University of Alberta

Impacts of Environmental Temperature and Dietary Energy on Core Body Temperature and Efficiency in Broiler Breeder Females

by

Dulal Chandra Paul

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ABSTRACT

The effects of environmental temperature, dietary energy, housing system, time of feeding and day length on core body temperature (CBT) dynamics in broiler breeder females was studied in a series of experiments. Environmental temperatures within the range of 15 to 27°C resulted in a CBT of 39.8 to 42.1°C. Environmental temperature affected feed intake, growth, CBT dynamics and efficiency in pullets, but not the egg production, egg weight or feed efficiency in hens. Low energy diet-fed hens laid heavier eggs. Free-run and caged hens had similar egg production but free-run hens produced heavier eggs. However, free-run hens required by 17.2% more energy than caged hens, likely to support activity level. Feeding twice per day delayed oviposition relative to morning-fed hens. Photoperiod effects were seen in diurnal CBT patterns. Peak CBT occurred soon after feeding and could be shifted by changing feeding time, and may have potential for heat stress mitigation.

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DEDICATION

This thesis is dedicated to my parents (Bhakta M. Paul & Laxmi R. Paul) for their blessing throughout my life.

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FORMAT OF THESIS

This thesis is organized in accordance with the Journal style of Poultry Science and format allowing for independent chapters to be suitable for submission to the journal. Four papers have been prepared from research data collected from the project at the University of Alberta to partially fulfill the requirements for the degree of Master of Science. Each paper is complete in itself containing an abstract, introduction, materials and methods, results and discussion, and reference section.

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LIST OF ABBREVIATIONS

ADFI	Average daily feed intake
ADG	Average daily gain
BW	Body weight
BW^b	Metabolic body weight
CBT	Core body temperature
СР	Crude protein
DE	Digestible energy
EM	Egg mass production
FE	Fecal energy
GE	Gross energy
HI	Heat increment
HR	Heat retention
ME	Metabolizable energy
MEI	Metabolizable energy intake
ME _m	Metabolizable energy requirement for maintenance
NE	Net energy
NE _m	Net energy for maintenance
Nadg	Negative average daily gain
Padg	Positive average daily gain
RFI	Residual feed intake
RME _m	Residual maintenance ME requirement
THP	Total heat production
UE	Urinary energy
8	Residual error

1 CHAPTER 1: GENERAL INTRODUCTION

2 **1.1. BACKGROUND**

The foundation of the broiler industry is the broiler parent stock. Broiler breeders are 3 unique in that they produce rapidly growing offspring for meat production and, at the same time, 4 they need a high rate of egg production to supply the next generation of broiler chicks (Renema 5 et al., 2008). The management of broiler breeders is more challenging than that of other 6 commercial poultry due to the need to control body weight (BW) to optimise production 7 (Robinson et al., 1993; Renema and Robinson, 2004). Most of the feed eaten by broiler breeders 8 9 supports maintenance requirements (70 to 85%; Sakomura, 2004; Romero et al., 2009b), leaving a very small proportion of dietary energy for growth and production. Small changes in ME_m, 10 dramatically affect growth and production in feed-restricted broiler breeders. Therefore, feed 11 allocation is a big challenge in maintaining the target BW and maximising egg production. 12

Environmental temperature plays an important role in the use of feed energy because 13 birds are homoeothermic animals and they need to maintain a relatively constant core body 14 temperature (CBT). The rate of heat exchange with the environment is proportional to the 15 difference between the surface body temperature and the environmental temperature (National 16 17 Research Council, 1981). Estimates of the thermoneutral zone in which laying hens do not have to increase or decrease the rate of metabolic heat production to thermoregulate (National 18 Research Council, 1981) ranges from 16 to 28°C (Barott and Pringle, 1946), and 18 to 31°C 19 20 (Pereira et al., 2007) in broiler breeder hens. This zone is bound by the lower critical temperature below which birds must expend energy to increase their CBT, and the upper critical temperature 21 above which birds must expend energy to keep their CBT from increasing. If environmental 22 23 temperature is below the lower critical temperature, birds dissipate an increasing amount of body

1

24 heat to the environment (McDonald, 1978). Heat production and heat loss becomes imbalancd, and that affects CBT. In cold environments, birds require more energy to maintain CBT, which 25 means the energy requirement for maintenance increases, decreasing efficiency. Conversely, 26 above the upper critical environmental temperature birds are unable to passively dissipate 27 adequate heat to the environment. In this situation, they need to either expend energy to cool 28 themselves by wing flapping or panting, or reduce heat production by reducing feed intake, 29 which often leads to decreased growth or egg production. Broiler breeder hens give priority to 30 maintenance; growth, and egg production receive nutrients after maintenance is taken care of 31 32 (Reves et al., 2011). Since the energy required for growth is relatively small (Sakomura, 2004), any changes in maintenance energy requirements can quickly affect growth rates. Since 33 managing growth is important for reproductive success, accurate feed allocation that takes 34 environmental temperature into account is necessary. 35

Housing systems can affect energy requirement for chickens. Free run hens have more 36 spacious housing systems allowing them increased levels of activity compared to caged hens. 37 Over 70% of ME intake is expended for the maintenance requirement (Sakomura, 2004) and 20 38 to 25% of energy was used for activity in free run laying hens (Boshouwers and Nicaise, 1985). 39 40 In addition, Li et al. (1991) reported that 19% of total ME intake is used for activities in free run laying hens; whereas, Rabello et al. (2004) reported that a free-run broiler breeder hen required 41 21.8% higher ME intake for activities than a caged broiler breeder hen. Because maintenance 42 requirements are dramatically affected by activity level, feed rationing needs to take the broiler 43 breeder housing system into account to achieve target BW and egg production. 44

The main objective of this chapter is to review the effects of environmental temperature on CBT dynamics, heat loss to the environment, efficiency, energy partitioning and effects of activity on energy utilization under different housing systems in broiler breeder females.

48 **1.2. A BRIEF HISTORY OF BROILER BREEDERS**

49 **1.2.1. General history**

Poultry have been domesticated for thousands of years. The red jungle fowl of Indian 50 origin was first domesticated in 8000 BC in Asia, Africa and Europe mainly for cock fighting 51 (Alders and Pym, 2009; Lundeen, 2010). Once the fowl was domesticated, people used them as a 52 source of nutrition in the form of meat and eggs. Early in the 20th century, cockerels were used as 53 a meat bird in the USA. Some sporadic reports were found that the term "broiler" (meat 54 producing chicken) was used from 1920, and chicken companies and interested academia 55 worked to breed chickens for the development of broilers. Poultry enthusiasts have made many 56 genetic changes during the process of domestication by establishing local varieties and selecting 57 for various traits like growth rate, feed conversion rate, survivability etc. (Alders and Pym, 58 59 2009).

60 **1.2.2. History of selection – priority traits**

The history of breeding programs for meat producing birds (broiler) is relatively recent. In practice, a broiler breeding program was initiated in the 1957 in USA and came to light as a commercial broiler in the year 1976 (Lundeen, 2010). Primarily, growth rate was the first criterion for selection of broiler traits. The growth rate of broilers increased steadily based on rapid growth rate, short harvesting period, lower feed conversion rate and strong consumer demand (poultry meat is acceptable for almost all major religions), ensuring affordability for consumers. An annual growth rates by 2.4%, feed conversion ratio by 1.2% and breast muscle

yield by 1.1% increased from 1976 to 1999 due to respective selection traits in broiler breeders 68 (McKay et al., 2012). Over a 32-year period, the average BW target of Hubbard broilers 69 increased by 34.8 g/year/bird (Renema et al., 2007). The continually increasing growth rate of 70 modern broilers allows each new generation to potentially reach market weight between half-a-71 day (Havenstein et al., 2003) and one day less each year (Gyles, 1989). An important criterion for 72 73 economic efficiency is the amount of feed required to produce 1 kg of breast meat. The ratio of feed to breast meat yield was 20:1 (kg/kg) in 1976 and it is now around 7:1 (kg/kg) (McKay et 74 al., 2009). To make the broiler industry profitable and sustainable, emphasis has been given to 75 selection traits such as growth rate, breast meat yield, feed conversion rate, fertility, hatchability, 76 egg production, egg weight, skeletal integrity, feathering, and mortality (McKay et al., 2012). 77

78

1.2.3. What is feed restriction?

Feed restriction means to reduce feed allocation compared to full-fed animals. When 79 broiler breeder females are fed *ad libitum*, they become obese resulting in increased rates of 80 81 lameness, skeletal disorder and heart failure as well as metabolic disorders like ascites (Savory et al., 1993). This results in reduced settable egg production, egg quality and chick production (Yu 82 et al., 1992; Robinson et al., 1993; Renema and Robinson, 2004) and increased multiple 83 84 ovulation, abnormal eggs, and irregular oviposition time (Hocking et al., 1996). Ad libitum feeding is detrimental to health, welfare, and reproductive efficiency (Bokkers and Koene, 2003; 85 86 Hocking, 2004). Feed restriction has many positive effects on broiler breeders including control 87 of BW, delayed sexual maturity, reduced metabolic disorders and mortality, and maximized egg 88 production, fertility, hatchability and chick production (Katanbaf et al., 1989; Renema et al., 2007). The greatest economic benefit of feed restriction in broiler breeders is maximizing egg 89 90 and chick production (Renema et al., 2007).

91

1.2.4. Importance of feed restriction for egg and chick production

Growth and production are negatively correlated in broiler breeder hens (Renema et al., 92 2007). The reproductive performance of broiler breeders is evaluated by considering the total 93 number of settable eggs, age at onset of lay, average egg weight, and offspring produced per hen 94 (Renema and Robinson, 2004; Richards et al., 2010). Broiler breeder females fed *ad libitum* have 95 96 increased health disorders (Savory et al., 1993; Aviagen, 2007), leading to reduced settable egg production, egg quality, fertility, hatchability, and chick production (Jaap and Muir, 1968; Yu et 97 al., 1992; Hocking et al., 1996; Robinson et al., 1993; Renema and Robinson, 2004). Egg 98 99 production was highly affected by feed allocation, with 166, 159, and 137 settable eggs produced by the 100, 120, and 140% feed allocation groups, relative to primary breeder recommendations, 100 respectively (Renema et al., 2006). Feed allocation in broiler breeder females decreased by 8%, 101 102 16% and 24% based on primary breeder's guideline and did not affect settable egg production, egg weight, fertility and hatchability (Fattori, et al., 1991). Severe feed restriction in broiler 103 breeder pullets (25% of ad libitum) delayed lay until 40 wk of age and decreased egg production 104 and egg weight (Hocking, 2004). Moderately feed restricted (primary breeder's recommended 105 feed allocation) broiler breeder hens produced a higher number of settable eggs and chicks than 106 107 full-fed or severely feed restricted broiler breeder hens (Yu et al., 1992). Feed restriction in broiler breeders has been increasing every year since the mid-1970s resulting in an increased 108 health condition, egg production and chick production (Renema et al., 2007). Every year feed 109 110 allocation decreased by around 3% to maximize the total number of settable eggs and chicks. Egg production increased from 145 to 170 eggs and chick production increased from 128 to 150 111 112 chicks per hen housed from 1973 to 2005 (Renema et al., 2007). It means an increase of more 113 than one chick every two years.

114 **1.2.5.** Importance of feed restriction for chick quality

Moderate feed restriction has a positive correlation to egg and chick production, and it 115 also influences chick quality. Physically, quality chicks should be uniform with a higher length, 116 more alert and active, and well hydrated; additionally they should have good reflexes and well-117 healed navels with normal mortality of less than 0.5% by 5 d of age (Tona et al., 2003). Quality 118 chicks perform well during incubation (33% of a broiler's life) and maximize post-hatch 119 performance (growth rate, feed conversion rate, breast yield with minimum mortality) up to the 120 121 end of the harvesting cycle. Chick quality mainly depends on maternal nutrition, maternal health, 122 egg shape, egg storage time and management during incubation (Osman et al., 2010). It is necessary to provide enough but not excessive or inadequate nutrient intake to broiler breeder 123 hens for maximizing settable egg production and minimizing health disorders. A balanced diet 124 and feed allocation in broiler breeders influence chick weight. Summers (2010) reported that 125 hatchling weight was higher with a maternal diet content of 5.52 g of protein per 100 kcal energy 126 compared to any other ratios either higher or lower. Feed restriction (primary breeder guideline) 127 in broiler breeders during rearing and breeding phases increased fertility (13%), hatchability 128 (21%) and viability (13%) compared to full-fed broiler breeder females (Yu et al., 1992). Thus, 129 130 feed restriction in broiler breeder hens contributed to chick quality as well as offspring performance including BW. Feed allocation in breeder hens is a big challenge to maximize 131 reproductive output. 132

133

1.3. THE INCREASING DEGREE OF FEED RESTRICTION

In the history of feed restriction in laying chickens, Heywang (1940) was one of the first 134 135 to investigate the effects of feed restriction (a laying hen was not allowed to consume all feed 136 that she would normally consume) on egg production, egg weight and BW. Feed restriction in broiler breeders was reported first to control BW growth for increasing laying performance in the 1950s and 1960s (Novikoff and Byerly, 1945). A high fibre diet was used as a treatment to reduce growth rate of pullets BW around 8 wk of age but they did not observe any effect on layer performance (Issacks et al., 1960). Emphasis on feed restriction was given in pullets to delay sexual maturity (Lee et al., 1971) in order to maximize reproductive output.

142 The industry focus on feed restriction led to the development of a system of skip-a-day feeding to control broiler breeder BW. Due to increased growth potential in broiler breeders, the 143 degree of feed restriction has increased in order to maintain egg production traits (Bruggeman et 144 145 al., 1999). To optimize production, an average 3 g/year decrease in broiler breeder BW at 6 wk has occurred since the year 1978 guidelines (Renema et al., 2007). The recent female breeder 146 guides provide the lowest BW recommendations for 6 and 12 wk for most strains, allowing 147 greater gains between 18 and 24 wk of age for the nourishment and development of reproductive 148 organs, which enhanced subsequent egg production (Renema et al., 2007). Nowadays feed 149 restriction has increased compared to the primary breeder's recommended feeding guideline. A 150 151 little decreasing rate of feed allocation to the broiler breeders does not show yet any sign of slowing down the growth potential of broilers. Every year feed allocation decreased 3 g resulting 152 in a decreased BW in broiler breeders; at the same time, broiler BW increased 37g/year/bird 153 (Renema et al., 2007). The degree of feed restriction in broiler breeders directly contributes to 154 high productivity (Robinson et al., 1991) and indirectly to their offspring performances (Al-155 156 Murrani, 1978). The current levels of feed restriction are believed to maximize chick production.

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1.3.1. Scarcity of feed as a resource

With increasing rates of growth in broilers, and no increase in BW targets for their parent stock, the relative rate of feed intake continues to decrease. Feed has become a scarce resource for modern commercial broiler breeders. This has increased competition for feed among broiler breeders within a flock, resulting in great problems with flock uniformity (Hudson et al., 2001).

162 **1.3.2. Management of feed restriction in broiler breeders**

Feed restriction is applied about 25 to 50% of ad libitum in broiler breeder during the 163 rearing phase to control BW to avoid obesity from 4 to 20 wk of age (Savory et al., 1993; 164 165 Ducuypere et al., 2006). Feed restriction from 7 to 15 wk of age is very critical because of its association with long term increases in reproductive performance (Bruggeman et al., 1999). 166 After this period feed restriction begins to relax to allow rapid growth of body and reproductive 167 organs up to sexual maturity (Hocking et al., 1993), especially from 20 to 24 wk of age when 168 getting ready for onset of lay at 40 wk of age. Thereafter, feed restriction is much more relaxed 169 with about 70% of ad libitum feed to supply adequate nutrient for production as well as growth 170 and maintenance (Aviagen, 2007). This strategy will help to control body weight to maximize 171 settable egg production. Renema et al. (2007) reported that the degree of feed restriction 172 173 increased slowly each year to keep broiler breeders on a healthy growth curve and maximize reproductive output. 174

175 **1.4. ENERGY SYSTEMS**

176 **1.4.1.** Overview from Gross energy to Net energy

Energy is defined as potential capacity to perform work (Leeson and Summers, 2001). Energy partitioning in chickens is presented in Figure 1.1. The total energy contained in ingested feed is referred to as a gross energy (GE). Digestible energy (DE) is the remaining energy after 180 subtracting the fecal energy from GE. Fecal energy is the gross energy in the feces. Fecal energy 181 can be portioned into energy from undigested food and energy from compounds of metabolic origin. Metabolizable energy (ME) is the energy available after subtracting urinary energy and 182 gaseous energy losses from DE. Urinary energy is the total gross energy in urine. It includes 183 energy from non-utilized absorbed compounds from the food, end products of metabolic 184 processes, and end products of endogenous origin. Net energy (NE) is the ME of the feed 185 corrected for the energy losses due to heat increment (heat of fermentation, heat of digestion and 186 absorption, heat of product formation, heat of waste formation and excretion). The remaining NE 187 188 is used for maintenance and production. The energy requirement for maintenance is used energy 189 for basal metabolism, thermoregulation and normal activities of birds (Emmans, 1994). The remaining energy after maintenance requirement of birds is productive energy, used for growth 190 191 and other products such as eggs in chickens or milk in mammals. The proportion of ME for maintenance can vary based on the physiological status of an animal and environmental 192 temperature (Sakomura, 2004). 193

194 **1.4.2.** Metabolizable energy vs. Net energy

The ME is the standard measure of energy in both energy requirements and diets for 195 poultry (Lopez and Leeson, 2007. The ME partitioning in broiler breeders is complex and 196 critical, because they are feed restricted. The daily ME requirement of birds depends on age, 197 BW, body composition, growth rate, rate of egg production (Sakomura, 2004) and environmental 198 temperature (Ahmad et al., 1974). The ME partitioning model is $MEI = aW^{b}(T) + c\Delta W + dEM$, 199 where MEI is daily ME intake, W^{b} is metabolic body weight derived from $(BW)^{0.75}$, ΔW is body 200 weight change, EM is egg mass output per bird per day, T is environmental temperature, a, c, and 201 202 d are the coefficients of maintenance requirement, growth and production, respectively

(Sakomura et al., 2003). The energy requirement for maintenance is about 70 to 85% of the total ME intake in broiler breeders (Sakomura, 2004; Romero et al., 2009b). That means a very small proportion of energy remains available for growth and production. In any situation, if the energy requirement for maintenance increases then growth and production are hampered seriously because broiler breeders are feed restricted. Although broiler breeders have the capacity to increase feed intake to compensate for increased maintenance requirements, they cannot do so voluntarily.

The ME includes the total heat increments of digestion and absorption, product formation, fermentation, and waste formation and excretion. NE is the energy used by animal after subtracting the total heat increment from ME. In poultry, metabolizable energy is used for diet formulation.

214 *1.4.2.1. ME calculation*

Measurement of feed intake and excreta collection of chickens is difficult because excreta become contaminated with spilled feed and fallen feathers, scale, and down shed of birds. Inert indicators or markers such as chromic oxide, silicon, celite, ferric oxide, or barium sulphate are used to avoid this contaminating problem. In the current study, the ME was determined using celite as a marker according to the formula developed by Olukosi et al. (2007) and all values are expressed on a dry matter basis.

ME (kcal/g) of feed = GE per g of feed – [GE per g of excreta * (Concentration of marker
 in diet/ Concentration of marker in excreta)].

223 1.4.2.2. Body heat transfer systems

The amount of energy in the form of heat transferred from one subject to another depends on the difference of temperatures between two subjects. There are four ways to transfer heat

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226 from chickens' bodies to the environment (National Research Council, 1981). Conduction is the 227 process of heat transferring from homoeothermic animals by direct contact to litter and walls. Convection is a process of heat dissipation from the chickens' body to the surrounding 228 229 environment depending on air movement. Radiation is another process of heat transferring from the animal body to the environment by radiation, if the surface body temperature of a bird is 230 higher than the environmental temperature. Conversely warmer walls and roofs may radiate heat 231 to the bird surfaces. About 95% of heat in birds is transferred by conduction and convection, and 232 5% is transferred by radiation (Wolf and Walsberg, 2000). Evaporation is an important method 233 234 to dissipate body heat to the environment at above the upper critical environmental temperature, birds lose heat through evaporative cooling and it depends on the panting rate of a chicken 235 (Richards, 1976; Mack et al., 2013). Water evaporation cools the liquid remaining because it 236 must extract the necessary heat of vaporization from that liquid in order to make the change from 237 a liquid to a gaseous state. 238

239

1.4.2.3. Metabilic source of heat production

240 The heat increment is the increase in heat production following consumption of food by an animal (Smith et al., 1978). The following four heat increments can be referred to as the total 241 242 heat increments of maintenance and production (National Research Council, 1981). Heat of fermentation is the heat produced in the digestive tract by microbial action. The heat of 243 fermentation was established as an additional indicator of metabolic activity. Microbial 244 245 fermentation occurs mainly in cellulose-eating animals. In poultry, microbial fermentation is negligible. However, caeca take part in the digestion of cellulose particularly in geese because 246 they are good forager. The role of caeca in chickens is limited due to less intake of cellulose in 247 their diet. Heat of digestion is the heat produced by the action of digestive enzymes on the feed 248

within the digestive tract. Heat of absorption is produced by moving digesta and it is a byproduct of absorption of nutrients through the digestive tract. Heat of product formation is a byproduct of metabolic process of product formation from absorbed metabolites. This is the heat produced by biosynthetic pathways. Heat of waste formation and excretion is the heat production associated with the synthesis and excretion of waste products. Synthesis of urea from ammonia is an energy-costly process in mammals and results in an increased in total heat production.

255 1.4.3. Implications of feed restriction for nutrient partitioning

256 1.4.3.1. The energy requirement for maintenance

The energy requirement for maintenance is the energy required to maintain basal metabolism, activity and thermoregulation. The energy requirement for maintenance in chickens depends on body size, activities, and environmental temperature. Sakomura (2004) and Romero et al. (2009b) reported that around 70 to 85% of the total ME intake is required for maintenance in feed restricted broiler breeders. The energy requirement for maintenance includes energy required for basal metabolism, normal activities, and thermoregulation (Sakomura et al., 2005).

Basal metabolism is the basic energy used by a healthy animal without changing BW in the fasting, awake and resting states under thermoneutral temperature. This energy is required to maintain vital cellular activity, respiration, and blood circulation. The energy used for basal metabolism in laying chickens was calculated using the equation 79*BW^{0.75} ME kcal/kg metabolic body weight per day developed by Carpenter (2005).

Energy is required for normal activities of chickens such as standing, walking, wing flapping, and lying down. Energy required for normal activities in full fed laying hens by 26 to 270 29% of maintenance energy (Sakomura et al., 2005) in floor housing systems. However, Rabello et al. (2004) reported that activities required about 22% more energy for broiler breeder hens in a
floor housing system than in a cage housing system. Activities in chickens shared a significant
amount of maintenance requirement.

Endothermic animals like chickens need energy for thermoregulation. Chappell et al. (1990) reported that the energy required for thermoregulation in adult penguins was about 10 to 16% of basal metabolism at an environmental temperature of 5 to 6°C. However, the energy requirement for thermoregulation could vary in chickens with changing environmental temperature below the lower or above the upper critical temperature.

279 *1.4.3.2. The energy requirement for growth*

Broiler breeder pullets use only around 20% of the total ME intake for growth 280 (Sakomura, 2004). In cold environments below the lower critical temperature, birds dissipate an 281 282 increasing amount of heat to the environment due to the greater temperature difference between the surface body and the environment. As the energy requirement for maintenance increases to 283 maintain their CBT, the remaining small proportion of energy is further decreased, which 284 dramatically affect growth in birds (Sakomura, 2004). Because of feed restriction, broiler 285 breeders cannot increase voluntary feed intake to compensate for increased energy used for 286 287 maintenance. For example, if the energy requirement for maintenance would increase only 1% in a cold environment, it would reduce growth in feed-restricted broiler breeder pullets. Assuming 288 the total metabolizable energy intake was 192 kcal/day, 85% of total ME intake (163.2 kcal/day) 289 290 would be used for maintenance requirement; a 1% increase would be total 164.83 kcal/day; the remaining small proportion of energy further decreased by about 10.67% (1.63 kcal/day) which 291 would decrease growth 1.63 g/bird/day in feed restricted breeder pullets. An example for 292 293 broilers, assuming the total ME intake was 496 kcal/day, 50% of the total ME intake was 248

kcal/day for maintenance requirement (Sakomura, 2004). If the energy requirement for maintenance would increase by 1%, the energy requirement for maintenance would be total 250.5 kcal/day. On the other side, if energy would decrease only by 1% (2.48 kcal/day) and this energy would decrease growth (0.81 g/bird/day) in a full-fed broiler. It seems negligible and they can minimize the effect by increasing feed intake because feeds were supplied *ad libitum* to broilers.

300 1.5. ENVIRONMENTAL TEMPERATURE: IMPLICATIONS IN FEED RESTRICTED 301 ANIMALS

Heat production and heat loss in birds depend on environmental temperature. This can affect core body temperature, feed intake, heat production, heat retention, egg production and efficiency (Figure 1.2).

1.5.1. Role of CBT in heat exchange with the environment

The thermoneutral zone is the range of environmental temperatures (Bligh and Johnson, 306 1973) in which a homeothermic animal does not have to increase or decrease normal metabolic 307 heat production for maintaining CBT (National Research Council, 1981). The CBT of adult full-308 fed layer chickens ranges from 40.6 to 41.4°C (Deeb and Cahaner, 1999). Savory et al. (2006) 309 reported that the CBT of feed-restricted broiler breeder females ranges from 39.6 to 41.2°C, 310 while full-fed broiler breeder hens exhibited a range in CBT from 40.8 to 41.2°C. Diet-induced 311 thermogenesis occurs more or less continuously in full-fed birds, compared to once per day for 312 feed restricted broiler breeder hens (Savory et al., 2006). This is likely the reason for both the 313 higher mean and the lower range in CBT in full fed hens. 314

Environmental temperature has a great impact on birds' physiology including CBT dynamics and energy requirements (Khalil et al., 2004). In environmental temperatures below 317 the lower critical temperature, when the difference between the surface body temperature and environmental temperature is high ($\Delta T > 25^{\circ}C$), birds lose an increasing amount of heat to the 318 environment and eat more to increase the rate of metabolic heat production to maintain a 319 320 relatively constant CBT (National Research Council, 1981). In addition, a decrease in CBT can lead to behavioural responses, including huddling, reducing body surface area, and adjusting 321 322 feathers to increase insulation and to minimise body heat loss (Richards, 1971). Birds also try to maintain their CBT by reducing heat dissipation through vasoconstriction (Wolfenson, 1983). 323 Initially, birds can regulate all physiological functions but when environmental temperature 324 325 continuously decreases after 15°C, then birds increase feed intake as much as possible. Presumably, egg production could be sustained due to higher amount of feed intakes. In this 326 situation, egg production and egg weight may not be affected by a colder environment but 327 increased feed intake will lead to decreased efficiency. Heat retention decreased due to a higher 328 amount of heat dissipation to the colder temperature resulting in decreased CBT (Figure 1.2). 329 When the temperature difference increases between the surface body and environment (e.g. 330 331 environmental temperature less than 4°C), birds cannot control core body temperature by increasing the rate of metabolic heat production due to higher rate of heat dissipation. 332 333 Presumably, the CBT in chickens would decrease if CBT dropped down to lower lethal point at 22°C and then they would die (Robert and Shafner, 1951). Heat retention and egg mass 334 production also would decrease in severe cold weather (Figure 1.2). 335

Conversely, at environmental temperatures above the upper critical temperature, the difference between the surface body temperature and the environmental temperature decreases (< 12°C), and birds cannot dissipate adequate heat to the environment. Birds show behavioural responses to increase heat dissipation including separating from each other, increasing surface 340 area by standing, trying to stay near window (presumably looking for a colder area or finding a way to lose heat through convection (air movement) and shadow, wing drooping and spreading, 341 and eventually dissipating excess heat by panting (a faster rate of breathing to promote 342 343 evaporative water loss; Freeman, 1965; Donkoh and Atuahene, 1988). They also try to maintain their CBT by increasing body heat loss through vasodilation in hot environments (Brody, 1945). 344 When they fail to control from their CBT increasing, birds decrease the rate of metabolic heat 345 production by decreasing feed consumption (May. and Lott, 1992). Sufficiently reduced feed 346 consumption results in reduced productive outputs such as growth or egg production or both. 347 348 Feed intake, total heat production, egg mass production and efficiency decrease with increasing environmental temperatures above the upper critical temperature (Figure 1.2). However, heat 349 retention increases due to decreased rate of heat dissipation with increasing environmental 350 temperature. Increased heat retention above the upper critical temperature may raise CBT in 351 chickens. When CBT is reached at the upper lethal point at 45 to 47°C, birds would die (Robert 352 and Shafner, 1951). Heat stress has a greater negative impact on production than cold stress. 353

354 1.5.1.1 Diurnal CBT pattern

Diurnal CBT patterns in birds depend on controlling physical heat loss and chemical heat 355 356 production (Freeman, 1966). Environmental temperature and feed consumption influence physical heat loss and chemical heat production, respectively. As birds are homoeothermic 357 animals, they balance energy in their bodies by controlling heat production and heat loss 358 (Monteith, 1974). Generally, during the day birds increase feeding related activity and feed 359 metabolism (Khalil et al., 2004). These activities produce heat, leading to increased CBT 360 compared to night time CBT in birds. The CBT elevated before lights were turned on and peaked 361 362 after feeding; thereafter CBT gradually decreased and dropped suddenly after lights were turned

363 off (de Jong et al., 2002). Boiler breeder management is more difficult than commercial layers and broilers, because broiler breeders are feed restricted and they are unable to increase the rate 364 of diet induced heat production to maintain their CBT. An accurate feed allocation and a proper 365 feeding time may optimize energy partitioning for maximizing production using limited 366 resource. Diurnal CBT patterns generate a clear idea of heat production during the day. Studying 367 CBT patterns in broiler breeders can enrich fundamental knowledge for future research related to 368 core body temperature dynamics, and aid feed management in different environmental 369 temperatures. 370

371 1.5.1.2 Energy sparing strategies

Homeothermic animals can save energy by physiological and physical changes in both 372 hot and cold environments (Richards, 1971; Mustaf et al., 2009). The comb and wattles act as a 373 374 radiator in birds (Wilson and Plaister, 1951). Vasoconstriction is the mechanism in homeothermic animals under the lower critical temperature to constrict the muscular wall of the 375 arterial blood resulting in a decreased blood flow to the skin, wattle and comb, and they become 376 377 pale in color (Wolfenson, 1983). This mechanism reduces the blood flow to the skin, comb and wattles resulting in reduced heat loss to the environment. Vasoconstriction in homoeothermic 378 379 animals minimizes heat loss mainly by constricting blood vessels which increase tissue insulation under cold stress (below the lower critical temperature; Ames et al., 1970). In addition, 380 in colder environments, animal increase muscular movement and shivering to maintain core 381 body temperature (Richards, 1971; Khalil et al., 2004). Cold environments produce a reduced 382 blood flow and decreased heat loss through the comb and wattles to the environment; thus 383 384 chickens thermoregulate in cold environments.

385 The smaller the temperature difference between birds and the environment, the less heat will be exchanged (Richards, 1971). For an example, ducks and many other animals can swim or 386 walk on ice; they have a counter-current heat exchange system between the arteries and veins in 387 their legs (Thomas and Fordyce, 2007). Arteries carry warm blood from the heart to extremities 388 including the feet. Similarly, veins carry cold blood from the feet. The arterial blood warms up 389 390 the venous blood when they pass each other through the upper part of legs. The relatively cold arterial blood circulates through the feet. Thus, ducks reduce the temperature difference between 391 the feet and the ice resulting in a reduced heat loss and they can swim or walk on ice. However, 392 393 food and oxygen are supplied to the feet tissues through arterial blood, making them just warm enough to avoid frostbite. In addition, the lower legs and feet have less soft tissue, mainly tendon 394 connected with bone, which require less warm blood. 395

Interestingly, this same system can work when a bird is standing in excessively warm 396 water. Overheated venous blood returns from the feet to the heart and comparatively cold arterial 397 blood from the heart heats up while passing each other through the upper part of legs and this 398 399 warm arterial blood circulates to the feet. This counter current heat exchange reduces the temperature of venous blood before entering the core organs of the body; comparatively warm 400 401 arterial blood keeps the temperature of the feet above normal. In this case, the small temperature difference between the feet and environment reduces the heat exchange from the environment 402 into the feet (Midtgard, 1981). Thus, birds also resist hot environments. 403

404

1.5.1.3 Energy releasing strategies

The initial response of homoeothermic animals exposed to environmental temperatures above the upper critical temperature (under heat stress) is to increase heat loss by vasodilation (Hammel et al., 1963; Mustaf et al., 2009). Vasodilation is a mechanism which enlarges blood

vessels resulting in relaxation of smooth muscle in the large veins and arteries. Blood circulation 408 409 increases and blood pressure decreases in the peripheral part of skin, wattle and comb, and heat dissipation is increased to the environment. The skin, comb and wattles become red due to 410 increased blood flow. In hot environments, blood flows increase and dissipate heat through un-411 feathered skin, combs and wattles to the environment, and thus chickens regulate the 412 413 thermoregulation in hot environments.

414 **1.5.2.** Effects of heat loss on maintenance requirements

Heat loss of homoeothermic animals depends on environmental temperature. When 415 environmental temperature is below the lower critical temperature, they dissipate heat to the 416 417 environment and they eat more to increase the rate of metabolic heat production to maintain a relatively constant core body temperature. The energy requirement for maintenance thus 418 419 increases in colder environments, and that decreases energetic efficiency. When environmental 420 temperature goes above the upper critical temperatures, birds increase blood flow to the surface and decrease tissue insulation resulting in increased heat dissipation by vasodilation (Brody, 421 1945) and increase body surface area by changing posture. Thus, they facilitate the rate of heat 422 loss to the environment. When environmental temperature rises above the upper critical 423 424 temperature, the homeothermic animals start to dissipate heat to the environment by evaporative 425 cooling (Richards, 1976). Birds lose energy through evaporative cooling and they increase the 426 energy requirement for maintenance in the hot environments.

427

1.5.3. Effects of environmental temperature on production

428 The balance between heat production and heat loss in chickens is an important factor to ensure optimal performance. Heat loss to the environment increases the energy requirements of 429

the birds for thermogenesis; hence they increase their feed intake to meet their increased energy
requirements (Teeter et al., 2005). Above thermoneutral environmental temperatures, laying hens
reduce feed intake to maintain a relatively constant CBT by reducing heat production; however,
reduced feed intake may reduce feed efficiency and production (May. and Lott, 1992).

In laying hens, every 1°C decrease in temperature from 30 to 22°C increased feed intake 434 by 0.85% and energy intake by 0.86% (Ahmad et al., 1974). This might be due to less heat 435 dissipation to the higher environmental temperature. Donkoh (1989) reported that feed intake 436 decreased by 0.7%, 8.7%, and 12.9% in broilers during 3 to 7 wk of age, when the environmental 437 438 temperature was 25, 30 or 35°C respectively, compared to 20°C. As a consequence, BW gain decreased by 1.6%, 21.6%, or 32.4%, respectively, at 7 wk of age. Sakomura (2004) concluded 439 that below the lower critical temperature, ME_m increased by 6.73 kcal (6%) in laying-type pullets 440 for every 1°C decrease of environmental temperature; above the upper critical temperature the 441 ME_m decreased by 0.88 kcal per 1°C increase of environmental temperature. This decrease in 442 feed and energy requirement with an increase in environmental temperature compels birds to eat 443 444 less, resulting in decreased growth, egg production and egg weight (Donkoh and Atuahene, 1988). De Andrade et al. (1977) reported a decrease in egg production by 17% and egg weight 445 446 by 8% when environmental temperature increased from 21 to 31°C in laying hens. Al-Bashan and Al-Habibi (2010) also observed a decrease in egg production by 11% when the 447 environmental temperature increased from 30 to 35°C in laying hens. 448

The published literature leads us to conclude that environmental temperatures have a large effect on feed intake and maintenance energy requirements of chickens, which can dramatically change the amount of energy remaining to support growth and egg production. The role of change in environmental temperature in broiler breeders has not been studied extensively. A major difference in modern broiler breeders compared to full-fed chickens is that feed intake is controlled by the flock manager. Therefore, voluntary changes in feed intake in feed-restricted broiler breeders to meet growth and productivity targets do not occur. A very small proportion of the total ME intake of broiler breeders is productive energy, and small changes in energy partitioned to maintenance can dramatically affect the amount of energy partitioned to growth. This emphasizes the need to conduct research on the implications of environmental temperature for broiler breeder feed allocation decisions.

460 **1.6. EFFECTS OF HOUSING SYSTEMS ON ENERGY REQUIREMENT FOR**

461 MAINTENANCE

Chickens are housed either in cages or free-run systems. Peterman (2003) reported that 462 cage housing systems account for approximately 90% of all commercial layer farms in the world. 463 Traditionally, broiler breeders are reared in free-range housing to allow natural mating for fertile 464 eggs (Fuquay and Renden, 1980) to reduce labor cost. It will be interesting to note the effects of 465 different housing systems on energy requirement for maintenance in broiler breeders to maintain 466 467 the target BW. Energy intake for free-run laying hens was higher than those of caged laying hens. Sakomura (2004) reported that the energy requirement for maintenance in free-run broiler 468 469 breeder hens was above 70% of total ME intake. Li et al. (1991) suggested that 19% of total ME intake was used for activities in laying hens; whereas Rabello et al. (2004) reported that a free-470 run broiler breeder hen required 21.8% higher ME intake for activities than a caged broiler 471 472 breeder hen. Energy required for normal activities of birds are integral part of maintenance. Feed allocation decisions need to be adjusted to meet the energy requirement for maintenance 473 474 according to the housing system for maintaining a target BW in broiler breeder hens.

475 **1.7. IMPLICATIONS FOR EFFICIENCY**

476 **1.7.1. The concept of efficiency**

In general terms, efficiency can be defined as the ratio of the output to the input of any
system (Wang and Kim, 2011). Generally, biological efficiency and energetic efficiency are used
in the evaluation of livestock production.

480 *1.7.1.1. Biological efficiency*

Biological efficiency is the degree to which a conversion takes place to change physical inputs (feed) into saleable product (meat, egg or both) under a particular production environment (Wang and Kim, 2011). The definition of biological efficiency can be applied at both the individual animal level and at the industry level. Feed efficiency could be an evaluation criterion for the performance of an individual animal and as a whole flock.

486 1.7.1.2. Energetic efficiency

The current research compared the input-output relationships using chickens from an 487 efficiency perspective. The chemical energy of feed is used by a biological unit to do work 488 489 (Leeson and Summers, 2001). The transfer of heat and work in thermodynamic processes is governed by thermodynamic laws that were determined in the 19th century (Ebeling et al., 2005). 490 491 The first law of thermodynamics states that the total amount of energy in a system always remains constant (Lehninger, 1971). It cannot be created or destroyed. This law allows for an 492 accounting of the flow of energy within any energetic system, including chicken production, in 493 494 which energy inputs and outputs are equal. The second law of thermodynamics states that the total entropy will increase over time when energy is transferred from beginning to the end of the 495 process (Lehninger, 1971). Living organisms like chickens preserve their internal order by taking 496 497 in chemical energy (useful energy) of nutrients or sunlight and returning to their surroundings an equal amount of energy into a less usable form (Lehninger, 1982). Birds lose more heat to the
environment and they need more energy to maintain a relatively constant CBT, when the
environmental temperature drops below the lower critical temperature (National Research
Council, 1981). The amount of energy required based on the rate of heat lost to the environment.
Energetic efficiency in chickens will decrease where environmental temperature drops below the
lower critical temperature and it will seriously affect feed restricted broiler breeders.

504 1.7.1.3. Measuring energetic efficiency in broiler breeder hens

A relationship between input and output (feed efficiency) is a direct measure of energetic 505 efficiency in animals (Skinner-Noble and Teeter, 2004; Orejano-Dirain et al., 2004). The energy 506 507 requirement for maintenance in animals is an important measure of energetic efficiency; a metabolic BW is considered as a scaling factor determining part of the heat expenditure. The 508 509 approach may be taken to assess energetic efficiency in poultry: quantification of residual variability in energy balance models (Romero et al., 2009a). The approach is generally used, and 510 is known as residual feed intake (RFI). The RFI is defined as the difference between observed 511 and predicted ME intakes. Bordas and Merat (1981), Johnson et al. (1999), and Herd et al. 512 (2003) reported that RFI has been used to determine energetic efficiency in different species 513 since the 1980s. An efficient animal is one that consumes less energy than the theoretical 514 515 requirement (Romero et al., 2009a). Estimations of ME requirements in RFI calculations have incorporated some of the assumptions of ME models of energy partitioning. The following 516 equation was used to determine the RFI of broiler breeders (Romero et al., 2009a): 517

518 $\mathcal{E}=MEI-\{(a-bT) BW^{0.75}+cADG+dEM\}$

where \mathcal{E} is the residual feed (energy) intake, MEI is ME intake kcal/d, BW^{0.75} is metabolic BW in Kg, T is temperature in °C, ADG is average daily gain (g/d), EM is daily egg mass production (g/d) and a, b, c and d are the coefficient of body weight, temperature, growth and egg mass respectively. The ME requirement for maintenance (ME_m) is generally considered to be dependent on environmental temperature, activity, BW, and includes heat increment of feeding.

524 **1.8. PROBLEM STATEMENT**

Management of genotype, nutrition and environmental temperature are key factors for the 525 productivity and sustainability of broiler breeders (Renema et al, 2008). Feed alone accounts for 526 approximately 60 to 70% of total costs of poultry production (Steiner et al., 2008). Feed 527 allocation and maintenance energy requirements for the broiler breeder are big challenges when 528 attempting to maximize production and increase efficiency under different environmental 529 530 temperatures. The energy requirement for maintenance is the first priority for an animal especially in broiler breeders because they are feed restricted. Environmental temperature 531 significantly affects the ME_m and thus efficiency (Zuidhof et al., 2012). Researchers have 532 533 observed that for laying hens in cold environments (environmental temperature is below the lower critical temperature), the difference between the surface body temperature and 534 535 environmental temperature increases, and the birds dissipate an increasing amount of heat to the environment (McDonald, 1978). They may eat more to increase the rate of metabolic heat 536 production to maintain a relatively constant CBT (National Research Council, 1981). In hot 537 538 environments (environmental temperature goes above the upper critical temperature but not exceeds the surface body temperature), when the difference between the surface body 539 temperature and the environmental temperature decreases birds are unable to dissipate necessary 540 541 heat to the environment resulting in a decreased the dependency on diet-induced thermogenesis

542 and they reduce feed intake. The larger proportion of ME intake is used for maintenance (70 to 85%; Sakomura, 2004; Romero et al., 2009b) and the remaining very small proportion of ME 543 intake is used for growth and production. When the energy requirement for maintenance 544 increases, then the energy available for growth and production becomes severely deficit resulting 545 in a decreased growth and production in broiler breeder females because they are supplied a 546 limited amount of feed. Thus, cold or hot environments decrease efficiency in broiler breeder 547 hens. Feed allocation needs to be adjusted with changing environmental temperature. However, 548 the effects of environmental temperature vary the energy requirement in broiler breeder hens to 549 maintain a relatively constant CBT. The relationship among the diurnal CBT pattern and 550 different dietary energy levels needs to be more precisely quantified for broiler breeder hens. 551 Therefore, to address the problem of an accurate estimation of feed allocation for the energy 552 553 requirement for maintenance with respect to change in environmental temperatures the current research was conducted. 554

555 **1.9. OBJECTIVES**

556 **1.9.1. General objective**

557 The main purpose of the current research was to investigate the effects of environmental 558 temperature and dietary ME level on core body temperature dynamics and efficiency in broiler 559 breeder females.

560 **1.9.2. Specific objectives**

To determine the relationship between environmental temperatures, core body
temperature dynamics and growth efficiency in broiler breeder pullets (Chapter 2).

563	• To investigate the effects of environmental temperatures and dietary energy levels on		
564	core body temperature dynamics and reproductive efficiency in broiler breeder hens		
565	(Chapter 3).		
566	• To identify the impacts of feeding time, photoperiod, and dietary ME level on core body		
567	temperature dynamics and oviposition time in broiler breeder hens (Chapter 4).		
568	• To assess the maintenance energy requirement, core body temperature dynamics and		
569	efficiency of broiler breeder hens in cage versus free run housing systems (Chapter 5).		
570	1.10. HYPOTHESES		
571	Birds in low environmental temperature will increase heat loss to the environment leading to		
572	increased feed intake to maintain core body temperature and reduced efficiency.		
573	A low energy diet will increase feed intake resulting in an increased core body temperature led to		
574	greater heat loss to the environment and decreased efficiency.		
575	1.11. APPROACH		
576	Four studies using Ross 708 broiler breeders were conducted to investigate		
577	i).Effects of four different environmental temperatures (15, 19, 23 and 27°C) on core body		
578	temperature dynamics and performance of broiler breeder pullets (4 to 20 wk);		
579	ii). Impacts of four different environmental temperatures and two dietary energy (High:2,912 ME		
580	kcal/kg; Low: 2,872 ME kcal/kg) levels on core body temperature dynamics and performance of		
581	broiler breeder hens (25 to 41 wk);		
582	iii) Effects of four feeding times (07:30, 11:30, 15:30 and Split: 07:30 and 15:30) with two		
583	photoperiods (16L:8D and 24L:0D) and two dietary energy levels on core body temperature		
584	dynamics and oviposition times of broiler breeder hens (44 wk); and		

iv) Effects of cage versus free run housing systems on the energy requirement for maintenance, core body temperature dynamics, and performance in broiler breeder hens (25 to 41 wk). Feed allocation was readjusted with changing environmental temperature to achieve the target BW of broiler breeder females at chamber level. Data on core body temperature, ADFI, egg production, and egg weight were collected. This offered a better understanding of core body temperature dynamics, the energy requirement for maintenance and efficiency in broiler breeder females.

592 **1.12. REFERENCES**

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- Ahmad, M. M., F. B. Mather, and E. W. Gleaves. 1974. Environmental temperature and dietary
 energy on dwarf and normal hens and normal roosters. Poult. Sci. 53:927-935.
- Al-Bashan, M. M., and M. S. Al-Habibi. 2010. Effects of ambient temperature flock age and
 breeding stock on egg production and hatchability of broiler hatching eggs. Eur. J. Biol.
 Sci. 2:55-66.
- Al-Murrani, W. K. 1978. Maternal effects on embryonic and post-embryonic growth in poultry.
 Br. Poult. Sci. 19:277-281.
- Alders, R. G., and R. A. E. Pym. 2009. Village poultry: still important to millions, eight thousand
 years after domestication. World's Poult. Sci. J. 65:181-190.
- Ames, D. R., J. E. Nellor, and T. Adams.1970. Biothermal vasomotion in sheep. J Anim. Sci.
 31:80-84.
- Aviagen. 2007. Ross 708 parent stocks performance objectives. Aviagen Ltd.
 www.aviagen.corn/docs/Ross%20708%20PS%20Performance%20Objectives.pdf
 Accessed June 2010.
- Barott, H. G., and E. M. Pringle. 1946. Energy and gaseous metabolism of the chicken from
 hatch to maturity as affected by temperature. J. Nutr. 31:35-50.
- Bligh, J., and K. G. Johnson. 1973. Glossary of terms for thermal physiology. J. Appl. Physiol.
 35:941-961.
- Bokkers, E. A. and, P. Koene, 2003. Eating behaviour, and preprandial and postprandial
 correlations in male broiler and layer chickens. Br. Poult. Sci. 44:538-544.
- Bordas, A., and P. Merat. 1981. Genetic variation and phenotypic correlations of food
 consumption of laying hens corrected for body weight and production. Br. Poult. Sci.
 22:25-33.
- Boshouwers, F. M. G., and E. Nicaise. 1985. Automatic gravimetric calorimeter with
 simultaneous recording of physical activity for poultry. Br. Poult. Sci. 26:534-541.
- Brody, S. 1945. Bioenergetics and growth. Pages 274-275. Reinhold Publishing Corporation.
 N.Y.
- Bruggeman, V., O. Onagbesan, E. D'Hondt, N. Buys, M. Safi, D. Vanmontfort, L. Berghman, F.
 Vandesande, and E. Decuypere. 1999. Effects of timing and duration of feed restriction
 during rearing on reproductive characteristics in broiler breeder females. Poult. Sci.
 78:1424-1434.

636 637	Carpenter, J. W. 2005. Exotic animal formulary. Page 559, 3 rd Ed. Elsevier Saunders, St. Louis.
638 639 640	Chappell, M. A., K. R. Morgan, and T. L. Bucher. 1990. Weather, microclimate and energy costs of thermoregulation for breeding Adelie penguins. Oecologia. 83:420-426.
641 642 643 644	De Andrade, A. N., J. C. Rogler, W.R. Featherston, and C. W. Alliston. 1977. Interrelationships between diet and elevated temperatures (cyclic and constant) on egg production and shell quality. Poult. Sci. 56:1178-1188.
645 646 647 648	Deeb, N., and A. Cahaner. 1999. The effects of naked neck genotypes, ambient temperature, and feeding status and their interactions on body temperature and performance broilers. Poult. Sci. 78:1341-1346.
649 650 651 652	de Jong, I. C., S. Van Voorst, D. A. Ehlhardt, and H. J. Blokhuis. 2002. Effects of restricted feeding on physiological stress parameters in growing broiler breeders. Br. Poult. Sci. 43: 157-168.
653 654 655	Donkoh, A., and C. C. Atuahene. 1988. Management of environmental temperature and rations for poultry production in the hot and humid tropics. Int. J. Biometeorol. 32:247-253.
656 657 658	Donkoh, A. 1989. Ambient temperature a factor affecting performance and physiological response of broiler chickens. Int. J. Biometeorol. 33:259-265.
659 660 661 662	Ebeling, W., I. M. Sokolov, and L. Schimansky-Geier. 2005. On history of fundamentals of statistical thermodynamics. Pages 2-3 in Statistical Thermodynamics and Stochastic Theory of Nonequilibrium Systems. World Scientific Publishing. Hackensack NJ, USA.
663 664 665	Emmans, G. C. 1994. Effective energy: a concept of energy utilization applied across species. Br. J. Nutr. 71:801-821.
666 667 668 669	Fattori, T. R., H. R. Wilson, R. H. Harms, and R. D. Miles. 1991. Response of broiler breeder females to feed restriction below recommended levels. 1. Growth and reproductive performance. Poult. Sci. 70:26-36.
670 671 672	Freeman, B. M. 1965. The relationship between oxygen consumption, body temperature and surface area in the hatching and young chick. Br. Poult. Sci. 6:67-72.
673 674 675	Freeman, B. M. 1966. Physiological responses of the adult fowl to environmental temperature. World's. Poult. Sci. J. 22:140-145.
676 677 678	Fuquay, J. I., and J. A. Renden. 1980. Reproductive performance of broiler breeders maintained in cage or on floors through 59 weeks of age. Poult. Sci. 59:2525-2531.
679 680	Gyles, N. R. 1989. Poultry, people and progress. Poult. Sci. 68:1-8.

681	Hammel, H. T., D. C. Jackson, J. A. J.Stolwijk, J. D.Hardy, S. B. Stromme. 1963.
682	Temperature regulation by hypothalamic proportional control with adjustable set
683	temperature. J. Appl. Physiol. 18:1146-1154.
684	
685	Havenstein, G. B., P. R. Ferket, and M. A. Qureshi. 2003. Growth, livability, and feed
686	conversion of 1957 versus 2001 broilers when fed representative 1957 and 2001 broiler
687	diets. Poult. Sci. 82:1500-1508.
688	
689	Herd, R. M., J. A. Archer, and P. F. Arthur. 2003. Reducing the cost of beef production through
690	genetic improvement in residual feed intake: opportunity and challenges to application. J.
691	Anim. Sci. 81(E. Suppl. 1):E9-E-17.
692	
693	Heywang, B. W. 1940. The effect of restricted feed intake on egg weight, egg production,
694	andbody weight. Poult. Sci. 19:29-34.
695	
696	Hocking, P. M. 2004. Roles of body weight and feed intake in ovarian follicular dynamics in
697	broiler breeders at the onset of lay and after a forced molt. Poult. Sci. 83:2044-2050.
698	
699	Hocking, P. M., M. H. Maxwell, and M. A. Mitchell. 1996. Relationship between the degrees of
700	feed restriction and welfare indices in broiler breeder females. Br. Poult. Sci. 37:263-278.
701	
702	Hocking, P. M., M. H. Maxwell, and M. A. Mitchell. 1993. Welfare assessment of broiler
703	breeder and layer females subjected to food and water restriction during rearing. Br.
704	Poult. Sci. 34:443-458.
705	
706	Hudson, B. P., R. J. Lien, and J. B. Hess. 2001. Effects of body weight uniformity and pre-peak
707	feeding programs on broiler breeder hen performance. J. Appl. Poult. Res. 10:24-32.
708	
709	Issacks, R.E., B. L. Reid., R. E. Davies, J. H. Quisenberry, and J. R. Couch. 1960. Restricted
710	feeding of broiler type replacement stock. Poult. Sci. 39:339-346.
711	
712	Jaap, R. G., and F. V. Muir. 1968. Erratic oviposition and egg defects in broiler-type pullets.
713	Poult. Sci. 47:417-423.
714	
715	Johnson, Z. B., J. J. Chewning, and R. A. Nugent. 1999. Genetic parameters for production traits
716	and measures of residual feed intake in large white swine. J. Anim. Sci. 77:1679-1685.
717	
718	Katanbaf, M. N., E. A. Dunnington, and P. B. Siegel, 1989. Restricted feeding in early and late-
719	feathering chickens.2. Reproductive responses. Poult. Sci. 68:352-358.
720	
721	Khalil, A. M., K. Matsui, and K. Takeda. 2004. Diurnal and oviposition-related changes in heart
722	rate, body temperature and locomotors activity of laying hens. Anim. Sci. J. 75:169-174.
723	
724	Lee, P. J. W., A. L. Gulliver, and T. R. Morris. 1971. A quantitative analysis of the literature
725	concerning the restricted feeding of growing pullets. Br. Poult. Sci. 12:413-437.

726 727 728	Leeson, S., and J. D. Summers. 2001. Pages 34-99 in scott's nutrition of the chicken. 4 th Ed. University Books. Guelph, ON.
729 730	Lehninger, A. L. 1971. Bioenergetics. 2nd Ed. W. A. Benjamin. Pages 18-36, Menlo Park CA.
731 732 733	Lehninger, A. L. 1982. The ATP cycle and cell bioenergetics. Pages 361-396 in Principles of Biochemistry. 1st Ed. Worth Publishers. New York, NY.
734 735 736	Li, Y.; T. Ito., and S. Yamamoto. 1991. Diurnal variation of heat production related to some physical activities in laying hens. Br. Poult. Sci. 32:821-827.
736 737 738 739	Lopez, G. and Leeson, S. 2007. Relevance of nitrogen correction for assessment of metabolizable energy with broilers to forty-nine days of age. Poult. Sci. 86:1696-1704.
740 741	Lundeen, T. 2010. How wild chicken became modern broiler. www.feedstuffsfoodlink.com.
742 743 744	May, J. D., and B. D. Lott.1992. Feed and water consumption patterns of broilers at high environmental temperatures. Poult. Sci. 71:331-336.
745 746 747 748	Mack, L.A., J.N. Felver-Gant, R. L. Dennis, H. W. Cheng. 2013. Genetic variation alters production and behavioral responses following heat stress in 2 strains of laying hens. Poult. Sci. 92:285-294.
748 749 750	McDonald, M. W. 1978. Feed intake of laying hens. World's Poult. Sci. J. 34:209-221.
751 752 753	McKay, J. C. 2009. The genetics of modern commercial poultry. Pages 3-9 in Biology of Breeding Poultry. Ed. P.M. Hocking, University of Edinburgh, UK.
754 755 756 757	McKay, J. C., N. F. Barton, A. N. M. Koerhuis, and J. McAdam. 2012. The challenge of genetic change in the broiler chicken. Ross Breeders Limited, Newbridge, Midlothian EH28 8SZ, UK. http://bsas.org.uk/downloads/genchan/paper1.pdf. Accessed November 24' 2012.
758 759 760 761	Midtgard, U. 1981. The rete tibiotarsale and arterio-venous association in the hind limb of birds: a comparative morphological study on counter-current heat exchange systems, Acta Zoologica, 62:67-87.
762 763 764	Monteith, J. L. 1974. The concept of thermal neutrality, page 425, in heat loss from animals and man, ed J. L. Monteith and L. E. Mount, Butterworth, London.
765 766 767	Mustaf, S.; N. S. Kahraman, and M. Z. Firat. 2009. Intermittent partial surface wetting and its effect on body-surface temperatures and egg production of white brown domestic laying hens in Antalya (Turkey). Br. Poult. Sci. 50:33-38.
768 769 770	National Research Council. 1981. Effect of environment on nutrient requirements of domestic animals. Natl. Acad. Press, Washington D. C.

771 Olukosi, O. A., A. J. Cowieson and O. Adeola. 2007. Age-related influence of a cocktail of 772 xylanase, amylase, and protease or phytase individually or in combination in broilers. Poult. Sci. 86:77-86. 773 774 Novikoff, M., and J. Byerly. 1945. Observations on two methods of feeding chickens from one 775 776 day to twelve months of age. Poult. Sci. 24:245-251. 777 778 Orejano-Dirain, C. P., M. Iqbal, D. Cawthon, S. Swonger, T. Wing, M. Cooper, and W. Bottje. 779 2004. Determination of mitochondrial function and site-specific defects in electron 780 transport in duodenal mitochondria in broilers with low and high feed efficiency. Poult. Sci. 83:1394-1403. 781 782 783 Osman, A. M. R., H. M. A. Wahed, and M. S. Ragab. 2010. Effects of supplementing laying 784 hens diets with organic selenium on egg production, egg quality, fertility and hatchability. Egypt. Poult. Sci. 30:893-915. 785 786 787 Pereira, D. F., I. A. Naas, C. E. B. Romanini, D. D. Salgado, and G. O. T. Pereira. 2007. Broiler breeder behavior and egg production as function of environmental temperature. Braz. J. 788 Poult. Sci. 9:9-16. 789 790 791 Peterman, S. 2003. Laying hens in alternative housing systems-practical experiences. 792 Dtsch.Tierarztl. Wochenschr, 110:220-224. 793 794 Rabello, C. B. V., N. K. Sakomura, F. A. Longo, and K. T. de. Resende. 2004. Effect of 795 the environmental temperature and rearing systems on metabolizable energy requirements for maintenance of broiler breeder hens. R. Bras. Zootec. 33:382-390. 796 797 Renema, R. A. and F. E. Robinson. 2004. Defining normal: comparison of feed restriction and 798 799 full feeding of female broiler breeders. World's Poult. Sci. J. 60:508-522. 800 Renema, R. A., F. E. Robinson, and M. J. Zuidhof. 2006. Role of broiler breeder genetics on 801 802 breeder chick quality and sensitivity to overfeeding. Aust. Poult. Sci. Symp. 18:34-38. 803 Renema, R. A., M. E. Rustad, and F. E. Robinson. 2007. Implications of changes to commercial 804 broiler and broiler breeder body weight targets over the past 30 years. World's Poult. Sci. 805 J. 63:457-472. 806 807 Renema, R. A., V. R. Sikur, F. E. Robinson, D. R. Korver, and M. J. Zuidhof. 2008. Effects 808 of nutrient density and age at photostimulation on carcass traits and reproductive 809 efficiency in fast- and slow-feathering turkey hens. Poult. Sci. 87:1897-1908. 810 811 812 Richards, S. A. 1971. The significance of changes in the temperature of the skin and body core of the chicken in the regulation of heat loss. J. Physiol. 216:1-10. 813 814

815 816	Richards, S. A. 1976. Evaporative water loss in domestic fowls and its partition in relation to ambient temperature. J Agric. Sci. 87:527-532.
817	
818	Richards, M. P., R. W. Rosebrough, C. N. Coon, and J. P. McMurtry. 2010. Feed intake
819	regulation for the female broiler breeder: In theory and in practice. J. Appl. Poult. Res.
820	19:182-193.
821	
822	Robert, E. M., and C. S. Shafner. 1951. Lethal internal temperature for the chicken, from fertile
823	egg to mature bird. Poult. Sci. 30:255-266.
824	Debiner E.E. N.A. Debiner and T.A. Statt 1001 Denne better ne fermione and the
825	Robinson, F. E., N. A. Robinson, and T. A. Scott. 1991. Reproductive performance, growth rate
826	and body composition of full-fed versus feed-restricted broiler breeder hens. Can. J.
827	Anim. Sci. 71:549-556.
828	Debinson E.E. I.I. Wilson M.W. Vu C. M. Essenke and P. T. Hardin 1002. The
829	Robinson, F. E., J. L. Wilson, M. W. Yu, G. M. Fasenko, and R. T. Hardin. 1993. The
830	relationship between body weight and reproductive efficiency in meat-type chickens.
831	Poult. Sci.72:912-972.
832 833	Remove I E M I Zuidhof R A Renome A Nacime and E E Robinson 2000a
833 834	Romero, L. F., M. J. Zuidhof, R. A. Renema, A. Naeima, and F. E. Robinson. 2009a. Characterization of energetic efficiency in adult broiler breeder hens. Poult. Sci. 88:227-
835	235.
835 836	255.
837	Romero, L. F., M. J. Zuidhof, R. A. Renema, F. E. Robinson, and A. Naeima. 2009b. Nonlinear
838	mixed models to study metabolizable energy utilization in broiler breeder hens. Poult.
839	Sci. 88:1310-1320.
840	561. 00.1510-1520.
841	Reyes, M. E., C. Salas, and C. N. Coon. 2011. Energy requirements for maintenance and egg
842	production of broiler breeder hens. Int. J. Poult. Sci. 10:913-920.
843	production of brother breeder hend. Int. 5. 1 out. Bei. 10.913 920.
844	Sakomura, N. K., R. Silva, H. P. Couto, C. Coon, and C. R. Pacheco. 2003. Modeling
845	metabolizable energy utilization in broiler breeder pullets. Poult. Sci. 82:419-427.
846	
847	Sakomura, N. K. 2004. Modeling energy utilization in broiler breeders, laying hens and broilers.
848	Braz. J. Poult. Sci. 6:1-11.
849	
850	Sakomura, N. K., R. Basaglia, C. M. L. Sa-Fortes, and J. B. K. Fernandes. 2005. Model for
851	metabolizable energy of laying hens. Braz. J. Anim. Sci. 34:557-567.
852	
853	Savory, C. J., K. Maros, and S. M. Rutter. 1993. Assessment of hunger in growing broiler
854	breeders in relation to a commercial restricted feeding programme. Anim. Welf. 2:131-
855	152.
856	
857	Savory, C. J., L. Kostal, and I. M. Nevison. 2006. Circadian variation in heart rate, blood
858	pressure, body temperature and EEG of immature broiler breeder chickens in restricted-
859	fed and <i>ad libitum</i> -fed states. Br. Poult. Sci. 47:599-606.

- Schwab, R. G., and V. F. Schafer. 1972. Avian thermoregulation and its significance in starling
 control. Proceedings of the 5th Vertebrate Pest Conference. Paper 25.
 http://digitalcommons.unl.edu/vpc5/25.
- Skinner-Noble, D. O., and R. G. Teeter. 2004. Components of feed efficiency in broiler breeding
 stock: The use of fasted body temperature as an indicator trait for feed conversion in
 broiler chickens. Poult. Sci. 83:515-520.
- Smith, R. R., G. L. Rumsey, and A. L. Scott. 1978. Heat increment associated with dietary protein, fat, carbohydrate and complete diets in salmonids 1: Comparative energetic efficiency. J. Nutr. 108:1025-1032.
- Steiner, Z., M. Domacinovic, Z. Antunovic, Z. Steiner, D. Sencic, J. Wagner, and D. Kis. 2008.
 Effect of dietary protein/energy combinations on male broiler breeder performance. Acta agriculturae Slovenica Suppl. 2:107-115.
- 876 Summers, J. D. 2010. Meeting the nutrient requirement of broiler breeders.
 877 http://www.cfo.on.ca/_pdfs/AugustNL-FN.pdf nov21.12
- Teeter, R. G., L. McKinney, and A. Baker. 2005. An accounting of broiler energy expenditure
 Feed info website, August.
- Thomas, D. B., and R. E. Fordyce. 2007. The heterothermic loophole exploited by penguins.
 Aust. J. Zool. 55:317-321.
- Tona, K., F. Bamelis, B. De Ketelaere, V. Bruggeman, V. M. B. Moraes, J. Buyse, O.
 Onagbesan, and E. Decuypere. 2003. Effects of egg storage time on spread of hatch,
 chick quality and chick juvenile growth. Poult. Sci. 82:736-741.
- Wang, J. P., and I. H. Kim. 2011. Effect of caprylic acid and yucca schidigera extract on
 production performance, egg quality, blood characteristics, and excreta microflora in
 laying hens. Br. Poult. Sci. 52:711-717.
- Wilson, W. 0., and T. H. Plaister. 1951. Skin and feather temperatures of hens kept at
 constant environmental conditions. Am. J. Physiol. 166:572-577.
- Wolf. B. O., and G. E. Walsberg. 2000. The role of plumage in heat transfer process of birds.
 Am. Zool. 40:575-584.
- Wolfenson, D. 1983. Blood flow through arteriovenous anastomoses and its thermal function
 in the laying hen. J. Physiol. 334:395-407.
- 901 902

867

871

875

878

881

884

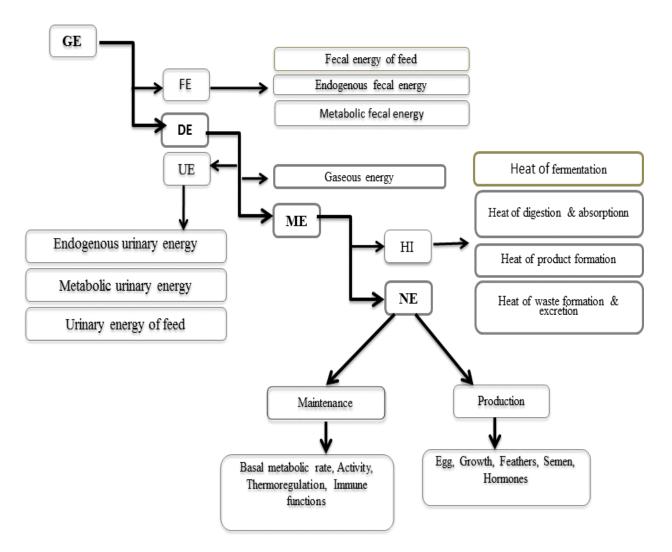
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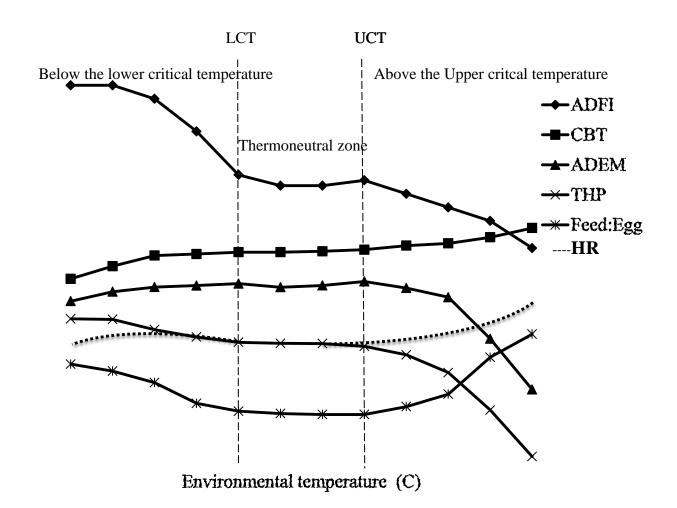
895

898

Yu, M. W., F. E. Robinson, R. G. Charles, and R. Weingardt. 1992. Effect of feed allowance
 during rearing and breeding on female broiler breeders. 2. Ovarian morphology and
 production. Poult. Sci. 71:1750-1761.



- 906 Figure 1.1 Partition of feed energy in animals
- Adapted from Leeson and Summers (2001) and Teeter et al. (2005).
- 908 GE=Gross Energy; FE=Fecal Energy; DE=Digestible Energy; UE=Urinary Energy;
- 909 ME=Metabolizable Energy; HI=Heat Increment; NE=Net Energy.



911 Figure 1.2 Theoretical effects of environmental temperature on core body temperature (CBT),

912 average daily feed intake (ADFI), total heat production (THP) , heat retention (HR), average

913 daily egg mass output (ADEM) and efficiency in broiler breeder hens.

In the thermoneutral zone, heat production and heal loss are balanced. ADFI= average daily

915 feed intake; CBT= core body temperature; ADEM= average daily egg mass production;

916 THP= total heat production; HR= heat retention; Feed: egg (g/g) = feed to egg conversion

917 ratio; LCT= lower critical temperature; UCT= upper critical temperature.

919 CHAPTER 2: EFFECTS OF ENVIRONMENTAL TEMPERATURE ON

920 PERFORMANCE AND CORE BODY TEMPERATURE IN BROILER BREEDER 921 PULLETS

922 ABSTRACT: An experiment was conducted to investigate the effects of environmental 923 temperature on average daily feed intake, average daily gain, feed efficiency, and core body temperature (CBT) dynamics in 800 feed restricted Ross 708 broiler breeder pullets from 4 to 924 20 wk of age. Pullets were housed in 8 climate-controlled environmental chambers, each 925 containing 4 pens. Standard breeder BW targets were achieved using feed restriction (daily) 926 927 from 14 d of age. Every 14 d, temperature treatments (15, 19, 23, or 27°C) were randomly reassigned to the chambers. Telemetric devices that transmitted CBT data every 10 min were 928 surgically implanted in the right abdominal cavity of 64 pullets (n=2/pen) at 13 wk of age. 929 930 Average daily feed intake was the highest in the 15°C treatment followed in decreasing order by the 19, 27, and 23°C treatments. Average daily gain was 0.91 g higher in the 27°C than in 931 the 15°C treatment. Feed, ME and CP conversion rates were higher in the 15°C than in the 23 932 and 27°C groups. Average CBT was lower in the 19°C treatment group than in the 23 and 933 27°C treatment groups. Feed restricted pullets had a clear diurnal CBT pattern, with a range 934 935 of approximately 2.47°C regardless of environmental temperature. The birds maintained a steady low CBT of 40.27°C at night, the CBT increased gradually from 1 h pre-dawn, pre-936 feed and reached a peak of 41.90°C at 0-1 h post-feed. After a postprandial CBT peak, CBT 937 938 decreased gradually to 1 h pre-dusk. Pullet CBT quickly dropped in the hour after the lights were turned off. Birds maintained a relatively constant CBT in different environmental 939 temperatures. In colder environments, the difference between body temperature and the 940 941 environmental temperature was greater and birds lost more heat to the environment. Therefore, they needed more energy to maintain homeostasis. Accurate broiler breeder pullet 942 feed allocation decisions need to accommodate effects of environmental temperature on 943

maintenance requirement because the expected growth rate can be affected by changes inenvironmental temperature.

946 **Keywords**: Environmental temperature, core body temperature, feed allocation, efficiency.

947 2.1. INTRODUCTION

Feed restriction is a common practice used with broiler breeders to achieve the target 948 BW and maximize reproductive efficiency (Robinson and Wilson, 1996). The target BW, 949 feed allocation and environmental temperature are crucial factors in the performance of 950 broiler breeders. A target BW is recommended for each strain by the breeding companies and 951 952 established based on field results and supported by experimental observations (Renema et al., 2007). Feed allocation is a key factor for controlling intake over time to achieve target BW in 953 954 feed-restricted broiler breeders. The relationship between BW and reproduction is a very 955 complex one in feed restricted broiler breeder females (Bornstein et al., 1984; Hocking and 956 Robertson, 2000; Joseph et al., 2002; de Beer and Coon, 2007). Excess energy consumption makes broiler breeders obese (Renema et al., 1994; Romero et al., 2009). In particular, 957 researchers have noted a negative relationship between growth and reproductive performance 958 in broiler breeder hens (Maloney et al., 1967; Jaap and Muir, 1968; Yu et al., 1992). Hocking 959 (2004) reported that the maintaining a target BW led to a uniformity in the birds for the age at 960 onset of lay and egg weight. However, broiler breeder obesity leads to decreased production, 961 962 fertility, hatchability, liveability and feed efficiency (Wilson and Harms, 1984), and erratic 963 ovulations, abnormal eggs and irregular oviposition times (Jaap and Muir, 1968; Yu et al., 1992). Indeed, Hocking (2004) reported that severely feed restricted (25% of ad libitum) 964 breeders produced underweight conditions relative to their recommended BW targets, which 965 966 resulted in delayed sexual maturity, poor egg production and low egg weight. So achieving the target BW in broiler breeder females through accurate feed allocation is of primary 967 968 concern in optimising reproductive efficiency.

969 Feed allocation should decide depend upon the environmental temperature. Temperature is one of the major environmental factors that influences the efficiency of 970 energy use by poultry (National Research Council, 1981). The energy requirement for 971 972 maintenance in broiler breeder pullets is about 80% of total ME intake (Sakomura, 2004). The CBT in feed restricted broiler breeder hens ranged from 39.0 to 41.2°C (Savory et al., 973 2006; de Jong et al., 2002). In cold environments, the difference between the surface body 974 temperature and environmental temperature increases, birds dissipate more body heat to the 975 environment, and they use higher proportion of ME intake to maintain core body 976 temperature. Since the first priority for nutrient utilization is maintenance (Reves et al., 977 2011), the amounts of nutrients available for growth diminish at lower environmental 978 979 temperatures. In other words, cold thermogenesis increases maintenance energy 980 requirements, as an animal must increase its feed intake and rate of metabolic heat production in order to maintain CBT (Green et al., 2009). This may result in a reduced BW gain because 981 feed-restricted broiler breeders cannot voluntary feed intake. Conversely, birds show 982 983 symptoms of heat stress above the upper critical temperature (above thermoneutral temperature; approximately 30°C); when the difference between the surface body temperature 984 and environmental temperature decreases (< 10°C). Under this condition, birds are unable to 985 dissipate adequate heat to the environment resulting in a rise of CBT. They may reduce the 986 rate of metabolic heat production by eating less, resulting in a decrease of nutrient intake 987 988 (National Research Council, 1981). Thus, poultry BW gain and egg production efficiency is adversely affected by high environmental temperature. The relationship between 989 environmental temperature and CBT affects the amount of energy used for thermoregulation 990 991 (Geraert et al., 1996). The energy requirement for maintenance of birds changes under high or low environmental temperature such that birds require more feed in cold environmental 992 temperature than hot environmental temperature. In addition, Howlider and Rose (1987) 993

994 reported that heat stressed broiler chickens became lighter but fatter with an increase in total 995 and abdominal fat by 0.8% and 1.6% respectively with each degree of environmental 996 temperature over 21°C. As broiler breeders' CBT rise, feed consumption, growth rate, feed 997 efficiency, and survivability decrease (Wilson and Harms, 1984). Therefore, environmental 998 temperature is a critical factor for chicken rearing in terms of maintaining CBT, body weight 999 gain, feed intake, and reproductive efficiency.

1000 The present study was carried out to explore the effects of different environmental 1001 temperatures on CBT dynamics, ADFI, ADG, and feed efficiency in feed restricted broiler 1002 breeder pullets. This information can be useful for understanding maintenance energy 1003 requirements, and is important for feed allocation decisions for broiler breeder pullets in 1004 varying environmental temperatures.

1005 **2.2. MATERIALS AND METHODS**

1006 **2.2.1. Animal Care Approval**

1007 This study was carried out in compliance with the guideline of the Care and Use of 1008 Experimental Animals (Canadian Council on Animal Care, 1993) and was approved by the 1009 Animal Care and Use Committee for Livestock at the University of Alberta.

1010 2.2.2. Experimental Design

A completely randomized design was used to evaluate the effects of environmental temperatures on ADFI, ADG, feed efficiency and core body temperature dynamics in broiler breeder pullets. Each of four temperature treatments (15, 19, 23, and 27°C) was assigned to 2 environmentally controlled chambers for 8 consecutive 14 d period from 4 to 20 wk of age. The temperature treatments were randomly preassigned using the PLAN procedure of SAS.

1016 2.2.3. Stocks and Management

1017 A total of 800 Ross 708 (Aviagen Inc., Huntsville, AL) 1 d old broiler breeder pullets 1018 were individually identified by bar-coded neck tags (Heartland Animal Health, Fair Play,

1019 MO) and placed randomly into eight climate controlled chambers. Each chamber was divided into four floor pens with 25 chicks per pen (5 chicks/ m^2) under the recommended brooding 1020 temperature (Aviagen, 2007). At 7 d of age, individual chicks were identified using bar coded 1021 1022 wing bands with the same digits in both wings. Feed was provided *ad libitum* for the first 14 d. At 15 d of age, daily feed restriction was imposed to maintain the target BW according to 1023 1024 the primary breeder's guidelines (Aviagen, 2007). Group feed allocation decisions were made twice per week, based on the mean BW of each pen. Pullets within each pen received the 1025 same amount of feed on any given day based on different environmental temperature 1026 1027 treatments. Environmental temperature treatments (15, 19, 23, and 27°C) were randomly assigned every 14 d period after 28 d of age. Each temperature treatment was replicated twice 1028 1029 in each of 8 time periods. Pullets were reared in the floor pens until 20 wk of age. The 1030 photoperiod was 23L:1D for the first 3 d and 8L:16D from 4 d to 20 wk of age. They were 1031 fed wheat- and soybean-based mash diets (Appendix A): Starter (2,900 kcal ME, 19% CP) from 0 to 2 wk and Grower (2,700 kcal ME, 15.0% CP) from 3 to 20 wk of age. 1032

1033 2.2.4. Surgical Implantation of Temperature Sensors

1034 Surgical procedure for implanting telemetry sensor in the abdominal cavity of 1035 chickens was performed according to Nain (2011). A total of 64, 13-wk old pullets were randomly selected (two pullets from each of 32 pens) for surgical implantation of CBT 1036 1037 sensors (ATS, Inc., Isanti, MN) into the right side of the abdomen. The CBT sensors had an 1038 accuracy rating of ± 0.1 °C, and were calibrated using a water bath procedure. Prior to surgery, the pullets were moved to individual cages and were fasted overnight. Each pullet 1039 was anaesthetized with 0.75% isoflurane at a rate of 1.5 L/min through an inhalation mask 1040 1041 just before surgery. The right ventral abdominal area was plucked and cleaned for surgery with Hibitane TM antiseptic (Chlorhexidine 2%, Ayerst Veterinary Laboratories). The CBT 1042 implants were placed in the right abdominal cavity after a 3 cm incision was made in the right 1043

1044 ventral abdominal flank. After implantation, the muscle layers and skin were sutured using synthetic absorbable material (3/0 Polydioxone Suture). A long term acting analgesic 1045 (meloxicam: 0.1 mg/kg; Metacam, Boehringer Ingelheim) and a short term acting analgesic 1046 1047 (buprenorphine: 0.01 mg/kg: Buprenex®, Norwich Eaton) were injected subcutaneously to prevent pain and discomfort during and after surgery. A broad spectrum antibiotic 1048 (Ampicillin: 50 mg/kg) was administered intramuscularly to prevent infection. Birds were 1049 returned to their respective cages after surgery and feed and water was provided ad libitum. 1050 1051 Each bird was observed for 15 to 20 minutes to ensure that they were able to stand up and 1052 had normal appetite. Pullets were returned to their original pens 24 h after surgery.

1053 2.2.5. Data Collection

1054 Average daily feed intake and average daily BW gain, based on twice per week group 1055 weights, was recorded throughout the experiment. Abdominally-implanted temperature 1056 sensors transmitted the CBT of each bird every 10 min from 13 to 20 wk of age. The following terminologies were used: Maximum CBT was the average of the highest daily 1057 1058 body temperature of individual chickens. Mean CBT was the daily average of all recorded body temperatures of individual chickens. Minimum CBT was the average daily lowest body 1059 1060 temperature of individual chickens. Range of CBT was the average of the difference between the daily maximum and minimum body temperatures of individual chickens. Night was 1061 1062 defined as the time when lights were turned off (15:00), excluding 1 h post-dusk and 1 h pre-1063 dawn; 1 h pre-dawn was the one hour time period before lights were turned on (07:00); prefeed was the time period from lights on to feeding time; 0-1 h post-feed was the first hour 1064 after feeding; 1-2 h post-feed was the second hour after feeding; 2-3 h post feed was the third 1065 1066 hour after feeding; 3-4 h post-feed was the fourth hour after feeding; >4 h post-feed was the remainder of time that the lights were on following the fourth hour post-feed, excluding pre-1067

1068 dusk; 1 h pre-dusk was the one hour time period before lights were turned off; 1 h post-dusk1069 was the one hour time period after lights were turned off.

1070 **2.2.6. Statistical Analysis**

Data were analyzed using the Mixed procedure of SAS 9.2 (SAS Institute, Cary, NC) and the treatment means were differentiated using Tukey's test. Environmental temperature treatments were considered fixed effects. The individual pen was the experimental unit for feed intake, body weight, and efficiency data. Correlated repeated measures of CBT were accounted for using age as a random effect with hen as a subject. Unless otherwise noted, differences between means were considered significant at $P \le 0.05$.

1077 2.3. RESULTS AND DISCUSSION

1078 2.3.1. Feed intake and BW gain

1079 An increase in environmental temperature led to a stepwise decrease in ADFI from 15 to 23°C treatment groups (Table 2.1). The target body weight was similar among the 1080 1081 treatments throughout experimental period. However, the ADFI of the 27°C treatment was not different from the 23 and 19°C treatments. Theoretically, the rate at which birds 1082 1083 dissipated heat to the environment was directly proportional to the difference between the 1084 surface body temperature and environmental temperature. Therefore, at cold temperatures 1085 birds require more feed to maintain body temperature (National Research Council. 1981). When the environmental temperature drops below the lower critical temperature; the 1086 1087 temperature difference between the surface body and the environment increases, birds dissipate more heat to the environment and they eat more to increase the rate of metabolic 1088 heat production for maintaining a relatively constant CBT. When the environmental 1089 temperature is above the upper critical temperature, the temperature difference between the 1090 surface body and the environment decreases, birds are unable to dissipate adequate heat to the 1091 1092 environment and they try to dissipate heat through panting (evaporative cooling) in the

1093 primary phase of the upper critical temperature (National Research Council, 1981). In both 1094 scenarios, birds expend more energy to maintain their CBT. The ADFI was lower in the 23°C treatment compared to the 15 and 19°C treatments. This might be due to less body heat being 1095 1096 lost to the environment, therefore requiring a lower metabolic rate of heat production. Conversely, ADFI was highest in the 15°C treatment group; probably because birds lost more 1097 heat to the environment and they required more energy for maintaining CBT. In hot 1098 environments, birds spend energy for cooling themselves, through activities such as panting 1099 1100 (Brody, 1945; Mack et al., 2013). This is consistent with the observation in the current study that pullets in the 27°C group did not eat less feed than birds in the 23°C treatment group. In 1101 the current study, ADG was higher in the 27°C treatment group (P = 0.06) compared to the 1102 1103 15°C treatment group. The ADG was similar in the 19, 23, and 27°C treatment groups and in 1104 the 15, 19, and 23°C treatment groups. By design, average daily gains were expected to be the 1105 same in each treatment. This observation is likely the result of birds growing more quickly and efficiently than expected at higher environmental temperatures. Feed, CP and ME 1106 1107 conversion rates were higher in the 15°C treatment compared to the 23 and 27°C treatments. This is consistent with the hypothesis that birds in the 15°C treatment lost more energy as 1108 1109 heat to the colder environment.

1110 **2.3.2.** Core body temperature dynamics

Average daily maximum CBT was higher by approximately 0.12°C in the 23 and 27°C environmental temperature treatments than in the 15 and 19°C treatments (Table 2.2). The mean CBT was similar when environmental temperature increased from 15 to 19°C. The CBT increased by 0.03°C with each subsequent environmental temperature increase. The mean CBT was significantly lower in the 19°C, compared to the 23 and 27°C treatment groups. Kadono and Besch (1978) recorded similar mean CBT in full-fed chickens under environmental temperatures within the range of 23 to 32°C. However, the birds were feed 1118 restricted in the current study, and therefore would have less of a continuous source of heat available from the processes associated with feed intake and digestion. Thus feed restricted 1119 birds must be more active in the physiological regulation of CBT. In the current study, 1120 1121 average daily minimum CBT increased with each subsequent increase in environmental temperature from 15 to 27°C treatment group, indicating that the minimum CBT may be the 1122 most sensitive to environmental influence. Being homoeothermic, birds may be conserving 1123 energy by maintaining a slightly lower CBT in cooler environment (15 to 19°C), and slightly 1124 higher CBT in warmer environment (23 to 27°C). This was in agreement with Zuidhof et al., 1125 1126 (2012). This response would reduce heat loss to the environment by reducing the temperature differential between body and environment. Although we did not measure surface 1127 1128 temperature, it would seem reasonable that in an attempt to thermoregulate (control the rate 1129 of heat loss to the environment) the difference in bird skin temperatures between treatments would have been greater than the 0.03°C difference observed in CBT. For example, Richards 1130 (1971) reported shank skin temperatures of 34.1, 39.2 and 40.2°C in 20, 30 and 40°C 1131 1132 environmental temperature, respectively. A similar range of CBT (2.47°C) was observed in all temperature treatment groups. Energy efficiency could be reduced in cold temperatures 1133 because of increased heat loss to the environment. 1134

1135 2.3.3. Diurnal CBT Rhythms

In all treatments, the CBT followed a clear diurnal rhythm (Figure 2.1; Table 2.3). The CBT started increasing before dawn, peaked at feeding time (0-1 h post-feed), and then slowly declined during the remainder of the day. There was a rapid post-dusk drop in CBT (1 h after lights were turned off), and CBT remained relatively low during the night (Table. 2.3; Figure. 2.1). The CBT was likely the highest after feeding because of the combined effects of the feeding activity of birds and feed metabolism (Kadono et al., 1981; Khalil et al., 2004). The CBT increased about by 0.61°C 1 h pre-dawn (one hour before lights on), probably in

1143 relation to changes from a state of sleep to wakefulness and activity. This may be associated with heat generated by muscle movement during activity (Khalil et al., 2004). In the pre-feed 1144 period (lights on to feeding time), CBT increased about by 0.67°C compared to 1 h pre-dawn, 1145 1146 which might be due to continued general increases in activity, as well as sudden visual due to lights on and auditory stimuli (Richards, 1971) such as the sound of a door opening. Other 1147 factors might have influenced CBT 1 h pre-dawn and pre-feed, including habituation to the 1148 lights coming on, and to feeding which was done fairly consistently about an hour after the 1149 lights were turned on. Just after feeding (0-1 h), CBT peaked. This was probably due to a 1150 1151 combination of diet-induced thermogenesis and increased feeding-related activity. After feeding (1-2 h), the gradual decline in CBT reflected a rate of metabolic heat production that 1152 1153 presumably decreased gradually after feeding. However, during the day time (from pre-feed 1154 to 1 h post-dusk), the CBT was higher than at night, likely due to a higher activity level. 1155 Khalil et al. (2004) reported activities such as body shaking, litter pecking, beak wiping, head scratching and preening during the day. In the 1 h pre-dusk period (one hour before lights 1156 off), CBT decreased by 0.02°C from daytime CBT. In the 1 h post-dusk period, CBT dropped 1157 by about 0.76°C. This was likely because of drastically reduced activity levels, resting and 1158 sleeping (Blokhuis, 1984; Khalil et al., 2004). This was in agreement with de Jong et al. 1159 (2002), who reported that higher activity level increased CBT in the day time compared to 1160 1161 night time. Similarly, Kadono and Besch (1978) reported that for full-fed broiler chickens, 1162 CBT started to increase 2 to 4 h before lights were turned on and decreased 2-4 h before lights were turned off. The CBT was higher in the day time and lower at night time in all 1163 treatment groups. These results were in agreement with those of others (Fronda, 1921; 1164 1165 Heywang, 1938; Winget et al. 1965). In the current study, a strong diurnal rhythm was observed in CBT of broiler breeder pullets, responding to feeding activities and photoperiod. 1166 Environmental temperature within the range from 15 to 27°C did not dramatically affect 1167

diurnal CBT rhythm in feed restricted broiler breeder pullets. There is some evidence, however, from slight reductions in CBT in cooler environments that broiler breeder pullets may be actively conserving energy to dissipate less heat to the environment by reducing the temperature difference between the body surface and the environment.

1172

2.3.4. Economic evaluation of pullets feed

Assuming a feed cost of \$380/T, the feed cost increased by \$0.02, \$0.09 and \$0.22 per pullet from 4 to 20 wk of age in the 23, 19 and 15°C treatment respectively relative to the 27°C treatment. The average feed intake was increased by decreasing environmental temperature from 27 to 15°C.

Colder environmental temperatures (15°C) decreased feed efficiency, CP and ME 1177 1178 utilization efficiency, and increased feed intake considerably compared to warmer 1179 environmental temperatures (27°C) in feed restricted broiler breeder pullets. Though there were subtle differences, CBT was maintained relatively constant across environmental 1180 temperatures, suggesting that heat loss from broiler breeder pullets, and therefore energy 1181 1182 requirements for maintenance, were higher at low environmental temperatures. Zuidhof et al. (2012) suggested that feed allocations for target growth rates would be expected to be about 1183 5% lower at environmental temperatures of 27°C compared to 15°C. To achieve a uniform 1184 rate of growth in broiler breeder pullets, feed allocation decisions should consider 1185 environmental temperature because the energy requirement for maintenance increases with 1186 1187 decreasing temperature.

2.4. REFERENCES

1190	Aviagen. 2007. Ross 708 parent stocks performance objectives. Aviagen Ltd.
1191	www.aviagen.com/docs/Ross 708 PS Performance Objectives.pdf. Accessed June 2010.
1192	
1193	Blokhuis, H. J. 1984. Rest in poultry. Appl. Anim. Behav. Sci. 12:289-303.
1194	
1195	Bornstein, S., I. Plavnik, and Y. Lev. 1984. Body weight and/or fatness as potential determinants
1196	of the onset of egg production in broiler breeder hens. Br. Poult. Sci. 25:323-341.
1197	
1198	Brody, S. 1945. Bioenergetics and Growth: with special reference to the efficiency complex in
1199	domestic animals. New York: Reinhold Publishing Co.
1200	
1201	Canadian Council on Animal Care. 1993. Guide to the use of experimental animals. Vol. 1. Can.
1202	Counc. Anim. Care, Ottawa, Ontario, Canada.
1203	
1204	de Beer, M., and C. N. Coon. 2007. The effect of different feed restriction programs on
1205	reproductive performance, efficiency, frame size, and uniformity in broiler breeder hens.
1206	Poult. Sci. 86:1927-1939.
1207	
1208	de Jong, I. C., Van Voorst, S., A. D. Ehlhardt, and H. J. Blokhuis. 2002. Effects of restricted
1209	feeding on physiological stress parameters in growing broiler breeders. Br. Poult. Sci.
1210	43:157-168.
1211	
1212	Fronda, F. M. 1921. A comparative study of the body temperature of the different species and
1213	some representative breeds of poultry- A preliminary report. Poult. Sci. 1:16-22.
1214	
1215	Geraert, P. A., J. C. F. Padilha, and S. Guiliaumin. 1996. Metabolic and endocrine changes
1216	induced by chronic heat exposure in broiler chickens: growth performance, body
1217	composition and energy retention. Br. J. Nutr. 75:195-204.
1218	
1219	Green, J. A., L. G. Halsey, R. P. Wilson, and P. B. Frappell. 2009. Estimating energy
1220	expenditure of animals using the accelerometry technique: activity, inactivity and
1221	comparison with the heart-rate technique. J. Exp. Biol. 212:471-482.
1222	
1223	Heywang, B. W. 1938. Effects of some factors on the body temperature of hens. Poult. Sci.
1224	17:317-323.
1225	
1226	Hocking, P. M. 2004. Roles of body weight and feed intake in ovarian follicular dynamics in
1227	broiler breeders at the onset of lay and after a forced molt. Poult. Sci. 83:2044-2050.
1228	bronor breeders at the onset of hay and after a foreed more. Fourt. Set. 05.2011 2030.
1229	Hocking, P. M., and G. W. Robertson. 2000. Ovarian follicular dynamics in selected and control
1225	(relaxed selection) male- and female-lines of broiler breeders fed ad libitum or on
1230	restricted allocations of food. Br. Poult. Sci. 41:229-234.
1231	restricted anocations of 1000. D1. 1 001. 301. 41.227-234.
1232	

1233 1234	Howlider, M. A. R., and S. P. Rose. 1987. Temperature and growth of broilers. World's Poult. Sci. J. 43:228-237.
1235 1236 1237	Jaap, R. G., and F. V. Muir. 1968. Erratic oviposition and egg defects in broiler-type pullets. Poult. Sci. 47:417-423.
1238	
1239 1240 1241	Joseph, N. S., A. A. J. Dulaney, F. E. Robinson, R. A. Renema, and M. J. Zuidhof. 2002. The effects of age at photostimulation and dietary protein intake on reproductive efficiency in three strains of broiler breeders varying in breast yield. Poult. Sci. 81:597-607.
1242	
1243 1244	Kadono, H., and E. L. Besch. 1978. Telemetry measured body temperature of domestic fowl at various ambient temperatures. Poult. Sci. 57:1075-1080.
1245	
1246 1247	Kadono, H., E. L. Besch, and. E. Usami. 1981. Body temperature, oviposition, and food intake in the hen during continuous light. J. Appl. Physiol. 51:1145-1149.
1248	
1249	Khalil, A. M., K. Matsui, and K. Takeda. 2004. Diurnal and oviposition-related changes in heart
1250	rate, body temperature and locomotors activity of laying hens. Anim. Sci. J. 75:169-174.
1251	
1252	Mack, L.A., J.N. Felver-Gant, R. L. Dennis, H. W. Cheng. 2013. Genetic variation alters
1253	production and behavioral responses following heat stress in 2 strains of laying hens.
1254	Poult. Sci. 92:285-294.
1255	
1256	Maloney, M. A., J. C. Gilbreath, J. F. Tierce, and R. D. Morrison. 1967. Divergent selection
1257	twelve-week body weight in the domestic fowl. Poult. Sci. 46:1116-1127.
1258	
1259	Nain, S. 2011. Improving the effectiveness of laying hens for use in value-added egg production.
1260	M. Sc. Diss. Univ. Alberta, Edmonton.
1261	
1262	National Research Council. 1981. Effect of environment on nutrient requirements of domestic
1263	animals. Natl. Acad. Press, Washington D. C.
1264	
1265	Renema, R. A., M. E. Rustad, and F. E. Robinson. 2007. Implications of changes to commercial
1266	broiler and broiler breeders' body weight targets over the past 30 years. World's Poult.
1267	Sci. J. 63:457-467.
1268	
1269	Renema, R. A., F. E. Robinson, V. L. Melnychuk, R.T. Hardin, L.G. Bagley, D.A. Emmerson,
1270	and J.R. Blackman, 1994. The use of feed restriction in improving reproductive traits in
1271	male line large white turkey hens. 1. Growth and carcass characteristics. Poult. Sci.
1272	73:1724-1738.
1273	
1274	Reyes, M. E., C. Salas, and C. N. Coon. 2011. Energy requirements for maintenance and egg
1275	production of broiler breeder hens. Int. J. Poult. Sci. 10:913-920.
1276	

1277	Richards, S. A. 1971. The significance of changes in the temperature of the skin and body core
1278	of the chicken in the regulation of heat loss. J. Physiol. 216:1-10.
1279	
1280	Robinson, F. E., and J. L. Wilson. 1996. Reproductive failure in overweight male and female
1281	broiler breeders. Anim. Feed Sci. Tech. 58:143-150.
1282	
1283	Romero, L. F., M. J. Zuidhof, R. A. Renema, F. E. Robinson, and A. Naeima. 2009. Nonlinear
1284	mixed models to study metabolizable energy utilization in broiler breeder hens. Poult.
1285	Sci. 88:1310-1320.
1286	
1287	Sakomura, N. K. 2004. Modeling energy utilization in broiler breeders, layer hens and broilers.
1288	Braz. J. Poult. Sci. 6:1-11.
1289	SAS 2008 SAS 0.2 @ 2002 2008 by SAS Institute Inc. Comp. NC. USA
1290	SAS. 2008. SAS 9.2 © 2002-2008 by SAS Institute, Inc., Cary, NC, USA.
1291 1292	Savory, C. J., L. Kostal, and I. M. Nevison. 2006. Circadian variation in heart rate, blood
1292	pressure, body temperature and EEG of immature broiler breeder chickens in restricted-
1293	fed and ad libitum-fed states. Br. Poult. Sci. 47:599-606.
1294	red and ad nortalin-red states. Dr. 1 outl. Set. 47.577-000.
1295	Wilson, H. R, and R. H. Harms. 1984. Evaluation of nutrient specifications for broiler breeders.
1297	Poult. Sci. 63:1400-1406.
1298	
1299	Winget, C. M., E. G. Averkin, and T. B. Fryer. 1965. Quantitative measurement by telemetry of
1300	ovulation and oviposition in the fowl. Anim. J. Physiol. 209:853-858.
1301	1
1302	Yu, M. W., F. E. Robinson, and A. R. Robblee. 1992. Effect of feed allowance during rearing
1303	and breeding on female broiler breeders. 1. Growth and carcass characteristics. Poult. Sci.
1304	71:1739-1749.
1305	
1306	Zuidhof, M. J., D. C. Paul, A. Pishnamazi, I. I. Wenger, R. A. Renema, and V. L. Carney. 2012.
1307	Temperature and protein: energy ratio linkages between breeder and broiler energetics,
1308	performance, and carcass quality. Final Report to Alberta Livestock and Meat Agency:
1309	Project #2008F138R. February 5.

Table 2.1 Average daily feed intake, average daily gain, and feed conversion rates of broilerbreeder pullets (4 to 20 wk) in different environmental temperature treatments.

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-	-	-	~

	l Temperature (°C	·	ADG	Feed:Gain	ME:Gain	CP:Gain
Set	Actual \pm SD	(g/d)	(g/d)	(g/g)	(kcal/g)	(g/g)
15	17.3 <u>+</u> 2.2	52.70 ^a	13.28 ^b	3.99 ^a	11.15 ^a	0.69 ^a
19	20.4 ± 1.1	52.03 ^b	13.81 ^{ab}	3.77 ^{ab}	10.52^{ab}	0.64 ^{ab}
23	23.9 <u>+</u> 1.0	51.14 ^c	13.92 ^{ab}	3.65 ^b	10.20^{b}	0.62 ^b
27	26.7 <u>+</u> 2.2	51.51 ^{bc}	14.19 ^a	3.61 ^b	10.09 ^b	0.61 ^b
SEM	-	0.17	0.25	0.08	0.23	0.01
Probability	-	0.0001	0.0626	0.0048	0.0048	0.0048

1313 ^{a-c}Means within column with no common superscript are significantly different ($P \le 0.05$)

1314 Set environmental temperature= designated temperature to each chamber through computer programing

1315 Actual environmental temperature = actual chamber temperature was recorded by data loggers

Table 2.2. Average daily maximum, mean, minimum, and range of core body temperature (CBT)of broiler breeder pullets (13 to 20 wk) in different environmental temperature treatments.

1319

Environme	ental Temperature		M					
Set	Actual \pm SD	Maximum	Mean	Minimum	Range			
	(°C)		CBT (°C)					
15	17.3 <u>+</u> 2.2	42.36 ^b	40.75 ^{bc}	39.87 ^d	2.48			
19	20.4 <u>+</u> 1.1	42.39 ^b	40.74 ^c	39.94 ^c	2.44			
23	23.9 <u>+</u> 1.0	42.47 ^a	40.77 ^{ab}	39.99 ^b	2.48			
27	26.7 <u>+</u> 2.2	42.52 ^a	40.80 ^a	40.03 ^a	2.49			
SEM	-	0.029	0.023	0.029	0.032			
Probability	-	< 0.0001	< 0.0001	< 0.0001	0.2616			

1320 ^{a-d}Means within column with no common superscript are significant different ($P \le 0.05$)

1321 Set environmental temperature = designated temperature to each chamber through computer

1322 Actual environmental temperature= actual chamber temperature was recorded by data loggers

1323 Maximum = average of daily highest CBT of individual hens during the study period

1324 Mean = average of daily mean CBT of individual hens during the study period

1325 Minimum = average of daily lowest CBT of individual hens during the study period

1326 Range = average of difference between daily highest and lowest CBT of individual hens during the study

1327 period

1328	Table 2.3. Core body temperatures of broiler breeder pullets (13 to 20 wk) at different times of
1329	day under different environmental temperature treatments.

1330

Time of day ¹	Environmental temperature (°C)				
	15	19	23	27	overall
	CBT(°C)				
Night	40.24 ^{hC}	40.21 ^{hD}	40.28^{jB}	40.37 ^{jA}	40.27 ^j
1 h pre-dawn	40.87 ^f	40.90 ^f	40.88 ^h	40.89 ^h	40.88 ^h
Pre-feed	41.49 ^{dD}	41.54 ^{cC}	41.57 ^{cB}	41.60 ^{cA}	41.55 ^c
0-1 h post-feed	41.88 ^{aBC}	41.86 ^{aC}	41.89 ^{aB}	41.98 ^{aA}	41.90 ^a
1-2 h post feed	41.64 ^{bAB}	41.61 ^{bB}	41.61 ^{bB}	41.65 ^{bA}	41.63 ^b
2-3 h post feed	41.54 ^{cAB}	41.53 ^{cAB}	41.52 ^{dB}	41.57 ^{dA}	41.54 ^d
3-4 h post feed	41.48 ^d	41.46 ^d	41.46 ^e	41.49 ^e	41.47 ^e
>4 h post-feed	41.44 ^{eA}	41.42 ^{eAB}	41.40 ^{fB}	41.43 ^{fA}	41.42^{f}
1 h pre-dusk	41.42 ^{eA}	41.40 ^{eAB}	41.37 ^{gC}	41.39 ^{gBC}	41.40 ^g
1 h post-dusk	40.63 ^{gB}	40.65 ^{gAB}	40.63 ^{iB}	40.67 ^{iA}	40.64 ⁱ
SEM	0.006	0.006	0.006	0.006	0.005
Source of variation	Probability				
Temperature	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001
Diurnal	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001
Diurnal*Temperature	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001

1331 ^{A-D}Means within row with no common superscript are significantly different ($P \le 0.05$)

1332 ^{a-j}Means within column with no common superscript are significantly different ($P \le 0.05$)

¹Night= lights off time period (16:00 to 06:00; excluding 1 h post-dusk and 1 h pre-dawn); 1 h pre-dawn=
one hour time period before lights on (at 07:00); Pre-feed= time period from lights on to feeding time; 0-1
h post-feed= first hour after feed; 1-2 h post-feed= second hour after feed; 2-3 h post feed= third hour
after feed; 3-4 h post-feed= fourth hour after feed; >4 h post-feed= remainder of the lights on period
following the fourth hour post-feed, excluding pre-dusk; 1 h pre-dusk= one hour time period before lights
off; 1 h post-dusk= one hour time period after lights off (at 15:00).

1339 Note: Normally, feeds were supplied to the birds at 07:30 and it little delayed at weighing days.

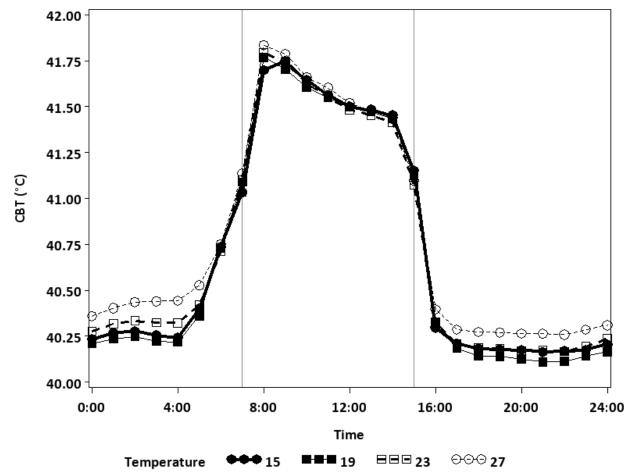


Figure 2.1 Diurnal core body temperature (CBT) pattern of broiler breeder pullets (13 to 20 wk)
in 15, 19, 23 and 27°C environmental temperature treatment. Vertical reference lines in the graph
indicate lights on at 07:00 and lights off at 15:00

1348 CHAPTER 3: IMPACT OF ENVIRONMENTAL TEMPERATURE AND DIETARY

1349 ENERGY ON CORE BODY TEMPERATURE AND REPRODUCTIVE

1350 PERFORMANCE OF BROILER BREEDER HENS

ABSTRACT: An experiment was conducted using 192 Ross 708 broiler breeder females from 1351 25 to 41 wk of age to evaluate the effects of environmental temperatures and dietary ME levels 1352 1353 on core body temperature (CBT) and reproductive performance. At 22 wk of age, pullets were 1354 randomly distributed among six climate-controlled environmental chambers (n=32/chamber) 1355 including 48 temperature sensor-implanted birds to measure the CBT. The experiment was a 4 x 1356 2 factorial arrangement, with four environmental temperatures (15, 19, 23, and 27°C) and two 1357 dietary ME levels (high: 2,912; or low: 2,786 kcal/kg). For every 2 wk period, environmental temperatures were randomly reassigned to the chambers with the constraint that each treatment 1358 1359 was represented at least once per period and each chamber had equal exposure to each temperature treatment by the end of the experiment. The CBT dynamics, ADFI, ADG, feed 1360 1361 efficiency, egg production, and egg weight were determined. The highest maximum, mean and 1362 minimum CBT occurred in the 27°C treatment, followed by the 23°C treatment, and then the 19 and 15°C treatments, which did not differ. Dietary energy levels had a little effect on maximum, 1363 minimum, or mean CBT. A higher CBT range (1.88°C) was recorded in the 15 and 19°C than in 1364 the 23 and 27°C (1.83°C) treatments. The CBT increased gradually from 1 h pre-dawn and 1365 peaked at 1 h after feeding. After peaking, CBT decreased gradually for the rest of the day. The 1366 1367 CBT quickly dropped in anticipation of the lights turning off and was lowest during the night. Average daily feed intake was higher at 15°C than at 19 or 23°C. Feed intake was 6% higher in 1368 the low energy treatment than in the high energy treatment, resulting in a higher intake of CP, 1369 1370 which contributed to higher egg weight in the low energy treatment. Efficiency (Feed, ME or

1371 CP) was lower in the low energy diet than in the high energy diet. Egg production was not 1372 affected by temperature or dietary energy treatments. In feed restricted broiler breeder hens, the 1373 feed allocation needs to increase with decreasing environmental temperature to maintain CBT 1374 and the target BW for maximizing reproductive output.

1375 KEYWORDS: Environmental temperature, core body temperature, dietary energy, performance.

1376 **3.1. INTRODUCTION**

Accurate feed allocation for broiler breeders to achieve target BW is a key to maximize reproductive output (Yu et al., 1992; Renema et al., 2007). Excess feed intake makes breeders obese resulting in multiple ovulations, deformed eggs, decreased egg production and increased irregular oviposition (Jaap and Muir, 1968; Renema and Robinson, 2004). Conversely, severe feed restriction made birds underweight, delayed sexual maturity and also decreased egg production (Hocking, 2004). Excessive or insufficient feed allocation results in sub-optimal production in broiler breeder hens.

Excess dietary protein must be deaminated prior to being used as an energy source and 1384 1385 therefore the heat increment from dietary protein is greater compared to dietary fat or carbohydrates (Musharaf and Latshaw, 1999). Excess dietary protein may increase CBT and 1386 1387 birds are unable to dissipate required heat to a hot environment. As a result, birds reduce feed intakes in warm environments (Mack et al., 2013), which is detrimental for production and 1388 product quality. Reducing the protein content in broiler diets may reduce heat production that 1389 1390 helps to reduce heat stress-related mortality and increase performance, especially in hot environmental conditions (Furlan et al., 2004). However, Spratt and Leeson (1987) reported that 1391 1392 high dietary protein contributed to increased egg production, egg weight, hatchability, and 1393 offspring performance in broiler breeders.

1394 Homoeothermic animals maintain a relatively constant core body temperature (CBT) over a wide range of thermoneutral temperatures (Deeb and Cahaner, 1999; Lacey et al., 2000a). 1395 Birds' ability to maintain their CBT depends on the difference between the internally produced 1396 1397 heat and the rate of heat dissipation to the environment. Romero et al. (2009) reported that in broiler breeder hens about 70% of ME intake was used for maintenance. The energy requirement 1398 1399 for maintenance in birds increases with decreasing environmental temperature below the critical temperature approximately less than 16°C because birds lose energy in the form of heat to the 1400 environment; they need more feed to increase the rate of metabolic heat production to maintain 1401 1402 their CBT (National Research Council, 1981) and this increased feed intakes decrease efficiency. 1403 In addition, in colder weather, concentrations of Thyroxin (T_3) and active forms of uncoupling proteins (UCPs) in the blood increased, which increased heat production resulting in increased 1404 1405 CBT in homoeothermic animals (Collin et al., 2005). When environmental temperature is above the upper critical temperature, the difference between the surface body temperature and the 1406 environmental temperature decreases, and birds are unable to dissipate adequate heat to the 1407 1408 environment (National Research Council, 1981). As a result, CBT may increase. At the primary stage of the upper critical temperature, birds try to dissipate heat to the environment though 1409 1410 evaporative cooling (Mack et al., 2013) to maintain a relatively constant CBT and thus they increase the energy requirement for maintenance. Wilson and Harms (1984) observed that such 1411 an increase in CBT reduced dependency on diet-induced metabolic heat production and birds 1412 1413 reduce their feed intake. The reduced feed intake resulted in a decrease of available nutrients, which causes poor production and product quality (National Research Council, 1981). 1414

1415 In broiler breeders, feed allocation decisions need to be readjusted based on 1416 environmental temperature specially, at above the upper and below the lower critical 1417 environmental temperature to maintain a relatively constant CBT for maximizing reproductive 1418 output. Birds need more energy for maintenance in a colder environment than in a thermoneutral zone because they lose more heat (energy) to the environment (Geraert et al., 1988). The 1419 1420 maintenance requirement is the first energy partitioning priority, which is to regulate the vital physiological functions including normal activities and thermoregulation (Reves et al., 2011). 1421 Thereafter, the remaining energy is used for growth and production. Pishnamazi et al. 1422 (unpublished data) found a quadratic relationship of environmental temperature on maintenance 1423 energy requirements in feed restricted broiler breeder hens: the ME requirement for maintenance 1424 1425 decreased with increasing temperatures from 15 to 24°C, and increased above 24°C. In broiler 1426 breeder hens, a large proportion of total ME intake is used for maintenance (more than 70%) and the remaining small proportion of ME intake is used for growth and production (Romero et al., 1427 1428 2009). The energy requirement for maintenance increases in colder environmental temperatures and that could seriously affect growth and production, especially when birds are feed restricted. 1429 Therefore, the feed allocation needs to be readjusted accurately with the change in environmental 1430 1431 temperature to maintain CBT, the target BW and reproduction.

1432 The objective of the current research was to investigate the effects of environmental 1433 temperatures and dietary energy on CBT, feed intake, egg production, egg weight, and efficiency 1434 of broiler breeder hens.

1435 **3.2. MATERIALS AND METHODS**

1436 **3.2.1. Animal Care Approval**

1437 The chickens in this research project were managed according to the Guide to the Care1438 and Use of Experimental Animals (Canadian Council on Animal Care, 1993). The experimental

protocol was approved by the Animal Care and Use Committee for Livestock of the Universityof Alberta.

1441 **3.2.2. Experimental Design**

1442 A 4 x 2 factorial experiment was conducted using four temperature treatments and two dietary energy levels (low: 2,786 kcal/kg; and high: 2,912 kcal/kg) in wheat and soybean based 1443 1444 diets (Appendix A). Both diets contained 15% CP and were provided in mash form from 21 to 41 wk. Four environmental temperature treatments (15, 19, 23, and 27°C) were randomly assigned 1445 to 6 chambers for consecutive 2 wk periods from 25 to 41 wk of age. Each treatment was 1446 1447 represented in at least one chamber per period, and over the entire experiment each chamber had equal exposure to all treatments. The individual hen was the experimental unit and age was used 1448 1449 as a random effect.

1450 **3.2.3. Stocks and Management**

A total of 192 Ross 708 (Aviagen Inc., Huntsville, AL) broiler breeders were selected 1451 randomly from a population of 800 birds, and housed in individual laying cages at 23 wk of age 1452 1453 in six climate-controlled environmental chambers. Feed allocations were provided to achieve the 1454 standard BW target recommended by the primary breeder (Aviagen, 2007). Individual hen BW, 1455 egg production, egg weights and CBT data were collected from 25 to 41 wk to investigate production performance of broiler breeder hens under different temperature and dietary 1456 treatments. The photoperiod was 12L:12D at 21 wk, and it was increased by one additional h/wk 1457 1458 to 16L:8D by 25 wk.

1459 **3.2.4 Core Body Temperature Sensors**

A total of 48 hens with a surgically implanted telemetric core body temperature sensor in its right abdomen were equally distributed among the 6 chambers, with 16 on each experimental diet. A detailed surgical procedure for sensor implantation was described in Chapter 2.

1463 **3.2.5. Data Collection**

1464 Body weight data were recorded twice per week throughout the experiment using an electronic balance (BW-1050, Weltech Agri Data, Charlotte, NC). Average BW, feed intake and 1465 1466 daily BW gain were recorded. Eggs were collected daily at 15:00, individually weighed and 1467 categorized as total, normal, abnormal or settable eggs. Abnormal eggs included membranous, soft shell, broken shells, and double yolked eggs. Total eggs was defined as all eggs including 1468 abnormal eggs; normal eggs included the total eggs minus abnormal eggs; and settable eggs were 1469 1470 normal egg minus eggs that weighed less than 52 g. The incidence of broken eggs was recorded and missing egg weight values were replaced by an estimate of egg weight on an individual hen 1471 basis by fitting a nonlinear regression of egg weight as a function of the hen age (wk) in the 1472 form: $EggWt = a \cdot be^{-c^*age}$ Where a = weight asymptote; b and c were estimated coefficients. Egg 1473 mass (EM) was defined on a hen basis as the sum of all eggs weights per 2 wk period divided by 1474 1475 the number of days in the period. Average egg weight was calculated per hen per period from 25 1476 to 41 wk.

Forty eight temperature-humidity loggers with a resolution of 0.1° C and an accuracy of $\pm 0.06^{\circ}$ C (Microlog EC650, Fourier Systems, New Albany, IN) were equally distributed and uniformly placed on birds level in the six chambers and were used to log environmental temperature every half hour. Core body temperature measurements were recorded every 10 min. The first day of lay was recorded as the age of sexual maturity. 1482 Average daily maximum CBT was the average of the highest daily body temperature of 1483 individual chickens. Daily mean CBT was the daily average of all recorded body temperatures of individual chickens. Average daily minimum CBT was the average daily lowest body 1484 1485 temperature of individual chickens. Average daily range of CBT was the average of the difference between the daily maximum and minimum body temperatures of an individual 1486 1487 chicken. The diurnal CBT pattern was classified as 1 h pre-dawn, pre-feed, 0-1 h post-feed, 1-2 h post-feed, 2-3 h post-feed, 3-4 h post-feed, >4 h post-feed, 1 h pre-dusk, 1 h post-dusk and night, 1488 and classified captions were described in detail in Chapter 2. 1489

1490 **3.2.6. Statistical Analysis**

The Mixed Procedure of SAS 9.2 (SAS Institute, Cary, NC) was used to analyze data and unless otherwise noted, differences between treatment means were at $P \le 0.05$. Each individually caged bird was an experimental unit. Treatments (environmental temperature and dietary energy level) were considered fixed effects, while age was a random effect. LS-mean separation was conducted using Tukey's test.

1496 **3.3. RESULTS AND DISCUSSION**

1497 **3.3.1.** Body weights, sexual maturity and egg production

The average BW, age at sexual maturity and age at 52 g eggs were similar between high and low energy diets (Table 3.1). This was in agreement with Bennett and Leeson (1990) who reported that high (3081 kcal/kg) or low (2550 kcal/kg) energy diets did not affect age at sexual maturity in broiler breeder hens. First egg weight was also similar between the diets. This was expected according to the experimental design since birds were maintained on the target BW recommended by the primary breeder (Aviagen, 2007). Egg production (total, normal and settable) was similar among the temperature treatments (Table 3.2). However, total egg 1505 production was higher in the interaction of high energy diet and high environmental temperature (27°C; Table 3.2) compared to any other interactions. Within the 27°C temperature treatment, 1506 birds fed high energy diet had higher total egg production. There was no difference in total egg 1507 production in the low energy diet. However, in the high energy diet, birds in the 27°C had greater 1508 1509 total egg production than the 23°C temperature treatment group. Normal egg production was 2.1 1510 eggs higher in the high energy treatment group than in the low energy treatment group. Egg production was expected to be similar because individual hens were not allowed to acclimatize to 1511 temperature treatments, since the treatments rotated every 14 d. This prevented any confounding 1512 1513 effects of body composition changes due to long-term exposure to specific temperatures, and is likely the reason we did not observe temperature treatment differences in egg production rates. 1514

1515 **3.3.2. Feed intake, body weight gain, egg weight and feed efficiency**

Feed intake was higher in the 15°C treatment group than those of 19 and 23°C treatment 1516 groups (Table 3.3). Presumably, birds dissipate more heat to lower environmental temperatures 1517 1518 because the difference between the surface body temperature and environmental temperature increases (National Research Council, 1981). In this case, birds need more feed to increase the 1519 rate of metabolic heat production to maintain their CBT. Environmental temperature ranging 1520 1521 from 15 to 27°C in broiler breeder hens had a quadratic effect on feed intakes (Pishnamazi et al., unpublished data). However, feed intake was similar among the 19, 23 and 27°C treatment 1522 1523 groups. It means total heat production could be similar as well as heat retention within the range 1524 of 19 to 27°C environmental temperature. Feed intake was also similar between the 15°C and 27°C temperature treatment groups. The ADG was higher in the 15 and 23°C than the 19 and 1525 1526 27°C treatment groups. This observation is consistent with the theory that birds in the 23°C 1527 treatment had dissipated less body heat to the environment compared to the 19°C treatment.

1528 At 15°C, presumably, birds raised feed intake to meet the maintenance requirement 1529 because they dissipate more heat to environment compared to other temperature treatments. De Andrade et al. (1977) reported that egg weight decreased by 8% when environmental 1530 1531 temperature increased from 21 to 31°C in full-fed laying hens due to decreased feed intake. In contrast, birds were not eating ad libitum in the current experiment. A precise feed allocation 1532 decision was made to maintain the target BW in broiler breeder hens within the range of 1533 environmental temperature from 15 to 27°C and that similar BW of hens could be the possible 1534 reason to produce similar egg weight. Egg production and egg weights were similar regardless of 1535 1536 temperature treatments. Environmental temperatures in broiler breeder hens within the range of 15 to 27°C had no effect on feed, ME and CP efficiency, and egg weight. 1537

Dietary energy levels were negatively correlated ($R^2 = -.69$) with feed intake, as well as 1538 the indices of efficiency: ME, CP, and feed per gram of egg weight. The average daily feed 1539 intake was 6% higher in the low energy diet than in the high energy diet group. This higher 1540 amount of feed intake did not affect daily BW gain but it did result in higher egg weight in low 1541 1542 energy birds. The low energy treatment decreased feed, ME and CP efficiency, and birds required a higher amount of feed (0.27 g), ME (0.26 kcal) and CP (0.03 g) for producing a one g 1543 of egg compared to the high energy treatment (Table 3.3). The low energy birds ate more. 1544 Presumably, they spent more energy to digest the higher amount of intake feed compared to high 1545 energy birds and the efficiency may decrease in the low energy birds. 1546

In the current study, average daily feed intake was 135 g and 143 g in high (2,912 ME kcal/kg) and low (2,786 ME kcal/kg) energy treatments, respectively, which was equivalent to 393 kcal ME and 22 g CP and 398 kcal ME and 23 g CP per day in the high and low energy treatments, respectively. Spratt and Leeson (1987) reported that 385 kcal ME and 19 g CP were required by individually-caged broiler breeder hens for maintaining normal egg production. Average egg weight (58.0 g) was higher in the low energy diet group compared to the egg weight (57.3 g) in the high energy diet group. Presumably, higher amount of CP intake contributes a heavier egg in low energy diet group than high energy diet group. This was in agreement with Spratt and Leeson (1987) who suggested that egg weight increased in broiler breeder hens due to higher amount of protein intake per day.

In the current study, the feed cost for a broiler breeder hen from 25 to 41 wk of age increased by \$0.14 per hen with decreasing environmental temperature from 27 to 15°C assuming a feed cost of \$380/T. However, feed costs did not decrease in the 23°C but decreased by \$0.04 per hen in the 19°C treatment based on feed to egg mass ratio relative to the 27°C treatment group.

1562 **3.3.3. Relationship between environmental temperature, dietary energy and CBT**

Birds maintained a minimum CBT (39.77°C) among treatments regardless of the 1563 environmental temperature to which they were exposed (Table 3.4). The daily average 1564 1565 maximum, minimum and mean CBT was similar among the 15 and 19°C treatments. The daily average maximum, minimum and mean CBT in broiler breeder hens increased significantly with 1566 1567 each subsequent increase in environmental temperature from 19 to 27°C (Table 3.4). Compared to the 19°C treatment, the maximum CBT increased at 23 and 27°C by 0.15 and 0.40°C, 1568 respectively; mean CBT increased by 0.11 and 0.32°C; and minimum CBT increased by 0.19 and 1569 0.47°C, respectively. The average range of CBT (1.88°C) was higher in the lower environmental 1570 1571 temperature (15 and 19°C) compared to the higher environmental temperature (23 and 27°C; 1.83°C) but only in the low energy treatment. Presumably, birds maintained lower body 1572 1573 temperature in the lower environmental temperature because they try to decrease the temperature difference between the body and the environment, to dissipate less heat to the environment, thusconserving heat in the body.

Overall, CBT were similar between the high and low energy treatments, suggesting that 1576 presumably, heat production was higher in the low energy treatment due to higher feed and 1577 protein intake (Musharaf and Latshaw, 1999). However, birds were likely able to cope with the 1578 1579 change in heat production within the environmental temperature ranging from 15 to 27° C. The daily average maximum, minimum and mean CBT were high in the 27°C treatment group 1580 regardless of high and low energy diets. The current study revealed that the trend of increases in 1581 1582 CBT was in a linear fashion with the increase of environmental temperatures ranging from 19 to 27°C in feed restricted broiler breeder hens. Presumably, birds reduce heat dissipation with 1583 increasing environmental temperature from 19°C onward. This was in agreement with Teeter et 1584 1585 al. (1992), who reported that the CBT in broiler chickens increased linearly with increasing environmental temperature from 24 to 35°C. The CBT increased linearly with increasing 1586 environmental temperature from 19 to 27°C regardless of dietary energy in feed restricted broiler 1587 1588 breeder hens.

1589 **3.3.4.** Diurnal core body temperature pattern in broiler breeder hens

Overall, CBT increased by 0.15°C suddenly in the pre-dawn period (one hour before lights on), because chickens woke up from sleep and were active (Cain and Wilson, 1974). The CBT increased by about 0.47°C after lights on in the current study (Table 3.5). This was in agreement with Richards (1971), who reported that CBT in chickens rose by about 0.42°C with a sudden response to visual stimuli of light in the morning. The CBT was highest during the first hour of feeding due to the higher rate of metabolism and heat increment increase due to the feeding activity (van Kampen, 1976; Cain and Wilson, 1974). Thereafter, the CBT gradually 1597 decreased over time, possibly because of a reduced rate of metabolism and lower activities 1598 (Kadono and Besch, 1978). The CBT decreased 1 h pre-dusk (one hour before lights off) and quickly dropped 1 h post-dusk (one hour after lights off), probably due to reduced locomotors 1599 1600 activities. de Jong et al. (2002) also reported that the CBT of broiler breeder dropped suddenly after lights turned off. The CBT was higher in the day time because of diet-induced 1601 thermogenesis (Khalil et al., 2004), feeding related activities and behavioural activities (body 1602 shaking, head shaking, preening, cage pecking, beak wiping, crouching; de Jong et al., 2002). 1603 The CBT was lower at night time because of rest, sleep, minimal metabolic rate and a reduction 1604 in other physiological activities in the absence of light (Cain and Wilson, 1974; Khalil et al., 1605 2004). In the current study, CBT was higher by 0.70°C in the day than at night in feed restricted 1606 broiler breeder hens (Table 3.5). Similarly, Lacey et al., (2000b); Kadono and Besch, (1978); 1607 1608 Cain and Wilson, (1974) reported that CBT was higher in day time than in night time in full-fed chickens. The CBT peaked in the first hour after feeding in feed-restricted broiler breeder hens. 1609

Diurnal CBT patterns in broiler breeder hens were similar trend in different 1610 1611 environmental temperatures (Figure 3.1). The difference in CBT between day and night increased linearly with decreasing environmental temperature from 27 to 15°C. The CBT in the 1612 0-1 h post-feed, 1-2 h post-feed, 2-3 h post-feed, 3-4 h post-feed, over 4 h post- feed, 1 h pre-1613 dusk, 1 h pre-dawn, pre-feed, 1 h post-dusk, and night were higher in the 27°C treatment group 1614 (Table 3.5). The CBT was highest in the 27°C, intermediate in the 23°C and lowest in the 19 and 1615 15°C temperature treatment (Table 3.4; Figure 3.1). Presumably, birds in the 27°C treatment 1616 1617 group dissipated less heat to the environment compared to other treatments. However, postprandial CBT was higher at 0-1 h post feed regardless of treatments but higher in 23 and 1618 1619 27°C treatments. In the current study, average CBT was 41.08 and 40.15°C in the day and the

night time respectively. These results were in agreement with those of others researchers who
found higher CBT in the day time compared to night time in full fed layer chickens (Fronda,
1921; Heywang, 1938; Winget et al., 1965). Similar diurnal CBT pattern in broiler breeder hens
was recorded in day and night time between the high and low energy diets (Figure 3.2).
However, the CBT in night time was higher in the low energy diet than in the high energy diet
(Table 3.5). Possibly, birds conserved more body heat due to higher feed intake.

The present study indicated that the CBT peaked in the first hour after feeding, 1626 suggesting a strategy for feeding time. If birds will be fed in the warmer time of day, they might 1627 1628 be unable to dissipate adequate heat to the environment leading less dependency on diet induced 1629 thermogenesis. As a consequence, birds may reduce feed intake, including other nutrients such as protein, amino acids, minerals and vitamins that will negatively affect the egg production and 1630 1631 egg size (National Research Council.1981). Generally, in a hot summer, mid-morning to midafternoon is a warmer period of day. Shifting feeding time in the cooler period of day likes early-1632 morning or late afternoon or a half meal in the early morning and a half meal in the late 1633 1634 afternoon could be a better approach to avoid heat stress in the hot summer. Birds can take enough feed for requirement and thus, they sustain their production. 1635

3.4 REFERENCES

1638 1639 1640	Aviagen. 2007. Ross 708 parent stocks performance objectives. Aviagen Ltd. www.aviagen.com/docs/Ross 708 PS Performance Objectives.pdf. Accessed June 2010.
1641 1642	Cain, J. R., and W. O. Wilson. 1974. The influence of specific environmental parameters on the circadian rhythms of chickens. Poult. Sci. 53:1438-1447.
1643 1644 1645	Canadian Council on Animal Care. 1993. Guide to the use of experimental animals. Vol. 1. Can. Counc. Anim. Care, Ottawa, Ontario, Canada.
1646 1647 1648 1649 1650	Collin, A., S. Cassy, J. Buyse, E. Decuypere, and M. Damon. 2005. Potential involvement of mammalian and avian uncoupling proteins in the thermogenic effect of thyroid hormones. Domest. Anim. Endocrin. 29:78-87.
1651 1652 1653	Bennett, C. D., and S. Leeson. 1990. Influence of energy intake on development of broiler breeder pullets. Can. J. Anim. Sci. 70:259-266.
1653 1654 1655 1656 1657	De Andrade, A. N., J. C. Rogler, W. R. Featherston, and C. W. Alliston. 1977. Interrelationships between diet and elevated temperatures (cyclic and constant) on egg production and shell quality. Poult. Sci. 56:1178-1188.
1658 1659 1660 1661	Deeb, N., and A. Cahaner. 1999. The effects of naked neck genotypes, environmental temperature, and feeding status and their interactions on body temperature and performance of broilers. Poult. Sci. 78:1341-1346.
1662 1663 1664 1665	de Jong, I. C., S. Van Voorst, D. A. Ehlhardt, and H. J. Blokhuis. 2002. Effects of restricted feeding on physiological stress parameters in growing broiler breeders. Br. Poult. Sci. 43:157-168.
1665 1667 1668	Fronda, F. M. 1921. A comparative study of the body temperature of the different species and some representative breeds of poultry- a preliminary report. Poult. Sci. 1:16-22.
1669 1670 1671	Furlan, R. L., DE. de. Fario Filho, P. S. Rosa, and M. Macari. 2004. Does low protein diet improve broiler performance under heat stress condition? Braz. J. Poult. Sci. 6:71-79.
1672 1673 1674	Heywang, B. W. 1938. Effects of some factors on the body temperature of hens. Poult. Sci. 17:317-323.
1675 1676 1677	Hocking, P. M. 2004. Roles of body weight and feed intake in ovarian follicular dynamics in broiler breeders at the onset of lay and after a forced molt. Poult. Sci. 83:2044-2050.
1678 1679 1680	Geraert, P. A., M. G. Macleod, and B. Leclercq. 1988. Energy metabolism in genetically fat and lean chickens: Diet-and cold-induced thermogenesis. J. Nutr. 118:1232-1239.

Kadono, H., and E. L. Besch. 1978. Telemetry measured body temperature of domestic fowl at
various ambient temperatures. Poult. Sci. 57:1075-1080.
Khalil, A. M., K. Matsui, and K. Takeda. 2004. Diurnal and oviposition-related changes in heart rate, body temperature and locomotors activity of laying hens. Anim. Sci. J. 75:169-174.
Lacey, B., T. K. Hamrita, M. P. Lacy, and G. L. Van Wicklen. 2000a. Assessment of poultry
deep body temperature responses to environmental temperature and relative humidity
using an on-line telemetry system. J. Trans. ASAE. 43:717-721.
Lacey, B., T. K. Hamrita, M. P. Lacy, G. L. Van Wicklen, and M. Czarick. 2000b. Monitoring
deep body temperature responses of broilers using biotelemetry. J. Appl. Poult. Res. 9:6-
12.
Mack, L.A., J.N. Felver-Gant, R. L. Dennis, H. W. Cheng. 2013. Genetic variation alter
production and behavioral responses following heat stress in 2 strains of laying hens.
Poult. Sci. 92:285-294.
Musharaf, N. A., and J. D. Latshaw. 1999. Heat increment as affected by protein and amino acid
nutrition. World's Poult. Sci. J. 55:233-240.
National Research Council. 1981. Effect of environment on nutrient requirements of domestic
animals. Natl. Acad. Press, Washington D. C.
Renema, R. A., and F. E. Robinson. 2004. Defining normal: Comparison of feed restriction and full feeding of female broiler breeders. World's Poult. Sci. J. 60:511-525.
Renema, R. A., M. E. Rustad, and F. E. Robinson. 2007. Implications of changes to commercial
broiler and broiler breeders' body weight targets over the past 30 years. World's Poult.
Sci. J. 63:457-467.
Reyes, M. E., C. Salas, and C. N. Coon. 2011. Energy requirements for maintenance and egg
production of broiler breeder hens. Int. J. Poult. Sci. 10:913-920.
Richards, S. A. 1971. The significance of changes in the temperature of the skin and body core
of the chicken in the regulation of heat loss. J. Physiol. 216:1-10.
Romero, L. F., M. J. Zuidhof, R. A. Renema, F. E. Robinson, and A. Naeima. 2009. Nonlinear
mixed models to study metabolizable energy utilization in broiler breeder hens. Poult.
Sci. 88:1310-1320.
SAS. 2008. SAS 9.2 © 2002-2008 by SAS Institute, Inc., Cary, NC, USA.

1726	Spratt, R. S., and S. Leeson. 1987. Broiler breeder performance in response to diet protein and
1727	energy. Poult. Sci. 66:683-693.
1728	
1729	Teeter, R. G., M. O. Smith, and C. J. Wiernusz. 1992. Research note: Broiler acclimation to heat
1730	distress and feed intake effects on body temperature in birds exposed to thermoneutral
1731	and high ambient temperatures. Poult. Sci. 71:1101-1104.
1732	
1733	Van Kampen, M. 1976. Activity and energy expenditure in laying hens: 3. The energy cost of
1734	eating and posture. J. Agric. Sci. 87: 85-88.
1735	
1736	Wilson, H. R., and R. H. Harms. 1984. Evaluation of nutrient specifications for broiler breeders.
1737	Poult. Sci. 63:1400-1406.
1738	
1739	Winget, C. M., E. G. Averkin, and T. B. Fryer. 1965. Quantitative measurement by telemetry of
1740	ovulation and oviposition in the fowl. Anim. J. Physiol. 209:853-858.
1741	
1742	Yu, M. W., F. E. Robinson, R. G. Charles, and R. Weingardt. 1992. Effect of feed allowance
1743	during rearing and breeding on female broiler breeders. 2. Ovarian morphology and
1744	production. Poult. Sci. 71:1750-1761.
1745	

Table 3.1 Impacts of dietary energy level on BW, age at sexual maturity, first egg weight, andage at 52 g egg weight in broiler breeder females.

Dietary ME (kcal/kg)	BW at sexual maturity (g)	Age at sexual maturity (d)	First egg weight (g)	BW at 52 g egg (g)	Age at 52 g eggs (d)
2,912	2,891	186.8	48.2	2,944	192.4
2,786	2,864	186.3	48.1	2,919	191.1
SEM	23.84	0.67	0.46	18.7	0.66
Probability	0.4351	0.6038	0.8573	0.3466	0.1475

Table 3.2 Impacts of environmental temperature and dietary energy level on total, normal and settable egg production in broiler breeder females from 25 to 41 wk of age.

1	7	5	3

Environmental temperature	Dietary	Total	Normal	Settable Egg
(°C)	ME	Egg	Egg Production	Production ³
Set Actual \pm SD	(kcal/kg)	Production ¹ (%)) (%)	(%)
15 17.5 ± 1.3		76.1	75.5	68.2
$19 20.4 \pm 1.0$		76.5	75.1	66.6
$23 \qquad 23.5\pm0.8$		75.0	73.1	66.4
27 27.3 ± 1.0		78.1	73.0	64.3
SEM		1.01	1.01	1.22
	2,912	77.3	75.3 ^a	66.6
	2,786	75.6	73.2 ^b	66.1
SEM		0.71	0.70	0.86
15	2,912	76.5 ^{ab}	76.1	68.7
19	2,912	77.8 ^{ab}	75.6	66.1
23	2,912	73.6 ^b	73.2	65.3
27	2,912	81.2 ^a	75.8	66.4
15	2,786	75.7 ^{ab}	74.9	67.7
19	2,786	75.2 ^{ab}	74.6	67.1
23	2,786	76.3 ^{ab}	73.0	67.4
27	2,786	75.0 ^b	70.1	62.2
SEM		1.73	1.43	1.72
Sources of variation		Probability		
Environmental temperature		0.1794	0.1564	0.1708
Dietary energy		0.0885	0.0446	0.6715
Interaction		0.0198	0.1905	0.2828

^{a-b}Means within column and main effect with no common superscript are significantly different (P < 0.05) ¹Total eggs= all eggs, including abnormal eggs ²Normal eggs= all eggs minus abnormal eggs

³Settable eggs= normal eggs < 52 g

Enviro (°C)	onmental temperature	Dietary energy	ADFI	ADG	Egg	Feed:Egg ¹	ME:Egg ²	CP:Egg ³
Set	Actual ± SD	(ME, kcal/kg)	(g)	(g)	wt (g)	(g/g)	(kcal/g)	(g/g)
15	17.5 <u>+</u> 1.3		141.87 ^a	7.73 ^a	57.74	3.23	9.20	0.53
19	20.4 <u>+</u> 1.0		137.83 ^b	5.67 ^b	57.74	3.18	9.06	0.52
23	23.5 <u>+</u> 0.8		137.82 ^b	8.48 ^a	57.59	3.16	9.00	0.51
27	27.3 <u>+</u> 1.0		139.78 ^{ab}	6.03 ^b	57.51	3.16	8.99	0.51
SEM			0.73	0.45	0.12	0.04	0.12	0.01
		2,912	135.30 ^b	6.95	57.32 ^b	3.07 ^b	8.93 ^b	0.50^{1}
		2,786	143.35 ^a	7.01	57.97 ^a	3.30 ^a	9.19 ^a	0.53
SEM			0.52	0.32	0.12	0.03	0.09	0.01
Source of variation				Pro	bability			
Enviro	onmental temperature		0.0001	< 0.0001	0.1450	0.5919	0.5900	0.590
Dietar	y energy		< 0.0001	0.8890	0.0002	< 0.0001	0.0377	0.000
Interac	ction		0.1304	0.9520	0.7696	0.6237	0.6259	0.624

Table 3.3 Impacts of environmental temperature and dietary energy level on average daily feed intake (ADFI), average daily gain
(ADG), and feed efficiency in broiler breeder females from 25 to 41 wk of age.

1761 ^{a-b}Means within column and main effect with no common superscript are significantly different (P < 0.05)

1762 ¹Feed:Egg (g/g) = ADFI divided average daily egg mass

1763 2 ME:Egg (kcal/g of egg) = average daily ME intake divided by average daily egg mass

1764 3 CP:Egg (g/g of egg) = average daily CP intake divided by average daily egg mass

1766	Table 3.4 Effects of environmental	temperature and dietary	energy level on core body

temperature (CBT) in broiler breeder hens from 25 to 41 wk of age.

1768

Environmental	Dietary	Maximum	Maan	Minimum	Damaa
temperature (°C)	energy	Maximum Mean Minimun		Minimum	Range
			(CBT(C)	
15		41.65 ^c	40.54 ^c	39.77 ^c	1.89 ^a
19		41.66 ^c	40.54 ^c	39.78 ^c	1.87 ^a
23		41.81 ^b	40.65 ^b	39.97 ^b	1.84 ^b
27		42.06 ^a	40.86 ^a	40.25 ^a	1.81 ^b
SEM		0.02	0.02	0.03	0.03
	2,912	41.80	40.65	39.93	1.88
	2,786	41.78	40.65	39.96	1.83
SEM		0.03	0.04	0.05	0.04
15	2,912	41.61 ^e	40.53 ^{de}	39.73 ^d	1.88^{ab}
19	2,912	41.64 ^{de}	40.53 ^{de}	39.76 ^d	1.88^{ab}
23	2,912	41.84 ^{bc}	40.66 ^{bc}	39.96 ^{bc}	1.88^{ab}
27	2,912	42.12 ^a	40.87^{a}	40.24 ^a	1.88^{ab}
15	2,786	41.70 ^{ce}	40.56 ^{ce}	39.80 ^{cd}	1.90^{a}
19	2,786	41.67 ^e	40.54 ^{ce}	39.80 ^{cd}	1.87 ^{ab}
23	2,786	41.77 ^{bd}	40.65 ^{bd}	39.98 ^b	1.79 ^b
27	2,786	42.00^{a}	40.85 ^a	40.25 ^a	1.74 ^b
SEM		0.04	0.04	0.05	0.04
Source of variation				Probability	
Temperature		<.0001	<.0001	<.0001	<.0001
Dietary energy		0.6786	0.9150	0.6497	0.3560
Interaction		<.0001	0.0118	0.0388	<.0001

1769 ^{a-d}Means within column and main effect with no common superscript are significantly different (P < 0.05)

1770 Maximum= daily highest CBT of individual hens

1771 Mean= average of daily CBT of individual hens

1772 Minimum= average of daily lowest CBT of individual hens

1773 Range= average of daily range of difference between the highest and lowest in CBT of individual hens

Time of day^1 Temperature (°C) Dietary energy² Overall 15 19 23 27 HE LE -CBT(°C)---40.02^{iD} 40.20^{hB} 40.07^{iC} 40.36^{iA} 40.15^{hB} 40.18^{hA} Night 40.16^{i} 40.19^{hD} 40.24^{hC} 40.37^{fB} 40.46^{gA} 40.30^{fB} 40.33^{fA} 1h pre-dawn 40.31^g 40.77^{eB} 40.75^{fC} 40.81^{dA} 40.81^{fA} 40.77^{dB} 40.80^{dA} 40.78^{e} Pre-feed 41.37^{bA} 41.28^{aB} 41.26^{aB} 41.36^{aA} 41.33^{aA} 41.30^{aB} 0-1 h post-feed 41.32^a 41.23^{bC} 41.21^{bC} 41.35^{aB} 41.43^{aA} 41.29^{aB} 41.32^{aA} 41.31^b 1-2 h post-feed 41.21^{bB} 41.16^{cC} 41.12^{cD} 41.32^{cA} 41.21^b 41.19^b 41.20° 2-3 h post-feed $41.05d^{C}$ 41.03^{dD} 41.08^{cB} 41.20^{dA} 41.09^c 41.09° 41.09^d 3-4 h post-feed 40.72^{fC} 40.76^{eB} 40.77^{eB} 40.85^{eA} 40.77^{f} >4 h post-feed 40.76^{e} $40.79^{\rm e}$ 40.22^{gD} 40.29^{gB} 40.43^{hA} 40.26^{gC} 40.29^g 40.31^g 40.30^h 1 h pre-dusk 39.97^{jD} 40.05^{jC} 40.17^{iB} 40.34^{jA} 40.12^{iB} 40.15^{iA} 40.13^j 1 h post-dusk 0.004 SEM 0.005 0.005 0.005 0.005 0.003 0.005 Source of variation-----------Probability------Time of day < 0.0001 < 0.0001 < 0.0001

Table 3.5 Effects of environmental temperature and dietary energy level on diurnal core bodytemperature (CBT) in broiler breeder hens from 25 to 41 wk of age.

1776 A-C Means across the row within each treatment with no common letters are significantly different at $P \le 0.05$.

1777 ^{a-j}Means within column within each treatment with no common letters are significantly different at $P \le 0.05$.

¹Night= lights off time period (22:30 to 04:30; excluding post-dusk and pre-dawn); 1 h pre-dawn= one hour time

1779 period before lights on (at 05:30); Pre-feed= time period from lights on to feeding time; 0-1 h post-feed= first hour

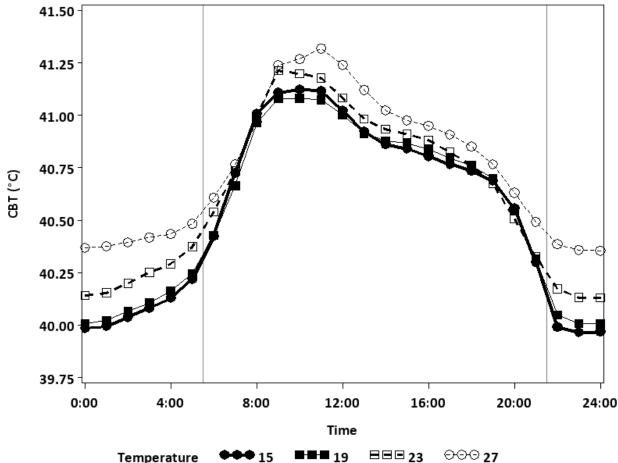
after feed, 1-2 h post-feed= second hour after feed, 2-3 h post feed= third hour after feed, 3-4 h post-feed= fourth

hour after feed, >4 h post-feed= remainder of the lights on period following the fourth hour post-feed, excluding pre-

dusk 1 h pre-dusk= one hour time period before lights off, 1 h post-dusk= one hour time period after lights off (at

1783 21:30). Note: Normally, feeds were supplied to the birds at 07:30 and it little delayed at weighing days.

1784 2 HE= High energy: 2,912 kca/kg; LE= Low energy: 2,786 kcal/kg



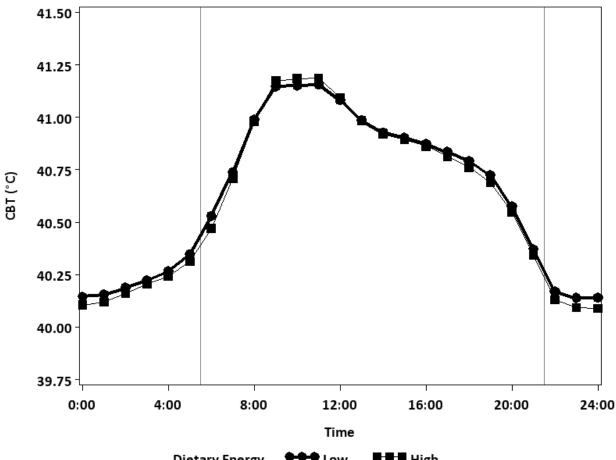
1785

Figure 3.1 Diurnal core body temperature (CBT) pattern of broiler breeder hens (25 to 41 wk of

1786 age) in different environmental temperatures. Vertical reference lines in the graph indicate lights 1787 on at 05:30 and lights off at 21:30 (Y axis indicates CBT in degree Celsius and X axis indicates 1788 time in hour). 1789

Night= lights off time period (22:30 to 04:30; excluding post-dusk and pre-dawn); 1 h pre-1790 1791 dawn= one hour time period before lights on (at 05:30); Pre-feed= time period from lights on to feeding time; 0-1 h post-feed= first hour after feed, 1-2 h post-feed= second hour after feed, 2-3 1792 h post feed= third hour after feed, 3-4 h post-feed= fourth hour after feed, >4 h post-feed= 1793 1794 remainder of the lights on period following the fourth hour post-feed, excluding pre-dusk 1 h pre-dusk= one hour time period before lights off, 1 h post-dusk= one hour time period after lights 1795

1796 off (at 21:30).



1797

Dietary Energy 📤 Low 🔳 🖿 High

Figure 3.2 Diurnal core body temperature pattern (CBT) of broiler breeder hens
(25 to 41 wk of age) in high and low dietary energy level. Vertical reference lines
in the graph indicate lights on at 05:30 and lights off at 21:30 (Y axis indicates
CBT in degree Celsius and X axis indicates time in hour).

Night= lights off time period (22:30 to 04:30; excluding post-dusk and pre-dawn); 1802 1 h pre-dawn= one hour time period before lights on (at 05:30); Pre-feed= time 1803 period from lights on to feeding time; 0-1 h post-feed= first hour after feed, 1-2 h 1804 post-feed= second hour after feed, 2-3 h post feed= third hour after feed, 3-4 h 1805 1806 post-feed= fourth hour after feed, >4 h post-feed= remainder of the lights on period following the fourth hour post-feed, excluding pre-dusk 1 h pre-dusk= one 1807 hour time period before lights off, 1 h post-dusk= one hour time period after lights 1808 1809 off (at 21:30).

1811 CHAPTER 4: IMPACT OF FEEDING TIMES, PHOTOPERIODS AND 1812 DIETARY ENERGY LEVELS ON CORE BODY TEMPERATURE AND 1813 OVIPOSITION IN BROILER BREEDER HENS

ABSTRACT: The effects of feeding time, photoperiod, and dietary energy level 1814 1815 on core body temperature (CBT) and oviposition time were investigated using 1816 Ross 708 broiler breeder hens at 44 wk (7 d) of age. A total of 192 hens, including 48 CBT temperature sensor-implanted hens, were equally and randomly 1817 1818 distributed over six climate-controlled environmental chambers (n=32/chamber). The experiment was a 4 x 2 x 2 factorial arrangement, with four feeding times 1819 1820 (07:30; 11:30; 15:30; or split feeding: 07:30 and 15:30); two dietary ME levels (high, 2,912 or low, 2,786 kcal/kg) and two photoperiods (standard, 16L:8D or 1821 1822 continuous, 24L:0D). Oviposition time was recorded at 10 min intervals using 1823 video cameras. Mean $(39.73 \pm 0.08^{\circ}C)$ and minimum $(40.38 \pm 0.05^{\circ}C)$ CBT was 1824 higher in the low energy fed chickens than in the high energy fed chickens. In 1825 general, CBT were low at night, and increased anticipatorily prior to the lights 1826 turning on. The CBT of breeder hens peaked within 2 h after feeding, after which 1827 CBT gradually decreased in anticipation of lights turning off. Diurnal CBT patterns were similar between the photoperiod treatments, but CBT of birds on the 1828 1829 continuous photoperiod did not drop as much at night as birds on the standard 1830 photoperiod. Feeding time, dietary energy level and photoperiod had no influence 1831 on egg production and egg weight. Average daily feed intake was similar among 1832 feeding time and photoperiod treatments. Oviposition time was later in split fed hens than in morning fed hens. Dietary energy level and photoperiod did not 1833

1834 affect oviposition times. Shifting feeding time in the early morning or late1835 afternoon especially colder part of day may be a strategy to mitigate heat stress.

1836 Keywords: Feeding time, photoperiod, core body temperature dynamics,

1837 oviposition, broiler breeder hens

1838 4.1. INTRODUCTION

Control of feed intake and lighting are important practices for optimizing 1839 broiler breeder management at the industry level (Renema and Robinson, 2004; 1840 Backhouse and Gous, 2005; Gibson et al., 2008; Romero et al., 2009). Feed is 1841 1842 normally provided to birds once daily, in the morning. However, this practice 1843 does not supply nutrients at peak demand times that coincide with the timing of requirements for egg shell development, and may reduce shell quality (Bootwalla 1844 1845 et al., 1983). Cave (1981) observed that feeding later in the day or splitting the single feed allocation across more frequent feeding throughout the day decreased 1846 weight gain and increased egg mass, suggesting that this strategy enhanced 1847 1848 nutrient availability for egg production and regulated excessive body tissue deposition. Taherkhani et al. (2010) likewise reported that split feeding broiler 1849 1850 breeder hens increased egg production. Moreover, Spradley et al. (2008) suggested that egg production was higher in split-fed broiler breeder hens than in 1851 one meal fed broiler breeder hens. Afternoon feeding in cage reared broiler 1852 1853 breeder hens increased the shell quality of eggs (Backhouse and Gous, 2005). Harms (1991) reported that medium and light BW hens had increased egg weight 1854 1855 and egg production when they were fed in the afternoon instead of morning. This 1856 finding was not supported by Brake (1988). It appears clear, meanwhile, that restricted-fed broiler breeders experienced hunger and frustration due to high
motivation for feeding (Hocking et al., 1996; de Jong et al., 2002). Split feeding
thus offers a potential method of reducing frustration and hunger because birds
get feed two times a day, increasing the duration of nutrient metabolism.
However, two time feedings may also increase labour costs and delay oviposition
resulting in increased management hazard.

Feeding time may change the oviposition time, though the conclusions of 1863 researchers in this respect appear inconsistent. Wilson and Keeling (1991) and 1864 1865 Backhouse and Gous (2005) reported that oviposition time was delayed due to 1866 feeding broiler breeder hens in the afternoon (16:00 to 18:00) due to late ovulation or prolonged egg formation time in the oviduct. Conversely, Samara et al. (1996) 1867 noted no difference in the oviposition time between morning- and afternoon-fed 1868 broiler breeder hens. Similarly, Lewis and Perry (1988) reported no difference in 1869 oviposition time between broiler breeder hens fed a single allocation of feed in the 1870 1871 morning or half the daily feed allocation twice each day.

Photoperiod can also affect oviposition patterns. Oviposition time was 1872 1873 delayed by 30 min per hour of photoperiod increase, with no difference after 14 h of photoperiod (Lewis et al., 2004). Full fed hens under 24L:0D photoperiod had 1874 a clear diurnal CBT rhythm. Presumably, the CBT increased due to feeding 1875 1876 activity and feed metabolism. This was in agreement with Kadono et al. (1983). They reported that CBT in full fed layer chickens was higher during the waking 1877 1878 phase (feeding and drinking time) than during the sleeping phase (decreased 1879 feeding and drinking time).

1880 Environmental temperature can change feed intake, weight gain, egg 1881 production, egg quality, and CBT dynamics in chickens. Core body temperature patterns follow diurnal patterns, increased by photoperiod (Fronda, 1921; 1882 1883 Heywang, 1938) and physiological events such as oviposition and ovulation (Winget et al. 1965). The CBT ranged from 39.0 to 41.2 °C in feed-restricted 1884 1885 broiler breeder hens (Savory et al., 2006). The CBT was increased due to feeding activity and feed metabolism during the photoperiod and decreased following feed 1886 removal (Skinner-Noble and Teeter, 2003). Variation in core body temperature is 1887 1888 likely an adaptation to regulate the rate of heat transfer from the body of chickens 1889 to their environment, and influences production efficiency (NRC, 1981). Previous studies (Chapter 2 and 3) suggest that the diurnal CBT pattern is mostly similar 1890 1891 between pullets and hens under a standard photoperiod. The current experiment was designed to compare the diurnal CBT pattern between a standard and 1892 continuous photoperiod, between dietary energy levels, and to systematically 1893 1894 determine whether feeding-related CBT dynamics could be detected by observing CBT patterns following feeding at different times in the day. Limited research 1895 1896 exists comparing the combined effects of feeding time, photoperiod and dietary energy level on CBT dynamics and oviposition time in broiler breeder hens. This 1897 1898 information can help us to understand the mechanisms of efficiency in response to 1899 management.

1900 **4.2. MATERIALS AND METHODS**

1901 **4.2.1. Animal Care Approval**

1902 This research project was managed in compliance with the Guide to the 1903 Care and Use of Experimental Animals (Canadian Council on Animal Care, 1993) 1904 and the experimental protocol was approved by the Animal Care and Use 1905 Committee for Livestock of the University of Alberta.

1906 **4.2.2. Experimental Design**

A 4 x 2 x 2 factorial experiment in a Completely Randomized Design was performed using four feeding times in which 100% of the daily feed allotment was provided at either 07:30, 11:30, or 15:30, or split, in which 50% of the daily feed allotment at both 07:30 and 15:30; two photoperiods (16L:8D and 24L:0D) and two dietary energy levels (2,912 kcal/kg and 2,786 kcal/kg; Appendix A).

1912 **4.2.3. Stocks and Management**

At 44 wk (7 d; 308 to 316 d) of age, 192 Ross 708 broiler breeder hens (Aviagen Inc., Huntsville, AL) in individual laying cages in six climate-controlled environmental chambers were used for seven days. Photoperiod in 3 of the chambers was 24L:0D, and 16L:8D in the remaining 3 chambers. Half of the birds in each chamber were fed the high energy diet, and half the low energy diet. The feeding time treatments were applied to equal numbers of birds in all treatments in all chambers.

4.2.4 Surgical Implantation Temperature Sensors

1921 A total of 64 broiler breeder females were implanted at 13 wk of age with 1922 temperature sensor transmitters in their right abdominal cavity. Details of the surgical procedures are provided in Chapter 2 of the current thesis. A total of 48
of these implanted hens were randomly and equally distributed among the
treatments in the six environmental chambers in this experiment. Core body
temperature measurements were recorded every 10 min.

1927 **4.2.5. Data Collection**

1928 Body weight of hen was recorded at the beginning and the end of the experiment and daily feed allocation was made according to the primary breeder's 1929 management guide line based on target BW and production rate. Eggs were 1930 1931 collected daily at 15:00, and individually weighed using a digital balance. The average daily maximum, mean, minimum, and range of individual CBT were 1932 determined according to the procedure described in Chapter 2 of the current 1933 1934 thesis. Temporal diurnal CBT categories were classified as; Night was defined as the time when lights were turned off at 21:30 only in 16L:8D treatment, excluding 1935 1 h post-dusk and 1 h pre-dawn; 1 h pre-dawn was the one hour time period 1936 1937 before lights on; pre-feed was the time period from lights turned on at 05:30 only in 16L:8D to feeding time; 0-1 h post-feed was the first hour after feeding; 1-2 h 1938 1939 post-feed was the second hour after feeding; 2-3 h post feed was the third hour after feeding; >3 h post-feed was the remainder of time that the lights were on 1940 1941 following the third hour post-feed, excluding pre-dusk; 1 h pre-dusk was the one 1942 hour time period before lights turned off; 1 h post-dusk was the one hour time period after lights were turned off. These categories were based on the lights 1943 1944 turning on at 05:30 and off at 21:30 in both photoperiod treatments; this was an 1945 arbitrary classification for the 24L:0D treatment.

1946 4.2.6. Webcam Video Camera

All hens were monitored by webcam for determination of oviposition time. Time-stamped images were taken at 10 min intervals for groups of 16 caged hens to determine oviposition time. Daily oviposition times for each hen were determined by video observation.

1951 **4.2.7. Statistical Analysis**

The Mixed Procedure of SAS 9.2 (SAS Institute, Cary, NC) was used for analysis and treatment means were differentiated using Tukey's test with a critical value of $P \le 0.05$. Feeding times, photoperiods and dietary energy levels were treated as main effects and age as a random effect. Each individually-caged bird was considered an experimental unit.

1957 **4.3. RESULTS AND DISCUSSION**

1958 4.3.1. Relationship of feeding time, photoperiods, dietary energy, and CBT

The maximum, mean and minimum CBT in broiler breeder hens were not 1959 1960 different according to feeding time (Table 4.1). Mean and minimum CBT were higher in the low energy diet-fed broiler breeder hens than the high energy diet-1961 1962 fed broiler breeder hens. Possibly, heat increment increased due to a higher amount of total feed intake (and therefore protein intake), and protein metabolism 1963 (Musharaf and Latshaw, 1999). The high energy treatment had a greater range in 1964 1965 CBT (1.87°C) than that of the low energy treatment (1.57°C). The intake of larger amount feed in low energy diet-fed birds might have produced more metabolic 1966 heat and required longer time for metabolism (Almirall and Steve-Garcia, 1994) 1967 1968 and thus heat increment prolonged resulting in a narrow range of CBT.

Photoperiod treatment had no effect on the maximum, mean, and minimum CBT in broiler breeder hens. The 16L:8D photoperiod had a greater range in CBT (1.82°C) than the continuous photoperiod (1.61°C). This was likely due to decreased activity after lights were turned off at night (Khalil et al., 2004) in the 16L:8D treatment. However, CBT was higher likely due to increased activity level in the 24L:0D treatment against night period (from 21:30 to 05:30) in the 16:8D treatment.

1976 **4.3.2. Diurnal core body temperature pattern**

1977 Across treatments, CBT was lowest $(39.97 \pm 0.01^{\circ}C)$ at night, increased in anticipation of the lights coming on (by 0.16°C one hour before lights on) and 1978 peaked at 40.87°C within 2 hours of feeding (Table 4.2). The CBT gradually 1979 1980 decreased after 2 h of feeding, and dropped substantially before lights were turned off (1 h pre-dusk), dropping quickly to night time CBT in the hour after lights 1981 were turned off (Figure 4.1). The CBT peaked after feeding regardless of feeding 1982 1983 time (Figure 4.1). The post-prandial CBT peak was likely due to heat increment related to feeding activity (Khalil et al., 2004), and feed metabolism and 1984 1985 absorption (Wilson et al., 1989). Interestingly, van Kampen (1976) observed that heat production increased during eating by an average of 37% due to the physical 1986 activities related to eating. Core body temperature increased in the current 1987 1988 experiment because the heat generated by the flurry of eating activity built up in the body, and took several hours to dissipate fully. The low CBT observed at 1989 night was likely due to reduced activity level after lights were turned off, although 1990 1991 activity level was not measured in the current experiment. Lacey et al. (2000);

Khalil et al. (2004), and Savory et al. (2006) reported similar diurnal CBT
patterns in broiler and broiler breeder chickens. However, CBT fluctuation was
higher in broiler breeder chickens than in broilers due to feed restriction.

1995 Night-time CBT were approximately 0.22°C lower in morning- and splitfed hens compared to noon- and afternoon-fed hens. The highest CBT was 1996 1997 observed in the noon-fed treatment. This would suggest that noon feeding may cause heat stress in breeders in hot environment; in this condition, the temperature 1998 difference between birds and the environment decreases. Birds are unable to 1999 2000 dissipate adequate amounts of heat to the environment and they reduce their dependency on diet-induced thermogenesis to maintain their CBT (Swennen et 2001 al., 2007) resulting in a decreased feed intake; decreased production and 2002 2003 efficiency. Conversely, morning- and split-feeding may be an appropriate way for keeping CBT control because morning is cooler than noon of a day and split-2004 feeding may produce comparatively less heat than one meal feeding. At cold 2005 2006 environment like in the morning, birds can dissipate adequate amount of heat to the environment for energy balance in open housing management. 2007

Day and night time CBT were 0.12°C and 0.28°C higher respectively in the low energy treatment than in the high energy treatment (Table 4.2; Figure 4.2). Feed intake was also higher in the low energy treatment (Table 4.3). The increase in CBT during day was likely due to a combination of increased feeding behaviour (van Kampen, 1976) and heat increment associated with digestion, absorption, and protein metabolism (Musharaf and Latshaw, 1999). Intakes of feed as well as protein and other nutrients were higher in the low energy group

resulting in higher heat increment during metabolism compared to the high energy
group. Presumably, the higher heat production resulting from the low energy diet
may increase CBT in both day and night time period.

2018 Interestingly, in spite of the absence of a day/night cue in the continuous photoperiod, the diurnal CBT pattern was similar to the 16L:8D photoperiod 2019 2020 (Figure 4.3). The CBT likely decreased during resting and sleeping time in feedrestricted broiler breeders in the continuous photoperiod, this was in agreement 2021 2022 with Kadono et al. (1981) who reported that CBT decreased during the sleeping 2023 phase in full-fed laying hens in the continuous photoperiod (24L:0D). Activity 2024 levels in breeder hens that could be stimulated by human activity, including feeding and egg collecting, which occurred during the day, but they were absent 2025 2026 during the night. During the night (21:30 to 05:30), CBT was 0.20°C higher in the continuous photoperiod group compared to the standard photoperiod group (Table 2027 4.2). This difference may be due to a smaller drop in night-time activity levels in 2028 2029 the continuous photoperiod than in the standard photoperiod. Birds used in the 2030 continuous photoperiod treatment group could have been influenced by the pre-2031 established diurnal pattern, sleeping pattern and other activity of the standard photoperiod (Birds were reared under the standard photoperiod (16L:8D) before 2032 starting this experiment). In addition, similar morning feeding cycle, feeding 2033 2034 related activity and feed metabolism resulted in a similar trend of diurnal CBT patterns in the standard and in the continuous photoperiod. 2035

However, feed-associated increases in CBT were lower (approximately 0.1°C) in the continuous photoperiod compared to the standard photoperiod

(Table 4.2), likely due to higher activity level increased CBT in standard photoperiod. In the standard photoperiod after lights were turned on, birds increased activity in addition to feeding activity and diet induced thermogenesis increased CBT, which was higher than the CBT in the continuous photoperiod.

2042 **4.3.3. Feeding times, photoperiods and dietary energy levels on egg**

2043 production

Feeding times did not significantly affect average daily feed intake 2044 (P=0.0508), egg production, and egg weight (P=0.0860) in the feed restricted 2045 2046 broiler breeder hens (Table 4.3). This was consistent with Backhouse and Gous 2047 (2005), who reported that feeding time had no effect on egg production and egg weight. However, Lewis and Perry (1988) and Wilson and Keeling (1991) 2048 2049 suggested that egg production decreased with split feeding. In addition, de Avila et al. (2003) reported that feeding in the afternoon reduced egg production in 2050 2051 chickens. Bootwalla et al. (1983) suggested that feeding times did not affect egg 2052 weight. Conversely, Farmer et al. (1983) observed that egg weight was higher 2053 with afternoon feeding than with morning feeding in caged broiler breeder hens. 2054 In afternoon feeding, birds may directly use feed energy for egg formation, whereas in morning feeding, birds store energy first and thereafter birds use stored 2055 energy for egg formation. Presumably, birds lose energy through this process. As 2056 2057 a result, egg weight decreases in feed restricted broiler breeder hens.

In the current study, ADFI was 3.5 g higher in the low energy diet fed broiler breeder hens than in the high energy diet fed broiler breeder hens. Dietary energy had no effects on daily BW gain, egg production and egg weight (Table

4.3). Photoperiods did not affect ADFI, ADG, egg production, and egg weight inbroiler breeder hens.

4.3.4. Feeding time, photoperiod and dietary energy level on oviposition time

2064 Oviposition time in broiler breeder hens was similar across the morning, noon and afternoon feeding treatments. This was in agreement with the results of 2065 Samara et al. (1996). However, the morning feeding resulted in oviposition 2066 2067 occurring over one hour earlier than split feeding. Several researchers have reported that split feeding delayed oviposition time in broiler breeder hens 2068 (Wilson and Keeling, 1991; Harms 1991; Samara et al., 1996; Backhouse and 2069 Gous, 2005). The reasons may be associated with the effects of feeding time on 2070 the timing of the open period for LH release (Backhouse and Gous, 2005). Birds 2071 2072 may have rest after a second time feeding that may increase release of LH for ovulation within the open period and that may be the reason for the delay in 2073 2074 oviposition time in split fed hens. In the current study, the continuous photoperiod 2075 delayed oviposition time (P = 0.0673; nearly significant) compared to the standard photoperiod. However, Lewis et al. (2004) observed that oviposition time was 2076 2077 advanced in a shorter photoperiod.

4.4. REFERENCES

2080	Almirall, M. and E. Esteve-Garcia, 1994. Rate of passage of barley diets with
2081	chromium oxide: Influence of age and poultry strain and effect of ß-
2082	glucanase supplementation. Poult. Sci. 73:1433-1440.
2083	
2084	Aviagen. 2007. Ross 708 parent stocks performance objectives. Aviagen Ltd.
2085	www.aviagen.com/docs/Ross 708 PS Performance Objectives.pdf.
2086	Accessed June 2010.
2087	
2088	Backhouse, D., and R. M. Gous. 2005. The effect of feeding time on shell quality
2089	and oviposition time in broiler breeders. Br. Poult. Sci. 46:255-259.
2090	•
2091	Bootwalla, S. M., H. R. Wilson, and R. H. Harms. 1983. Performance of broiler
2092	breeders on different feeding systems. Poult. Sci. 62:2321-2325.
2093	
2094	Brake, J. 1988. Relationship of time and strain to egg shell quality and
2095	hatchability in broiler breeders. Poult. Sci. 67:538-543.
2096	
2097	Cain, J. R., and W. O. Wilson. 1971. Multichannel telemetry system for
2098	measuring body temperature: circadian rhythms of body temperature,
2099	locomotor activity and oviposition in chickens. Poult. Sci. 50:1437-1443.
2100	
2101	Canadian Council on Animal Care. 1993. Guide to the Use of Experimental
2102	Animals. Vol. 1. Can. Counc. Anim. Care, Ottawa, Ontario, Canada.
2103	
2104	Cave, N. A. 1981. Effect of diurnal programs of nutrient intake on the
2105	performance of broiler breeder hens. Poult. Sci. 60:1287-1292.
2106	r
2107	de Avila, V. S., A. M. Penz Jr., P.A.R. de. Brum, P. S. Rosa, A. L. Guidoni, and
2108	E. A. P. de Figueiredo. 2003. Performance of female broiler breeders
2109	submitted to different feeding schedules. Braz. J. Poult. Sci. 5:197-202.
2110	č
2111	de Jong, I. C., S. van Voorst, D. A. Ehlhardt, and H. J. Blokhuis. 2002. Effects of
2112	restricted feeding on physiological stress parameters in growing broiler
2113	breeders. Br. Poult. Sci. 43:157-168.
2114	
2115	Farmer, M., Sr. D. A. Ronald, and M. K. Eckman. 1983. Calcium metabolism in
2116	broiler breeder hens. 2. The influence of time feeding on calcium status of
2117	the digestive system and egg shell quality in broiler breeders. Poult. Sci.
2118	62:465-471.
2119	
2120	Fronda, F. M. 1921. A comparative study of the body temperature of the different
2121	species and some representative breeds of poultry- a preliminary report.
2122	Poult. Sci. 1:16-22.

2123 2124	Gibson, L. C., J. L. Wilson, and A. J. Davis. 2008. Impact of feeding program after light stimulation through early lay on the reproductive performance
2124	of broiler breeder hens. Poult. Sci. 87:2098-2106.
2125	of broner breeder nens. Fourt. Sci. 87.2098-2100.
	Harms, R. H. 1991. The influence of changing time of feeding on performance of
2127 2128	broiler breeder hens. Poult. Sci. 70:1695-1698.
2129	
2130 2131	Heywang, B.W. 1938. Effects of some factors on the body temperature of hens. Poult. Sci. 17:317-323.
2132	
2133	Hocking, P. M., M. H. Maxwell, and M. A. Mitchell. 1996. Relationship between
2134	the degrees of feed restriction and welfare indices in broiler breeder
2135	females. Br. Poult. Sci. 37:263-278.
2136	
2137	Kadono, H., E. L. Besch, and E. Usami. 1981. Body temperature, oviposition, and
2138	food intake in the hen during continuous light. J. Appl. Physiol. 51:1145-
2139	1149.
2140	
2141	Kadono, H., and E. Usami. 1983. Ultradian rhythm of chicken body temperature
2142	under continuous light. Jpn. J. Vet. Sci. 45:401-405.
2143	
2143	Khalil, A. M., K. Matsui, and K. Takeda. 2004. Diurnal and oviposition-related
2144	changes in heart rate, body temperature and locomtor activity of laying
2145	hens. Anim. Sci. J. 75:169-174.
2147	
2148	Lacey, B., T. K. Hamrita, M. P. Lacy, and G. L. Van Wicklen. 2000. Assessment
2149 2150	of poultry deep body temperature responses to environmental temperature and relative humidity using an on-line telemetry system. J. Trans. ASAE.
2150	43:717-721.
2152	
2152	Lewis, P. D., D. Backhouse, and R. M. Gous. 2004. Photoperiod and oviposition
2154	time in broiler breeders. Poult. Sci. 45:561-564.
2155	
2155	Lewis, P. D., and G. C. Perry. 1988. Effect of a single or double daily allocation
2157	of food on shell weight and oviposition time of broiler breeder hens.
2158	Proceedings of the 4th International Poultry Breeders' Conference. Ayr.
2150	72-78.
2160	72.76.
2160	Musharaf, N. A., and J. D. Latshaw. 1999. Heat increment as affected by protein
2161	and amino acid nutrition. World's Poult. Sci. J. 55:233-240.
2162	
2165	Renema, R. A., and F. E. Robinson. 2004. Defining normal: Comparison of feed
2164	restriction and full feeding of female broiler breeders. World's Poult. Sci.
2165	J. 60:511-525.
2100	$J, 00.J11^{-}J2J,$

2167 2168 2169 2170	Romero, L. F., R. A. Renema, A. Naeima, M. J. Zuidhof, and F. Robinson. 2009. Effect of reducing body weight variability on the sexual maturation and reproductive performance of broiler breeder females. Poult. Sci. 88:445- 452.
2171 2172 2173 2174 2175	Samara, M. H., K. R. E. Robbins, and M. O. Smith. 1996. Interaction of feeding time and temperature and their relationship to performance of the broiler breeder hen. Poult. Sci. 75:34-41.
2175 2176 2177	SAS. 2008. SAS 9.2 © 2002-2008 by SAS Institute, Inc., Cary, NC, USA.
2177 2178 2179 2180 2181 2182	Savory, C. J., L. Kostal, and I. M. Nevison. 2006. Circadian variation in heart rate, blood pressure, body temperature and EEG of immature broiler breeder chickens in restricted-fed and ad libitum-fed states. Br. Poult. Sci. 47:599-606.
2183 2184 2185 2186	Skinner-Noble, D. O., and R. G. Teeter. 2003. Components of feed efficiency in broiler breeding stock: energetics, performance, carcass composition, metabolism, and body temperature. Poult. Sci. 82:1080-1090.
2187 2188 2189 2190	Spradley, J. M., M. E. Freeman, J. L. Wilson, and A. J. Davis. 2008. The influence of a twice-a-day feeding regimen after photostimulation on the reproductive performance of broiler breeder hens. Poult. Sci. 87:561-568.
2191 2192 2193 2194	Taherkhani, R., M. Zaghari, M. Shivazad, and A. Z. Shahneh. 2010. A twice-a- day feeding regimen optimizes performance in broiler breeder hens. Poult. Sci. 89:1692-1702.
2195 2195 2196 2197	van Kampen, M. 1976. Activity and energy expenditure in laying hens: 3. The energy cost of eating and posture. J. Agric. Sci. 87:85-88.
2198 2199 2200 2201	Wilson, H. R., and L. J. Keeling. 1991. Effect of time of feeding on oviposition time and production performance in broiler breeders. Poult. Sci. 70:354-259.
2202 2203 2204 2205	Wilson, H. R., F. B. Mather, R. L. Brigmon, E. L. Besch, V. P. Dugan, and N. Z. Boulos. 1989. Feeding time and body temperature interactions in broiler breeders. Poult. Sci. 68:608-616.
2206 2207 2208 2209	Winget, C. M., E. G. Averkin, and T. B. Fryer. 1965. Quantitative measurement by telemetry of ovulation and oviposition in the fowl. Am. J. Physiol. 209:853-858.

Feeding	Dietary	Photoperiod ³	Core body temperature (°C)					
time ¹	Energy ²		Maximum	Minimum	Mean	Range		
Morning			41.26	39.47	40.25	1.79		
Noon			41.36	39.73	40.37	1.63		
Split			41.27	39.41	40.24	1.86		
Afternoon			41.34	39.74	40.35	1.60		
SEM			0.07	0.11	0.07	0.10		
	2,912		41.32	39.45 ^b	40.23 ^b	1.87 ^a		
	2,786		41.30	39.73 ^a	40.38 ^a	1.57 ^b		
SEM			0.05	0.08	0.05	0.07		
		16L:8D	41.31	39.49	40.29	1.82 ^a		
		24L:0D	41.30	39.69	40.32	1.61 ^b		
SEM			0.05	0.08	0.05	0.07		
Source of variation			Probability					
Feeding time			0.6765	0.0522	0.4166	0.224		
Dietary energy			0.7928	0.0110	0.0198	0.004		
Photoperiod			0.9093	0.0526	0.6981	0.039		
Feeding time* Dietary energy			0.2990	0.8075	0.2949	0.986		
Feeding time* Photoperiod			0.6880	0.3835	0.3609	0.537		
Dietary energy* Photoperiod			0.9379	0.2744	0.4369	0.260		
Feeding time* Dietary energy*			0.2823	0.3555	0.1963	0.694		
Photoperiod								

Table 4.1 Core body temperature (CBT) of broiler breeder hens (44 wk of age)
fed two dietary energy levels at different times, and subjected to standard and
continuous photoperiods.

2213

^{a,b}Means within column with no common superscript are significantly different ($P \le 0.05$), ¹Morning feeding: birds fed entire daily feed allocation at 7:30; Noon feeding: birds fed entire daily feed allocation at 11:30; Split feeding: birds fed 50% daily feed allocation at 7:30 and 50% at 15:30; Afternoon feeding: birds fed entire daily feed allocation at 15:30.

2218 ²High energy= 2,912 kcal/kg; Low energy= 2,786 kcal/kg.

³16L:8D=standard photoperiod; 24L:0D=continuous photoperiod.

Time of day ¹	Feeding time ²			Photoperiod		Dietary energy level		Overall	
	Morning	Noon	Afternoon	Split	Standard	Continuous	High	Low	
	(07:30)	(11:30)	(15:30)	(07:30 &	(16L:8D)	(24L:0D)	(2,912	2,786	
				15:30)			kcal/kg)	kcal/kg	_
					CBT (°C)				
Night	39.73 ^{fB}	39.94 ^{fA}	39.95^{hA}	39.70^{hB}	39.73 ^{hB}	39.93 ^{gA}	39.69 ^{gB}	39.97^{hA}	39.83 ^h
1 h pre-dawn	39.90 ^{eB}	40.10^{eA}	40.11^{fA}	39.84 ^{gB}	39.93 ^{gB}	40.05^{eA}	39.88 ^{fB}	40.09^{gA}	39.99 ^g
Pre-feed	40.18^{dB}	40.40^{dA}	40.45^{dA}	40.15 ^{eB}	40.34 ^{eA}	40.24^{dB}	40.21 ^{eB}	40.37 ^{eA}	40.29 ^e
0-1 h post-feed	40.74^{bB}	40.78^{bB}	40.93^{aA}	40.84^{aAB}	40.88^{bA}	40.78^{aB}	40.80^{b}	40.85^{b}	40.83 ^b
1-2 h post-feed	40.83 ^{aB}	40.98^{aA}	40.88^{bAB}	40.78^{bB}	40.93 ^{aA}	40.81 ^{aB}	40.84^{a}	40.90^{a}	40.87^{a}
2-3 h post-feed	40.78^{bB}	40.93^{aA}	40.65^{cBC}	40.59^{cC}	40.78°	40.69^{b}	40.69 ^{cB}	40.79 ^{cA}	40.74°
>3 h post-feed	40.47^{cB}	40.62^{cA}	40.36^{eB}	40.43^{dB}	40.50^{d}	40.44°	40.37^{dB}	40.57^{dA}	40.47^{d}
1 h pre-dusk	39.96 ^{eB}	40.12^{eA}	40.12^{fA}	39.93 ^{fB}	40.01^{f}	40.05^{e}	39.92^{fB}	40.14^{fA}	40.03^{f}
1 h post-dusk	39.70^{fB}	39.98^{fA}	40.01 ^{gA}	39.66 ^{hB}	39.72^{hB}	39.96 ^{fA}	39.70 ^{gB}	39.97^{hA}	39.84 ^h
SEM	0.03	0.03	0.03	0.03	0.02	0.02	0.02	0.02	0.01
Source of variation					Pro	bability			
Time of day	< 0.0001				< 0.0001		< 0.0001		< 0.0001

2221	Table 4.2 Diurnal core body temperature (CBT) patterns of broiler breeder hens (44 wk of age) fed two dietary energy
2222	levels at different times, and subjected to standard and continuous photoperiods.

^{A-C}Means across rows within treatment with no common superscript are significantly different ($P \le 0.05$).

^{a-h}Means within column with no common superscript are significantly different ($P \le 0.05$).

¹Night= time when lights were off (22:30 to 04:30: excluding post-dusk and pre-dawn); 1 h pre-dawn= one hour time period

before lights on (at 05:30); Pre-feed= time period from lights on to feeding time; 0 to 1 h post-feed= first hour after feed; 1 to 2 h

2227 post-feed= second hour after feed; 2 to 3 h post feed= third hour after feed; >3 h post-feed= remainder of the light period

following the third hour post-feeding; excluding pre-dusk; 1 h pre-dusk= one hour time period before lights off (at 21:30); 1 h
 post-dusk= first hour of darkness after lights off.

²Morning feeding: birds fed entire daily feed allocation at 07:30; Noon feeding: birds fed entire daily feed allocation at 11:30;

2231 Split feeding: birds fed 50% daily feed allocation at 07:30 and 50% at 15:30; Afternoon feeding: birds fed entire daily feed

2232 allocation at 15:30.

2233	Table 4.3 Oviposition time and production performance of broiler breeder hens
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2234 (44 wk of age) fed two dietary energy levels at different times, and subjected to

FeedingDietaryPhotoptime1energy2eriod3	ADFI (g/d)	ADG (g/d)	Egg production (%)	Egg wt (g)	Oviposition (h)
Mornin	138.13	-16.51	69.79	64.36	10:55 ^b
g Noon	138.13	1.99	69.79	63.52	11:32 ^{ab}
Split	137.98	-3.93	70.31	65.60	12:00 ^a
Aftern	138.17	5.60	72.92	64.67	11:26 ^{ab}
oon SEM	0.70	5.98	3.47	0.77	0:15
2,912	136.38 ^b	-8.97	70.31	64.20	11:29
2,786	139.83 ^a	2.54	71.09	64.87	11:24
SEM	0.51	4.22	2.45	0.66	0:10
16L:8D	137.46	-0.40	72.92	64.33	11:13
24L:0D	138.75	-6.02	68.49	64.75	11:41
SEM	0.54	4.22	2.45	0.85	0:10
Source of variation			-Probability		
Feeding time	0.9973	0.05	08 0.9058	0.0860	0 0.0405
Dietary energy	0.0001	0.34	83 0.8220	0.242	1 0.7436
Photoperiod	0.1611	0.05	59 0.2035	0.7484	4 0.0673
Feeding time*Dietary energy	0.9973	0.26	34 0.8532	0.5259	9 0.8164
Feeding time*Photoperiod	0.9983	0.61	18 0.2302	0.5042	2 0.4210
Dietary energy*Photoperiod	0.7367	0.43	38 0.7078	0.9994	4 0.4442
Feeding time*Dietary energy*Photoperiod	0.9983			0.328	

2236 ^{a,b}Means within column with no common superscript are significantly different ($P \le 0.05$),

¹Morning feeding: birds fed entire daily feed allocation at 07:30; Noon feeding: birds fed entire daily feed allocation at 11:30; Split feeding: birds fed 50% daily feed allocation at 07:30 and 50% at 15:30; Afternoon feeding: birds fed entire daily feed allocation at 15:30.

2241 ²High energy= 2,912 kcal/kg; Low energy= 2,786 kcal/kg

³16L:8D=standard photoperiod; 24L:0D=continuous photoperiod

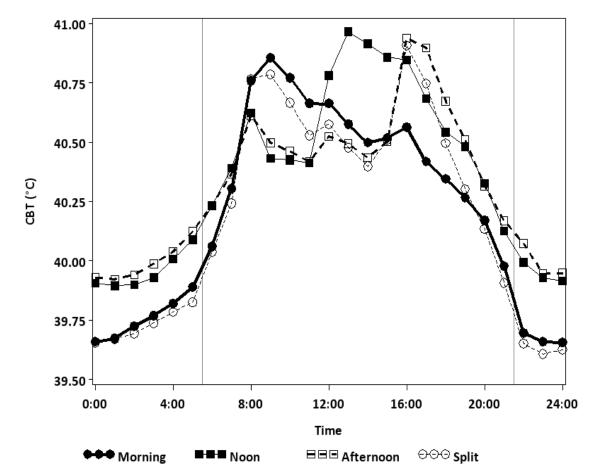
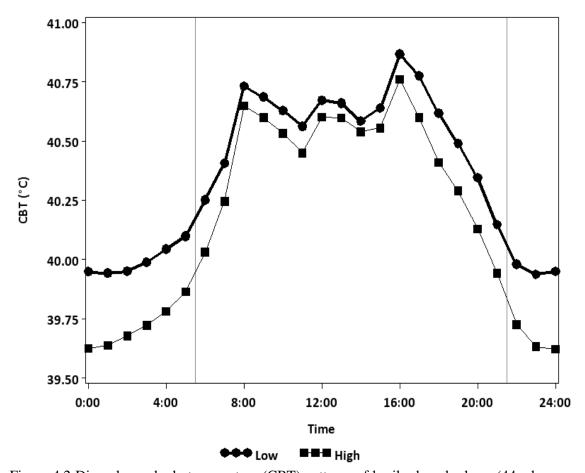


Figure 4.1 Diurnal core body temperature (CBT) patterns of broiler breeder hens (44 wk of age: 7 d) in different feeding times. Y axis indicates CBT in degree Celsius and X axis indicates time in hour.

Morning feeding: birds fed entire daily feed allocation at 07:30; Noon feeding: birds fed entire daily feed allocation at 11:30; Split feeding: birds fed 50% daily feed allocation at 07:30 and 50% at 15:30; Afternoon feeding: birds fed entire daily feed allocation at 15:30. Vertical reference lines in the graph indicate lights on at 05:30 and lights off at 21:30.

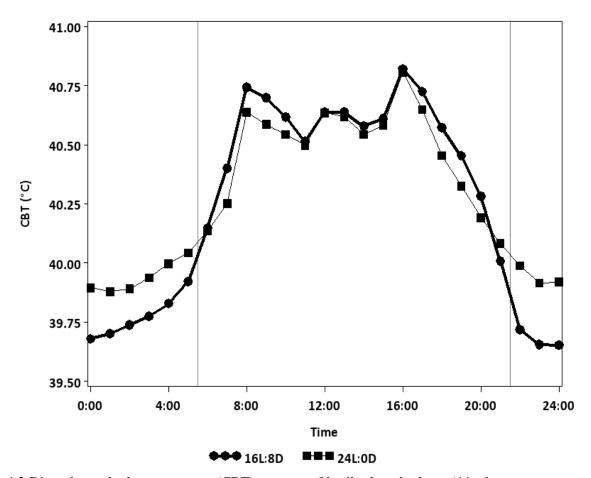
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2254 2255

Figure 4.2 Diurnal core body temperature (CBT) patterns of broiler breeder hens (44 wk of age: 7 d) fed high and low energy diets. Y axis indicates CBT in degree Celsius and X 2256

2257 axis indicates time in hour. Vertical reference lines in the graph indicate lights on at 05:30 2258 and lights off at 21:30.



2260

Figure 4.3 Diurnal core body temperature (CBT) patterns of broiler breeder hens (44 wk

of age: 7 d) in standard (16L:8D) and continuous (24L:0D) photoperiods. Y axis indicates

2263 CBT in degree Celsius and X axis indicates time in hour. Vertical reference lines in the

graph indicate lights on at 05:30 and lights off at 21:30.

2266 CHAPTER 5: ENERGETIC EFFICIENCY, PRODUCTION 2267 PERFORMANCE AND CORE BODY TEMPERATURE DYNAMICS OF 2268 BROILER BREEDER HENS IN CAGE VERSUS FREE-RUN HOUSING 2269 SYSTEMS

ABSTRACT: A study was conducted using 172 Ross 708 broiler breeder hens 2270 2271 from 25 to 41 wk of age to determine the ADFI, ADG, egg production, egg 2272 weight, feed efficiency, and core body temperature (CBT) dynamics in cage versus free-run housing systems. A total of 140 hens were randomly distributed 2273 among 4 free-run pens within two chambers (n=35 birds per pen). The remaining 2274 2275 32 birds were randomly allocated to individual cages in two chambers (n=16 birds 2276 per chamber). Twenty-four temperature sensor-implanted broiler breeder hens were equally distributed among six locations (two chambers and four pens), 2277 2278 which transmitted CBT every 10 min. Birds were provided the same diet under a constant housing temperature (19°C) and relative humidity (60%). Egg production 2279 did not differ between the cage and free-run housed broiler breeder hens. 2280 2281 However, egg weight and daily feed intake was higher in free-run housed hens than in caged hens. The maximum and minimum CBT were also higher in the 2282 2283 free-run hens. The highest diurnal CBT was recorded in the first hour after feeding, averaging 41.2°C and 41.4°C in caged and free-run hens, respectively. In 2284 free-run hens, CBT at night was 0.3°C higher than in the caged hens. Free-run 2285 birds expended 17.2% more energy for maintenance than cage housed birds, 2286 translating into a 17.0% higher feed intake. The current study provides 2287 2288 information about feed consumption and energy requirements that can contribute to appropriate interpretation of cage based breeder nutrition research to free-runbirds.

2291 Keywords: Productive traits, energy requirement, diurnal CBT pattern, ADFI,2292 housing systems, broiler breeders.

2293 **5.1. INTRODUCTION**

Commercially, free-run housing systems are popular for broiler breeders 2294 because these systems allow chickens to mate naturally and reduce the cost of 2295 managing reproduction. Broiler breeder research is often done in cages. 2296 2297 Application of cage housed broiler breeder research to commercial free-run housing, therefore, may not translate directly. In free-run housing, broiler breeders 2298 are feed restricted and usually feed is supplied daily in the morning. Competition 2299 for a limited amount of feed results in poor BW uniformity, contributing to poor 2300 production and egg quality (Petitte et al., 1982). In laying hens, cages facilitate 2301 even feed distribution, easier determination of sick chickens through individual 2302 2303 observation, clean egg collection, less feed consumption and the maintenance of BW uniformity (Farooq et al., 2002). 2304

Hatching egg weight is important because it increases subsequent broiler weight (Vieira and Moran, 1998). Many researchers (Petitte et al., 1982; Mohan et al., 1991; Anderson and Adams, 1994; Leyendecker et al., 2001) noted that egg weight was higher in cage housing systems than in free-run housing systems. In contrast to those studies, Tumova and Ebeid (2005); Singh et al. (2009) recorded higher egg weight with higher feed intake in free-run housed layer chickens than in caged layers. Other researchers (Basmacioglu and Ergul, 2005; Thomas and

Ravindran, 2005) reported that egg weight was not influenced by the housing
system. The energy requirement for maintenance was higher in free-run hens than
in caged hens, and therefore feed intake being higher in free-run systems, and CP
being consumed at higher rates that may contribute to increase egg weight.

2316 Fertility and hatchability are also indicators of breeder performance, and 2317 very important for sustainable production systems. Several researchers (Fuquay and Radden, 1980; Petitte et al., 1982; Petitte et al., 1983; Leeson and Summers, 2318 1985) have reported inconsistent fertility and hatchability rates due to housing 2319 2320 type. High rates of fertility and hatchability can be achieved in both systems, but good management practice for both natural mating and artificial insemination are 2321 2322 important for success. In addition, efficient technical know-how of artificial 2323 insemination is an important for success.

The energy requirement for maintenance in birds is also an important 2324 2325 difference related to housing systems for broiler breeders. Anderson and Adams 2326 (1994); Muthusamy and Viswanathan (1998); Farooq et al. (2002) reported that daily energy intake was 25 kcal/bird higher in full-fed, free-run commercial layers 2327 2328 than in caged layers. In broiler breeders, Rabello et al. (2004) suggested that freerun hens required more energy, likely for increased activity level, than caged 2329 hens. A substantial amount of research has been conducted to investigate the 2330 2331 effects of cage versus free-run housing systems on egg production, egg weight, and energy requirement of birds, but inconsistent conclusions were drawn. The 2332 2333 CBT dynamics of feed restricted broiler breeder hens has not been studied in cage

versus free-run housing systems, but could be a tool for better understanding ofphysiological or metabolic status of feed restricted broiler breeder hens.

The objective of the current study was to determine the effect of cage versus free-run housing systems on energy requirements, CBT dynamics, and production efficiency in broiler breeder hens.

2339 **5.2. MATERIALS AND METHODS**

2340 **5.2.1. Animal Care and Approval**

The current experiment was compliant with the Guide to the Care and Use of Experimental Animals (Canadian Council on Animal Care, 1993) and was approved by the Animal Care and Use Committee for Livestock of the University of Alberta.

2345 **5.2.2. Experimental Design**

The effect of cage and free-run housing systems on average daily feed intake, egg production, egg weight, feed efficiency, energetic efficiency, and CBT dynamics in broiler breeder hens was examined using two treatments (caged and free-run) in a completely randomized design. For all variables except CBT where the experimental unit was the individual hen in all treatments, the experimental units were the individual hen in the cage housing system and the pen in the freerun housing system.

2353 **5.2.3. Stocks and Management**

At hatch, a total of 800 Ross 708 pullets were individually identified by bar-coded neck tags (Heartland Animal Health, Fair Play, MO), weighed and randomly allocated to 1 of 8 environmental chambers. Each chamber was divided

into four floor pens with 25 pullets per pen (5 pullets/ m^2) in a climate controlled 2357 2358 facility under recommended brooding temperature. At 7 d of age, each pullet was also tagged with matching bar coded wing bands on each wing. Feed was 2359 2360 provided ad libitum for the first 14 d of age. From 15 d, pullets were feed restricted to maintain breeder recommended BW target (Aviagen, 2007). Pullets 2361 2362 were reared in the pens until 20 wk of age. The photoperiod was 23L:1D for the first 3 d and 8L:16D from 4 d to 20 wk of age. The photoperiod was changed to 2363 12L:12D at 21 wk, and the light was increased by one hour per week until 16L:8D 2364 at 25 wk of age. The light intensity was 60 lux. At 21 wk of age, 32 pullets were 2365 placed in individual laying cages (0.135 m²/hen) in temperature-controlled 2366 environmental chambers, and 140 pullets were placed in 4 free-run pens within 2 2367 chambers (35 hens/pen; 0.2025 m²/hen). Environmental temperature and relative 2368 humidity in the chambers were set at 19°C and 60 %, respectively, during the 2369 experimental period. Photoperiod was 16L:8D with lights on at 05:30. Water was 2370 2371 supplied *ad libitum* using nipple drinkers. Feed was supplied to birds at 07:30 except weighing days. The time of feeding was recorded daily. Data loggers 2372 2373 (Microlog EC650, Fourier Systems, New Albany, IN) were used to record actual room temperature at feeder height. Wheat-and soybean-based diets in mash form 2374 were given: Starter (2,900 kcal ME, 19% CP) from 0 to 2 wk; Grower (2,700 kcal 2375 2376 ME, 15% CP) from 3 to 20 wk; Breeder (2,912 kcal ME, 16.4% CP) from 25 to 41 wk of age (Appendix A). 2377

2378 **5.2.4 Surgical Implantation Temperature Sensors**

At 13 wk of age, 24 birds had temperature telemetry devices surgically implanted into the abdominal cavity (see chapter 2 for details). The implants were approximately the size of an 'AA' battery. Implanted birds were randomly distributed among the four pens and two chambers (4 birds each).

2383 **5.2.5. Data Collection**

Feed allocation, egg production, and egg weight were recorded from 25 to 2384 41 wk of age. Individual (caged) or group (free-run) BW were recorded twice per 2385 2386 week. Average daily gain (ADG) was calculated from the difference between 2387 initial and final BW for each weighing interval. The temperature sensors transmitted CBT of implanted birds at approximately 10 min intervals. Eggs were 2388 collected daily at 15:00, and weighed individually. The total eggs variable was 2389 defined as all eggs including broken, double yolk and deformed; normal eggs as 2390 2391 total eggs minus broken, double yolked and deformed eggs; and settable eggs as 2392 normal eggs 52 g or greater in weight. Feed efficiency was measured based on the ratio of average daily feed intake per hen and average daily egg mass per hen 2393 2394 (Flock, 1998). The following terminologies were used in this thesis: Maximum CBT was the average highest daily body temperature of individual hens. Mean 2395 CBT was the average daily body temperature of individual hens. Minimum CBT 2396 2397 was the average daily lowest body temperature of individual hens. Range of CBT was the average difference between the daily highest and lowest body 2398 2399 temperatures of an individual hen. Night was defined as the time period when 2400 lights turned off excluding post-dusk and pre-dawn (22:30 to 04:30), 1 h pre-dawn

2401 was the one hour time period before lights turned on; pre-feed was the time period 2402 from lights on to feeding time; 0-1 h post-feed was the first hour time period after feeding; 1-2 h post-feed was the second hour time period after feeding; 2-3 h post 2403 2404 feed was the third hour time period after feeding; 3-4 h post-feed was the fourth hour time period after feeding; >4 h post-feed was the remainder of time period 2405 that the lights were on following the fourth hour post-feed, excluding pre-dusk; 1 2406 h pre-dusk was the one hour time period before lights turned off; 1 h post-dusk 2407 was the one hour time period after lights were turned off. 2408

2409 **5.2.6. Statistical Analysis**

The Mixed procedure of SAS 9.2 (SAS Institute, Cary, NC) was used to compare the treatment means using Tukey's test with a significance level of $P \le$ 05. Housing system was considered as a fixed effect within all dependable variables and date was used as a random effect. Nonlinear mixed procedure of SAS 9.2 was used to develop an energy partitioning (energetic efficiency) model

2415 for caged and free-run broiler breeder hens. The model was in the form of;

2416 MEI = $((a+u) + c^{*}Te)^{*}BW^{0.35} + g^{*}padg - (ng)^{*}nadg + e^{*}eggmass + \varepsilon$,

Expected energy requirement for maintenance (a + u); where $u \sim N(0, Vu)$ associated with each hen in cage and each pen in free-run housing system was estimated from 25 to 41 wk of age using a mixed nonlinear model, Te = environmental temperature; padg= positive average daily gain; nadg= negative average daily gain; eggmass= average daily eggmass; \mathcal{E} = error.

2422 5.3. RESULTS AND DISCUSSION

2423 **5.3.1. Feed Efficiency**

Average daily feed intake and ADG were higher in free-run housed broiler 2424 2425 breeder hens compared to caged hens (Table 5.1). This was in agreement with 2426 Farooq et al. (2002) who reported that feed intake was higher in full-fed layers in 2427 a free-run system than caged hens. The higher feed intake in free-run housed broiler breeder hens did not affect egg production (Table 5.2). The higher feed 2428 intake might be due to increased activity level of hens in the spacious free-run 2429 2430 housing system compared to caged hens (Rabello et al 2004). Egg weight was heavier in free-run hens than in caged hens, higher intake of feed as well as CP 2431 may have contributed to increased egg weight in free-run hens (Singh et al., 2432 2009). Feed, ME, and CP efficiency decreased in free-run hens compared to caged 2433 hens. Possibly because free-run hens required higher energy for higher activity 2434 2435 level in large floor area compared to caged hens. Farooq et al. (2002) reported that 2436 feed efficiency increased as egg production was higher with lower feed intake in caged hens than in free-run hens. The ADFI increased by 17% in free-run hens 2437 2438 compared to caged hens in maintaining target BW. Some of this feed contributed 2439 to higher ADG, but most was used to fuel activity.

2440 **5.3.2. Egg Production in Cage versus Free-run Housing Systems**

Total, normal and settable egg production was similar between cage and free-run housed broiler breeder hens (Table 5.2). This was in agreement with Petitte et al. (1982); Roll et al. (2009), who also reported a similar egg production in cage and free-run housed commercial hens. However, Anderson and Adams (1994) stated that the normal egg production was higher in caged hens compared
to free-run housed broiler breeder hens. In commercial layers, Yousaf and Ahmed
(2006) also reported higher egg production in caged hens compared to free-run
hens. Good management systems were provided to both caged and free-run hens.
There is no biological reason that housing types (cage vs free-run) should
influence egg production.

Egg weight was higher in free-run hens than in caged hens (Table 5.1). 2451 This result was in agreement with Petitte et al. (1982); Anderson and Adams 2452 2453 (1994); Pistekova et al. (2006), who reported that egg weight was heavier in free-2454 run housed layer chickens compared to caged layer chickens. Conversely, Yakubu et al. (2007) reported that egg weight was higher in caged hens than in free-run 2455 2456 hens. Several researchers (Basmacioglu and Ergul, 2005; Yousaf and Ahmed, 2006; Thomas and Ravindran, 2005; Zemkova et al., 2007; Roll et al., 2009) 2457 indicated that the housing system did not affect egg weight in layer chickens. In 2458 2459 the current study, BW of broiler breeder hens was higher in free-run hens than caged hens. In addition, ADFI as well as other nutrients intake including CP was 2460 2461 higher in free-run hens than in caged hens and those excess nutrients may contribute to heavier eggs. This was in agreement with the result of Halaj et al. 2462 (1998); Basmacioglu and Ergul (2005), who reported that egg weight was 2463 2464 influenced by nutrition and age.

5.3.3. CBT of Broiler Breeder Hens in Different Housing Systems

The daily average maximum and minimum CBT were higher in free-run broiler breeder hens than in caged hens (Table 5.3), which might be increased by

2468 higher daily feed intake (feed metabolism) and possibly, due to increased activity 2469 level. The mean CBT and the range of CBT were similar in both cage and freerun housed broiler breeder hens. The range of CBT in caged and free-run broiler 2470 2471 breeder hens was from 39.8 to 41.6°C and 40.0 to 41.8°C respectively. Similarly, Savory et al. (2006) reported that CBT in broiler breeder hens ranged from 39.6 to 2472 2473 41.2°C. However, Deeb and Cahaner (1999) reported the CBT ranged from 40.4 to 41.6°C in full fed chickens. In the current study, the result may indicate that 2474 heat increment in broiler breeder hens did not differ in various housing systems to 2475 2476 maintain a relatively constant CBT. However, when the temperature difference 2477 between the surface body and the environment increases, birds dissipate heat to the environment and they require more feed to increase the rate of metabolic heat 2478 production to maintain CBT (National Research Council, 1981). 2479

2480 5.3.4. Diurnal CBT Patterns

Diurnal CBT patterns were closely related between caged and free-run 2481 2482 hens. The CBT increased by 0.2°C one hour before lights on and it continued increasing after lights on until 1-2 h post-feeding time (Table 5.4). This increased 2483 2484 CBT may be associated with the increased activity level 1 h prior to lights on possibly due to biological response of birds and after lights turned on, birds' 2485 response to sudden visual and auditory stimuli of lights on and sounds by 2486 2487 attendants respectively (Richards, 1971). This was in agreement with the results of Lacey et al. (2000), who reported CBT increased before lights on, rising till 2488 noon in full fed birds. Moreover, Kadono and Besch (1978) reported that CBT 2489 2490 started to increase 2 to 4 h before lights on in full fed chickens. The highest

2491 diurnal CBT was recorded at the 0-1 h post-feed (one hour after feeding) followed 2492 by 1-2 h post-feed, 2-3 h post-feed, 3-4 h post-feed, > 4 h post-feed, pre-feed, 1 h pre-dusk, 1 h pre-dawn, night and 1 h post-dusk of day (Table 5.4). The overall 2493 2494 highest CBT (41.3°C) was recorded at the feeding time (0-1 h post feed) and the lowest CBT (40.1°C) was at the 1 h post-dusk (Table 5.4). In the current study, 2495 2496 the CBT increased at the day time due to light, activity and feed metabolism (Khalil et al., 2004). The CBT gradually decreased in both caged and free-run 2497 broiler breeder hens from 1-2 h post-feed until lights off. Presumably, it may be 2498 2499 due to reduced rate of metabolic heat production. Kadono and Besch (1978) 2500 suggested that CBT decreased 2 to 4 h before lights off in full-fed chickens. The CBT dropped quickly after lights turned off, possibly due to sharply reduced 2501 2502 activity level (Cain and Wilson, 1974; Khalil et al., 2004). The diurnal CBT was higher in the day time period than in the night time period in both caged and free-2503 run broiler breeder hens. These results were supported by several researchers 2504 2505 (Lacey et al., 2000; Fronda, 1921; Heywang, 1938; Winget et al. 1965); they reported that CBT was higher at day time periods than at night time periods. In 2506 2507 the current study, the peak CBT of broiler breeder hens during 0-1 h post-feed (feeding time) may be the combined effects of nutrients metabolism and feeding 2508 activity. Therefore, feed intake and activity levels played a major role for diurnal 2509 2510 CBT dynamics in broiler breeder hens.

The diurnal CBT was higher at night, 1 h pre-dawn, 0-1 h post-feed, 1-2 h post-feed, 1 h pre-dusk, and 1 h post-dusk in free-run hens compared to caged hens (Table 5.4). In the current study, CBT patterns were similar between cage

2514 and free-run housed broiler breeder hens. However, the diurnal CBT was very 2515 close between free-run and caged hens during the day time period, and CBT was 2516 higher during night time period in free-run hens than in caged hens (Figure 5.1). It 2517 may be due to extra activity of broiler breeder hens at night in the free-run housing system. This activity possibly increases the rate of basal metabolism 2518 2519 resulting in higher heat increment in the body (Boshouwers and Nicaise, 1985) Free-run hens also increased insulation by sleeping together on the floor resulting 2520 in reduced heat dissipation to the environment. However, caged birds may lose 2521 2522 more heat through convection (air flow surrounding the bird) and conduction (birds contact with metal wires of the cage), and that could be the reason of lower 2523 CBT at night. 2524

2525 **5.3.5. Energetic Efficiency**

The energy requirement for maintenance was determined for caged and 2526 free-run broiler breeder hens. Body weights of broiler breeder hens were higher in 2527 2528 free-run hens than in caged hens (Table 5.1). The mean residual feed intake (RFI) did not differ between cage and free-run broiler breeder hens (Table 5.5). 2529 2530 Swennen et al. (2007) suggested that a high RFI indicated less efficient resulting in a higher feed intake in cockrels and a greater postprandial thermogenesis. The 2531 RME_m was higher in the free-run hens compared to cage hens. This result 2532 2533 demonstrates that broiler breeder hens in the free-run housing system were less efficient than in cage housing system. The concept of RME_m was defined as the 2534 2535 residual of estimated maintenance requirement, and RFI as the residual of 2536 predicted feed intake (Romero et al., 2009). In the current study, the energy

2537 requirement for maintenance in free-run housed broiler breeder hens was 17.2% (kgBW^{0.35}) higher than in cage housed broiler breeder hens (Table 5.5). This 2538 higher energy expenditure was possibly due to extra activitiy level in the large 2539 2540 floor area in the free-run housing system. This was in agreement with Rabello et al. (2004), who reported that the requirement for ME_m was 21.8 % higher in free-2541 2542 run hens compared to caged breeder hens. BW of broiler breeder hens was slightly higher in free-run hens compared to caged hens. However, in the current 2543 study, free-run broiler breeder hens were less efficient than caged hens, possibly 2544 2545 free-run hens expended more energy for activity level.

A similar range of CBT in both cage and free-run housing system may 2546 indicate that broiler breeder hens were energy balanced (total heat 2547 2548 production=total heat loss) in either cage or free-run housing systems. In addition, diurnal CBT pattern was closely related in both the cage and the free-run housed 2549 broiler breeder hens. The ADFI and the ME_m was 17.0 % and 17.2 % higher in 2550 2551 free-run housed broiler breeder hens compared to cage housed broiler breeder 2552 hens due to higher activity levels in spacious free-run housing systems. Egg 2553 production was similar in both the cage and the free-run housing system. Heavier egg weight and lower feed efficiency was observed in free-run hens compared to 2554 caged hens. Birds raised in cages were more efficient than free-run hens. Feed 2555 2556 allocation decision needs to be readjusted, when cage research data is applied to commercial free run housing systems. 2557

5.5. REFERENCES

2560	
2561	Anderson, K. E., and A. W. Adams. 1994. Effects of cage versus floor housing
2562	environments and cage floor mesh size on bone strength, fearfulness, and
2563	production of single comb white leghorn hens. Poult. Sci. 73:1233-1240.
2564	
2565	Aviagen. 2007. Ross 708 parent stocks performance objectives. Aviagen Ltd.
2566	www.aviagen.com/docs/Ross 708 PS Performance Objectives.pdf.
2567	Accessed June 2010.
2568	
2569	Basmacioglu, H., and M. Ergul. 2005. Research on the factors affecting
2570	cholesterol content and some other characteristics of eggs in laying hens-
2571	the effect of genotype and housing system. Turk. J. Vet. Anim. Sci.
2572	29:157-164.
2573	
2574	Boshouwers, F. M. G., and E. Nicaise. 1985. Automatic gravimetric calorimeter
2575	with simultaneous recording of physical activity for poultry. Br. Poult. Sci.
2576	26:531-541.
2577	
2578	Canadian Council on Animal Care. 1993. Guide to the Use of Experimental
2579	Animals. Vol. 1. Can. Counc. Anim. Care, Ottawa, Ontario, Canada.
2580	
2581	Cain, J. R., and W. O. Wilson. 1974. The influence of specific environmental
2582	parameters on the circadian rhythms of chickens. Poult. Sci. 53:1438-
2583	1447.
2584	
2585	Deeb, N., and A. Cahaner. 1999. The effects of naked neck genotypes,
2586	environmental temperature, and feeding status and their interactions on
2587	body temperature and performance of broilers. Poult. Sci. 78:1341-1346.
2588	
2589	Fuquay, J. I., and J. A. Renden. 1980. Reproductive performance of broiler
2590	breeders maintained in cages or on floors through 59 weeks of age. Poult.
2591	Sci. 59:2525-2531.
2592	
2593	Farooq, M., M. A. Mian, F. R. Durrani, and M. Syed. 2002. Feed consumption
2594	and efficiency of feed utilization by egg type layers for egg production.
2595	Livestock Research for Rural Development. 14(1). Published by
2596	Fundación CIPAV, Cali, Colombia.
2597	http://www.lrrd.org/lrrd14/1/cont141.htm. Accessed August 2011.
2598	
2599	Flock, D. K. 1998. Genetic-economics aspects of feed efficiency in laying hens.
2600	World's Poult. Sci. J. 54:225-239.
2601	

2602	Fronda, F. M. 1921. A comparative study of the body temperature of the different
2603	species and some representative breeds of poultry- a preliminary report.
2604	Poult. Sci. 1:16-22.
2605	
2606	Halaj, M., J. Benkova, and J. Baumgartner. 1998. Parameters of hen egg quality in
2607	various breeds and strains. Czech. J. Anim. Sci. 43:375-378.
2608	
2609	Heywang, B.W. 1938. Effects of some factors on the body temperature of hens.
2610	Poult. Sci.17:317-323.
2611	
2612	Kadono, H., and E. L. Besch. 1978. Telemetry measured body temperature of
2613	domestic fowl at various ambient temperatures. Poult. Sci. 57:1075-1080.
2614	
2615	Khalil, A. M., K. Matsui, and K. Takeda. 2004. Diurnal and oviposition-related
2616	changes in heart rate, body temperature and locomotor activity of laying
2617	hens. Anim. Sci. J. 75:169-174.
2618	
2619	Lacey, B., T. K. Hamrita, M. P. Lacy, and G. L. Van Wicklen. 2000. Assessment
2620	of poultry deep body temperature responses to environmental temperature
2621	and relative humidity using an on-line telemetry system. J. Trans. ASAE.
2622	43:717-721.
2623	
2624	Leeson, S., and J. D. Summers. 1985. Effect of cage versus floor rearing and skip-
2625	a- day versus every feed restriction on performance of dwarf broiler
2626	breeders and their offspring. Poult. Sci. 64:1742-1749.
2627	
2628	Leyendecker, M., H. Hamann, J. Hartung, J. Kamphues, C. Ring, G. Gluender, C.
2629	Ahlers, I. Sander, U. Neumann, and O. Distl. 2001. Analysis of genotype-
2630	environment interactions between layer lines and housing systems for
2631	performance traits, egg quality and bone breaking strength-1st
2632	communication: Performance traits. Zuchtungskunde. 73:290-307.
2633	
2634	Mohan, B., V. Mani, and S. Nagarajan. 1991. Effect of different housing system
2635	on the physical qualities of commercial chicken eggs. Indian J. Poult. Sci.
2636	26:130-131.
2637	
2638	Muthusamy, P., and K. Viswanathan. 1998. Effect of rearing systems on
2639	performance of commercial layers. Indian J. Poult. Sci. 33:264-267.
2640	1
2641	National Research Council. 1981. Effect of environment on nutrient requirements
2642	of domestic animals. Natl. Acad. Press, Washington D. C.
2643	
2644	Petitte, J. N., R. O. Hawes, and R. W. Gerry. 1983. The influence of cage versus
2645	floor pen management of broiler hens on subsequent performance of cage
2646	reared broilers. Poult. Sci. 62:1241-1246.

2647 2648	Petitte, J. N., R. O. Hawes, and R. U. Gerry. 1982. The influence of flock uniformity on the reproductive performance of broiler breeder hens housed in cases and floor page. Doubt. Sei. (1):2166-2171
2649 2650	in cages and floor pens. Poult. Sci. 61:2166-2171.
2650	Pistekova, V., M. Hovorka, V. Vecerek, E. Strakova, and P. Suchy. 2006. The
2652	quality comparison of eggs laid by laying hens kept in battery cages and in
2653	a deep litter system. Czech. J. Anim. Sci. 51:318-325.
2654	Distante C. A. 1071 The significance of showers in the temperature of the ship
2655 2656	Richards, S. A. 1971. The significance of changes in the temperature of the skin and body core of the chicken in the regulation of heat loss. J. Physiol.
2657	216:1-10.
2658	
2659	Roll, V. F. B., R. C. Briz, and G. A. M. Levrino. 2009. Floor versus cage rearing:
2660	effects on production, egg quality and physical condition of laying hens
2661	housed in furnished cages. Ciencia Rural, Santa Maria. 39:1527-1532.
2662	
2663	Rabello, C. B. V., N. K. Sakomura, F. A. Longo, and K. T. de. Resende. 2004.
2664	Effect of the environmental temperature and rearing systems on
2665	metabolizable energy requirements for maintenance of broiler breeder
2666	hens. R. Bras. Zootec. 33:382-390.
2667	
2668	Romero, L. F., M. J. Zuidhof, R. A. Renema, A. Naeima, and F. E. Robinson.
2669	2009. Characterization of energetic efficiency in adult broiler breeder
2670	hens. Poult. Sci. 88:227-235.
2671	
2672	SAS. 2008. SAS 9.2 © 2002-2008 by SAS Institute, Inc., Cary, NC, USA.
2673	
2674	Savory, C. J., L. Kostal, and I. M. Nevison. 2006. Circadian variation in heart
2675	rate, blood pressure, body temperature and EEG of immature broiler
2676	breeder chickens in restricted-fed and ad libitum-fed states. Br. Poult. Sci.
2677	47:599-606.
2678	
2679	Singh, R., K. M. Cheng, and F. G. Silversides. 2009. Production performance and
2680	egg quality of four strains of laying hens kept in conventional cages and D_{1} by D_{2} is a 256 264
2681	floor pens. Poult. Sci. 88:256-264.
2682	
2683	Swennen, Q., P. J. Verhulst, A. Collin, A. Bordas, K. Verbeke, G. Vansant, E.
2684	Decuypere, and J. Buyse. 2007. Further investigations on the role of diet-
2685	induced thermogenesis in the regulation of feed intake in chickens:
2686	Comparison of adult cockerels of lines selected for high or low residual
2687	feed intake. Poult. Sci. 86:1960-1971.
2688	
2689	Thomas, D.V., and V. Ravindran. 2005. Comparison of layer performance in cage
2690	and barn systems. J. Anim. Vet. Adv. 4:554-556.
2691	

2692	Tumova, E., and T. Ebeid. 2005. Effect of time of oviposition on egg quality
2693	characteristics in cages and in a litter housing system. Czech J. Anim. Sci.
2694	50:129-134.
2695	
2696	Vieira, S. L., and E. T. Moran, JR. 1998. Broiler yields using chicks from egg
2697	weight extremes and diverse strains. J. Appl. Poult. Res.7:339-346.
2698	
2699	Winget, C. M., E. G. Averkin, and T. B. Fryer. 1965. Quantitative measurement
2700	by telemetry of ovulation and oviposition in the fowl. Am. J. Physiol.
2701	209:853-858.
2702	
2703	Yakubu, A., A. E. Salako, and A. O. Ige. 2007. Effect of genotype and housing
2704	system on the laying performance of chickens in different seasons in semi-
2705	humid tropics. Int. J. Poult. Sci. 6:434-439.
2706	
2707	Yousaf, M., and N. Ahmed. 2006. Effects of housing systems on productive
2708	performance of commercial layers following induced molting by
2709	aluminium oxide supplementation. Pak. Vet. J. 26:101-104.
2710	
2711	Zemkova, L., J. Simeonovova, M. Lichovníkova, and K. Somerlíkova. 2007. The
2712	effects of housing systems and age of hens on the weight and cholesterol
2713	concentration of the egg. Czech J. Anim. Sci. 52:110-115.
2714	

2715	Table 5.1 Body weight, average daily feed intake, average daily gain and feed
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2716	efficiency of broiler breeder	s (25 to 41 wk)) in cage and free-run	housing systems.

Housing system	Body weight (kg)	Average daily feed intake (g)	Average daily gain (g)	Average egg weight (g)	Feed:Egg (g/g)	ME:Egg (kcal/g)	CP:Egg (g/g)
Cage	3.31 ^b	133.42 ^b	5.68 ^b	57.38 ^b	3.05 ^b	8.89 ^b	0.50 ^b
Free-run	3.37 ^a	156.06 ^a	9.71 ^a	59.00 ^a	3.77 ^a	10.98 ^a	0.62 ^a
SEM	0.07	1.75	1.28	0.53	0.13	0.38	0.02
Probability	0.0029	<.0001	0.0316	0.0275	0.0047	0.0047	0.0047

^{a-b}Means within column with no common letters are significantly different ($P \le 0.05$)

Feed:Egg (g/g) = average daily feed intake divided average daily egg mass

ME:Egg (kcal/g of egg) = average daily ME intake divided by average daily egg mass CP:Egg (g/g of egg) = average daily CP intake divided by average daily egg mass

2723	Table 5.2 Egg production and egg weight of broiler breeders (25 to 41 wk) in cage
2724	and free-run housing systems.

Housir system	0		al egg ction (%)	ettable roduction %)
Cage	76.38	73.34	6	7.22
Free-ru	un 69.50	69.20	60	6.65
SEM	2.91	2.75	2	4.56
Probab	oility 0.1071	0.156	54 (0.7228

^{a-b}Means within column with no common letters are significantly different ($P \le 0.05$) Total egg production = all eggs including abnormal eggs Normal egg production = total eggs minus abnormal eggs

Settable eggs = normal eggs minus <52g eggs

Housing system	Maximum	Mean	Minimum	Range
			-CBT (°C)	
Cage	41.64 ^b	40.54	39.77 ^b	1.87
Free-run	41.82 ^a	40.62	39.99 ^a	1.83
SEM	0.04	0.04	0.06	0.05
Probability	0.0009	0.1256	0.0096	0.6522

Table 5.3 Core body temperature (CBT) of broiler breeders (21 to 41 wk) in cageand free-run housing systems.

2732 ^{a-b}Means within column with no common letters are significantly different ($P \le 0.05$)

2733 Maximum= daily highest CBT of individual hens

2734 Mean= average of daily CBT of individual hens

2735 Minimum= average of daily lowest CBT of individual hens

2736 Range= average of daily range of difference between the highest and lowest CBT of

2737 individual hens

Time of day ¹	Treatment		Overall
	Cage	Free-run	
		CBT(°C)	
Night	40.00^{iB}	40.30 ^{hA}	40.15 ⁱ
1 h pre-dawn	40.17^{hB}	40.44 ^{gA}	40.30 ^g
Pre-feed	40.68^{f}	40.71 ^f	40.69 ^f
0-1 h post-feed	41.20 ^{aB}	41.35 ^{aA}	41.28 ^a
1-2 h post-feed	41.18 ^{bB}	41.24 ^{bA}	41.21 ^b
2-3 h post-feed	41.09 ^c	41.08 ^c	41.09 ^c
3-4 h post-feed	41.00 ^d	41.00 ^d	41.00 ^d
>4 h post-feed	40.71 ^e	40.71 ^e	40.71 ^e
1 h pre-dusk	40.21 ^{gB}	40.27 ^{iA}	40.24 ^h
1 h post-dusk	39.97 ^{jB}	40.22^{jA}	40.10 ^j
SEM	0.008	0.006	0.005
Probability	< 0.0001	< 0.0001	< 0.0001

Table 5.4 Diurnal core body temperature (CBT) rhythm of broiler breeders (25 to41 wk) in cage and free-run housing systems.

^{A-B}Means across rows with no common letters are significantly different ($P \le 0.05$). 2741 ^{a-j}Means within column with no common letters are significantly different ($P \le 0.05$) 2742 2743 ¹Night= lights off time period (22:30 to 4:30; excluding post-dusk and pre-dawn); 1 h pre-dawn= one hour time period before lights on (at 05:30); Pre-feed= time period from 2744 2745 lights on to feeding time; 0-1 h post-feed= first hour after feed, 1-2 h post-feed= second hour after feed, 2-3 h post feed= third hour after feed, 3-4 h post-feed= fourth hour after 2746 feed, >4 h post-feed= remainder of the lights on period following the fourth hour post-2747 2748 feed, excluding pre-dusk 1 h pre-dusk= one hour time period before lights off, 1 h postdusk= one hour time period after lights off (at 21:30). 2749 2750

Table 5.5 Energetic efficiency and the energy requirement for maintenance of broiler breeders (25 to 41 wk) in cage and free-run housing systems.

2753

Housing system	RFI^1	RME_m^2	ME _m ³
	Kcal of ME/d	kcal of ME/kg ^{0.35}	
Cage	-8.94	-1.29 ^b	234.65 ^b
Free-run	1.35	32.14 ^a	274.93 ^a
SEM	7.80	2.48	1.59
Probability	0.3796	<0.0001	<0.0001

^{a-b}Means within column with no common letters are significant different ($P \le 0.05$)

¹Residual feed intake (RFI) was refered to the diffrence between observed and predicted

2756 ME intake. The predicted MEI was calculated for each hen in cage and each pen in free-

run housing system from 25 to 41 wk of age using the mixed nonlinear model: MEI =

2758 $((a+u) + c^*Te)^*BW^{0.35} + g^*padg - (ng)^*nadg + e^*eggmass + \mathcal{E}$, Te = environmental

temperature; padg= positive average daily gain; nadg= negative average daily gain;
eggmass= average daily eggmass; E= error.

2761 ²Residual maintenance requirement (RME_m). Residual of the regression between ME_m

and MEI for each hen in cage and each pen in free-run housing system: $MEI_m = 218.31 + 0.06*MEI + \varepsilon$;

 $^{3}ME_{m}$ = predicted maintenance requirement (kcal/kg BW^{0.35}); MEI = average ME intake

2765 (kcal/d) from 25 to 41 wk of age. Expected maintenance requirement (a + u); where

 $u \sim N(0, Vu)$ associated with each hen in cage and each pen in free-run housing system was

estimated from 25 to 41 wk of age using a mixed nonlinear model, which is defined in thefirst footnote.

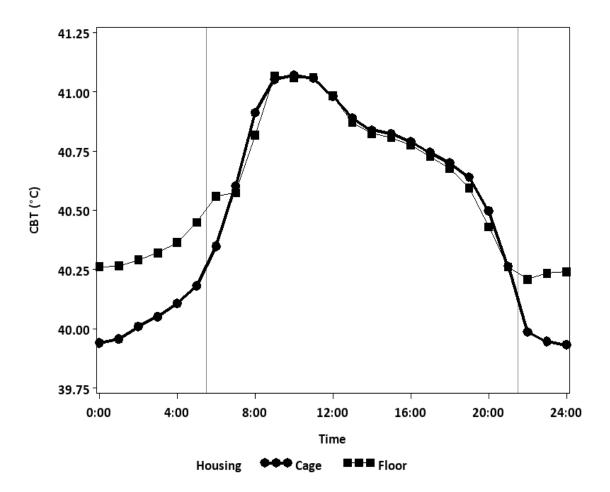


Figure 5.1 Diurnal core body temperature pattern of broiler breeder hens (25 to 41 wk) in

cage and free-run housing system. Y axis indicates CBT in degree Celsius and X axis

2773 indicates time in hour. Vertical reference lines in the graph indicate lights on at 05:30 and

lights off at 21:30.

2777 CHAPTER 6: CORE BODY TEMPERATURE DYNAMICS:

2778 IMPLICATIONS FOR BROILER BREEDER PRODUCTION

2779 **6.1. INTRODUCTION**

2780 Chickens are homoeothermic animals that maintain a relatively narrow 2781 range of core body temperature (**CBT**) from 40.6 to 41.4°C (Deeb and Cahaner, 2782 1999). However, feed-restricted broiler breeders exhibited a range in CBT from 2783 39.6 to 41.2°C (Savory et al. 2006). They are capable of maintaining an energy 2784 balance between heat production and heat loss to keep a relatively constant CBT 2785 in thermoneutral conditions (18 to 27°C; National Research Council, 1981).

When birds are housed in a cold environment (below the lower critical 2786 2787 temperature) and the difference between the surface body temperature and the 2788 environmental temperature increases; birds lose an increasing amount of body heat to the environment. They need more feed to increase the rate of metabolic 2789 heat production to maintain their CBT. In contrast, when birds are housed in a hot 2790 2791 environment (above the upper critical temperature), the difference between the 2792 surface body temperature and the environmental temperature decreases; they are 2793 unable to dissipate adequate body heat to the environment. Above the upper 2794 critical temperature, birds try to dissipate body heat (energy) to the environment 2795 through evaporative cooling to maintain their CBT (Richards, 1971; National Research Council, 1981). Thus, the bird's energy requirement for maintenance 2796 increases in both the lower and the upper critical temperature. Sakomura (2004) 2797 2798 reported that the energy requirement for maintenance is around 80% of total ME 2799 intake for broiler breeder pullets. Romero et al. (2009) reported that the energy

2800 requirement for maintenance is around 70% of total ME intake for broiler breeder 2801 hens. In a cold environment, the large proportion of the energy requirement for maintenance further increases to maintain their CBT. The remaining small 2802 2803 proportion of energy available for growth and production dramatically declines resulting in decreased growth and production, particularly in feed restricted 2804 2805 broiler breeders, where available ME cannot increase due to increased voluntary intake. Efficiency decreases as energy requirement for maintenance to maintain 2806 CBT increases. In the current study, feed efficiency was determined by the ratio 2807 2808 between input (feed intake) and output (BW gain or egg mass) according to Wang and Kim (2011). 2809

Broiler breeders are feed restricted to maximize reproductive output by 2810 maintaining a target BW profile (Hocking, 2004; Aviagen, 2007; Renema et al., 2811 2007) through an accurate feed allocation. Full-fed broiler breeders became obese, 2812 leading to multiple ovulations, deformed eggs and irregular oviposition (Renema 2813 2814 and Robinson, 2004). Severely feed-restricted (25% of ad libitum) broiler breeders led to delayed onset of lay and decreased egg production and egg weight 2815 2816 (Hocking, 2004). Both full-feeding and severely feed-restricted broiler breeders had reduced reproductive output. An accurate feed allocation decision is a big 2817 challenge which depends on the energy requirement of chickens, and that in turn 2818 2819 depends on BW, age, rate of lay and environmental temperature (Sibbald, 1980).

Feed cost per bird increases in a cold environment depending on desired barn temperature. Heating costs may be reduced if birds are placed at lower environmental temperature. In the current study, birds were reared in four

environmental temperatures (15, 19, 23 and 27°C). Feed cost, heating cost and savings were calculated relative to 27°C of barn temperature (Appendix B).

This thesis examined ways in which environmental temperature, dietary energy levels, feeding times, photoperiods and housing systems influence CBT dynamics and efficiency. Real time CBT was recorded using telemetry devices at 10-min intervals in broiler breeders to determine the CBT dynamics suggesting a feeding program to avoid heat stress. In addition, financial savings were assessed based on feed costs and heating costs of broiler breeder females reared in different barn temperatures.

2832 6.2. CORE BODY TEMPERATURE IN CHICKENS RELATIVE TO 2833 ENVIRONMENTAL TEMPERATURE.

In general, when environmental temperature is above or below the 2834 thermoneutral point, birds initially apply physiological mechanisms and physical 2835 2836 posture changes to maintain their CBT. The temperature difference increases between the surface body and environment, when the environmental temperature 2837 2838 drops below the lower critical temperature. Birds then dissipate an increasing amount of heat to the environment and they eat more to increase the rate of 2839 metabolic heat production to maintain their CBT (National Research Council, 2840 2841 1981). In addition, birds conserve body heat to increase insulation by sitting, huddling, reducing body surface area, and adjusting feathers. Birds also try to 2842 dissipate less heat to the environment by reducing blood flow to the skin through 2843 2844 vasoconstriction. Initially, birds can regulate all physiological functions but when environmental temperature continuously decreases below the lower critical 2845

temperature, then birds increase feed intake as much as possible. When the energy
requirement for maintenance increases in feed restricted broiler breeders; a
reduced proportion of energy is left for growth and production. This decreases
growth, production and efficiency.

2850 Conversely, when an environmental temperature is above the upper 2851 critical temperature, the temperature difference between the surface body and the environment decreases, and birds cannot lose adequate heat to the environment. 2852 Birds change physical posture to increase heat dissipation including separating 2853 2854 from each other, increasing surface area by standing, trying to stay near window (presumably looking for a colder area or finding a way to lose heat through 2855 convection (air movement) and shadow, wing drooping and spreading, and 2856 2857 eventually dissipating excess heat by panting (a faster rate of breathing promotes evaporative water loss; Freeman, 1965). They also try to maintain their CBT by 2858 increasing blood flow to extremities resulting in body heat loss through 2859 2860 vasodilation in hot environments. When they cannot dissipate adequate heat, CBT increases and birds decrease the rate of metabolic heat production by decreasing 2861 2862 feed consumption. Sufficiently reduced feed consumption results in reduced productive outputs such as growth or egg production or both resulting in 2863 decreased efficiency. Heat stress has a greater negative impact on efficiency than 2864 2865 cold stress.

2866 6.3. DIURNAL CORE BODY TEMPERATURE PATTERNS

Feed-restricted broiler breeder females had a distinct diurnal CBT pattern.The CBT peaked within 1 h after feeding in both broiler breeder pullets and hens

regardless of environmental temperature, feeding time, photoperiod and housing
system. This was likely due to feeding activity and feed metabolism (Khalil et al.,
2004). The CBT decreased in the remainder of day and was minimum at night
during sleep and rest (Chapter 2: Figure 2.1; Chapter 3: Figure 3.1; Chapter 4:
Figure 4.1; Chapter 5: Figure 5.1). Feeding activity, feed metabolism and normal
activity could increase day time CBT compared to night time CBT.

The CBT in birds was lower at night time in the standard photoperiod due 2875 to sleep and rest (Cain and Wilson, 1974; Khalil et al., 2004) compared to day 2876 2877 time. However, CBT was lower in the continuous photoperiod during same time period against night time compared to day time of standard photoperiod. The CBT 2878 in broiler breeder hens dropped (0.3°C) quickly after lights turned off in the 2879 16L:8D treatment group and slightly dropped (0.1°C) in the 24L:0D at the same 2880 time, and remained similar in the next 8 h light period in both the 24L:0D and 2881 16L:8D treatment groups. It might be due to synchronized feeding times; birds in 2882 2883 both photoperiods were fed in the morning every day. Possibly, birds developed a biological habit due to repetitive feeding cycle resulting in activity prior to 2884 2885 feeding leading to increased CBT. The CBT patterns were similar in the common 16 h light period in both treatments. The CBT was higher during the hours from 2886 21:30 to 05:30, which corresponded to dark period in the 16L:8D treatment, 2887 2888 because birds were likely more active when exposed to continuous lighting. This was a short study (7 d) and birds in the 24L:0D treatment group could be 2889 2890 influenced by the pre-established diurnal pattern of the standard photoperiod

(Birds were reared under the standard photoperiod (16L:8D) before starting thisexperiment), and regular morning feeding cycle.

The CBT at night was higher in free run hens than caged hens. 2893 2894 Presumably, activity levels of broiler breeder hens in the spacious free run housing system could contribute to a residual higher rate of metabolic activity, 2895 2896 leading in turn to higher CBT at night compared to caged hens. In addition, freerun hens sleep together and sit on the floor resulting in increased insulation that 2897 could reduce heat dissipation to the environment. Conversely, caged birds may 2898 2899 lose more heat through convection (air flow surrounding the bird) and conduction 2900 (birds contact with metal wires of the cage) than free-run hens. Increased feed intake in free run hens is mainly used for activity level and activity increases heat 2901 2902 production in the body. Possibly, vasomotor activity (vasodilation) could increase peripheral blood flow and expanded blood vessel especially in comb, wattles, and 2903 feet resulting in heat dissipation to environment at a higher rate to maintain a 2904 2905 homeostatic CBT in free run hens. However, the CBT peaked due to feeding related activity and diet induced thermogenesis within 1 h after feeding, and CBT 2906 2907 was lower at night than at day regardless of environmental temperature and 2908 housing systems.

Breeder hens consumed about 6% more feed in the low energy diet compared to the high energy diet as their CP intake was 1 g higher per day in the low energy diet than in the high energy diet. Theoretically, heat production was higher in low energy diet-fed birds because of higher volume of feed and a higher amount of CP intake. When birds use CP as an energy source, heat production

would be higher about 30% compared to fat or carbohydrate (Geraert et al., 1996).
Low energy diet-fed birds likely lose heat as soon as possible through
vasodilation (Mustaf et al., 2009); presumably, birds expand blood vessels,
resulting in increasing temperature difference between surface body and
environmental temperature, and increase heat loss to the environment.

2919 A significant rise in CBT was observed within 1 h after feeding. In the hot summer, mid-morning to mid-afternoon is the hottest time of the day. If birds 2920 were fed at hot times of day, the difference between the surface body temperature 2921 2922 and the environmental temperature would decrease. Birds would be unable to dissipate adequate heat to the environment resulting in a rise of CBT. In such 2923 2924 conditions, birds decrease feed intake resulting in a decrease in growth, egg 2925 production and egg quality (National Research Council.1981; Randall and Hiestand, 1939). As a consequence, birds decrease the rate of metabolic heat 2926 production to maintain CBT. Shifting feeding time from a hot time to a cooler 2927 2928 time of day, like early morning or late night or late afternoon just before the sun sets or a half meal early morning and another half meal in the late afternoon, 2929 2930 could be an appropriate approach to mitigate heat stress of birds in the hot 2931 summer.

2932 6.4. MAINTENANCE ENERGY REQUIREMENTS

The metabolizable energy requirement for maintenance (ME_m) includes the total heat of digestion and absorption, product formation, fermentation, and waste formation and excretion. Heat production and heat loss are balanced based on the difference between the surface body temperature and environmental 2937 temperature (Monteith, 1974). The energy requirement for maintenance in 2938 chickens increased with decreasing environmental temperature below the lower 2939 critical temperature (National Research Council, 1981). Thus, ME_m in chickens 2940 can vary depending on environmental temperatures. The net energy for 2941 maintenance (NE_m) is the energy used by animal after subtracting the total heat 2942 increment from the ME_m . Thus, NE_m is not affected by environmental temperature 2943 or feed intake.

Feed intake was decreased linearly in feed-restricted breeder pullets with 2944 2945 increasing environmental temperature from 15 to 27°C. Zuidhof et al. (2012) 2946 demonstrated a 5% decrease feed intake in broiler breeder pullets when changing 2947 the environmental temperature from 15 to 27°C, possibly due to severe feed 2948 restriction resulting in less heat production in a higher environmental temperature. The degree of feed restriction is higher in pullets than in hens due to their higher 2949 growth potential (de Beer and Coon, 2007). Theoretically, the energy requirement 2950 2951 for maintenance would decrease linearly with increasing environmental temperature (Figure 6.1) because breeder pullets would eat less feed due to higher 2952 2953 feed restriction and they would have less diet induced thermogenesis. Pishnamazi et al., (unpublished) reported that environmental temperature ranging of 15 to 2954 27°C had a quadratic effect on the energy requirement for maintenance in broiler 2955 2956 breeder hens (Figure 6.2). The energy requirement for maintenance increased with 2957 decreasing the environmental temperature from 24 to 15°C and with increasing the environmental temperature from 24 to 27°C. The energy requirement for 2958 2959 maintenance was minimal at 24°C, likely heat production and heat loss is

2960 comparatively balanced than other environmental temperatures. Above 24°C, the 2961 energy requirement for maintenance increases because birds expend energy to cool themselves (Pishnamazi et al., unpublished). Presumably, pullets produce 2962 2963 less heat depending on severity of feed restriction, ME_m in relaxed feed restriction pullets would decrease linearly until 27°C (Figure 6.1) and this linear relationship 2964 could be extended beyond a 27°C environmental temperature with a severe feed 2965 restriction. Severe feed restricted breeder pullets consume less feed than relaxed 2966 feed restricted pullets. The ME_m in severe feed restricted pullets would decrease 2967 2968 linearly until a higher environmental temperature (Figure 6.1). This gradual decreasing of ME_m indicates that the thermal point could further extend in 2969 severely feed restricted pullets than in relaxed feed restriction pullets. As a result, 2970 we could hypothesize that the thermoneutral zone could be wider in pullets than 2971 broiler breeder hens. 2972

2973 **6.5. APPLICATION TO COMMERCIAL BROILER BREEDER**

2974 **FARMING (RECOMMENDATION)**

According to the law of thermodynamics, heat production (H_p) and heat 2975 loss (H_l) will be equal in the poultry barn. That is, $H_p = H_l$ within a barn. Heaters 2976 2977 and animals are two sources of heat production. The sources of heat loss are ventilation, walls, ceiling and floor of the building. Where, H_p = heat production, 2978 H_l = heat loss; $H_p = f$ (heater, birds), $H_l = f$ (celling, floor, ventilation, wall); 2979 2980 f(heater) = f(celling, floor, ventilation, wall) - f(birds). Heat loss through the 2981 building depends on the difference $\{\Delta T = (inside - outside temperature)\}$ between the inside and outside temperature and the insulation status of the building. We 2982

assumed an outside temperature was -15°C. Total and sensible heat productions
were calculated for an individual bird in different environmental temperatures (15,
19, 23 and 27°C) with the following equation developed by Pedersen and
Thomsen (2000);

2987
$$q_t = 9.84*m_a^{0.75}(4*10^{-5}(20 - T_{ts})^3 + 1); q_s = 0.83q_t (0.8 - 1.85*10^{-7}(T_{ts} + 10)^4);$$

2988 where: q_t = Total heat production, J s⁻¹; q_s = Sensible heat produced, J s⁻¹; m_a =
2989 Mass per animal, live animal kg; T_{ts} = Dry bulb temperature, °C.

2990 Heat production from heaters was calculated as follows:

2991 f(heater) = f(ceiling + floor + ventilation + walls) - f(birds) (Appendix B). Feed 2992 cost was calculated for individual birds in different environmental temperatures 2993 during the rearing period (4 to 20 wk) and the breeding period (25 to 41 wk).

We assumed feed cost was \$380/T and natural gas cost was \$3.25/GJ 2994 (Equal Energy, 2010). The effects of changes in environmental temperature on 2995 feed intake and heating cost was applied to calculate saving cost for broiler 2996 2997 breeder females. Feed cost, heat cost and savings were calculated relative to 27°C scenario. Feed cost increased by \$0.02, \$0.09 and \$0.22 per pullet; \$0.00, \$0.04, 2998 2999 0.14 per hen) and heat cost decreased by 0.11, 0.21 and 0.30 per pullet; \$0.23, \$0.43 and \$0.62 per hen in the 23, 19 and 15°C treatment respectively, 3000 compared to 27°C treatment (Figure 6. 3; Figure 6. 4). Net savings for each pullet 3001 from 4 to 20 wk of age were \$0.08, \$0.12, and \$0.09 in the 15, 19 and 23°C 3002 respectively relative to 27°C. Net savings for each hen from 25 to 41 wk of age 3003 were \$0.48, \$0.39 and \$0.23 in the 15, 19 and 23°C respectively. The highest 3004

savings were \$0.12 per pullet in the 19°C and \$0.48 per hen in the 15°C relative to
27°C.

Feed intakes increased as well as feed costs increased and heating cost 3007 3008 decreased with decreasing barn temperature from 27 to 15°C, when outside 3009 temperature was -15°C. Birds increased feed intake because they lose energy to 3010 the lower environmental temperature. On the other side, heating costs decreased with decreasing barn temperature because the temperature difference decreases 3011 3012 between the barn and outside. Thus, less heat was lost to outside environment. 3013 Decreased heat costs of broiler breeders were higher than increased feed costs in the 15°C relative to 27°C. 3014

Feed allocation would increase to maintain core body temperature in broiler 3015 3016 breeder females in cold environmental temperature (below the lower critical temperature). Consequently, birds could increase the rate of metabolic heat 3017 production. Inadequate feed allocations in feed restricted broiler breeders, 3018 3019 particularly in a cold environment, would leave a small proportion of energy for growth and production. A precise feed allocation decision based on environmental 3020 3021 temperature is a big challenge to optimize growth and production in feed restricted breeders. 3022

Adjustment of feed allocation with the change of environmental temperature is recommended. For each 1°C decrease in barn temperature from 23 to 15°C, feed consumption increased around 0.2 g and 0.5 g for each broiler breeder pullet and hen respectively. This extra feed allocation is mostly utilized for maintenance

requirements of birds, particularly to keep a relatively constant CBT in lowerenvironmental temperatures.

A 17.0 % increase in feed allocation would be needed to compensate the energy lost by activity in free-run hens, when research outcomes from caged broiler breeders are applied to the industry level.

3032 High energy diet-fed breeders were more energy efficient than low energy diet-fed breeders. High energy diet-fed birds had lower maintenance energy than 3033 low energy diet-fed birds because birds on a high energy diet required less 3034 3035 quantity of feed and possibly use less energy to digest this feed. Feed volume was 3036 18% higher in the low energy diet than in the high energy diet. When dietary CP 3037 is used as an energy source, heat production increased by 30% compared to fat and carbohydrate (Geraert et al., 1996). Thus, lower CP:ME ratio (0.055) diet fed 3038 birds can avoid heat stress. 3039

3040 Heat production in chickens was influenced by several factors like activity, feeding time, light intensity, dietary crude protein, housing system, and day 3041 length. Feeding related activity and feed metabolism in broiler breeders increased 3042 3043 CBT in peak within 1 h after feeding. Heat production in chickens would increase with increasing intake of dietary CP, at higher environmental temperature, birds 3044 3045 are unable to lose adequate amount of heat (National Research Council, 1981). 3046 Then, they would reduce the dependency on diet induced thermogenesis to maintain their CBT resulting in decreased feed intake. This results in decreased 3047 3048 production and efficiency.

3049 Reducing CP:ME ratio in the diet, birds could decrease heat production with 3050 reduced intake of CP, and birds can avoid heat stress in hot environment (Zuidhof et al., 2012). In general, birds increase activity level in the larger area in the free 3051 3052 run housing system. This increased activity level in birds result in high heat production compared to caged hens because caged hens were allowed a limited 3053 3054 area and they cannot increase activity like free run hens. Intake of small amount 3055 of dietary CP in caged hens could produce less heat, which could avoid heat stress in a high environmental temperature (above the upper critical temperature). 3056

3057 Environmental temperature generally increases with increasing day length and this temperature decreases at the end of day. If birds are fed in the early morning 3058 3059 of day when environmental temperature is normally lower than at noon time or at 3060 the end of day (late afternoon), when environmental temperature cools down, birds also can avoid heat stress because they can lose adequate heat to the cold 3061 3062 environment of day. Birds increase activity level with increasing light intensity 3063 resulting in increased heat production and they decrease heat production with decreasing light intensity (Boshouwers and Nicaise, 1987). So, birds with lower 3064 3065 light intensity also can mitigate heat stress in hot summer.

Reducing rearing space, light intensity, less intake of CP and decreasing CP:ME ratio in diet, shifting feeding time from a hot period to a cold period of day, likely early morning or late night or late afternoon just before the sun sets or a half meal early morning and another half meal in the late afternoon, could be an appropriate management approach to mitigate heat stress in the hot summer.

3071 **6.6. SOME BASIC FINDINGS OF THE RESEARCH**

Core body temperature ranged from 39.99 to 42.47°C in the 23°C
 treatment groups which seems to be normal CBT in broiler breeder pullets.
 Because, breeder pullets are more efficient in the 23°C than other
 environmental temperatures.

Core body temperature in broiler breeder hen ranged from 39.77 to
 41.06°C within the range of environmental temperature from 15 to 27°C.
 Because, broiler breeders are similar efficient within environmental
 temperature from 15 to 27°C.

3080 6.7. FUTURE RESEARCH

3081 Production, feed efficiency and efficient use of resources as well as fundamental information are key issues to make the poultry industry sustainable 3082 and profitable. Feed consumption was linearly decreased in broiler breeder pullets 3083 3084 and it had a quadratic relationship in broiler breeder hens with increasing 3085 environmental temperature from 15 to 27°C. The energy requirement for pullet maintenance linearly decreased until 27°C environmental temperature; possibly, 3086 3087 due to a higher feed restriction in broiler breeder pullets than in broiler breeder hens. This may indicate that the thermoneutral zone for broiler breeder pullets 3088 3089 could extend above 27°C. However, further experimentation is needed to identify 3090 the upper and lower critical environmental temperature and CBT for broiler breeder females' performance under dietary and housing conditions. The CBT 3091 3092 may act as an indicator of heat stress when CBT goes above the normal range and cold stress when CBT drops below the normal range. This could help broiler 3093

- 3094 breeder industry to setup guidelines for management in different environmental
- 3095 temperatures.

6.8. REFERENCES

3105

3109

3116

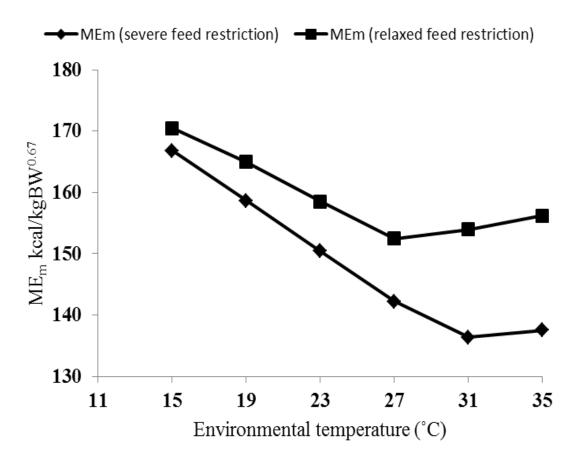
3120

3125

3129

- Aviagen. 2007. Ross 708 parent stocks performance objectives. Aviagen Ltd.
 www.aviagen.com/docs/Ross 708 PS Performance Objectives.pdf.
 Accessed June 2010.
- Boshouwers, F. M., and E. Nicaise. 1987. Physical activity and energy
 expenditure of laying hens as affected by light intensity. Br. Poult. Sci.
 28:155-163.
- Cain, J. R., and W. O. Wilson. 1974. The influence of specific environmental
 parameters on the circadian rhythms of chickens. Poult. Sci. 53:14381443.
- Deeb, N., and A. Cahaner. 1999. The effects of naked neck genotypes, ambient
 temperature, and feeding status and their interactions on body temperature
 and performance broilers. Poult. Sci. 78:1341-1346.
- 3113
 3114 Equal Energy. 2010. <u>www.equalenergy.ca/en/financials/2010_annual_report_v</u>
 3115 03282011.pdf. Accessed March 2013.
- Freeman, B. M. 1965. The relationship between oxygen consumption, body
 temperature and surface area in the hatching and young chick. Br. Poult.
 Sci. 6:67-72.
- Geraert, P. A., J. C. F. Padilha, and S. Guillaumin. 1996. Metabolic and endocrine
 changes induced by chronic heat exposure in broiler chickens: Growth
 performance, body composition and energy retention. Br. J. Nutr. 75:195204.
- Hocking, P. M. 2004. Roles of body weight and feed intake in ovarian follicular
 dynamics in broiler breeders at the onset of lay and after a forced molt.
 Poult. Sci. 83:2044-2050.
- Khalil, A. M., K. Matsui, and K. Takeda. 2004. Diurnal and oviposition-related
 changes in heart rate, body temperature and locomotors activity of laying
 hens. Anim. Sci. J. 75:169-174.
- Monteith, J. L. 1974. The concept of thermal neutrality, page 425, in heat loss
 from animals and man, ed J. L. Monteith and L. E. Mount, Butterworth,
 London.
- 3137
 3138 Mustaf, S.; N. S. Kahraman, and M. Z. Firat. 2009. Intermittent partial surface
 3139 wetting and its effect on body-surface temperatures and egg production of
 3140 white brown domestic laying hens in Antalya (Turkey). Br. Poult. Sci.
 3141 50:33-38.

3142 3143	National Research Council. 1981. Effect of environment on nutrient requirements of domestic animals. National academy press. Washington, D. C.
3144	
3145	Pedersen, S. and M. G. Thomsen. 2000. Heat and moisture production broilers
3146	kept in straw bedding. J. Agric. Eng. Res. 75:177-187.
3147	
3148 3149	Randall, W. C., W. A. Hiestand. 1939. Panting and temperature regulation in the chicken. Am. J. Physiol. 127:761-767.
3150	
3151	Renema, R. A., and F. E. Robinson. 2004. Defining normal: Comparison of feed
3152	restriction and full feeding of female broiler breeders. World's Poult. Sci.
3153	J. 60:511-525.
3154	
3155	Renema, R. A., M. E. Rustad, and F. E. Robinson. 2007. Implications of changes
3156	to commercial broiler and broiler breeders' body weight targets over the
3157	past 30 years. World's Poult. Sci. J. 63:457-467.
3158	
3159	Richards, S. A. 1971. The significance of changes in the temperature of the skin
3160	and body core of the chicken in the regulation of heat loss. J. Physiol.
3161	216:1-10.
3162	210.1-10.
3163	Romero, L. F., M. J. Zuidhof, R. A. Renema, F. E. Robinson, and A. Naeima.
3164	2009. Nonlinear mixed models to study metabolizable energy utilization in
3165	broiler breeder hens. Poult. Sci. 88:1310-1320.
3166	bioner breeder nens. I buit. Sei. 88.1510-1520.
3167	Sakomura, N. K. 2004. Modeling energy utilization in broiler breeders, laying
	hens and broilers. Braz. J. Poult. Sci. 6:1-11.
3168	hens and drohers. Braz. J. Poult. Sci. 0.1-11.
3169	
3170	Savory, C. J., L. Kostal, and I. M. Nevison. 2006. Circadian variation in heart
3171	rate, blood pressure, body temperature and EEG of immature broiler
3172	breeder chickens in restricted-fed and <i>ad libitum</i> -fed states. Br. Poult. Sci.
3173	47:599-606.
3174	
3175	Sibbald, I. R. 1980. Metabolizable energy in poultry nutrition. Bioscience.
3176	30:736-741.
3177	
3178	Wang, J. P., and I. H. Kim. 2011. Effect of caprylic acid and yucca schidigera
3179	extract on production performance, egg quality, blood characteristics, and
3180	excreta microflora in laying hens. Br. Poult. Sci. 52:711-717.
3181	
3182	Zuidhof, M. J., D. C. Paul, A. Pishnamazi, I. I. Wenger, R. A. Renema, and V. L.
3183	Carney. 2012. Temperature and protein: energy ratio linkages between
3184	breeder and broiler energetics, performance, and carcass quality. Final
3185	Report to Alberta Livestock and Meat Agency: Project #2008F138R.
3186	February05.





3187 3188 Figure 6.1 Theoretical ME requirements for maintenance (severe feed restriction and 3189 relaxed feed restriction) in broiler breeder pullets (4 to 20 wk of age) in different 3190 environmental temperatures. Y axis indicates the energy requirement for maintenance 3191 kcal per kg metabolic BW and X axis indicates environmental temperature in degree 3192 Celsius.

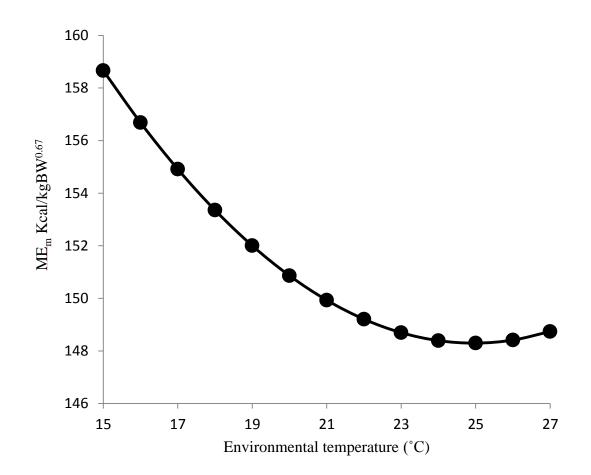


Figure 6.2 Predicted ME requirements for maintenance of broiler breeder hens (25 to 41
wk of age) in different environmental temperatures. Y axis indicates the energy
requirement for maintenance kcal per kg metabolic BW and X axis indicates
environmental temperature in degree Celsius.

3199 Source: Pishnamazi et al. (unpublished)

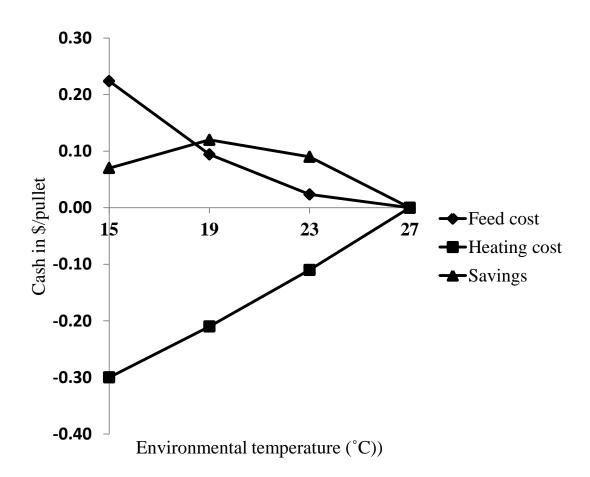


Figure 6.3 Relative feed and heating costs of broiler breeder hens reared in environmental temperatures of 15, 19, 23, and 27°C from 25 to 41 wk of age. All costs and savings relative to 27°C scenario, and assume that the outdoor temperature was 15°C. Y axis indicates cash in \$ per pullet and X axis indicates environmental temperature in degree Celsius.

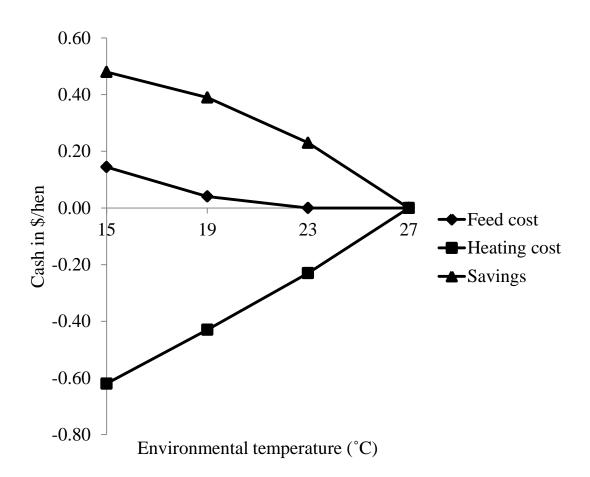


Figure 6.4 Relative feed and heating costs of broiler breeder hens reared in environmental temperatures of 15, 19, 23, and 27°C from 25 to 41 wk of age. All costs and savings relative to 27°C scenario, and assume that the outdoor temperature was -15°C. Y axis indicates cash in \$ per hen and X axis indicates environmental temperature in degree Celsius.

3214 Appendix A: Broiler Breeder diets

Items	Starter diet	Grower diet
Ingredients%		
Corn	15.000	33.450
Wheat	39.410	35.000
Soybean meal (SBM)	17.590	6.660
Oats	15.000	6.190
Canola meal	5.000	5.000
Canola oil	2.380	8.000
Dical phosphate	1.980	1.000
Calcium carbonate	1.580	0.000
Ground limestone	0.000	1.520
Broiler premix ¹	0.500	0.500
Salt	0.450	0.390
Choline chloride premix ²	0.500	0.500
L-lysine	0.355	0.122
DL-methionine	0.207	0.093
Avizyme 1302	0.050	0.050
Threonine	0.000	0.025
Total	100.002	100.000
Calculated (% unless otherwi	ise indicated)	
ME kcal/kg	2,900.000	2,865.000
Crude Protein%	19.000	15.000
Calcium%	1.100	1.000
Analyzed (% unless otherwis	e indicated)	
ME kcal/kg	NA	2,792.000
Crude Protein%	NA	16.990

3215 Table A.1 Starter and grower diet of broiler breeder (Ross 708)

3216 NA= not analyzed

¹The premix provided the following (per kg of diet): vitamin A (retinyl acetate),

3218 10,000 IU; cholecalciferol, 4,000 IU; vitamin E, 35 IU; vitamin K, 4.0 mg; pantothenic

3219 acid, 15 mg; riboflavin, 10 mg; folic acid, 0.2 mg; vitamin B₁₂, 0.02 mg; niacin, 65 mg;

thiamine, 4.0 mg; pyridoxine, 5.0 mg; biotin, 0.2 mg; choline, 2.63 mg iodine, 1.65

3221 mg; Mn, 120 mg; Cu, 20 mg; Zn, 100 mg, Se, 0.3 mg; and Fe, 80 mg.

²Provided choline chloride in the diet at a level of 100 mg/kg.

Items	High energy	Low energy
Ingredients%		
Corn	39.882	35.526
Wheat	30.000	30.000
Soybean meal (SBM)	17.000	15.143
Limestone	7.829	7.928
Wheat bran	0.000	7.585
Canola oil	2.295	1.000
Dical phosphate	1.414	1.212
Layer Vit/Mineral PMX ¹	0.500	0.500
Choline Chloride PMX ²	0.500	0.500
Common salt	0.386	0.380
DL Methionine	0.143	0.147
Avizyme 1302	0.050	0.050
Lysine HCl	0.000	0.028
Total	99.999	99.999
Calculated (% unless other	wise indicated)	
ME kcal⁄kg	2,900.000	2700.000
Crude Protein%	15.200	15.190
Calcium%	3.300	3.300
Analyzed (% unless otherw	vise indicated)	
ME kcal⁄kg	2,912.000	2,786.000
Crude Protein%	16.400	16.100

3223 Table A.2 Layer diet of broiler breeder (Ross 708)

¹The premix contained (per kg of diet): iron, 80 mg; zinc, 100 mg; manganese, 88 mg; copper, 15 mg; iodine, 1.65 mg; selenium, 0.3 mg; vitamin A, 12,500 IU; vitamin D₃, 3,125 IU; vitamin E, 40 IU; vitamin K (menadione), 2.5 mg; niacin, 37.5 mg; D-pantothenic acid, 12.5 mg; riboflavin, 7.5 mg; pyridoxine, 5 mg; thiamine, 2.55 mg; folic acid, 0.625 mg; biotin, 0.15 mg; vitamin B₁₂, 0.01875 mg; and choline, 2.767055 mg. ²Provided choline chloride in the diet at a level of 100 mg/kg.

APPENDIX B: CALCULATIONS USED FOR ECONOMIC ANALYSIS.

Heat production =Heat loss

Heater + Animal= ventilation+ building

Heater = (ventilation+ building)-Animal Assuming outside temperature= -15C Natural gas price=3.25\$/GJ

Rearer: 100birds/chamber Breeder: 48birds/chamber

Measurement of chamber (m) Length=4.45 Width=3.85 Height=3.00

Heat loss for ventilation (VHL)

 $Qv=m*Cp*\Delta T$ Qv= heat loss for ventilation in watt/s Cp= specific heat capacity usually given as 1 $\Delta T=$ temperature difference between inside and outside

m= air flow rate (m^3/h)

Heat loss for walls (HLW)

$Q=A\Delta T/r$	A= area of the wall	A=(length*height*2)+ (width*height*2)
	ΔT = temperature difference between inside and outside	
	r=resistance of wall	
	Q= heat loss for walls in watt/s	

Heat loss for ceiling (HLC)

$Q=A\Delta T/r$	A= area of the ceiling	A=length*width
	ΔT = temperature difference between i	inside and outside
	r=resistance of ceiling	
	Q= heat loss for ceiling in watt/s	

Heat loss for floor (HLF)

	P= perimeter of the room	P=2 (length + width)
Q=P∆TF	ΔT = difference of temperature betw	ween inside and outside
	F=1.42 (resistance of normal concr	ete floor
	Q= heat loss for floor in watt/s	
laulationa mana ann	morrow by Dr. I. Foddog, Datingd Drofosson	University of Alberto

3224 All calculations were approver by Dr. J. Feddes, Retired Professor. University of Alberta.

3225	APPENDIX C: SCHEMATIC DIAGRAM O	F CHICKENS
3226	USED IN DIFFERENT EXPERIME	INTS
3227	Experiment 1	Ň
3228	800 pullets (floor p Experimental perio	en) d from 4 to 20 wk of age
3229		\backslash
3230	Randomly selected	
3231	V selected	
3232	Experiment 2 192 hens (individually caged)	Experiment 3 140 hens (free-run)
3233	Experimental period from 25 to 41 wk of age	Experimental period from 25 to 41 wk of age
3234	Randomly selected	
3235		

Experiment 4 192 hens (individually caged) short experiment (7 d) at age of 44 wk