, 1970), and the availability of substrate suitable for colonization (Round, 1981). None of these studies however has employed the light/dark bottle technique to assay epilithic primary production in various microhabitats simultaneously.

When shore ( 15 cm depth) midstream (50-60 cm depth) and shaded midstream epilithic productivity were compared, shading reduced the mean rate of production as well as the photosynthetic index or efficiency. Interestingly however the quantity of chlorophyll a increased. This finding suggests a chromatic adaptation i.e. the blue green dominant epilithic algae may increase their chlorophyll a content. Evidence currently exists for this phenomenon among marine algae (Round, 1981), hence we could expect freshwater species particularly those adapted to survival in streams containing colored water (which changes seasonally) to maximize their utilization of incoming light. Towns (1981) shaded a lotic system sufficiently to allow only 6% of the outside light beneath

a canopy. He found that this degree of shading resulted in the disappearance of periphyton. This conclusion, however, specifically demands careful examination of the substrata to ensure that species not visible to the naked eye are not present.

The microhabitat study provided evidence that epilithic algal standing crops develop earlier and achieve higher values initially in the shallow river regions and although chlorophyll <u>a</u> never reaches the levels of those found for midstream sites, the epilithon are very productive. Midstream communities which receive less light due to absorbance and reflection from the overlying water, on the other hand, have a lower photosynthetic index. Shaded habitats respond less rapidly but their efficiency closely resembles that found in the shore region. Thus it would appear that the epilithon of the brown water alkaline streams of northeastern Alberta are particularly well adapted to variable incoming light. The heterogeneity of lotic epilithic communities therefore demands particular caution with respect to defining epilithic standing crops and productivity. All microhabitats must be examined during any attempt to discern quantitatively lotic epilithic algal standing crops and productivity.

## 5.2.4.4 Substrata Comparison

Evidence currently exists to suggest that size, texture and chemistry may all, at times, influence the distribution of benthic algae in streams (Whitton, 1975). Nielsen (et al, 1984) found that rock substrate showed higher absolute values of chlorophyll a (56%) than those collected from glass substrates. Wetzel (1975) believed that the number of discrepancies found between algal populations on natural and artificial substrates were sufficiently large to necessitate a thorough evaluation for each study.

Although artificial substrata were not employed to routinely monitor epilithic primary production, visual observations did suggest that discrepancies existed between naturally occurring bitumen and rock substrata. Moreover synthetic crude oil derived from bitumen was shown to have a stimulatory action upon bacterial, algal and macroinvertebrate organisms in the Muskeg River (Lock et al, 1981a, 1981b).

In the Steepbank River, microalgae were more abundant upon rock (limestone) substrate while macroalgae, particularly Nostoc commune, was more abundant upon bitumen substrate. However, epilithic primary productivity was 22% higher on the rock substrata than on the bitumen substrata. Hellebust  $\underline{et}$   $\underline{al}$  (1975) found that crude oil exposed to benthic algae initially caused bleaching of blue-green algae at superoptimal light intensities, however in long term experiments it was stimulatory to some blue green algae. In the Steepbank River the potential for bleaching is reduced by the brown coloration of the overlying water, moreover the benthic communities are continually exposed to weathered oils derived from local runoff as well as bituminous substrate erosion: Thus it would appear that rock substrata are better suited to epilithic community development, Bitumen substrata do however provide a selective advantage for the development of species such as Nostoc spp. where the individual filaments are protected from high concentrations of newly released oil products. During periods of high water temperatures, bitumen substrata were in fact found to be very soft, weak and brittle and continually contaminated equipment with tar. The soft easily eroded nature of the bitumen substrate may also be less suitable for colonization by micro-blue-green algae filaments. Conversely Nostoc commune may utilize the bitumen substrate as a source

of nutrients and hence outcompete other epilithic species. Further investigation is required to discern the specific reasons for higher numbers of <u>Nostoc commune</u> on bitumen substrate; it may in fact be simply related to bitumen providing a substrate more suitable for colonization by <u>Nostoc commune</u> reproductive structures.

#### 5.2.4.5 Diurnal Experiment

There is an obvious variation in photosynthetic activity between day and night, but less obvious is a disparity in the amount of carbon fixed during succeeding intervals in the light period (Round, 1981). Generally 2-4 hr experiments are performed in situ to discern phytoplankton carbon uptake and the data extrapolated to the whole light period. Round (1981) indicated that such extrapolation often yields misleading values hence daily rates should be computed only from a sequential series of short exposures.

Routine studies to estimate epilithic primary productivity were conducted from 1000 through 1400 hrs. Values averaged 26.2 mg C m<sup>-2</sup> hr<sup>-1</sup> for this period during the diurnal experiment. The average rate for the entire 24 hour period was 13.9 mg C m<sup>-2</sup> hr<sup>-1</sup>. The average rate however for the daylight period (0800-2000 hrs) was 27.4 mg C m<sup>-2</sup> hr<sup>-1</sup>. Thus the routine experiments conducted from 1000 hrs through 1400 hrs closely estimated mean epilithic primary productivity for the daylight period (0800-2000 hrs).

Each successive series of overlapping incubations utilized whole rock samples totalling 106 individual estimates of epilithic algal standing crop. Mean standing crop estimates per incubation set ranged from 5.4 mg chlorophyll  $\underline{a}$  m<sup>-2</sup> to 13.9 mg chlorophyll  $\underline{a}$  m<sup>-2</sup>. The grand mean standing crop estimate was 7.1 mg chlorophyll  $\underline{a}$  m<sup>-2</sup> having a

standard deviation of 5.13. Hence lotic epilithic algal standing crops are extremely heterogeneous.

The high degree of variability among samples can be attributed to not only physical and chemical regimes but also the species composition and physiological state of the epilithon. Evidence currently exists to indicate that chlorophyll <u>a</u> content of cells also exhibits a circadian rhythm (Round, 1981). The high degree of heterogeneity among epilithon, however render it virtually impossible to define whether or not this factor contributed to the het geneity observed during the diurnal study.

In general the Muskeg River exhibits a peak of carbon fixation as well as photosynthetic efficiency which occurs between mid-morning and the early afternoon. Measurement of carbon uptake during this period provided a representative measure of epilithic primary production. Caution however is warranted with respect to defining epilithic algal standing crops especially if small sample techniques are employed.

## 9.2.5 Temporal Nitrogen Fixation Changes

## 5.2.5.1 Epilithon

Seasonal studies of epilithic algal nitrogen fixation in five brown water rivers revealed distinct peaks of fixation during the symmer as well as much smaller rates when the water temperature fell below 506 and under ice-cover of winter. However, nitrogen fixation was detectable over the entire temperature range encountered in these rivers (Fogg and Stewart 1968, Horne 1972, Stewart 1970). Cyanophycean algae were numerically dominant throughout the year (Hickman et al 1978, 1980) with greatest development during the summer, thus nitrogen fixation paralleled their wax and wane in their biomass. Moreover, a direct correlation existed between mean Cyanophycean population size for each

river and both light and dark incubation. Light fixation rates were greater than those in dark confirming the light dependent nature of nitrogen fixation (Dugdale and Dugdale, 1962, Goering and Ness 1964, Horn and Fogg 1970). The high dark fixation rates found during the study were probably due to epilithic community exposure to light prior to the experiement which began at 1000 hours (Dugdale and Dugdale, 1962, and Fay, 1976).

Light nitrogen fixation among the five rivers ranged from 1.52 to 27.14 u moles N<sub>2</sub> hr<sup>-1</sup> m<sup>-2</sup>. These values are similar to those found by Horne and Carmiggelt (1975) and Horne (1975) for Californian streams. Both the Californian streams and the rivers (except the Muskeg) of northeastern Alberta supported extensive development of heterocystous Nostoc spp which formed variously sized gelatinous clumps on the stable rock substrata. In the Muskeg River both heterocystous and non-heterocystous blue-green algae which dominated the epilithon, fixed less nitrogen than in the other rivers. This routine study site however was exceptional. Only very large stable boulders supported Nostoc and samples could not be obtained for assay which included Nostoc spp.

Horn and Fogg (1970) found that rates of nitrogen fixation in aquatic systems are highest when nitrate concentrations are less than 0.3 mg  $L^{-1}$ . With the exception of the MacKay River nitrate-nitrogen concentrations averaged less than this value in all rivers. Hence bluegreen algae as well as other nitrogen-fixing components of the epilithic community are probably important contributors to the overall nitrate-nitrogen budgets of these northeastern Alberta rivers.

#### 5.2.5.2 Microhabitat Differences

As indicated earlier, blue-green algal species were dominant among all of the microhabitats investigated in the Muskeg River. Both heterocystous and non-heterocystous species were present however colonial species were dominant in the shaded midstream while filamentous species were dominant in the unshaded areas. Pronounced fluctuations of numbers were detected during the summer in the unshaded habitats however, only a fall peak was detected in the shaded habitat.

Artificial shading depressed the rate of nitrogen fixation as well as the amplitude of nitrogen fixation rates. Potts and Whitton (1977) also noted the marked influence of light on nitrogen fixation and reported that both heterocystous and win-heterocystous blue-green algae are capable of fixing elemental nitrogen. Rudd and Hamilton (1975) emphasized the need to include assays of low light regions when estimating aquatic nitrogen fixation.

This study provides further evidence of the impact of not only shading but also the need to assay both light and dark samples when defining lotic nitrogen fixation. Moreover the rates of nitrogen fixation varied spatially and were greatest in the midstream epilithic community. The marked spatial variability of nitrogen fixation therefore demands careful assessment of lotic characteristics prior to estimating daily or annual nitrogen inputs from biological sources.

The midstream unshaded community of the Muskeg river represents the lotic habitat which is the most metabolically active., Both heterotrophic and autorophic organisms operate simultaneously to maintain community stability.

### 5.2.5.3 Substrata Comparisor

Blinn (et al, 1980) suggested that microtopographic differences among lotic substrates and possibly the rate of substrate solubilization may affect colonization. A definitive example of the chemical interaction between substratum and colonizers was presented by Parker (et al 1973). They found that particular elements and substrate types were species-specific. In the Steepbank River Nostoc commune, a known nitrogen fixer, (Stewart, 1974) was particularly abundant on bitumen, substrata. Horne (1975) and Horne and Carmiggelt (1975) showed that Nostoc species grows on stones mainly in early summer and fixes nitrogen at a maximum rate during the day, is lowest in the evening but picks up during the night.

In the Steepbank River total blue green algal numbers were higher on rock substrates but nitrogen fixation was highest on the bitumen substrate where Nostoc commune was most abundant. Both light and dark assays indicated the significant contribution of Nostoc colonies toward the total rate of nitrogen fixation. During early September however nitrogen fixation and Nostoc colonies suddenly decreased exhibiting a temporal pattern similar to that reported by Horne and Carmiggelt (1975). They attributed this phenomenon to scour resulting from turbulent water flow which prevented autumn establishment of Nostoc colonies. This phenomenon also occurred in the Steepbank River. Therefore physical forces also exert an influence on potential lotic nitrogen fixation, the effect of which is more profound for bitumen substrata than rock substrata.

### 5.2.5.4 Diúrnal Experiment

The diurnal variation of nitrogen fixation and the effects of light upon this process are indeed complex but related. Although it is a light dependent process (Stewart, 1974); some authors have also shown that blue-green algae continue to fix nitrogen during the dark period (Hay, 1965, Horne and Fogg, 1970, Duong 1972). The highest rate of nitrogen fixation-(3.15 u moles  $N_2\ m^{-2}\ hr^{-1}$ ) was detected during the morning (after 0800 hrs). Horne (1979) similarly reported this phenomenon for Clear Lake, California and in both cases the rates decreased after the morning peak despite increasing irradiance. Nitrogen fixation efficiency (u moles N<sub>2</sub> mg $^{-1}$  chlorophyll  $\underline{a}$  m $^{-2}$  hr $^{-1}$ ) in clear glass bottles was maximum between 1800 and 2300 hr, whereas dark bottle efficiency was highest between 2300 and 0400 hr; thus it would appear that recent light exposure is critical for the production of enzymes and other products required for the process of nitrogen fixation after which the efficiency of nitrogen fixation is at a maximum. It would seem however that epilithon has a limited capacity to continue this process as nitrogen fixation rates declined to a minimum by early morning just prior to daybreak (0400 hrs). The occurrence of continual dark fixation during both the light and the dark period incubations suggests that nitrogen fixation is a characteristic activity associated with facultative heterotrophes which operate whether or not light is available. The specific contribution by algae apart from the role of bacterial nitrogen fixers however were not defined during this study. Teal and Berlo (1970) reported that bacterial nitrogen fixation in salt marsh sediment was more than ten times larger than algal fixation. Therefore further investigation is essential for a better understanding of the role of eplithic algae in relation to lotic nitrogen fixation.

# 5.2.6 Cluster and Principal Components Analysis

Although cluster analysis had several important disadvantages it proved to be a useful tool for evaluating such a large data base. Cluster analysis produced hierarchic clusters regardless of the structure of the original matrix. Hence data which may have been uniformly distributed in ecological hyperspace were clustered into a hierarchy even if no such structure existed in nature. This technique also gives equal weight to each variable included in the study. Despite these assumptions some relationships were discerned. It showed that the lotic systems located in northeastern Alberta are primarily affected by fluctuations in river discharge, velocity, temperature and intrients which profoundly influence the epilithic communities. Annual spates explain 22.93% of the data variance and season-related patterns of discharge, velocity and nitrate-nitrogen (lotic influence) act as major factors explaining an additional 16.58% of the data variability. 12.39% of the data variance remaining is primarily explained by the influence of nutrients specifically nitrate-nitrogen and phosphorus. Temperature is also important particularly in relation to diatom and Chlorophycean eplithic algae. Other authors have underlined the importance of the hydrologic regime for large tropical rivers (Stoli, 1975). Garcia de Emiliani and Depetris (1982) and on southern Alberta Rivers (Charlton et al 1985). Due however to a lack of studies quantifying the factors that control the functioning peculiar to these systems, much-needed comparisons are not yet available. Blum (1956) pointed out the frequently abrupt changes which occur monthly as well as weekly in lotic systems. Hence many difficulties surround attempts to correlate biological results with environmental causes.

#### 6. Conclusions

This study has described the major epilithic algal groups, species composition, succession of dominants, standing crops (chlorophyll a and cell numbers), primary productivity and nitrogen fixation fluctuations in relation to the physical-chemical regime of five brown water rivers. The factors influencing epilithic algae, their population wax and wane, standing crops, primary productivity and nitrogen fixation have been indicated. No one factor is responsible; rather, interacting factors both disruptive and non-disruptive and of these, current velocity and discharge are most important. The latter is also related to the chemical makeup of the water itself. Thus, discharge and to a lesser extent the biota, modify the nutrient status of the water. Current velocity, a physically disruptive force, is important in affecting and controlling population size, standing crops and primary productivity, the latter through controlling standing crop size. Naturally, some of the epilithic algae grow better in faster flowing water since they are, better adapted than others. Their growth is enhanced by increases in current velocity e.g., Cyanophyta compared to filamentous Chlorophyta.

Epilithic algae exhibit an extremely heterogenous spatial variability with respect to standing crop, primary productivity and nitrogen fixation. The midstream epilithic communities are the most stable followed by shore and shaded communities respectively. Even in the presence of 80% shading, eplithic algae are successful suggesting a specific adaptation to low light utilization. In addition, the epilithon of the five brown water rivers exhibited a preference for rock substrata with the exception of Nostoc commune which preferred bituminous substrata. There appears to be some relationship between

epilithic colonization, and substrate type. Cyanophycean algae are the dominant eplithic forms in northeastern Alberta Rivers followed by diatoms. The epilithon are important sources of new nitrogen to the overall nutrient regimes of the Muskeg, Steepbank, Hangingstone, MacKay and Ells Rivers.

Finally, this thesis has provided an extensive data base which documents the temporal patterns of physical, chemical and biotic variables contributing to the structure and function of lotic systems. It prowided evidence of the great dissimilarity which wists between lotic and lentic systems.

#### **BIBLIOGRAPHY**

- Akena, A.M. 1979. An intensive surface water quality study of the Muskeg River watershed. Volume I: Water Chemistry. Prep. for the Alberta Oil Sands Environmental Research Program by Alberta Environment. AOSERP Report 85. 187 pp.
- Allen, M.B., D.I. Arnon, 1955. Studies on nitrogen-fixing blue-green .algae. I. Growth and nitrogen fixation by Anabaena cylindrica Lemm. Plant Physiol., 30, 366-372.
- Anon, 1976. Standard methods for the examination of water and wastewater. American Public Health Association. 14th ed. Washington, D.C. 1193 pp.
- Antoine, S.E. (1983) Algal Flora of the River Tigris, Iraq. Nova Hedwigia, 30(7) '535-542'
- Blinn, D.W., A. Frederickson, V. Korte. 1980. Colonization rates and community structure of diatoms on three different rock substrata in a lotic system. Br. Phycol. Soc. 303-310
- Blum, J.L. 1963. The influence of water currents on the life functions of algae. Ann. N.Y. Acad. Sci. 108:353-358
- Blum, J.L. 1956. The ecology of river algae. Botanical Review, Vol XXII #5 291-341.
- Bombowna, M. 1972. Primary production of a montane river. Pages 661 to 671 in 2. Kajak and A. Hillbrichtllkowska, eds. Productivity problems of freshwaters. Proc. I.B.P. U.N.E.S.C.O. Symp. Prod. Prob. Freshwaters Kazimierz Dolny, 1970. Polish Scientific Publications, Warsaw and Krakow.
- Bond, W.A., K. Machniak. 1977 Interim report on an intensive study of the fish fauna of the Muskeg River watershed of northeastern Alberta. Prep. for Alberta Oil Sands Environmental Research Program by Fisheries and Environment Canada, Dep. of Fisheries. AOSERP Report 26. 137 pp.
- Boon, P.J., S.W. Shires, 1976. Temperature studies on a river system in north east England. Freshwat. Biology. 6,23-32.
- Bourrelly, P. 1970. Les Algues d'eau douce. Tome III. Les Algues bleues et rouges, Les Eugleniens, Pendinens et Cryptomonadines. N. Boubee & Cie, Paris. 512 pp..
- Bourrelly, P. 1968. Les Algues d'douce. Tome II. Les Algues jaunes et brunes, Chrysophycees, Pheophycees, Xanthophycees et Diatomees. N. Boubee & Cie, Paris. 483 pp.
- Bourrelly, P. 1966. Les Algues d'eau douce. Tome I. Les Algues Vertes. N. Boubee & Cie, Paris. 511 pp.

- Butcher, R.W. 1932. Studies in ecology of rivers. II The microflora of rivers with special reference to the algae on the river bed. Ann. Bot. 46:813-862.
- Cairns, J., R.L. Kaesler and R. Patrick. 1970. Occurrence and distribution of diatoms and other algae in the upper Potomac River. No. Nat. 436:2-12
- Carrigy, M.A., J.W. Kramers (eds.) 1973. Guide to the Athabasca Oil Sands area. Prep. for Canadian society of Petroleum Geologists Oil Sands Symposium. Alberta Research Council. Information Series 65. 213 pp.
- Cattaneo, A. Ghittori S., V. Vendegna. 1975. The development of benthonic phytocoenosis on artificial substrates in the Ticino River. Oecologia 19, 315-327.
- Charlton, S.E.D., H.R. Hamilton, P.M. Cross. 1985. The Limnological Characteristics of the Bow, Oldman and South Saskatchewan Rivers II The Primary Producers. Tech. Rept. (in prep).
- Charlton, S.E.D., M. Hickman. 1984. Seasonal, Physical, Chemical and Algal Changes in Five Rivers flowing through the oil sands region of north-eastern Alberta, Canada. Internat. Revue. ges Hydrobiologia. 69(3):297-332.
- Charlton, S.E.D., M. Hickman & C.G. Jenkerson. 1981. Longitudinal physio-chemical and algal surveys of rivers flowing through the oil sands region of northeastern Alberta, Canada. Nova Hedwigia XXXV. pp. 465-522.
- Cleve-Euler, A. 1951-1955. Die diatomeen von Schweder und Finnland. K. svensk Vetensk-Akad. Handl. Fjarde Ser., 2.1, 3.3, 4.1, 4.5, 5.4. 1172 pp.
- Crowther, R.A. 1980. Ecological Investigations of Hartley Creek, Alberta. PH.D. Thesis University of Calgary. 288 pp.
- Dillard, G.E., 1966 The Seasonal periodicity of Batrachospermum macrosporum Mont. and Audouinella Violacea (Kuetz.) Ham. in Turkey Creek, Moore County, North Carolina. Jr. of Mitchell Sci Society., 82 204-207.
- Douglas, B. 1958. The ecology of the attached diatoms and other algae in a stony stream. J. Ecol. 46:295-322.
- Duffer, W.R., T. Dorris. (1966) Primary productivity in a southern Great Plains Stream. Limnol. Oceanogr. 11:143-151.
- Dúgdale, V.A., R.C. Dugdal 1962. Nitrogen metabolism in lakes. 2. Labol. Oceanogr. 7:170-177.

- Duong, T.P. 1972. Nitrogen fixation and productivity in eutrophic, hardwater lakes. In situ and laboratory studies, Ph.D. Thesis, Michigan State University.
- Eaton, J.W. & B. Moss. 1966. The estimation of numbers and pigment content in epipelic algal populations. Limnol. Oceanogr., 11, 584-595.
- Edwards, A.M.C. 1974. Silica depletion in some Norfolk rivers. Freshwat. Biol. 4:267-274.
- Egborge, A.B. 1973. A preliminary check-list of the phytoplankton of the Oshun River, Freshwat. Biol. Vol. 3. 569-572.
- Egborge, A.B. 1974. The seasonal variation and distribution of phytoplankton in the River Oshun, Nigeria. Freshwat. Biol. Vol 4 177-191.
- Eichenberger, E., K. Wukrmann. 1975. Growth and photosynthesis during the formation of a benthic algal community.
- de Emiliani, M.O., P.J. Depetris. 1982. A principal components analysis of the phytoplankton from a pond in the Parana River Valley, Argentina. Hydrobiologia 89, pp 117-122.
- Fay, P. 1976. Factors influencing dark nitrogen fixation in a blue-green alga. Appl. Environ. Microbiol. 31: 376-379.
- Fay, P. 1965. Hetrotrophy and nitrogen fixation in <u>Chlorogloea</u> fritschii. J. Gen. Microbiol. 39:2-20.
- Fjerdingstad, E. 1950. The microflora of the River Mollea with special reference to the relation of the benthal algae to pollution Folia Limnologica Scandinavia, No. 5, 123 pp.
- Flett, R.J., J.N.M. Rudd, R.D. Hamilton.1975. Acetylene reduction assays for nitrogen fixation in freshwaters: a note of caution. Appl. microbiol. V.29:5 580-583.
- Fogg, G.E., W.D.P. Stewart. 1968. In situ determinations of biological nitrogen fixation in Antarctica. Br. Antarct. Survey Bull. 15:39-46.
- Fox, M., Ross W.A. 1979. The influence of oil sands development on trapping in the Fort McMurray region. Prep for the Alberta Oil Sands Environmental Design, University of Calgary. AOSERP Project LS 26.2 136 pp
- Gessner, F. 1955. VEB Deutscher Verlag für Wissenschaft, Hydrobotanik I. Energiehaushalt. Berlin. 517 pp.
- Goering, J.J., J.C. Neess. 1964. Nitrogen fixation in two Wisconsin Lakes. Limnol. Oceanogr. 9:530-539.

- Gruendling, G.K. 1971. Ecology of the epipelic algal communities in Marion Lake, British Columbia. J. Phycol. 7:239-49.
- Gumtow, R.B. 1955. An investigation of the periphyton in a riffle of the West Gallatin River, Montana. Trans. Am. Microsc. Soc. 74:278-92.
- Groendza, A.R., M.L. Brehmer. 1960. A quantitative method for the collection and measurement of stream periphyton. Limnol. Oceanogr.a 5:190-194.
- Haaslam, S.M. 1978. River plants. The macrophytic vegetation of watercourses. Cambridge Univ. Press. Cambridge, U.K. 396 pp.
- Hansmann, E.W. and H.K. Phinney. 1973. Effects of logging on periphyton in coastal streams of Oregon. Ecology 54:194-199.
- Happy, C.M. 1970. Some physio-chemical investigations of stratification in Abbot's Pool, Somerset, III. The distribution of some dissolved substances. J. Ecol. 58:621-624.
- Hardy, R.W., R.D. Holsten, E.K. Jackson, R.C. Burns, 1968. The acetylene ethylene assay for No fixation: Laboratory and field evaluation. Plant Physiol., 43:1185-207.
- Hellebust, J.A., B. Hanna, R.G. Sheath, M. Gergis, T.C. Hutchinson. 1975. Experimental crude oil spills on a small subartic lake in the MacKenzie Valley N.W.T.: Effects on phytoplankton, periphyton, and attached aquatic vegetation. Proceed. 1975 conf. "Prevention and Control of Oil Pollution" Mar. 25-27, San Francisco, California, U.S.A.
- Henriksson, E. 1971. Algal nitrogen fixation in temperate regions. Plant and Soil Special Vol. 415-419.
- Hickman, M. and F.E. Round. 1970. Primary production and standing crops of epipsammic and epipelic algae. Br. Phycol. J. 5:247-255.
- Hickman, M., S.E.D. Charlton, C.G. Jenkerson. 1982. A comparative study of bent algal primary productivity in the AOSERP study area. Prep. for the Alberta Oil Sands Environmental Research Program and Department of Botany. University of Alberta and Department of Plant Sciences, University of Western Ontario. AOSERP Report 128. 139 pp.
- Hickman, M., S.E.D. Charlton, C.G. Jenkerson. A longitudinal physico-chemical and algal survey of five rivers flowing through the AOSERP study area. Prep. for the Alberta Oil Sands Environmental Research Program by Department of Botany, University of Alberta. AOSERP Project WS 1.3.4. 113 pp.
- Hickman, M. 1969. Methods for determining the primary productivity of epipelic and epipsammic algal associations. Limnol. Oceanogr. 14:936-941.

- Hickman, M. 1971. The standing crop and primary production of the epiphyton attached to Equisetum fluviatile L. in Priddy Pool, North Somerset. Br Phycol. J. 6:51-59.
- Hickman, M. 1971b. Standing crops and primary productivity of the epipelon of two small ponds in North Somerset, ULK. Oecologia (Berl.) 6:238-253.
- Hickman, M. 1973. The standing crop and primary productivity of phytoplankton in Abbot;s Pond, North Somerset. J. Ecol. 61:269-287.
- Hickman, M. 1974. The standing crop and primary productivity of the epiphyton att dend to Chiloscypus polyanthus (L.) Chorda rivularis (Schrad.) Nees in a spring fed stream. Arch. Hydrobiol. 73:464-469.
- Hickman. M. 1976. Seasonal cycles of epipelic diatoms at a shallow littoral station in a large lake. Arch. Protistenkd. 118:365-375.
  - Hickman, M. and D.M. Klarer. 1975. The effect of the discharge of thermal effluent from a power station on the primary productivity of an epiphytic algal community. Br. Phycol. J. 10:81-91.
  - Hooper, F.F. 1947. Plankton collections from the Yukon and Mackenzie River systems. Trans. Am. Microsc. Soc. 66 (1) 74-85.
  - Hooper, N.M., G.G.C. Robinson. 1978. Seasonal dynamics of epiphytic algal growth in a marsh pond: composition metabolism and nutrient availability. Canadian Journal of Botany 56(19):2441-2448.
  - Horne, A.J. and C.F.W. Carmiggelt. 1975. Algal nitrogen fixation in Californian streams: seasonal cycles. Freshwater Biol. 5:461-470.
  - Horne, A.J. 1975. Algal nitrogen fixation in Californian streams: diel cycles and nocturnal fixation. Freshwater Biol. 5:471-477.
  - Horne, A.J. 1972. The ecology of nitrogen fixation on Signy Island, South Orkney Islands. Brit. Antarctic Surv. Bull. No. 27, 1-18.
  - Horne, A.J., G.E. Fogg. 1970. Nitrogen fixation in some English lakes.
  - Hustedt; F. 1930a. Die Susswasser flora Mitteleuropas. Bacillariophyta (Diatomeae). Jena. (Heft 10:1-466)
  - Hustedt, F. 1930b, 1959, 1961-1966. Die Kieselalger Deutschlands,
    Osterreich's und der Schweiz unter Besucksichtigung der ubrigen
    Lander Europas sowie der angresenden Meeresgebiete. In Dr. L.
    Raberhorts Kryptogamen-flora von Deutschland, Osterrich, und der
    Schweiz vii, Teil 1:953 pp., Teil 2:845 pp.; Teil 3:816 pp.

- Hutchinson, G.E. 1967. A Treatise on Limnology, Volume I. Geography, Physics and Chemistry. John Wiley & Sons, Inc. 1115 pp.
- Hynes, H.B. 1970. The ecology of running waters. Univ. Toronto press Toronto. 555 pp.
- Ilmavirta, V., R.I. Jones. 1977. Factors affecting the <sup>14</sup>C method measuring phytoplankton production. Ann. Bot. Fennici 14:97-101.
- Jones, J.G. 1978. Spatial variation in epilithic algae in a stony stream (Wilfin Beck) with particular reference to <u>Cocconeis</u> placentula. Freshw. Biol. 8:539-46.
- Jones, J.E. 1974. A method for observation and enumeration of epilithic algae directly on the surface of stones. Oceologia (Bal) 16:1-8.
- Khoja, T., B.A. Whitton. 1971. Heterotrophic growth of blue green algae. Arch. Microbiol. 79:280-282.
- Kilham, P. 1971. A hypothesis concerning silica and the freshwater planktonic diatoms. Limnol. Oceanogr. 16:10-18.
- Korte, V.L., D.W. Blinn. 1983. Diatom colonization on artificial substrata in pool and riffle zones studied by light and scanning electron microscopy. J. Phycol. 19:332-341.
- Langford, T.D. 1970. The temperature of a British river upstream and downstream of a heated discharge from a power station.

  Hydrobiologia 35:353-375.
- Lock, M.A., P.H. John. 1979. The effect of flow patterns on uptake of phosphorus by river periphyton. Limnol. Oceanogr. 24(2):376-383.
- Lock, M.A., R.R. Wallace, D.R. Barton, S.E.D. Charlton. 1981a. The effects of synthetic crude oil on microbial and macroinvertebrate benthic river communities: Part II The Response of an established community to contamination by synthetic crude oil. Environmental Poll. (Series A) 24:263-275.
- Lock, M.A., R.R. Wallace, D.R. Barton, S.E.D. Charlon. 1981b. The effects of synthetic crude oil on microbial and macroinvertebrate benthic river communities Part I: Colonization of synthetic crude oil contaminated substrata. Environmental Poll. (Series A) 24:207-217.
- Lund, J.W.G., C. Kipling and E.D. LeCren. 1958. The inverted microscope method of estimated algal numbers and the statistical basis of estimation of counting. Hydrobiologia 1:143-170.
- MacKereth, F.J.H. 1963. Some methods of water analysis for limnologists. Freshwater Biological Association (U.K.). Publication no. 21. 71 pp.

- Margalef, R. 1948. A new limnological method for the investigation of thin layered epilithic communities. Hydrobiologia 1:215-216.
- Marker, A.F.H. 1976a. The benthic algae of some streams in southern England. I. Biomass of the epilithon in some small streams. J. Ecol. 64:343-358.
- Marker, A.F.H. 1976b. The benthic algae of some streams in southern England. II. The primary production of the epilithon in a small chalk stream. J. Ecol. 64:359-373.
- Marker, A.F.H., R.J.M. Gunn. 1977. The benthic algae of some streams in southern England. J. Ecol. 65, 223-234.
- McConnell, W.J. and W.F. Sigler. 1959. Chlorophyll and productivity in a mountain river. Limnol. Gceanogr. 4:335-351.
- McIntire, C.D. 1966. Some effects of current velocity on periphyton communities in laboratory streams. Hydrobiologia 27:559-70.
- McIntire, C.D. 1968. Physiological-Ecological studies of benthic algae in laboratory stream. J.W.P.C.F. Vol. 40, #11, Part 1.
- McIntire, C.D., H.K. Phinney. 1965. Lab studies of periphyton production and community melabiolism in lotic environments. Ecol. Monogr. 35:237-258.
- Moore, J.W. 1979. Seasonal succession of phytoplankton in a large subarctic river: Hydrobiologia vol. 67:107-112.
- Moore, J.W. 1977. Some factors affecting algal densities in a eutrophic farmland stream. Oecologia (Berl.) 29:335-340.
- Moore, J.W. 1974. The Benthic algae of southern Baffin Island. I. Epipelic communities in rivers. Jr. of Phycology Vol. 10, No. 1, pp. 50-57.
- Moss, B. 1969. Algae of two Somersetshire pools: Standing crops of phytoplankton and epipelic algae as measured by cell numbers and chlorophyll a. J. Phycol. 5:158-163.
- Moss, B. 1967a. A spectrophotometric method for the estimation of percentage degradation of chlorophylls to pheopigments in extracts of algae. Limnol. Oceanogr. 12:335-340.
- Moss, B. 1967b. A note on the estimation of chlorophyll a in freshwater algal communities. Limnol. Oceanogr. 12:340-342.
- Murphy, M.L. 1984. Primary production and grazing in freshwater and intertidal reaches of a coastal stream, Southeast Alaska, Limnol. Oceanogr. 29(4):805-815.

- Neill, C.R., B.J. Evans. 1979. Synthesis of surface water hydrology. Prep. for the Alberta Oil Sands Environmental Research Program by Northwest Hydraulic Consultants Ltd. AOSERP Report 60, 84 pp.
- Nielsen, T.S., W.H. Funk, H.L. Gibbons, R.M. Duffner. 1984. A Comparison of Periphyton Growth on Artificial and Natural Substrates in the Upper Spokane River. Northwest Science, Vol. 58, No. 4, pp 243-248.
- Odum, H.T. 1956. Primary production in flowing waters. Limnol. Oceanogr. 1:102-117.
- Parker, B.C., G.L. Samsel Jr., G.W. Prescott. 1973. Comparison of Microhabitats of Microscopic Subalpine Stream Algae. Amer. Midland Naturalist 90(1) 143-153.
- Patrick, R. 1970. Benthic Stream communities. A discussion of the factors that affect their structure and how they function. Am. Sci. 58:546-9.
- Patrick, R. and C.W. Reimer. 1975. The diatoms of the United States, Volume II, Part I. Philadelphia Acad. Nat. Sci. Monogr. 13. 213 pp.
- Patrick, R. and C.W. Reimer. 1966. The diatoms of the United States. Volume 1. Philadelphia Acad. Nat. Sci. Monogr. 13. 866 pp.
- Patrick, R. 1961. A study of the numbers and kinds of species found in rivers in eastern United States. Proc. Acad. Nat. Sci. Phila. Vol. 113 #10, 215-258.
- Potts, M., B.A. Whitton. 1977. Nitrogen Fixation by Blue-green Algae Communities in the Intertidal zone of the Lagoon of Aldabra Atoll. Oceologia 27, 274-283.
- Pratt, L. 1976. The tar sands syncrude and the politics of oil. Hurtig Publishers. Edmonton, Alberta, Canada. 197 pp.
- Prescott, G.W. 1961. Algae of the western Great Lakes area. Wm. C. Brown Col, Dubugne, Iowa. 997 pp.
- Reisen, W.K., D.J. Spencer. 1970. Succession and current demand relationships of diatoms on artificial substrates in Prater's Creek, South Carolina. J. Phycol. 6:117-121.
- Riemann, B. 1978. Carotenoid interference in the spectrophotometric determination of chlorophyll degradation products from natural populations of phytoplankton. Limnol. Oceanogr. 23:1059-1065.
- Round, F.E. 1981. The ecology of algae. Cambridge University Press, Cambridge. 653 pp.

The same of

Round, F.E. 1964. The ecology of benthic algae. Algae and Man (Ed. by D.F. Jacksen) 138-184, Plenum Press, New York.

- Round, F.E. and M. Hickman. 1971. Phytobenthos sampling and estimation of primary production. Pages 169-196 in N.A. Holme and A.D. McIntire, eds. Methods of the study of the marine benthos. International Biological Programme Handbook No. 16, Part II. Blackwell Scientific Publications, Oxford. 334 pp.
- Rowe, J.S. 1972. Forest regions of Canada. Canadian Forestry Service Publication #1300. 170 pp.
- Ruttner, F. 1960. Fundamentals of Limnology. Univ. Toronto Press. 295 pp.
- Schindler, D.W, R.V. Schmidt, and R.A. Reid. 1972. Acidification and bubbling as an alternative to filtration in determining phytoplankton production by the carbon-14 method. J. Fish. Res. Bd., Can. 29:1627-1631.
- Schoeman, F.R. 1973. A systematical and ecological study of the diatom flora of the diatom flora of Lesotho with special reference to the water quality. V & R Printers. Pretoria. 355 pp.
- Schwartz, F.W. 1979. Interim report on a hydrogeological investigation of the Muskeg River Basin. Prep. for the Alberta Oil Sands Environmental Research Program by the Department of Botany, University of Alberta. AOSERP Project HG 1.1. 104 pp. Report #48.
- Sladeckova, A. 1962. Limnological investigation methods for the periphyton "Aufwuchs" community. Bot. Rev. 28:2037-2058.
- Smith, S.B. (ed.) 1979. Alberta Oil Sands Environmental Research Program Interim report covering the period April, 1975 to November, 1978. Prep. by A.S. Mann, R.A. Hursey, R.T. Seidner and B. Kasinska-Banas. Edmonton, Alberta. 101 pp.
- Sneth, P.H. and R.R. Sokal 1973. Numerical Taxonomy. W.H. Freeman and Co., San Francisco. 573 pp.
- Stewart, W.D.P. 1974. Algal physiology and Biochemistry, Univ. of Calif. Press. Berkeley and Los Angeles. 989 pp.
- Stewart, W.D.P., H.W. Pearson, 1971. Effects of aerobic conditions on growth and metabolism on blue green algae. Proc. Roy. Soc. Land. V. 175:293-311.
- Stewart, W.D.P. 1970. Nitrogen Fixation by blue-green algae in Yellowstone thermal areas. Phycologia, 9:261-8.
- Stewart, W.D.P. 1968. Nitrogen input into aquatic ecosystems. IN Algae, Man and the Environment, ed. Jackson D.F. Syracuse Univ. Press. 434 pp.
- Stewart, W.D.P., G.F. Fitzgerlad, R.H. Burris. 1967. In situ studies on N2 fixation using the acetylene reduction technique. Proced. Nat. Acad. Sci. 58(5)2071-2078:

- Swale, E.M.F. 1964. A study of the phytoplankton of a calcareous river. J. Ecol. 52:433-446.
- Teal, J.M. I.V. Berlo. 1979. Nitrogen Fixation by Rhizosphere and Free-living Bacteria in Salt Marsh Sediments. Limnol. Oceanogr. 23 (I) 126-132.
- Tippet, R. 1970. Artificial surfaces às a method for studying populations of benthic micro-algae in freshwater. Br. Phycol. J. 5:187-199.
- Towns, D.R. 1981. Effects of artificial shading on periphyton and invertebrates in a New Zealand stream. New Zealand Jr. of Marine Freshwat. Res. V 15:27 pp 185-192.
- Turchenek, L.W., J.D. Lindsay. 1978. Interim report on a soils inventory in the Athabasca Oil Sands area. A.O.S.E.R.P. report #28. 100 pp.
- Walmsley, J.L., D.L. Bagg. 1977. Calculations of annual averaged sulphur dioxide concentrations at ground level in the AOSERP study area. Prep. for the Alberta Oil Sands Environmental Research Program by Fisheries and Environment Canada, Atmospheric Environment Service. AOSERP Report 19. 40 pp.
- Wang, W.C. and R.L. Evans. 1969. Variations in silica and diatoms in a stream. Limnol'. Oceanogr. 14:941-944.
- "Waters. 1961. Notes on the chlorophyll method of estimating the photosynthetic capacity of stream periphyton. Limnol. Oceanogr. Vol. 6. 186-188.
- Wetzel, R.G. 1975. Limnology. W.B. Saunders Col. Toronto, 743 pp.
- Wetzel, R.G. 1964. A comparative study of the primary productivity of higher aquatic plants, periphyton and phytoplankton in a large shallow lake. Int. Revue ges. Hydrobiol. 49:1-61.
- Whitford, L.A. and G.J. Schumacher. 1963. Communities of algae in North Carolina streams and their seasonal relations. Hydrobiologia 22:133-196.
- Whitford, L.A. and G.J. Schumacher. 1963. Effect of a current on respiration and mineral uptake in spirogyra and oedogonium. Ecology 45(#1):168-170.
- Whitton, B.A. 1975. River ecology, Studies in Ecology, Vol. 2, Univ. Calif. Press. Berkeley, California, U.S.A. 725 pp.
- Wishart, D. 1975. Clustan. I.C. user manual, University College, London, 124 pp.
- Wright, J.C., I.K. Mills, 1967. Productivity studies on the Madison River, Yellowstone National Park, Limnol. Oceanogr. 12:568-77.

Appendix 1. The algae found at each site in the Muskey River (= present; - absent).

|                                  |            |                   |                | Site             |                |                   | •                |                |
|----------------------------------|------------|-------------------|----------------|------------------|----------------|-------------------|------------------|----------------|
| Algae.                           | 1          | 2                 | 3              | 4                | 5              | 6                 | . 7              | 8              |
| •                                |            |                   |                |                  |                |                   | •                |                |
| CYANOPHYTA                       |            |                   |                |                  |                |                   | •                |                |
| Anabaena affinis Lemm.           | -          | +                 | +              | -                | -              | +                 | -                | -              |
| Calothrix braunii Bornet &       | -          | -                 | -              | + `              | +              | +                 | +                | +              |
| Flahault                         |            |                   | •              |                  | • . •          |                   |                  |                |
| Gomphonema lacustris v. compacta | -          | + *,              |                | -                | -              | -                 | -                |                |
| Lemm.                            |            |                   |                |                  |                |                   |                  | h              |
| Lyngbnya aerugineo-caerulea      | +          | +                 | +              | +                | + .            | +                 | +                | +              |
| (Kutz.) Gomont                   |            |                   |                |                  |                |                   |                  |                |
| Merismopedia glauca (Ehr.)       | -          | +                 | -              | <b>759</b>       | · <del>-</del> | - '               | -                | -              |
| Naegeli                          |            |                   |                |                  | •              |                   |                  |                |
| Nostoc spp.                      | - '        | · <del>-</del> ., | <del>-</del> - | -                | -              | + .               | +                | +              |
| Oscillatoria sp.                 | -          | +                 | +              | +                | 7              | -                 | <b>.</b> .       |                |
|                                  |            | •                 |                |                  | (              |                   |                  |                |
| CHLOROPHYTA                      |            |                   | . * *          |                  |                |                   |                  |                |
| Ankistrodesmus falcatus (Corda)  | -          | +                 | -              | <b>-</b> .       | +              | + ,               | +                | +              |
| , Ralfs.                         |            |                   | •              | . 3              |                |                   |                  | •              |
| Chlamydomonas sp.                | + .        | <b>‡</b>          | +              | -                | ·<br>•         | -                 |                  | -              |
| Cladophora glomerata (L.) Kutz.  |            | -                 |                | +                | +              | +                 | +                | +              |
| Closterium sp.                   | -          | *; <b>+</b>       | -              | - ·              | -              | -                 | . · ·            | <del>-</del> , |
| Coelastrum scabrum Reinsch       |            | · +               | - (            | <b>-</b> ,       | -              |                   | - ( <del>`</del> | - ·            |
| Crucigenia quadrata Morren       | · <b>-</b> | -                 | -              | -                | -              |                   | -                | +              |
| Scenedesmus bijuga (Turp.) Lager | -          |                   | _              | <b>-</b> ',      | _              | • •               | -                | +.             |
| Spirogyra sp.                    | -          | +                 |                | -                | <b>-</b>       | <b>-</b> \ .      | -                | <b>-</b> .     |
| Stigeoclonium sp.                | _          |                   |                | . <b>-</b> .     | . 4            | ,: <sup>-</sup> - | •                | +              |
|                                  |            | 1 .               |                | •                |                | *                 | •                |                |
| <b>Д</b> ҮРТОРНҮТА               | . •        |                   |                | <b>.</b>         |                |                   | • <u>-</u>       |                |
| Cryptomonas erosa Ehr.           | +          | + .               | ·              | ·<br>- ,.        | -              | ., <del>-</del> , | -                | + .            |
| 6. ovata Ehr.                    | _          | -                 | ŧ              | -                | <b>.</b>       | , <b>-</b> '      |                  |                |
| odomonas minutum Skuja           | -          | -                 | +              | - <del>-</del> : | •              | -                 | <b>.</b>         | · -            |
|                                  | •          |                   |                |                  |                |                   | •                |                |
|                                  |            |                   |                |                  |                |                   |                  |                |

## Appendix 1. Continued.

| Append IX 11 continued.        |                      | •          | ٥.       |              |              |          |            |                     |
|--------------------------------|----------------------|------------|----------|--------------|--------------|----------|------------|---------------------|
|                                |                      |            |          | Site         |              |          | ,          | <u>.</u>            |
| Algae                          | 1'\                  | 2          | 3        | 4            | 5            | 6        | 7_         | 8                   |
|                                |                      |            |          | ÷            |              |          |            |                     |
| PYROPHYTA                      | -                    | + ,        | +        | `-           | -            | -        | -          |                     |
|                                |                      |            |          | •            |              |          | ٠,         |                     |
| CHRYSOPHYTA                    |                      |            |          |              |              |          |            |                     |
| Chromulina spp.                | -                    | -          | +        | -            | -            | -        | -          |                     |
| EUGLENOPHYTA                   | s                    | -          | -        |              |              |          | <b>.</b>   |                     |
| Euglena sp.                    | -                    | + .        | +        | _            | -            | .: - · · | ۸ - ۲ - ۱  | 10.16 <u>-</u><br>1 |
| Phacus sp.                     |                      | +          | +        | <b>-</b>     | <b>-</b> , . | -        |            | -                   |
| Trachelomonas sp.              | <b>-</b> .           | +          | •        |              | - '          | · •      |            | -                   |
|                                |                      |            | •        |              |              |          |            |                     |
| RHODOPHYTA                     |                      |            | •        | ٠.,          |              |          |            | <b>~</b> :          |
| Batrachospermum vagum (Roth.)  | -                    | <b>-</b>   | -        | -            | +            | +        |            | • •                 |
| C.A. Agardh.                   |                      |            |          |              |              |          |            | ing.                |
|                                | •                    |            |          |              |              |          | ,          |                     |
| BACILLARIOPHYTA                |                      |            | َدِ ''   | • •          |              |          | ٠.         |                     |
| Achnanthes                     | +                    | -          |          | +            | * -          | -        | ·          | +                   |
| A. lanceolata Breb.            | •                    |            | rs .     |              | ,            | á .      |            |                     |
| A. mínutissima Kutz.           | -                    | + 24       |          | +            | -            | -        |            | †                   |
| Amphipleura pellucida Kutz.    |                      | -          | +        | -            |              | +        | ,          | ±1<br>■ .           |
| Cocconeis pediculus Ehr.       | +                    | ,-         |          |              |              | - »·     | -          | _                   |
| C. placentula Ehr.             | . •                  | +          |          | * : <b>†</b> | <del></del>  | + ·      | +          | +                   |
| Cyclotella meneghiniana Kutz.  | <u>.</u>             |            |          |              | -            | :        | •          | 1                   |
| Cymatopleura solea (Breb.)     |                      | +          | . +      |              | -            |          | •          | ,,*=                |
| W. Sm                          |                      |            | 4        |              |              |          |            |                     |
|                                | ٠                    |            |          |              |              |          |            | *                   |
| BACILLARIOPHYTA                | ÿ.,                  |            | . ~      |              | •            |          |            | •                   |
| Cymbella prostrata (Berk.) Cl. | . 10 <b>-</b><br>. 1 | , <b>+</b> | -        | -            | <b>-</b>     | •        | •          |                     |
| C. sinuata Greg.               | • . •                |            | <b>-</b> | <b>4</b>     | +            | <b>-</b> | <u>.</u>   | -                   |
| C. ventricosa putz.            | <b>-</b>             | F          | +        | +            | <b>+</b>     | †v.      | <b>†</b> . | ÷                   |
| Epithemia argus Kutz.          | +                    | •          | <b>-</b> | ₹., •        | <b>.</b>     | <b>†</b> | - ;        | ₹<br>4              |
| F sarey Kutz                   | +                    | + .        | _ `      | ₹ .          | +            | - / %    | +          | + ;                 |

|                              |              |             |            |          |            |            | Sit      | e               |                 | -          |          |                  |                                       |     |
|------------------------------|--------------|-------------|------------|----------|------------|------------|----------|-----------------|-----------------|------------|----------|------------------|---------------------------------------|-----|
| Al gae                       |              | 1           |            | 2        |            | 3          | 4        |                 | 5               |            | 6        | 7                |                                       | 8   |
|                              |              |             |            |          |            |            |          |                 |                 |            | •        |                  |                                       |     |
| Eunotia lunaris (Ehr.) Grun  | + ′          |             | +          |          | +          | -          |          | -               |                 | -          | -        |                  | -                                     |     |
| Fragilaria capucina Desm.    | <del>-</del> |             | +          |          | -          |            |          |                 |                 | + '        | 1        | -                | , <del>-</del>                        |     |
| F. crotonensis Kitton        | -            |             | +          |          | -          | -          |          | -               |                 | - ,        | -        |                  | · <b>-</b>                            |     |
| F. pinnata Ehr.              | -            |             | +          |          | -          | -          |          | , <b>-</b> ^    |                 | -          | 4        |                  | -                                     |     |
| F. vaucheriae (Kutz.) Peters |              |             | +          |          | +          | +          |          | +               |                 | -          | -        | •                | +                                     |     |
| Gomphonema abbreviatum       | -            | ,           | -          | •        | -          | -          |          | +               |                 | -          | -        | •                | +                                     |     |
| (Agardh.) Kutz.              | ÷            |             |            |          |            |            |          |                 |                 | •          |          |                  |                                       |     |
| G. acuminatum Ehr.           | <b>-</b> ·   |             | +          |          | +          |            |          | -               |                 | ÷          | -        |                  | ·                                     |     |
| G. acuminatum v. coronata    | +            |             | -          |          | -          | -          |          | -               |                 | -          | •        |                  | , _                                   |     |
| (Ehr.) W. Sm.                |              |             |            |          |            |            |          |                 | ,               |            |          | •                |                                       |     |
| G. lanceolatum Ehr.          | +            |             | <i>c</i> - |          |            | -          |          | * -             |                 |            | ·        | ٠.               | +                                     |     |
| G. qlivaceum (Lyngb.) Kutz.  | +            | ,           | +          |          |            | • -        |          | +               | ,               | +          | 4        | ۲                | •                                     |     |
| G. parvulum Kutz.            |              |             | -          |          | <b>.</b> - | +          | -        | 7               |                 | - 1        | <b>.</b> | -                | <del>-</del> .                        |     |
| Melosira islandica D. Mull.  | _            |             | -          | <i>'</i> | +          | , <u>-</u> |          | -               | a.              | -          | -        | -                | -                                     |     |
| Meridion circulare Agardh.   | - i -        |             | -          |          | -          | -          |          | -               |                 | +          |          | -                | -                                     | ٠,. |
| Navicula cryptocephala Kutz. | +            |             | +          | ٠,       | +          | +          |          |                 |                 | ŧ          |          | - ,              | <b>-</b> .                            |     |
| N. cuspidata Kutz.           |              |             | +          |          | +          | -          |          | -               |                 | ` <u>.</u> | •        | <del>-</del> · . | -                                     |     |
|                              |              |             |            | • •      |            |            |          |                 |                 |            |          |                  |                                       |     |
| BACILLARIOPHYTA              |              |             |            | •        |            |            |          |                 | . • •           | 1          |          |                  |                                       |     |
| N. gracilis Ehr.             | +            |             | +          |          | +          | , ,        | •        | · -             |                 | · <u>-</u> | . 3      | - :              | +                                     |     |
| N. graciloides A. Mayer      | _            |             | +          |          | +          | . 1        | •        | <sup>√2</sup> ± |                 | <b>, +</b> |          | ÷ ,              | -                                     |     |
| N. radiosa Kutz.             |              | 'ؤ'<br>هسا. | ÷          |          | +          | _          |          | -               |                 | -          | ٠        | -                | +                                     | •   |
| Nitzschia acuta Hantzsch.    | -            | •           | · <u>-</u> |          | -          | -          | •        | +               |                 | -          |          | -                | -                                     |     |
| N. dissipata (Kutz.) Grun    | +            |             | _          |          | -          | -          | ١.       | _               | • • •           | - '        |          | 2                | +                                     |     |
| N. fonticola Grun.           | -            |             | +          |          | _          |            | -        | _               |                 | -          |          | <b>-</b> .       | -                                     | •   |
| N. gracilis Hantzsch.        |              |             | +          |          | <u>.</u> . | •          | -        | er- , , ;<br>-  |                 |            |          | _                |                                       |     |
| N. palea (Kutz.) W. Sm.      | +            |             | -          |          | _          | 4          | ·        | 1               |                 | +          |          | <del>†</del>     | +                                     |     |
| N. recta Hantzsch.           | _            |             | +          |          | -          |            | <b>.</b> | -               | -               | _          |          | <u>.</u>         | -                                     |     |
| N. sublinearis, Hust.        | -            |             | +          |          | _ ',       |            | •        | •               |                 | _          |          | _                | _                                     |     |
| Pinnularia gibba Ehr.        | / -          |             | +          | . !      | +          |            |          |                 |                 | -          |          | <u>.</u>         | • • • • • • • • • • • • • • • • • • • | •   |
| P. molaris /Srun.            |              |             | 4          |          | +          |            |          | _               | ag ari<br>¥a ga | _          |          | -                | -                                     |     |
| Fe motal 135 Var une         |              |             |            |          |            | 3          |          |                 |                 | i i        |          |                  |                                       | ٠.  |

Appendix 1. Concluded.

| : · · · · · · · · · · · · · · · · · · · |   |   |   |     |   |     |   |     |              |            |   |            |   |   | _ |
|---|---|---|---|-----|---|-----|---|-----|--------------|------------|---|------------|---|---|---|
|   | , |   |   |     |   |     | S | itε | <u> </u>     |            |   |            |   |   | _ |
| Algae                                   |   | 1 |   | 2   |   | 3   |   | 4   |              | 5          | 6 |            | 7 | 3 |   |
|   |   |   |   |     | • |     |   |     |              |            |   |            |   |   |   |
| Rhoicosphenia curvata (Kutz.)           | _ |   | - |     | - |     | + | ,   | <del>-</del> | -          |   | -          | - |   |   |
| Grun                                    | ^ |   |   | 1   |   |     |   |     |              | •          |   |            |   |   |   |
| Stauroneis phoenicenteron Ehr.          | - |   | - |     | + | :-  | - | ,   | •            | · <u>-</u> |   | -          | - | • |   |
| Surirella angustata Kutz.               | _ |   | - |     | - |     | - |     | -            | +          | • | -          | 4 |   |   |
| Synedra ulna (Nitzsch.) Ehr.            | + |   | + | , . | + |     | + |     | +            | +          |   | -          | - |   |   |
| Tabellaria fenestrata (Lyngb.)          | + | ₽ | + |     | - | . , | - |     | _            |            |   | <b>-</b> , | - | • |   |
| Kutz                                    |   |   |   |     |   |     | : |     |              |            |   | *          |   |   |   |

Appendix 2. The algae found—at each site in the Steepbank River (+ present; - absent).

|   |          | S          | ite        | ,          |                  |            |     |
|---|----------|------------|------------|------------|------------------|------------|-----|
| Algae                                   |          | 1          | 2          | 3          | 4 .              | 5          | •   |
| 9<br>19<br>10                           |          |            |            |            |                  |            |     |
| CÝANOPHYTA                              |          |            |            |            |                  |            |     |
| Anabaena affinis                        |          | +          | · -        | +          | +                | -          |     |
| Calothrix braunii                       | `        | -          | <b>-</b> . | +          | +                | •          |     |
| Lyngbya aerugineo-caerulea              |          | -          | +          | +          | +                | .+         |     |
| Nostoc sp                               |          | +          | +          | +          | +                | _          |     |
| Oscillatoria sp                         |          | -          | +          | -          | · +              | -          | .,  |
|   |          |            | ,          |            |                  | ø          |     |
| CHLOROPHYTA                             |          |            |            | -          | •                |            | ,   |
| Ankistorodesmus falcatus                |          | +          |            | -          | i +              | +          |     |
| Chlamydomonas                           |          | +          | , -        | +          | \$ +             | -          |     |
| Chlorella vulgaris                      |          | . <b>-</b> | -          | +          | +                | +          |     |
| Cladophora glomerata                    |          |            | + 1,       | +          | +                | -          |     |
| Cosmarium sp                            |          | . <b>-</b> | +          | - '        | , + <sup>"</sup> | +          |     |
| Hyalotheca sp                           |          | -          |            | +          | -                | _          | 4   |
| Microspora Loefgrenii                   |          | -          | +          | + ,        | ·                | -          | ž   |
| Microspora sp                           |          | -          |            | · -        | ¥<               | +          | •   |
| Pediastrum biradiatum_                  |          | غارب<br>   | • * * •    |            | -                | +          |     |
| <b>.</b>                                | <b>,</b> |            |            |            | . ,              |            |     |
| PHYRROPHYTA                             | . '      | +          | . =        |            | . <u>.</u>       | · -·       |     |
|   |          |            | •          | •          |                  |            |     |
| CHRYSOPHYTA                             | •        |            |            |            | •                |            |     |
| Chromulina sp                           | , t      | +          | -          | , <b>-</b> | <u> -</u>        | <b>-</b> ' | · , |
|   | . 1      | Q          | •          | 4          |                  | *          |     |
| EUGĻENOPHYŢA                            |          | •          | •          |            |                  |            | •   |
| Euglena sp                              | ,        | +          |            |            | _                |            |     |
|   |          |            |            |            | •                |            |     |
| RHODOPHYTA                              | •        |            | •          |            | •                |            |     |
| Batrachospermum vagum (Roth.) C.A.Agard | lh .     | _          | +5         | -          | -                | -          |     |

|  |              | Site          |            |     | · ·       |          |
|--|--------------|---------------|------------|-----|-----------|----------|
|  |              | ,             |            | ,   | •         |          |
| Algae  | 1            | 2             | 3          | 4   | 5         | • ,      |
|  |              | <u> </u>      |            |     |           |          |
|  |              |               |            |     |           |          |
| BACILLARIOPHYTA                              |              |               |            |     |           | e e      |
| Achnanthes sp.                               | -            | -             | • •        | -   |           |          |
| A. lanceolata Breb.                          | -<br>1       |               | +          | +   | + -       |          |
| A. minutissima Kutz.                         | -)           | +             | -          | -   | -         |          |
| Cocconeis pediculus Ehr.                     | <b>≠</b>     | +             | +          | +   | +         |          |
| C. placentula Ehr.                           | -            | +             | +          | +   | +         | •        |
| Cyclotella meneghiniana Kutz.                | -            | +             | +          | +   | +         |          |
| Cymbella cistula (Hemp.) Grun                | -            | -             | -          | -   | + ,       |          |
| C. ventricosa Kutz.                          | -            | - '           | +          | +   | +         |          |
| Diatoma vulgare Bory.                        | -            | +             | -          | -   | . +       | •        |
| D. vulgare v. grandis (Sm.) Grun             | -            | f             | . +        | - , | -         |          |
| Epithemia argus Kutz.                        | -            | +             | ,+         | -   | +         |          |
| E. sorex Kutz.                               |              |               | +,         | +   | +         |          |
| E. turgida Kutz.                             | -            | + ,           | -          | •   | +         |          |
| Fragilaria capucina Desm.                    | -            | -             | +          | +   | -         |          |
| F. pinnata Ehr.                              | -            | <b>-</b> ,    | +          | •   | -         |          |
| Fustrulia rhomboides v. amphileuroides Grun. | ->           |               | +          | •   | 4-        |          |
| Gomphonema acuminatum Ehr.                   | · <b>-</b>   | +             | -          | ž   |           |          |
| G. gracilé Ehr.                              | _            | +             | -          | . ~ | -         |          |
| N. graciloides A. Mayer                      | · , <b>-</b> |               | -          | +   | +         |          |
| N. radiosa Kutz.                             | _            | · -,          |            | . ~ | +.        |          |
| Nitzschia gracilis Hantzsch.                 | <b>-</b> _ · | +             |            | _   | + []      |          |
| N. hantzschiana Rabh.                        |              | -             | <b>+</b> . | _   |           |          |
| N. palea (Kutz.) W. Sm.                      | _            | +             | ; +        | •   | +         |          |
| Rhoicosphenia curvata (Kutz.) Grun.          |              | <b>-</b>      | +          |     | ,         |          |
| Rhopalodia gibba = (Ehr.) O. Mull.           | _            | +             | +.         | +   | +         | •        |
| Surirella angustata Kutz.                    |              |               | +          | _   | - · · · · | <b>*</b> |
| Synedra ulna (Nitzsch.) Ehr.                 | _ :          | · · · · · · · | +          | +   | +         |          |
| Tabellaria flocculosa (Roth.) Kutz.          | +            | +             | <b>.</b>   |     |           |          |
| Tabellaria Trocculosa (Nocii.) Nucz.         | •            | •             |            |     |           |          |

Appendix 3. The algae found at each site in the Hangingstone River ( + present; - absent).

|                                       | 6              |               |   |                  | 19                                    |                    |
|---------------------------------------|----------------|---------------|---|------------------|---------------------------------------|--------------------|
|                                       |                | S             | ite                                     |                  |                                       |                    |
| Algae                                 | 1              | 2             | 3                                       | 4                | 5                                     | 6 .                |
|                                       |                |               | · • · · · · · · · · · · · · · · · · · · | <del>-, ,</del>  | · · · · · · · · · · · · · · · · · · · |                    |
| CYANOPHYTA                            |                | •             |   |                  |                                       |                    |
| Anabaena affinis Lemm. l              | +              | <br>+         | _                                       | +                |                                       |                    |
| Calothrix braunii Bornet & Flahault   | _              | _             | +                                       | _                | -                                     | +                  |
| Lyngbya aerugineo-caerulea (Kutz.)    | +              | +             | +                                       | ,<br>+           | , <b>-</b>                            | +                  |
| Oscillatoria                          | -              | _             | +                                       | +                | +                                     | _                  |
|                                       |                |               | •                                       |                  |                                       | 1                  |
| CHLOROPHYTA                           |                |               |   | - 1              | •                                     | •                  |
| Chlamydomonas sp.                     | <b>a</b> +     | +             | , <del>-</del>                          | _                | _ 0                                   | <u>;</u> ,         |
| hlorella vulgaris Bever               | + -            | - 1           | +                                       | -                | -                                     | _                  |
| Closterium sp.                        | _              | _             | -,                                      | +                | + 1                                   | +                  |
| Pediastrum boryanum (Turp.) Meneghini | ·<br>-         | <i>.</i><br>- | Ţ.                                      | + -              | - 6                                   | -                  |
| Pleurotaenium spp.                    | . <del>-</del> | -             | *-·                                     | _                | : <u>-</u>                            | · '.               |
| Spirogyra sp.                         | ·<br>-         |               | <u>.</u> .                              | _                |                                       | +                  |
|                                       |                | ٠.            | -                                       |                  | • .                                   |                    |
| СКУРТОРНУТА                           |                |               |   |                  |                                       |                    |
| ryptomonas ovata Ehr.                 | + .            | _             | -                                       | <b></b>          | -                                     |                    |
| Rhodomonas minutum Skuja              | -              | 4.            | , <b>-</b>                              |                  | -<br>-                                | -                  |
|                                       | . d            |               | · ·                                     |                  |                                       | •                  |
| UGLENOPHYTA                           |                |               | •                                       |                  |                                       | ,                  |
| uglena sp. '                          | +              | +             | -                                       | -                | -                                     | - /                |
|                                       | •              | •             |   | -                |                                       |                    |
| HODOPHYTA :                           | •              |               | •                                       | · · · · ·        | ÷                                     | (                  |
| atrachospermum vagum (Roth.) C.A. Ag. | -              |               | 1<br>-                                  | - 33             | + , '                                 | +                  |
|                                       |                |               |   | * .              |                                       |                    |
| ACILLARIOPHYTA                        |                |               |   |                  | , . <u>.</u>                          |                    |
| chnanthes lanceolata Breb.            | (a)            | +             | +                                       | . <del>  †</del> | +                                     | +                  |
| mphipleura pellucida Kutz.            | •              | -             | -                                       |                  | _                                     | <b>+</b> · · · · · |
| occoneis pediculus Ehr.               |                | 4             | -                                       | •                | +,                                    |                    |
| <u>Placentula</u> Ehr.                |                | *_            | +                                       | _                | +                                     | +                  |
|                                       |                |               |   |                  |                                       |                    |

| Appendix | ** | Continued. |
|----------|----|------------|
| Appendix | ٥. | continued. |

| Append 1x 3. Conclinated.        |                                       |            |                                       | Site            |             | ·                                       |             |
|----------------------------------|---------------------------------------|------------|---------------------------------------|-----------------|-------------|---|-------------|
| Algae                            |                                       | 1          | 2                                     | . 3             | . 4         | . 5                                     | 6           |
|                                  |                                       |            | <del>,</del>                          |                 |             | <del></del>                             | <del></del> |
| Cyclotella meneghiniana Kutz.    | (a)                                   | +          | +                                     | +               | _           | *                                       |             |
| Cymatopleura solea (Breb.) W. Sm | •                                     | _          |                                       | +               | +           | ` <u>-</u> '                            |             |
| cymbe prostrata (Berk.) Cl.      | •                                     | +          | +                                     | +               | _           |   |             |
| C. sinuata Greg.                 | ••                                    | -          | '. <u>-</u>                           | -               | <b>4</b> ~  | ` -                                     | ٠.          |
| C. ventricosa Kutz.              | •                                     | +          | + ,                                   | +               | <b>†</b>    | <b>-</b> .                              |             |
| Diatoma vulgare Bory             |                                       | 7          | -                                     | -               | -           | +                                       |             |
| Epithemia argus Kutz             |                                       | · · ·      | • _                                   | -               | +           | `-                                      | ٠,          |
| E. sorex Kutz.                   | •                                     | -          | -                                     | . +             | +           | +                                       | ,           |
| E. turgida Kutz.                 | **                                    |            | <u>-</u>                              | -0              | +           | +                                       |             |
| Pragilaria capucina Desm.        |                                       | , <b>-</b> | +                                     | ~               | -           | + 2                                     |             |
| F. vaucheriae (Kutz.) Peters     | •                                     | +          | +                                     | +               | +           | ÷                                       | •           |
| Gomphonema abbreviatum (Ag.) Kut | .2                                    | +          | +4                                    | <b>-</b> .      | <b>-</b> ·  | - ' v                                   |             |
| G. lanceolatum Ehr.              | `.7                                   | -          | -                                     | -               | -           | +                                       | •           |
| G. olivaceum (Lyngb.) Kutz.      |                                       |            | -                                     | +,              | -           | •                                       |             |
| G. parvulum Kutz.                | · · · · · · · · · · · · · · · · · · · | ,+         | -                                     | , <b>-</b>      | _           | + .                                     |             |
| Gyrosigma acuminatum (Kutz.) Rab | ph.                                   | -          | · •                                   | +               | -           | -                                       | ı ,         |
| Melosira islandica O. Mull       | , ,                                   |            | -                                     | . +             | -           | - ·                                     |             |
| M. varians C.A. Ag.              |                                       |            | -                                     | +               | +           | · · · ·                                 | •           |
| Navicula cryptocephala Kutz.     |                                       | +          | •                                     | -               | +.          | + '                                     | -41         |
| N. graciloides A. Mayer          | • •                                   | + •        | +                                     | +               | * <b>*/</b> | +                                       |             |
| Nitzschia acuta Hantzsch.        |                                       | -          | +                                     | ;;; <b>-</b> ;; | - 1         |   | *.          |
| N. dissipata (Kutz.) Grun.       |                                       |            | +                                     | -               | +           | • +                                     |             |
| N. palea (Kutz.) W. Sm.          |                                       | +          | +                                     | +               |             |   |             |
| N. recta Hantzsch                |                                       | .+         | -                                     | , 💝             | +           | · • • • • • • • • • • • • • • • • • • • | f           |
| <u>Pinnularia gibba</u> Ehr.     | *                                     | -          | , <b>-</b>                            | , te            | -           | •                                       |             |
| Rhopalodia ibba (Ehr.) 0. Mull.  |                                       |            | • • • • • • • • • • • • • • • • • • • | +               | <b></b>     |   |             |
| R. gfbberula (Ehr.) O. Mull.     |                                       | _          | -                                     | +               |             | +                                       | •           |
| Synedra ulna (Nitzsch.) Ehr.     |                                       | +          | ŧ                                     | +               | +           | +                                       |             |
|                                  |                                       | . 24       | • • •                                 | -               |             |   | H           |

<sup>(</sup>a) Diatom sample lost in transit.

| Appendix 4. | The | algae | found | at | each | site | in | the | MacKay | River | (+ | present |
|-------------|-----|-------|-------|----|------|------|----|-----|--------|-------|----|---------|
| - absent).  |     |       |       |    |      |      |    |     |        |       |    |         |

|                                      |             |            |                   | Sit        | e                 |                  |            |                |             |
|--------------------------------------|-------------|------------|-------------------|------------|-------------------|------------------|------------|----------------|-------------|
| Al gae                               | 1)          | 2          | 3                 | 4          | 5 -               | 6                | 7          | 8              |             |
|                                      | <del></del> |            |                   |            |                   | <del></del>      |            | <del>`</del>   | <del></del> |
| CYANOPHYTA                           |             |            |                   |            |                   |                  |            |                |             |
| Anabaena affinis Lemm.               | <b>.</b>    | +          | . +               | *+         | _                 | . +              |            | +              | .4          |
| Chroococcus limneticus Lemm.         | _           | · .        | _                 |            | + 1               | م                |            | _              |             |
|                                      |             | _          | _                 |            | · .               |                  | · +        | _              |             |
|                                      | _           | Ī.         |                   | , ···      |                   |                  | +          | +              | •           |
| Lyngbya aerugineo-caerulea (Kutz.)   | •           | _          |                   | •          | •                 | •                | •          | •              |             |
| Gomont                               | ٠           | •          | _                 | 1          |                   |                  | <b>%</b> : |                | •           |
| Merismopedia glauca (Ehr.) Naegeli.  | -           |            | -                 | ,          |                   | •                | •          | -              |             |
| Nostoc spp                           | -           | -          |                   | *          | #                 | . <del>-</del> / | -          | -              |             |
| Oscillatoria amphibia C.A. Agardh.   | +           | -          | -                 | -,         | • -               | -                | . •        | · <del>-</del> |             |
|                                      | ,           |            |                   |            |                   |                  |            |                |             |
| CHLOROPHYTA                          |             |            |                   | -          | •                 |                  |            | •              |             |
| Ankistrodesmus falcatus (Corda)      | -           | -          | +                 | +          | . +               | +                | +          | ·              | •           |
| Ralfs.                               |             |            |                   |            |                   | ٠                |            | ,              |             |
| Chlamydomonas globosa Snow           | +           | +          |                   | • •        | -                 | - "              | -          | ,              | ,           |
| Chlamydomonas sp.                    | +           | +          | . +               | +          | -                 | +                | +          | . +            | ,           |
| Chlorella ellipsbidea Gerneck        | +           | <b>-</b>   |                   | -          | -                 |                  | -          | -              |             |
| C. vulgaris Beyer                    | 7+          | +          | -                 | +          | +                 | . , -            | +          | +              | •           |
| Claophora glemerata (L.) Kutz.       | +           | · -,,      | +                 | <u>(</u> + | +                 | +.               | +          | +              |             |
| Closterium sp.                       |             | -          | +                 | +          | +                 | +                | +          | "              |             |
| Coelastrum scabrum Reinsch.          | -           | -          | 3 <del>-</del> j. | +          | -                 | <u> </u>         | -          |                |             |
| Cosmartum spp.                       | <b>4</b> -  | -          | +.                | +          | . +               | +                | +.         | -              |             |
| Gloeocystis gigas (Kutz.) Lager      | -           |            | · .               | +          | +                 | _                | -          |                | •           |
| Microspora Loefgrenii (Norst.) Lager | _           | -          | +                 | •,         | <br><del></del> . | _                | -          | -              |             |
| Dedogonium sp.                       | _           | -          | +                 | +          | +                 |                  | +          | _              |             |
| Pediastrum biradiatum`v. emarginatum |             |            |                   |            |                   |                  |            |                |             |
| . convexum Prescott.                 | -           | <u>.</u> : | -                 | -          | +                 | -                |            | +              | •,          |
| Scenedesmus acutiformis Schroeder    | <b>-</b>    |            |                   | -          | +                 | +                | +          | +              |             |
| 5. bijuga (Turp.) Lager.             |             | -          | + •               | -          | +                 | +                | +          | + .            |             |
| . quadricauda (Turp.) de Breb.       |             | <u>.</u>   | _                 | <b>-</b> , | -<br>+            | _,               |            | +              |             |

Amphipleura lindheimeri Grun

A. pellucida Kutz.

Site 4 % 5 6 Al gae Spaerocystis schroeteri Chodat Sphaeroplea annulina (Roth.) C.A. Agardh. Stigeoclonium sp. Ulothrix. sp. CRYPTOPHYTA Cryptomonas ovata Ehr. Rhodomonas minutum Skuja **PYRROPHYTA** - CHRYSOPHYTA Chromulina spp. Dinobryon sertularia Mallomonas spp. EUGLENOPHYTA Euglena sp. Phacus sp. Trachelomonas RHODOPHYTA Batrachospermum vagum (Roth. C.A. Agardh. BACILLARIOPHYTA. Achnanthes lanceolata Breb.

| Appella IX 4. Coll Cilluea. | Appendix | 4. | Continued. |
|-----------------------------|----------|----|------------|
|-----------------------------|----------|----|------------|

| ·                                  |           |              |                  | Sit        | e          |           |              |              |
|------------------------------------|-----------|--------------|------------------|------------|------------|-----------|--------------|--------------|
| Algae                              | 1         | 2            | 3                | 4          | 5          | 6         | . 7          | 8            |
|                                    | <u> </u>  | <del></del>  | <u> </u>         | •          |            | - <u></u> | <del>,</del> |              |
| Cocconeis pediculus Ehr.           | - 🙀       | +            | +                | +          | + ´        | +         | +            | +.           |
| C. placentula Ehr.                 | +         | +            | + .              | +          | +          | +         | +            | + -          |
| Cyclotella meneghiniana (Ehr.)     | +         | · <b>-</b>   |                  | -          | ~          | -         | +            | -            |
| Kutz.                              |           |              |                  |            |            |           |              |              |
| Cymbella cistula (Hemp.) Grun.     | -         | <b>-</b> . · | -                | -          | +          | -         | -            | -            |
| C. lanceolata (Ehr.) V.H.          | +         | -            | -                | -          | -          | -         | -            | -            |
| C. prostrata (Berk.) Cl.           | -         | -            | -                | -          | -          | +.        | -            | -            |
| C. ventricosa Kutz.                | +         | -            | +                | <u>-</u>   | +          | +         | +            | +            |
| Epithemia argus Kutz.              | -         | -            | -                |            | +          | . +       | ,+           | + -          |
| E. Sorex Kutz.                     |           | -            | -                | -          | +          | +         | ` <b>+</b>   | + "          |
| E. turgida Kutz.                   | -         | -            | -                | -          | -          | +         | -            | -            |
| Eunoitia pecinalis v. minor (Kutz. | )         |              |                  |            |            |           |              |              |
| Rabh.                              | - '       | -            | +                | -          |            | -         | +            | +            |
| E. valida Hust.                    | -         | <b>-</b> .   | -                | -          | -          | ٠-        | +            | +,           |
| Fragilaria capucina Desm.          | -         | -            | +                | -          |            | -         | +- ,         | , <b>+</b>   |
| F. pinnata Ehr.                    | +         | -            | <b>-</b> ,       | -          | -          | -         | +            | +            |
| F. vaucheriae (Kutz.) Peters.      | -         | ,-           | -                | +          | +.         | +         |              | +            |
| Gomphonema lanceolatum Ehr.        | -         | -            | +                | + ~        | -          | +         | +            | , <b>+</b> . |
| G. olivaceum (Lyngb.) Kutz.        | -         | <b>-</b> ,   | •                | -          | + .        | +         | -            | -            |
| G. parvulum (Kutz.) Grun.          | -         | -            | +                | -          |            | <b>-</b>  | •,           | -            |
| Gyrosigma acuminatum (Kutz.) Rabh. | -         | -            | -                | -          | +          |           | +            | +            |
| Hantzschia amphioxys (Ehr.) Grun.  | _         | -            | -                | ′ <b>-</b> | ٠.         | -         | +            | +            |
| Meridion circulare Agardh.         | •         | -            | ÷                | +          | -          | -         | -            | -            |
| Navicula cryptocephala Kutz.       | +         | •            | +                | +          | +          | +,        | +            | +            |
| N. cuspidata Kutz.                 | -         | ٠_           | - ;              | ·-         | -          | +         | -            | •            |
| N. pupula Kutz.                    | <b>'+</b> | •            | - `              | <b>-</b> , | -          | ,<br>-    | -            | · ·          |
| N. radiosa Kutz.                   | +         |              | + '              | . <b>+</b> | +          | + .       | .+           | +            |
| Neidium affine (Ehr.) Cl.          | _         | , <b></b>    | + .              | _          | ·          |           | - "          | <b>-</b> *.  |
| N. affine v. amphirhynchus (Ehr.)  |           |              |                  |            |            |           | 1,           |              |
| cı.                                | + ` `     |              | _** <sub>5</sub> | -,         | <b>-</b> · | •         | <b>-</b> .   | - '          |

| Nitzschia acuta Hantzsch +  | 7 8 + + - + - + + + + + +   |                                    |           |            |            | •      | Sit           | e           |              | ,  |              |
|---|---|------------------------------------|-----------|------------|------------|--------|---------------|-------------|--------------|----|--------------|
| N. dissipata (Kutz.) Grun.  N. fonticola Grun.  N. palea (Kutz.) W. Sm.  N. recta Hantzsch.  N. sublincaris Hust.  Pinnularia gibba Ehr.  N. wiridis v. sudetica (Hilse)  Hust.  Rhopalodia gibba (Ehr.) O. Mull. + - + - + + + + + + + + + + + + + + +                   | + | gae                                |           | . 1        | - 2        | ÷ 3    | 4             | 5           | 6            | .7 | 8            |
| N. dissipata (Kutz.) Grun.  N. fonticola Grun.  N. palea (Kutz.) W. Sm.  N. recta Hantzsch.  N. sublincaris Hust.  Cinnularia gibba Ehr.  C. molaris Grun.  C. viridis v. sudetica (Hilse)  Hust.  Chopalodia gibba (Ehr.) O. Mull. + - + - + + + + + + + + + + + + + + + | +   |                                    |           |            |            |        | <del></del>   | <del></del> | <del></del>  |    | <del></del>  |
| N. dissipata (Kutz.) Grun.  N. fonticola Grun.  N. palea (Kutz.) W. Sm.  N. recta Hantzsch.  N. sublincaris Hust.  Cinnularia gibba Ehr.  C. molaris Grun.  C. viridis v. sudetica (Hilse)  Hust.  Chopalodia gibba (Ehr.) O. Mull. + - + - + + + + + + + + + + + + + + + | + + + + + + + + + + + + + + + + + + +   | tzschia acuta Hantzs               | sch       | · <u>-</u> | _          | +      | _             | _           |              | -  | _            |
| N. fonticola Grun.  N. palea (Kutz.) W. Sm.  N. recta Hantzsch.  N. sublincaris Hust.  Pinnularia gibba Ehr.  P. wiridis v. sudetica (Hilse)  Hust.  Rhopalodia gibba (Ehr.) O. Mull. + - + - + + + + + + + + + + + + + + +   | + + 1 +               | <del></del>                        | ,         | -          | _          | +      | -             | -           | -            | _  | -            |
| palea (Kutz.) W. Sm.  | +                         | · <del></del>                      | •         | ť          | -          |        | -             | -           | -            | _  | -            |
| recta   Hantzsch.   | +, + + + + +  |                                    | Sm.       | +          |            | +      | -             | *. <b>+</b> | +            | +  | + ;          |
| I. sublincaris Hust.  Innularia gibba Ehr.  I. molaris Grun.  I. viridis v. sudetica (Hilse)  Hust.  I. hopalodia gibba (Ehr.) O. Mull. + - + - + + + + + + + + + + + + + + +   | + + + + + + + + + + + + + + + + + +   | •                                  |           | +          | -          | + ,    | +             | , <b>-</b>  | -            | +. | <u>-</u>     |
| Pinnularia gibba Ehr.   | + -<br>+ -<br>+ +<br><br><br>+ +  | sublincaris Hust.                  |           |            | -          | +      | -             | -           | ÷ .          | -  | -            |
| Hust.  Hust.  Hopalodia gibba (Ehr.) 0. Mull. + - + - + + + + + + + + + + + + + + +   | + - + + + + + +   |                                    | * • _     | +          | -          | ·<br>- | - '           | -           | -            | +  | -            |
| Hust + + + + + + + + + +  | + +<br><br><br>+ +  | molaris Grun.                      | ,         | + '        | <b>-</b> . | -      | - '           | -           | -            | +  | -            |
| hopalodia gibba (Ehr.) 0. Mull. + - + + + + + + + + + + + + + + +   | + + +   | viridis v. sudetica                | (Hilse)   | -          |            | •      |               |             | =            |    |              |
| tauroneis anceps Ehr. +   | + +<br><br><br>+ +  | Hust.                              | •         | -          | -          | -      | . <del></del> | +           |              | -  | <b>-</b> · . |
| <pre>. phoenicentron Ehr</pre>  |   | <u>opalodia gibba</u> (Ehr.        | ) 0. Mull | . +        | -          | +      | -             | -           | +            | +  | +            |
| urirella angustata       Kutz.       + - + + + +         . ovalis       Breb.       + +         ynedra ulna       (Nitzsch.)       Ehr.       + + - + + + + + + + + + + + + + + +   | + + +   | <u>auroneis</u> <u>anceps</u> Ehr. |           | +.,        | -          | -      | -             | -           | -            | -  |              |
| . ovalis Breb + ynedra ulna (Nitzsch.) Ehr + + - + + + abellaria flocculosa (Roth.)   | + +   | phoenicentron Ehr                  | ,         | . <b>+</b> | -          | -      | -             | +           | •            | -  | -            |
| ynedra ulna (Nitzsch.) Ehr + + - + + + + dabellaria flocculosa (Roth.)  | + +   | <mark>rirella angustata</mark> Ku  | itz.      | +          | -          | +"     | + `           | +           | +            | -  | -            |
| abellaria flocculosa (Roth.)  |   |                                    | ŕ         | -          | ٠ -        | -      |               | + '         | %•           | -  | -            |
|   |   | <u>nedra ulna</u> (Nitzsch.        | ) Ehr.    | -          |            | +.     | <b>†</b> .    | -           | <b>+</b> ′ · | +  | <b>+</b>     |
| Kutz. +   | -, -,   | bellaria flocculosa                | (Roth.)   |            | c.         | -      | •             | ň           |              |    |              |
|   | · •   | Kutz.                              |           | +          |            | -      | -             | -           | -            | -, | <b>-</b> ,   |

. .



Appendix 5. The algae found at each site in the Ells River ( + present; - absent).

| CYANOPHYTA  Calothrix braunii  Lyngbya aerugineo-caerula (Kutz.)  Gomont  Nostoc spp. | + + +          | -+         | Site 3     | 4            | 5  | 6      |
|---|----------------|------------|------------|--------------|--|--------|
| CYANOPHYTA  Calothrix braunii  Lyngbya aerugineo-caerula (Kutz.)  Gomont  Nostoc spp. |                | -<br>+     | ~          | +            | -<br>-   | -<br>- |
| Calothrix braunii  Lyngbya aerugineo-caerula (Kutz.)  Gomont  Nostoc spp.             | + +            | -<br>+     |            | +            | ,<br>-<br>-  | -      |
| Lyngbya aerugineo-caerula (Kutz.) Gomont Nostoc spp.                                  | + + +          | -<br>+     |            | +            | -  | •      |
| Gomont Nostoc spp.  | +              | +          |            | +            | -  |        |
| Nostoc spp.   | +              | •          | 1          |              |  | _      |
|   | +              |            |            |              |  |        |
| 0.W 0000WT4   |                | -          | -          | -            | - ′  | • -,   |
| OUL ODODUVES  |                |            |            |              | * 1  |        |
| CHLOROPHYTA   |                |            | -          |              |  |        |
| Chlamydomonas sp.   | +              | , <b>+</b> | -          | -            | -  | +      |
| Chlorella vulgaris Beyer.   | +.             | +          | •          | -            | •  | -      |
| Cladophora glomerata (L.) Kutz.   | +              | +          |            | -            | ~  | -      |
| Cosmarium spp.  | +              | -          |            | -            | · -  | -      |
| Microspora pachyderma (Wille) Lager.  | +              | -          |            | -            | , <b>-</b>   | -      |
| Microspora sp.  | . +            | -          | -          | - ·          | -  | -      |
| Stigeoclonium sp.   | +              | -          | -          | -            |  | -      |
|   |                |            |            |              |  |        |
| СКУРТОРНУТА   |                |            |            | •            | *  |        |
| Cryptomonas ovata Ehr.  | -              | +          | +          | • •          | -  | -      |
|   |                |            | . to       |              | ,  | `      |
| CHRYSOPHYTA   |                |            |            |              |  |        |
| Chromulina spp.   | +              | -          |            | <b>-</b> . ' | -  |        |
|   |                |            |            |              |  |        |
| EUGLENOPHYTA  |                |            |            | ٧,           | . •  |        |
| Phacus sp.  | , <b>+</b>     | -          |            | -            | -  | -      |
| <u>Trachelomanos</u> sp.  | +,             | -          | . <b>-</b> | • :          | -  | -      |
|   | * *            |            |            |              |  |        |
| BACILLARIOPHYTA   | 5.             |            | 0          |              | in the state of th |        |
| Achnanthes lanceolata Breb.   | j <del>t</del> | -          | <b>-</b> . |              | +  | •      |
| A. minutissima Kutz.  | <del>.</del>   |            | . • .      | <b>-</b> ,   | -  | -,     |
| AmphipTeura lindheimeri Grun.   |                | -          | · -        | +            | . =  | •      |
| Asterionella formosa Hass.  | +              |            | <u>.</u>   | ; <b>-</b>   | -  | -      |

|  |            |               |                     |                                       |                  | •            |
|--|------------|---------------|---------------------|---------------------------------------|------------------|--------------|
|  |            | S             | ite                 | •                                     |                  |              |
| Algae                                    | 1          | , 2           | 3                   | - 4                                   | 5                | 6            |
|  |            | <del></del>   | <u></u>             | · · · · · · · · · · · · · · · · · · · | · · · · ·        |              |
| Cocconeis pediculus Ehr.                 | +          | -             | +                   | +                                     | -                | € 1 ± 1      |
| C. placentula Ehr.                       | +          | +             | +                   |                                       | +                | +,           |
| Cyclotella comuta (Ehr.) Kutz.           | +          | +1            | +                   | +                                     | <b>-</b> _       | -            |
| C. kutzingiana Thwaites                  |            | -             | +                   | -                                     | ·_               | -            |
| C. meneghiniana Kutz.                    | +          | + .           | +                   |                                       | - '              | *            |
| Cymatopleura solea (Breb.) W. Sm.        | +          |               |                     | -                                     | - '              |              |
| Cymbella prostrata (Berk.) A.            | +          | •             | -                   | +                                     | -                | · · ·        |
| G. sinuata Greg.                         | +          | +             | +                   | +                                     | +                | <del>-</del> |
| C. ventricosa Kutz.                      | +          | +             | +                   | +1                                    | +.               | -            |
| Diatòma elóngatum Agardh.                | -          | ~~ <b>+</b>   |                     | , <del>+</del>                        | . +              | . •          |
| D. vulgare Bory                          | +          |               | -                   | -                                     | -                | -            |
| D. vulgare v. grandis (Sm.) Grun.        | · · +      | <del>al</del> | + .                 | +                                     | +                | +            |
| Epithemia argus Kutz.                    | +          |               | -                   | -                                     | , , <del>-</del> | -            |
| E. sorex Kutz.                           | +          |               | ٠_                  | +                                     | ` - ' <u>-</u> ' | • •          |
| Fragilaria capucina Desm.                | -          | +             |                     | -                                     | -                | <b>-</b> ,   |
| F. construens (Ehr.) Grun.               | <b>+</b> . | + .           | -                   | + .                                   |                  | -            |
| F. construens v. binodis (Ehr.) Grun.    | +          | _ ·           | -                   | -                                     |                  | ·            |
| F. crotonensis Kitton.                   | -          |               | _                   | +                                     | . =              |              |
| F. pinnata Ehr.                          | _          | +             | +                   | +                                     | +                | • -          |
| V. vaucheriae (Kutz.) Peters.            | +          | -             | ·<br>2 <del>†</del> | _ ;                                   | l'               | + ,          |
| Frustulia rhomboides v. amphilpleuroides | s + *      |               | - ·                 | · ·-                                  | ``.<br>          |              |
| Grun.                                    | -          | 1             |                     | ,                                     |                  | • ,          |
| Gomphonema abbreviatum (Ag.) Kutz.       | +          | _             |                     | _                                     | _                |              |
| G. lanceolatum Ehr.                      | +          |               | × , <b>+</b>        | +                                     |                  | _            |
| G. olivacium (Lyngb.) Kutz.              | +          | +             | +                   | +                                     |                  | +            |
| Gryrosigma acuminatum (Kutz.) Rabh.      | -          | •             |                     |                                       | _                | ·            |
| Melosira varians C.A. Ag.                | -          | -             |                     | +                                     | ط<br>-           | ,.<br>       |
| Navicula cryptocephal Kutz.              | _          | +             | +                   | +                                     | +                | · , ,        |
| N. graciloides A. Mayer                  | _          |               | 1                   | +                                     | +                | +            |
| J. warra was nayer                       |            |               |                     |                                       | ,                | τ.           |

| Appendix | 5. | Concluded. |
|----------|----|------------|
|----------|----|------------|

|  |            | S     | ite    |            |     |                |
|--|------------|-------|--------|------------|-----|----------------|
| Algae                                      | 1          | 2     | 3      | 4          | 5   | 6              |
|  |            | ,     |        |            |     |                |
| N. minima v. <u>atomoides</u> (Grun.,) Cl. | -          | -     | -      |            | * + | -              |
| Nitzschia dissipata (Kutz.) Grun.          | +          | +.    | +      | , +        | +   | +              |
| N. fonticola Grun.                         | + ,        | .+    | +      | •          | -   |                |
| N. palea (Kutz.) W. Sm.                    | - :        | +.    | +      | +          | -   | ,+             |
| N. recta Hantzach.                         | + .        | -     | · +    | , <b>-</b> | +   |                |
| Pinnularia molaris Grun.                   | <b>-</b> · | • - • | +      | -          | . = | · <del>-</del> |
| Rhoicosphemia curvata (Kutz.) Grun.        |            |       | ;<br>T | , ·+       |     |                |
| Rhopalodia gibberula (Ehr.) O. Mull.       |            |       | - ,    | •          | +   | _              |
| Stephanodiscus astraea (Ehr.) Grun.        | -          | +     | -      | -          | -   | -              |
| Surirella angustata Kutz.                  | -          |       |        | - '        | . + | -              |
| Synedra ulna (Nitzsch.) Ehr.               |            | +     | , =    | +          | . + | -              |

Appendix 6. Summary of factors potentially influencing the growth of the major algal groups.

Algal Groups

Factors

DIATOMS

Muskeg River

.Entire period . Irradiance.

Open water 1978 Current velocity.

Open water 1979 Irradiance; NO3-N; Temperature.

Steepbank River

Entire period Irradiance; Temperature; Current velocity.

Open water 1978

Open water 1979 Temperature; Current velocity; Discharge; NO<sub>3</sub>-N.

Hangingstone River

Entire period SiO<sub>2</sub>; Discharge; Current velocity.

Open water 1978 ŞiO<sub>2</sub>

Opén water 1979 - Température; Current velocity; Discharge; Ca; Na.

MacKay River

Entire period Irradiance; NO<sub>3</sub>-N.

Open water 1978 Discharge; Current velocity; SiO2; Irradiance.

Open water 1979 Irradiance; PO<sub>4</sub>-P; SiO<sub>2</sub>.

Ells River

Entire period Na; Ca; Irradiance; PO<sub>4</sub>-P.

Open water 1978 Temperature; Irradiance; Current velocity; Discharge.

Open water 1979 Ca; Temperature; Po<sub>4</sub>-P; Discharge; Current velocity;

NO3-N.

Appendix 6. Continued.

Algal Groups

Factors

CHOROPHYTA

Muskeg River

Entire period Temperature.

Open water 1978

Calcium.

Open water 1979

Irradiance; NO3-N; Temperature; Discharge; SiO2.

Steepbank River

Entire period NO3-N; Temperature; SiO2.

Open water 1978 SiO2; Current velocity; Discharge.

Open water 1979 Temperature; NO<sub>3</sub>-N; PO<sub>47</sub>P; SiO<sub>2</sub>.

Hangingstone River

Entire period Temperature; Fe; SiO<sub>2</sub>; Irradiance; Current velocity;

Discharge.

Open water 1978 Current velocity; Discharge.

Open water 1979 S#O2; Na; NO3-N; PO4-P; Current velocity; Discharge.

MacKay River

Entire period Irradiance; Current velocity; Discharge; Temperature;

`SiO2; NO3-N.

Open water 1978 Current velocity; Discharge; Fe.

Open water 1979 Current velocity; Discharge; Temperature; PO<sub>4</sub>-P; Na.

Ells River

Entire period PO<sub>4</sub>-P; Na.

Open water 1978 Discharge; Current velocity.

Open water 1979 Temperature; PO<sub>4</sub>-P; Irradiance; Na; Ca.

## Appendix 6. Continued.

Algal Groups

Factors

CYANOPHYTA

Muskeg River

Entire period

SiO2; Discharge.

Open water 1978

Current velocity; Discharge.

Open water 1979

NO<sub>3</sub>-N; Irradiance; Temperature; SiO<sub>2</sub>; Discharge.

Steepbank River

Entire period

Irradiance.

Open water 1978

Current velocity; Discharge.

Open water 1979

Current velocity; Discharge; Temperature; Na; Ca;

5102; PO4-P; NO3-N.

Hangingstone River

Entire period

SiO2; K; Discharge; Current velocity; NO3-N.

Open water 1978

Current velocity; Discharge; NO3-N.

Open water 1979

SiO2; PO4-P; Na; Discharge; Current velocity.

MacKay River

Entire period

Irradiance; SiO2; Temperature; PO4-P.

Open water 1978

Discharge.

Open water 1979

Irradiance.

Ells River

Entire period

NO3-N; Irradiance; PO4-P.

Open water 1978

Open water 1979

Ca; Temperature; Na.

## Appendix 6. Concluded.

Open water 1979

Algal Groups

Factors

RHODOPHYTA

Muskeg River

Entire period SiO2; Temperature; Ca; Mg; Current velocity; Mn.

Open water 1978 Irradiance.

Open water 1979 Irradiance; NO3-N; PO4-P.

Steepbank River

Entire period Irradiance; Temperature; SiO2.

Open water 1978 NO3-N; Current velocity; Discharge.

Irradiance; NO<sub>3</sub>-N.

Appendix 6b. Summary of factors potentially influencing epilithtc algal standing crops.

| Study Period       | Factors  |
|--------------------|--|
| Muskeg River       |  |
| Entire period      | Temperature.   |
| Open water 1978    | Current velocity.  |
| Open water 1979-   | Temperature.   |
| Steepbank River    |  |
| Entire period      | Current velocity; Discharge.   |
| Open water 1978    | $SiO_2$ ; $NO_3$ -N; Irradiance; Current velocity; Discharge.  |
| Open water 1979    | Current velocity; Discharge; Temperature; Irradiance; Na; Ca; SiO <sub>2</sub> ; NO <sub>3</sub> -N; PO <sub>4</sub> -P. |
| Hangingstone River |  |
| Entire period      | Current velocity; Discharge; Temperature.  |
| Open water 1978    | NO3-N; Irradiance; SiO2; Fe; Current velocity; Discharge.  |
| Open water 1979    | Current velocity; Discharge; Temperature; Irradiance; Ca; Na; SiO <sub>2</sub> ; NO <sub>3</sub> -N; PO <sub>4</sub> -P. |
| MacKay River       |  |
| Entire period      | Current velocity; Discharge.   |
| Open water 1978    | Irradiance; PO <sub>4</sub> -P; NO <sub>3</sub> -N; Discharge; Temperature; Fe; SiO <sub>2</sub> ; Mn; Current velocity  |
| Open water 1979    | Current velocity; Fe; Discharge; NO <sub>3</sub> -N; Na.   |
| Ells River         |  |
| Entire period      | SiO <sub>2</sub> ; NO <sub>3</sub> -N; Ca; Na; Irradiance.   |
| Open water 1978    | Temperature; Irradiance; Mn; Current velocity; Discharge.  |
| Open water 1979    | Current velocity; Discharge; SiO <sub>2</sub> ; Mn.  |

Appendix 7. Summary of factors potentially influencing epidithic algal primary productivity.

Muskeg River

Entire period

Standing crop; Ca; SiO2; Mg.

Open water 1978

Standing crop.

Open water 1979

Mn; K; SiO<sub>2</sub>; Mg; Standing crop; Temperature;

PO4-P; Na; C1.

Steepbank River

Entire period

C1; Mg; K; NO3-N; Irradiance.

Open water 1978

Open water 1979

Standing crop; Irradiance; Temperature; NO3-N.

Hangingstone River

Entire period

NO<sub>3</sub>-N; Ca; Mg; Cl; K; Temperature.

Open water 1978

Standing crop.

Open water 1979

Standing crop.

MacKay River

Entire period.

NO3-N; Irradiance; Standing crop; SiO2; Mg.

Open water 1978

Irradiance; PO<sub>4</sub>-P; NO<sub>3</sub>-N; Standing crop; Fe; Mg; Ca; Temperature; SiO<sub>2</sub>; Mn.

Open water 1979

Temperature; Irradiance.

El/Is River

Entire period

PO4-P; Standing crop; Ca; Temperature.

Open water 1978

Fe; SiO<sub>2</sub>; Mn.

Open water 1979

Temperature; Irradiance; Standing crop.

Appendix 8. Correlations between epilithic algal standing crop and primary productivity for the entire study period and for the open water periods of 1972 and 1979.

| River        | . a )·   |                      | Open Wate           | r Period             |
|--------------|----------|----------------------|---------------------|----------------------|
| , **         |          | Entire Study         | Period 1978         | 1978                 |
| Muskeg       | •        | r ≈ 0.356<br>p •0.05 | r = 0.536<br>p 0.05 | r = 0.678<br>p 0.10  |
| Steepbank    |          | N.S.                 | N.S.                | r = 0.989<br>p 0.005 |
| Hangingstone | <b>:</b> | N.S.                 | r = 0.668<br>p 0.10 | r = 0.921<br>p 0.10  |
| MacKay       |          | r = 0.310<br>p 0.25  | r = 0.776<br>p 0.05 | N.S.                 |
| Ells -       | ta.      | r = 0.519<br>p -0.10 | N.S.                | r = 0.603<br>p 0.25  |

Appendix 9. Correlations between epilithic algal nitrogen fixation and nitrate nitrogen, chlorophyll <u>a</u>, numbers of Cyanophyceae and temperature.

| River             | Nitrate<br>Nitrogen | Cyanophycean<br>Numbers | Temperature     | Chlorophyll<br><u>a</u> |
|-------------------|---------------------|-------------------------|-----------------|-------------------------|
| Muskeg            | •                   | •                       |                 |                         |
| r                 | -0.304<br>p .05     | 0.274<br>p.08           | -0.202<br>p.18  | -0.410<br>p .01         |
| multiple r        | 0.518               | 0.522                   | 0.411           | 0.410                   |
| •                 | · ·                 | ٠.                      |                 |                         |
| Steepbank<br>r    | -0.258              | <b>€</b> 0.116          | 0.865           | 0.460                   |
|                   | p .18               | p .34                   | p .01           | p .05                   |
| multiple r        | 0.872               | 0.879                   | 0.855           | 0.879                   |
|                   |                     | •                       |                 |                         |
| Hangingstone<br>r | -0.397              | -0.279                  | 0.890           | 0.302                   |
|                   | p .08               | p .20,                  | p .01           | p .15                   |
| multiple r        | 0.905               |                         | 0.873           | 0.933                   |
| MacKay            |                     | •                       | •               |                         |
| r                 | -0.161              | 0.273                   | 0.438           | 0.089                   |
| multiple r        | p .27<br>0.373      | p .16                   | p .05<br>0.369  | p .38<br>0.250          |
| marerpre i        | 7                   |                         |                 |                         |
| Ells              |                     |                         |                 |                         |
| ៊ី <b>ក</b>       | -0.064              | -0.496                  | 0.796           | 0.429                   |
| multiple r        | p .41<br>0.815      | ₩ .04                   | p :01°<br>0.747 | p .07<br>0.813          |
| marerpie i        | 0.013               |                         |                 |                         |
| All Rivers        | -0.282              | 0.240                   | 0.398           | 0.036                   |
| Date Combined     | °p .01              | p .01                   | p .01           | p .37                   |
| multiple ré       | . 0.418             | 0.352                   | 0.464           | 0.471                   |

The mean and range for various physical and chemical factors for the five fivers 0.004 ×  $\frac{0.002 - 0.051}{x} = 0.009$ **t**~ \* MacKay Hangingstone Rivers 48.7 = 20.0 Steepbank mg:11.0 -44.8 Units Muskeg  $\text{mg.L-}^{-1} \, \underline{0}$ Conductance **Temperature** Potassium Sulphate Ca]cium Sodium

10.6

 $\frac{0}{x}$ .012 0.80 ×  $\frac{0}{x}$ 00.9 Ells  $\frac{0.008-0.157}{x} = 0.050$  $\frac{0.135}{x} = 0.515$ = 20.0 = 7.584.98 9.15 < **6.30 - 8.65** - 66.0 MacKay 0.45 ×١٥ 0.425  $\frac{0.006 - 0.600}{x} = 0.104$ Hangingstone  $\frac{30.0}{11.5}$ 9.75 6.50 - 8.50 5 - 66.0 x Rivers  $\frac{0.166}{x}$ 2.70 ×  $\frac{0}{x}$  $\frac{0.013-0.232}{x} = 0.049$ 0.345 = 0.224 $\frac{0.82}{x}$  - 5.30  $\frac{0.33}{\times} = 61.5$ 6.40 - 8.25Steepbank  $\frac{0}{x}$ .150- ( Ol×  $mg.L^{-1} \frac{0.006 - 0.352}{x} = 0.030$  $mg.L^{-1} = \frac{0.104 - 0.298}{x}$ - 35.6 = 10.3 6.20 -- 8.40 0.80 × ., Muskeg 0.48 ×i.× . 6.1× Appendix 10a. Concluded.  $m_{g_*}1^{-1}$ mg.L-1 Total meq.HCO3.L<sup>-1</sup> Alkalinity .Units Phosphate-phosphorus Discharge Dissolved **Chloride** Nitratenitrogen Silica

Appendix 10b. Relationships between various parameters and discharge

| Paramete         | Parameter Muskeg                       | Steepbank                               | Hangingstone                            | MacKay                                  | Ells                                   |
|------------------|--|---|---|---|--|
| N<br>a           | y=21.9-2.7 ln x<br>r=-0.368<br>p 0.025 | y=30.5-0.01 ln x<br>r=-0.894<br>p 0.005 | (.y=31.5-4.9 ln x<br>r=-0.562<br>p 0.01 | y=54.3-12.3 ln x<br>r=-0.834<br>p 0.005 | y=4.4-0.3 ln x<br>r=-0.695<br>p 0.005  |
| ×                | N.S.                                   | N.S.                                    | y=1.9-0.1 lm x<br>r=0.351<br>p 0.10     | N.S.                                    |  |
| æ<br>B           | y=10.1-2.1 ln x<br>r=-0.674<br>p 0.005 | y=12.2-2.0 lm x<br>r=-0.894<br>p 0.005  | y=9.6-2.6 ln x<br>r=-0.859<br>p 0.005   | y=11.6-2.3 ln x<br>r=-0.631<br>p 0.005  | y=4.1-0.05 ln x<br>r=-0.702<br>p 0.005 |
| E                | y=26.6-3.9 ln x<br>r=-0.618<br>p005    | y=24.6-3.9 ln x<br>r=-0.657<br>p 0.005  | y=22.1-4.2 ln x<br>r=-0.733<br>p 0.005  | y=29.96-4.8 ln x<br>r=-0.636<br>p 0.004 | N.S.                                   |
| -                | N.S.                                   | y=4.6-1.8 ln x<br>r=-0.754<br>p 0.005   | y=15.9-4.8 ln x<br>r=-0.836<br>p 0.005  | y=13.4-3.9 ln x<br>r=-0.806<br>p 0.005  | N.S.                                   |
| 504              | N.S.                                   | y=9.9-2.3 ln x<br>.r=-0.780<br>p 0.005  | y=22-4.8 ln x<br>r=-0.919<br>p 0.005    | y=38.4-7.5 ln x<br>r=-0.845<br>p 0.005  | N.S.                                   |
| Si0 <sub>2</sub> | y=6.5-0.99 ln x<br>r=-0.469<br>p 0.005 | y=6.0/1.1 ln x<br>r=-0.536<br>p 0.005   | y=6.7-1.1 ln x<br>r=-0.676<br>p 0.005   | N.S.                                    | .s.                                    |
| P04-P            | y=0.02-0.002 ln > r=-0.295 p 0.05      | x y=0.09-0.03 ln<br>r=-0.625<br>p 0.005 | × ×                                     | N.S.                                    | N.S.                                   |
| , NO3-N          | N.S.                                   | y=0.2+0.19 ln x<br>r=0.431              | S.N.                                    |   | N.S.                                   |
|                  | 4                                      |   | r k                                     | **N.S.=not significant                  | ant                                    |

Appendix 11. Correlations between epilithic algal standing crop and potentially limiting nutrients for the entire study period and for the open water periods of 1978 and 1979.

| Nutrie           | nt     | En                   | itire Study Perio    | od     |                       |
|------------------|--------|----------------------|----------------------|--------|-----------------------|
|                  | Muskeg | Steepbank            | Hangingstone         | MacKay | Ells                  |
| SiO <sub>2</sub> | N.S.a  | r = 0.385<br>p 0.10  | N.S.                 | N.S.   | r = -0.420 $p = 0.10$ |
| N03-N            | N.S.   | N.S.                 | N.S.                 | N.S.   | N.S.                  |
| P04-P            | N.S.   | N.S.                 | N.S.                 | N.S.   | r = -0.365 $p = 0.10$ |
| 1n               | N.S.   | r = 0.747<br>p 0.005 | r = 0.687<br>p 0.005 | N.S.   | r = 0.447<br>p 0.05   |
| Ca               | N.S.   | r = 0.371<br>p 0.10  | r = 0.412<br>p 0.05  | N.S.   | r = -0.442 $p = 0.05$ |
| Na               | N.S.   | N.S.                 | N.S.                 | N.S.   | r = -0.357<br>p 0.10  |

|                  |      | 0    | pen Water Period      | 1 (1978)             | <b>&amp;</b>                          |
|------------------|------|------|-----------------------|----------------------|---------------------------------------|
| SiO <sub>2</sub> | N.S. | •    | r = -0.590<br>p 0.10  | r = -0.854<br>p 0.01 | N.S.                                  |
| N03-N            | N.S. |      | r = -0.679 $p = 0.05$ | r = 0.918<br>p 0.01  | N.S.                                  |
| P04-P            | N.S. | N.S. | N.S.                  | r = -0.932<br>p 0.01 | N.S.                                  |
| Fe               | N.S. | N.S. | r = -0.591<br>p 0.10  | r = -0.875<br>p 0.01 | * * * * * * * * * * * * * * * * * * * |
| Mn .             | N.S. | N.S. | N.S.                  | r = -0.774<br>p 0.05 | r = -0.610<br>p 0.10                  |

Appendix 11. Concluded.

| Nutrie           | nt               |                     | Open Water Per      |                       |                     |
|------------------|------------------|---------------------|---------------------|-----------------------|---------------------|
| i.e              | Muskeg           | Steepbank           | Hangingstone        | MacKay                | Ells                |
| SiO <sub>2</sub> | N.S.             | r = 7.17<br>p 0.10  | r = 0.621<br>p 0.10 | N.S.                  | r = 0.594<br>p 0.25 |
| N03-N            | r=404<br>p -0.25 |                     | r = 0.479<br>p 0.25 | r = -0.605<br>p 0.25  | N.S.                |
| P04-P            | N.S.             | r = 0.385<br>p 0.25 |                     | N.S.                  | N.S.                |
| Fe               |                  | r = 0.406<br>p 0.25 | N.S.                | r = -0.679 $p = 0.10$ | N.S.                |
| Ca               | N.S.             | N.S.                | r = 0.828<br>p 0.05 | N.S.                  | N.S.                |
| Na               | N.S.             | N.S.                | r = 0.538<br>p 0.25 | r = 0.862<br>p 0.05   | N.S.                |
| :<br>Mn          | N.S.             | N.S.                | N.S.                | N.S.                  | r = 0.627<br>p 0.25 |
|                  |                  |                     |                     | 0 ~                   | *                   |

a N.S. = Not significant.

Appendix 11b. Correlations between epilithic algal standing crop and physical factors for the entire study period and for the open water periods of 1978 and 1979.

| Phŷšical<br>Factors |                    |                     | Entire Study Pe        | riod                 | f<br>ur              |
|---------------------|--------------------|---------------------|------------------------|----------------------|----------------------|
|                     | Muskeg             | Steepbank           | Hangingstone           | MacKay               | Ells                 |
| Temperature         | r=-0.265<br>p 0.10 | N.S.                | r = -0.191<br>p 0.25   | N.S.                 | N.S.                 |
| Irradiance          | N.S.               | N.S.                | N.S.                   | N.S.                 | r = -0.231<br>p 0.25 |
| Current<br>Velocity | r=-0.146<br>p 0.25 | r=-0.236<br>p 0.25  | r = -0.231<br>p 0.25   | r = -0.287<br>p 0.25 | N.S.                 |
| Discharge<br>/      | r=-0.168<br>p Q.25 | r=-0.202<br>p 0.25  | r = -0.217<br>p 0.25   | r = -306             | N.S. 77<br>p 0.10    |
|                     | • .                | -                   | Open Water Perio       | d (1978)             |                      |
| Temperature         | N.S.               | N.S.                | N.S.                   | r = 0.893<br>p 0.01  | r = -0.746<br>p 0.05 |
| Irradiance          | N.S.               | r=0.637<br>p 0.10   | r = 0.637<br>p 0.05    | r = 0.958<br>p 0.01  | r = -0.695<br>p 0.10 |
| Current<br>Velocity | r=-0.385<br>p 0.10 | r=-0.629<br>p 0.10  | r = -0.447 $p = 0.25$  | r = -0.720<br>p 0.05 | r = 0.416<br>p 0.25  |
| Discharge           | N.S.               | r=-0.596<br>p 0.25  | r = -0.413<br>p 0.25   | r = -0.909<br>p 0.01 | r =354<br>p 0.25     |
|                     | •                  |                     | Open Water Perio       | d (1979)             |                      |
| Temperature         | r=-0.625<br>p 0.10 |                     | r = -0.421<br>p 0.25   | r = -0.592<br>p 0.25 | N.S.                 |
| Irradiance<br>P     | r=-0.51<br>0.25    | 6 N.S.              | r = -0.44<br>p 0.25    | 18 N.S.              | N.S.                 |
| Current<br>/elocity | N.S.               | r = -0.65<br>p 0.10 | 1 r = -0.664<br>p 0.10 | r = -0.764<br>p 0.10 | r = 0.671<br>p 0.25  |
| Discharge           | N.S.               |                     |                        | r = -0.635 $0.25$    | r = 0.639<br>p 0.25  |