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UNIVERSITY OF ALBERTA

Synthetic studies on silyl cuprates, and synthesis of A58365B, an inhibitor of angiotensin-converting enzyme



A thesis submitted to the faculty of Graduate Studies and Research in partial fulfillment of the requirements for the degree of Doctor of Philosophy.

DEPARTMENT OF CHEMISTRY

Edmonton, Alberta SPRING, 1997



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Jan 6 1997.

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FACULTY OF GRADUATE STUDIES AND RESEARCH

The undersigned certify that they have read, and recommend to the Faculty of Graduate Studies and Research for acceptance, a thesis entitled **Synthetic studies on silyl cuprates** and synthesis of A58365B, an inhibitor of angiotensinconverting ensyme submitted by Yuanxi Zhou in partial fulfillment of the requirements for the degree of Doctor of Philosophy.

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Abstract

This thesis describes two projects. In the first, the response of vinyl epoxides **56** and **87** to dimethylphenylsilyl cuprates was examined. The results show that the course of the reactions is sensitive to the stoichiometry of the reagent.



The second section of the thesis describes the total synthesis of A58365B - an inhibitor of angiotensin-converting enzyme. This section also reports the preparation of optically active material of high, but, as yet, undetermined, optical purity.



A58365A

1





A58365B 2, and the related pyridone 1, are water-soluble metabolic products of *Streptomyces chromofuscus* NRRL 19098. Both substances inhibit angiotensin-converting enzyme.



The racemic form of A58365B was synthesized by a key process based on enyme radical cyclization. In this approach, the C(7)-C(8) unsaturation is introduced early but in the disguised form of a spirolactone. Compound 71 represents all the carbons that make up the C(6)-C(9) segment of the natural product, including the propionic side chain. The ring B portion of 2 was constructed from ε -hydroxynorleucine methyl ester (72). Compounds 71 and 72 were coupled, and subjected to further elaboration into 69. Enyme radical cyclization, followed by functional group manipulation, led to racemic A58365B. Optically active material was also obtained following an identical route, but using (L)- ε -hydroxynorleucine as the starting material.

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List of Abbreviations

AIBN	2,2'-azobisisobutyronitrile
BOC	tert-butoxycarbonyl
DCC	N,N'-dicyclohexylcarbodiimide
DDQ	2,3-dichloro-5,6-dicyano-1,4-benzoquinone
DMAP	4-(dimethylamino)pyridine
DME	1,2-dimethoxyethane
DMF	dimethylformamide
DMSO	dimethyl sulfoxide
EDCI	1-(3-dimethylaminopropyl)-3-
	ethylcarbodiimide
HOBT	1-hydroxybenzotriazole hydrate
LDA	lithium diisopropylamide
LTMP	lithium 2,2,6,6-tetramethylpiperidide
m-CPBA	<i>m</i> -chloroperoxybenzoic acid
NMM	N-methylmorpholine
PCC	pyridinium chlorochromate
TBAF	tetrabutylammonium fluoride
Tf	trifluoromethanesulfonyl (triflyl)
TFA	trifluoroacetic acid
THF	tetrahydrofuran
TIPS	triisopropylsilyl

.

 $S_N 2'$ Addition of Organocopper Reagents to Vinyloxiranes

Allylic systems, in addition to undergoing ordinary substitutions without rearrangement, can also react with rearrangement of the double bond, as indicated in **Scheme 1**. This transformation is called an S_N2' reaction.



Scheme 1



Scheme 2

The S_N2' mechanism, as shown above, involves the movement of three pairs of electrons, but there is no evidence that requires that bond making and breaking be concerted. The stereochemistry of the S_N2' reaction has been investigated and it is found that both $syn^{1,2}$ and $anti^{3,4}$ substitution can take place, depending on the nature of X and Y, though the syn pathway predominates in most cases (Scheme 2).



Scheme 3



scheme 4

Vinyloxiranes constitute a special subset of allylic electrophiles. The S_N2' addition of organocopper reagents to vinyloxiranes represents a potentially versatile route to a variety of allylic alcohols which are very important synthetic intermediates (**Scheme 3**). Allylic alcohols in turn, are readily and stereoselectively transformed into vinyloxiranes,⁵ thus allowing for iterative stereoselective chain elongation (**Scheme 4**). First observed in 1970 by Anderson and Johnson^{6,7} with butadiene epoxide and isoprene monoepoxide, this reaction was subsequently explored by others, using cyclic vinyloxiranes with endocyclic or exocyclic double bonds.

Acyclic vinyloxiranes



Scheme 5





In general, S_N2' displacements on allylic alcohol derivatives (**Scheme 5**) and cyclic vinyloxiranes (**Scheme 6**) by organocopper reagents have been found to proceed with inversion (*anti* pathway).^{8,9,10} Acyclic vinyloxiranes would be expected to follow an analogous course. However, unlike their cyclic counterparts, the acyclic system can react via either s-cis or s-trans conformers and thus yield mixtures of E and $Z S_N 2'$ products⁸ (Scheme 7).



Scheme 7

Additions of various methyl copper reagents to the optically pure acyclic vinyloxirane were performed by Marshall and Trometer¹¹ in order to evaluate E/Z and syn/anti preferences.

The unsubstituted vinyloxirane 17 (E) and 18 (Z) gave a mixture of S_N2 and S_N2' products with the four reagents examined, Me₂CuLi, MeCuCNLi, Me₂CuMgBr and MeLiCuI.BF₃. A high $S_N2':S_N2$ product ratio was obtained with MeCuCNLi, and Z-vinyloxirane 18 gave a better $S_N2':S_N2$ ratio than the E

.

isomer 17 (Scheme 8). The higher preference for the $E-S_N2^+$ product indicated the *s*-trans transition state conformations of 17 (*E*) and 18 (*Z*) were stereoelectronically favored (Scheme 8).



81 0 3

6

9

18 (*Z*)

83%

Scheme 8

They also examined other substrates in which stereochemical preferences might be more readily discerned.¹¹ Compounds 23 (E) and 24 (Z) seemed well suited on several counts. In the first place, the oxirane methyl substituent should increase the energy of s-cis conformers thereby diminishing the amounts of Z- S_N2' product. Secondly, the presence of additional substituents on the oxirane ring would be expected to disfavor S_N2 product formation. Finally, anti and syn S_N2' displacements would lead to diastereoisomers, not enantiomers, whose analysis would not depend upon assumptions regarding optical purity.

Addition of MeCuCNLi to the E-vinyloxirane 23 afforded an 88:12 mixture of the two $E-S_N2'$ products 25 (antiaddition) and 26 (syn-addition). The Z-vinyloxirane 24, on the other hand, yield a 1:99 mixture of isomer 25 (synaddition) and 26 (anti-addition) under the same conditions. In neither case was an S_N2 product detected (Scheme 9).



Scheme 9

The effect of substituents and geometry on stereoselectivity was further explored with the nonracemic vinyloxiranes 27, 30, 33 and $34.^{12}$ Anti-addition of MeCuCNLi to the E-vinyloxirane 27 afforded only two products, the anti-E 28 and anti-Z 29 as a 75:25 mixture (Scheme 10). 6

Anti-addition Z-vinyloxirane 30 also yielded two products, the syn-E 31 and syn-Z 32 in an isomer ratio of 16:84 (Scheme 11).





Another two vinyloxiranes, 33 and 34, were examined next. Anti-addition of MeCuCNLi to the E isomer 33 yielded syn-E ent-31 and syn-Z ent-32 in a 49:1 ratio (Scheme 12). Anti-addition Z-vinyloxirane 34 gave mainly anti-E ent-28 (Scheme 13).







The experimental facts reveals two interesting trends: (1) Acyclic vinyloxiranes 27, 30, 33 and 34 show a high anti preference in S_N2' addition of methylcuprate. (2) The E/Zratio is significantly influenced by the geometry of the vinyloxirane substrate.

-

The ratios reflect transition state energies related to the *s*-trans and *s*-cis conformers of the respective vinyloxiranes. Molecular modeling calculations on the *s*-cis and *s*-trans conformers were carried out^{12} and the results are shown in **Scheme 14**.

Two significant conclusions can be drawn from **Scheme** 14. (1) Compound 27 shows the smallest energy difference between the *s*-trans and *s*-cis conformers. Therefore S_N2' addition to 27 has the lowest E/Z selectivity. (2) Compounds 27, 33 and 34 were calculated to favor the *s*-trans, whereas 30 prefers the *s*-cis conformation. S_N2' addition to 30 therefore gives more of the Z product than similar addition to 27, 33 and 34.

Alkylidene Exocyclic Epoxides

Ziegler and Cady were the first to employ exocyclic epoxides for the introduction of stereocenters in a side chain by S_N2 ' alkylation.¹³

Treatment of oxirane **35** with Bu_2CuLi at -25 °C gave rise to **36** (S_N2' syn-addition) and **37** (S_N2' anti-addition) in a ratio of 95:5. Exposure of Z-oxirane **38** to Bu_2CuLi yielded both **36** (S_N2' syn-addition) and **37** (S_N2' anti-addition) in a ratio of 25:75. Treatment of **39** with Me₂CuLi gave the products derived from both modes of allylic addition (S_N2' and S_N2) to the vinyloxirane. Most (70%) of the reaction mixture consisted of isomers **36** (S_N2' anti-addition) and **37** 10

 $(S_N2' syn-addition)$ in an 85:15 ratio, and the remaining 30% consisted of the single compound **40** (anti-S_N2) (Scheme 15).



.



It can be argued that 35 and 38 have different conformational preferences, and that vinyloxirane 35 would exist in a twist conformation in transition state 40 where the axial C-O bond maintains overlap, and A-strain could be avoided. The preferred mode of Bu₂CuLi attack would occur from the α face of the double bond, syn to the oxygen. Oxirane 38, on the other hand, would preferentially react through conformation 41. Although A-strain must develop to some extent between the oxirane methane and ethylidene groups, this difficulty must be accommodated in order to attain overlap. Therefore, the approach of the cuprate is favored on the β face in an *anti*-mode and distal to the ring methyl group. The results obtained from 39 might be attributed to the transition state 42 which leads to the major regioisomer 36 via an anti approach to the vinyl oxirane system and from the β face, in spite of the axial methyl group (Scheme 16).



Scheme 16

Cuprate addition to the methylenecyclododecylidene

12

epoxide **43** was studied by Marshall and Flynn,^{14a} and it was found that epoxide **43** reacted cleanly with organocuprate via S_N2' substitution to give the *trans* S_N2' product **44** (Scheme **17**).



In several cases, in which the nature of R in RCu was varied, the $S_N^{2'}$ trans product predominated over the *cis* by 95:5. Presumably, the predominance of trans products reflects certain conformational preferences of the cyclododecylidene epoxide **43** in the transition state.¹⁵ This type of stereocontrol is of special interest in connection with macrocyclic natural product synthesis. In order to clarify the stereochemical results, the addition of butyl cuprates to the medium and large ring cycloalkylidene epoxides **46a-46f** was also studied,¹⁶ and the results are summarized in **Scheme 18**. In all cases, the trans $S_N^{2'}$ product **47** was formed as the major product. The addition thus appears to be relatively insensitive to the nature of the cuprate species.



46a-46f

.

47a-47f

48a-48f

	epoxide	cuprates	yield	47	48	
						-
8	46 a	BuMgBr/Cul	87%	85	15	
10	46b	BuMgBr/Cul	76%	98	2	
		-				
11	46c	Bu MgBr/Cul	52%	9 5	5	
12	46d	BuMgBr/Cul	82%	90	10	
		oungen our		••		
13	46e	BuMgBr/Cul	71%	93	7	
14	46f		87%	94	6	
17	401	BuMgBr/Cul	01%	34	0	
10	46b	LiBuCuCN	61%	95	5	
10	46b	LiBu ₂ Cu	77%	98	2	

Scheme 18

It is known that the addition of organocuprates to an allylic system involves initial π -complexation of the cuprate on the less hindered face of the double bond followed by S_N2' oxidative addition, leading to a σ -copper complex, which undergoes reductive elimination to produce the product with retention of stereochemistry.¹⁷ In the present case, S_N2' oxidative addition of the initial π -complex can occur via transition states **49** (*s*-*cis*) or **50** (*s*-*trans*) to give the *cis* or *trans* α -allyl intermediates **51** or **52**, respectively, and this pathway would lead to products **48** and **47**. The

calculations using Still's RINGMAKER/BAKMO¹⁶ program indicate that the *s*-trans conformation is favored over the *s*-cis conformation by 2-3 kcal/mole.¹⁶ Accordingly, the preference for trans products in S_N2' additions of organocuprate to epoxides **46a**-**46f** could come from a favored transition state involving the π -complex **50** of the lower energy *s*-trans conformation (**Scheme 19**).





Syn-anti preferences of these additions were determined through studies on the (R)-cycloalkylidene epoxides **53a** and **53b** (see Scheme 21).¹⁸ These cycloalkenes are chiral and, depending upon the syn or anti preference of the addition, an R or S enantiomer would be produced. In these cases, syn S_N2' addition to the (R)-epoxides **53** would afford the allylic alcohols (R)-**54** (see **Scheme 20**), whereas anti addition would give (S)-**54**. The foregoing analysis is based on the assumption that the bridging methylene chain effectively blocks attack on the double bond from within the ring cavity. Thus, the exo conformer must undergo syn S_N2' addition and the endo conformer can give only the anti S_N2' adduct. These two reaction pathways yield enantiomeric products (R)-54 and (S)-54 (Scheme 20). It should be noted that the two s-trans conformers 53-exo and 53-endo in Scheme 20 are calculated to be nearly equal in energy and lower than the s-cis¹⁸



Scheme 20

In fact, treatment of the 12-membered cycloalkylidene epoxide **53a** with *n*-butylmagnesium bromide-CuI afforded a 99:1 mixture of *trans* and *cis* allylic alcohols **54a** and **55a**; compound **54a** was determined to have the *R* configuration. Therefore, the foregoing addition must proceed via the *syn* S_N2' pathway from the *s*-trans exo conformer of **53a**, and the allylic alcohol (*R*)-**54a** is formed with high stereoselectivity (Scheme 21). The 14-membered cycloalkylidene epoxide 53b behaved analogously. Treatment with the butylcopper reagent gave a 94:6 mixture of allylic alcohols 54b and 55b. The sign of rotation of 54b is suggestive of the *R* configuration, as would be expected from syn S_N2' addition to the *s*-trans exo conformer of epoxide 53b (Scheme 21).



Scheme 21

The preferred syn S_N2' addition to epoxides 53 is thought to reflect differences in the ability of the epoxide oxygen to coordinate with the metal cation in the exo and endo s-trans conformers (Scheme 20). This coordination would be hampered by the bridging methylene chain in the endo conformer, thereby decreasing its reactivity toward S_N2' attack. The exo oxygen on the other hand, is readily accessible to the metal ion. 17

Cyclic 1,3-Diene Monoepoxides

Initial stereochemical studies on the S_N2' addition of cuprates to cyclic vinyloxiranes employed the monoepoxide of 1,3-cyclohexadiene (56) as the substrate.^{19a} Use of a rigid structure, such as 56, is potentially more informative from a mechanistic standpoint.

The results of the reaction of **56** with Gilman organocuprate compounds are summarized in **Scheme 22**. These results show that the conjugate and direct addition reactions are competitive processes, and a strong preference for *anti* addition is observed in both S_N2 and S_N2' reactions.



Scheme 22

Anti addition has been also observed in the case of 1,3cycloheptadiene monoepoxide (59),^{19b} a high yield of methyl transfer products being obtained, and the ratio of **61** (*anti* S_N2' product) and **62** (*anti* S_N2 product) was found to be 97:3

(Scheme 23).

.



Scheme 24

From this result, it was felt that the strongly electron-withdrawing acrylate ligand might be responsible for the highly specific anti 1,4-addition. In order to confirm this point, several mixed cuprates, containing a methyl group and an electron-withdrawing ligand, were examined, and the results are listed in **Scheme 24**. In fact, there is a progression of increasing regiospecificity from the homocuprate (R = Me) to the acrylate mixed cuprate. From a practical standpoint, the cyano methylcuprate is the most convenient to prepare, but the acrylate ligand exerts the greatest influence on the regiospecificity, followed closely by the cyano group.

A significant extension of this methodology followed the discovery that enol derivatives of cyclic α , β -epoxyketones underwent highly stereoselective *anti* S_N2' addition by cyano cuprate reagents.

Alam and Martin reported the facile introduction of three stereogenic centers and the overall functionalization of five carbon atoms of a six-carbon unit.²⁰ This methodology is based on the stereospecific functionalization of the readily available 1,3-cyclohexadiene monoepoxide (**56**) and the subsequent oxidative cleavage of the final cyclohexene derivative. This approach relies on repetitive stereocontrolled 1,4-openings of an epoxide, as described below.

Mixed cyanocuprates add stereospecifically (100% anti addition) and highly stereoselectively (the isomer ratio of 1,4-addition > 95%) to epoxide **56** to give the allylic alcohol 20

57, which can be transformed into the new epoxide 63 stereospecifically by MCPBA (Scheme 25).



a, R = Me, b, R = Ph







In order to repeat the 1,4-addition to an epoxy alkene system, **57b** was converted into epoxide **64** in a three-step procedure. When **64** was treated with lithium methyl cyanocuprate, cyclohexene **65** was produced stereospecifically (**Scheme 26**). An alternative strategy for the introduction of a second carbon substituent via an epoxy alkene is shown in **Scheme 27**. In this approach, **57** was oxidized first, and then the resulting product was silylated to silyl enol ether **67**, which underwent the 1,4-addition reaction with MeCuCNLi to produce **68** in essentially quantitative yield. As shown in **Scheme 28**, compound **68** can be either desilylated to the aldol **69** or oxidized to **71** by ozonolysis.



The α -substitution of ketones and other carbonyl

compounds is most directly accomplished by alkylation of enolates with an electrophile. This general approach fails with highly substituted and non aliphatic electrophiles. Also, α '-alkylation of α , β -unsaturated ketones can be complicated by the existence of multiple sites for deprotonation.

Marino, Jaén and Floyd reported a new strategy for α' substitution of α,β -epoxyketones.¹⁹ This approach is based on the stereospecific 1,4-addition of organocuprates to enol silyl ethers of α,β -epoxyketones or the enol esters, and represents an umpolung of reactivity for the carbon atom α to the ketone functionality (**Scheme 29**).



Scheme 29



Scheme 30

Since the organocopper reagents are not subject to the steric requirements of the electrophile or the reactivity profiles of the nucleophile, they provide a unique route for the introduction of vinyl, aryl, and highly substituted alkyl groups α to the ketone group. The organocopper method also allows stereochemical control of the newly introduced α' -substituent.
This reaction was found to be applicable to a variety of substituted epoxycyclohexenyl silyl ethers²¹ (Scheme 30). In all of the cases in Scheme 30, anti- S_N2' products 78 were obtained.

Marino applied the enol phosphate methodology to a synthesis of α -multistriatin (86), an aggregation pheromone of the European elm bark beetle.²¹

This synthesis is noteworthy for its use of sequential regio- and stereoselective S_N2' addition for elaboration of the carbon framework. Epoxyketone **66a**, available from 1,3-cyclohexadiene monoepoxide (**56**), as outlined in **Scheme 25** and **Scheme 26** (followed by oxidation), was converted into the enol phosphate **79**. Addition of MeCuCNLi gave the *anti* S_N2' product **80**. Hydrogenolysis of the phosphonate with Li/NH₃ yielded the allylic alcohol **81**, which underwent sequential hydroxyl-directed epoxidation and oxidation to the epoxyketone **82**. This was converted into the alkylidene epoxide **83** through Wittig methylenation. Compound **83**, when treated with MeCuCNLi, gave the S_N2' product **84** exclusively. Acetylation, followed by ozonolysis and reduction, afforded triol **85**, from which natural product (±)- α -multistriatin (**86**) could be elaborated (**Scheme 31**).



Scheme 31

The use of 1,3-cyclopentadiene monoepoxide would afford a 2,3-dialkylated cyclopentanone. A potential application is described in **Scheme 32**. However, it should be noted that the stereochemistry of the second S_N2' addition would have to be syn, in contrast to all known examples in cyclohexene systems. In the event, syn addition was found to occur.









This methodology has been successfully used in the total synthesis of prostaglandins by Marino and Pradilla.²² As shown in **Scheme 33**, epoxide **87** was treated with organocuprate **91**, and compound **92** was obtained in 80% yield. Several transformations led to the epoxy enol silyl ether **94**, which afforded an 8:1 mixture of S_N2 ' adducts in 80% yield, upon treatment with the cyanocuprate **95**. Separation of the C-15 diastereoisomers, followed by sequential deprotection and oxidation, led to racemic PGE1 (**97**) (**Scheme 33**).

Mechanistic Considerations

Cuprate reagents show remarkable anti-selectivity in S_N2 ' reactions for a wide variety of systems, the only cases of clear syn predominance that do not involve chelation of the cuprate to the leaving group can be attributed to overriding steric constraints. The strong anti selectivity and the high rates of reaction of organocopper reagents as compared to other carbon nucleophiles imply that some special structural feature of the cuprate may play a pivotal role in S_N2 ' displacement.

Corey and Boaz suggest that this preference derives from orbital symmetry.²³ Accordingly, nucleophilic displacements by d^{10} copper most likely will involve an electron pair in a sterically accessible high energy d orbital. The electron density in this orbital will have the usual binodal symmetry and, moreover, it will be exceedingly diffuse due to

electron-electron repulsion in d¹⁰ systems, and this characteristic leads to bidentate overlap in allylic systems so that the transition state for $S_N 2'$ attack has some $S_N 2$ character. The binding in this transition state can be described as resulting from the simultaneous interaction of a copper d orbital with the LUMO (π^*) at the γ carbon and, to a considerably smaller extent, with the antibonding orbital (σ^*) at the backside of the α carbon. Such binding is pictured in Scheme 34. If the phasing of the π^* (C=C) and σ^* (C-X) orbitals is appropriate for an $S_N 2'$ transition state, the symmetry of the d(Cu) orbital allows binding between d(Cu), π^* (C=C) and σ^* (C-X) orbitals, as shown in Scheme 34. Thus, these cuprate displacements possess $S_N 2'$ and $S_N 2$ character, with the latter stereoelectronically controlling the anti stereochemistry.



Scheme 34

(II) Results and discussion



Scheme 35

During studies on the degradation of mevinolin (98a) and compactin (98b) to the enone $99^{24,25}$ (Scheme 35), the response of vinyl epoxide 100a to (PhMe₂Si)₂CuLi^{26,27,28} (prepared from CuI and PhMe₂SiLi²⁹) was examined, and two isomeric products 101a and 102a were obtained. The ratio of these isomers was variable and the major product from one experiment could be the minor product in next run. For this reason, PhMe₂SiLi²⁹ was tried. Although the desired allylic silane 101a was produced, 24,30 the yields were erratic (40-91%) and also depended inversely on the scale of the reaction. When the compactin series (100b)²⁵ was examined later, a complex mixture was obtained with the lithium reagent; therefore, it was decided to try silyl cuprates, but made this time from CuCN.²⁹ Such reagents³¹ gave reproducible results and afforded the isomeric silanes 101b and 102b. The ratio of the isomers, which was established without

separation, by ¹H NMR measurements, depended on the stoichiometry of the cuprate mixture (Scheme 36).



Scheme 36

The stereochemistry of 101b and 102b was established by ¹H NMR measurements, the characteristic features being that $J_{7,8} = 0$ Hz and $J_{6,7} = 3.4$ Hz for 101b, while $J_{7,8} = 5.5$ Hz and $J_{6,7} = 0$ Hz for 102b. Examination of Dreiding models shows that the J-values are entirely consistent with the dihedral angles measured on the models.

It is not clear why the different reagent mixtures produce stereoisomeric products. Reaction of carbon organocuprates with vinyl epoxides generally^{32,33} proceeds by an *anti*-S_N2' pathway,³² as discussed above, but the mechanism is clearly complicated.^{23,32,33} In the present case, additional considerations are that the effective reagent in each of the PhMe₂SiLi/CuCN equilibrium mixtures^{29b} has different steric requirements and offers different opportunities for prior complexation with the epoxide oxygen.

The mevinolin series was reexamined next, but using the CuCN-derived reagents, and entirely comparable results were found (**Scheme 36**). Again, the multiplicity of the vinyl signal in the ¹H NMR spectra was characteristic, being a singlet for **101a** and a doublet (J = 4.5 Hz) for **102a**.

These unexpected results mean that the stereochemistry of the products can be controlled by using different relative amounts of PhMe₂SiLi and CuCN. In order to find if this method is general, we studied two other examples.

We have treated vinyl epoxide **87** with the three silyl cuprates (**Scheme 37**), and once more the outcome was found to be sensitive to reagent composition. However, the results are now complicated by direct epoxide opening, which was not observed in the mevinolin (**100a**) or compactin (**100b**) series. Addition of pure PhMe₂SiLi to compound **87** gave only anti-S_N2 product **105** in 27% yield. A 1:1 mixture of PhMe₂SiLi and CuCN yielded both anti and syn S_N2' products **103** and **104** (total yield 72%) in the isomeric ratio of 5.6:1. But the syn-S_N2' product **104** was not observed at all in the 2:1 and 3:1 mixtures; in both these cases, a mixture of anti-S_N2' product **103** and anti-S_N2 product **105** was obtained (79% yield for the 1:2 case and 70% for the 1:3 mixture) (**Scheme 37**).



reagents	103	104	105	total yield%
PhMe ₂ SiLi	x	X	27%	27%
1:1/PhMe ₂ SiLi:CuCN	5.6	1	x	72%
2:1/PhMe ₂ SiLi:CuCN	1.17	x	1	79%
3:1/PhMe ₂ SiLi:CuCN	1	x	1.5	70%

Scheme 37

We were unable to purify the anti- S_N^2 product 105, and so its *p*-nitrobenzoyl ester 106 was prepared (Scheme 38) in order to get good spectral data. The structure and stereochemistry of the product from 87 were readily apparent from the ¹³C and ¹H NMR spectra, taken together with

mechanistic considerations. We assume that direct epoxide opening of **87** with PhMe₂SiLi occurs by the normal *trans* pathway. Assignment of *cis* and *trans* stereochemistry to the S_N2' products **103** and **104** was guided by the characteristic chemical shift difference^{9a} between the two methylene protons, the *cis* isomer having the larger difference (0.57 ppm for *cis*-product **104** and 0.26 for *trans*-product **103** isomer).



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Scheme 38
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1,3-Cyclohexadiene monoepoxide (56) was also examined by using three different stoichiometries of the cuprate mixture, and the results are shown in Scheme 39. Addition of pure PhMe₂SiLi to epoxide 56 produced only the anti-S_N2 product 109. In the case of the 1:1 ratio (PhMe₂SiLi/CuCN), only the anti-S_N2' product 107 was obtained in 70% yield. The corresponding 2:1 and 3:1 cuprate reagents yielded a mixture of anti-S_N2' compound 107 (major product) and anti-S_N2 compound 109 in a yield of 94% for 2:1 reagent combination, and in 90% yield for the for 3:1 combination. In this example, the syn-S_N2' product 108 was not observed at all. Evidently, direct epoxide opening, in the case of simple sixmembered rings, is easier than the syn-S_N2' pathway.



Scheme 39

The stereochemistry of **107** and **109** could be assigned only after hydrogenation to the corresponding cyclohexanols **110** and **111** (Scheme 40).

Pure (¹H NMR, 200 MHz) trans-4-(dimethylphenylsilyl)cyclohexanol (**111**) has: ¹³C NMR (75.5 MHz, C₆D₆): δ -5.02 (CH₃Si), 24.67 (CHSi), 26.23 (CH₂), 37.53 (CH₂), 70.70 (CHOH), 128.00 (aryl CH), 129.11 (aryl CH), 134.14 (aryl CH), 138.24 (aryl CSi). ¹H NMR (C₆D₆): δ CHOH 3.2 (tt, J = 10.75, 3.8 Hz). For comparison purposes we note that *cis* and *trans* 4-*t*-butylcyclohexanol have δ CHOH at 3.75 and 3.24, respectively in C₆D₆. *Trans*-2-(Dimethylphenylsilyl)cyclohexanol (**110**) has: ¹³C NMR (75.5 MHz, C₆D₆) δ -3.24 (CH₃Si), -2.78 (CH₃Si), 25.41 (CH₂), 27.04 (CH₂, two overlapping signals), 34.80 (CHSi), 38.34 (CH₂), 72.86 (CHOH), 127.93 (aryl CH), 128.99 (aryl CH), 134.52 (aryl CH), 139.93 (aryl CSi). Four CH₂ signals in the ¹³C NMR spectrum of the saturated alcohol **110** indicated that the product **109** is generated though the *anti*-S_N2 pathway from **57**.





In summary, our results show that the course of reactions of dimethylphenylsilyl cuprates with vinyl epoxides is sensitive to the stoichiometry of the reagent and, in favorable cases, significant stereochemical control is possible in preparatively useful S_N2 ' experiments.

(III) General experimental procedures

Argon was purified by passage through a column $(3.5 \times 42 \text{ cm})$ of BASF R-311 catalyst and then through a similar column

of Drierite. Glassware was dried in an oven for at least 3 h before use (120 °C) and either cooled in a desiccator over Drierite, or assembled quickly, sealed with rubber septa, and allowed to cool under a slight static pressure of argon. Reaction mixtures were stirred by Teflon-coated magnetic stirring bars.

Solvents for chromatography or extractions were distilled before use.

Products were isolated from solution by evaporation under water pump vacuum at, or below, 30 °C, using a rotary evaporator.

Melting points were determined on a Kofler block melting point apparatus.

Commercial thin layer chromatography (TLC) plates (silica gel, Merck 60F-254) were used. Spots were detected by examination under UV light or by spraying the plate with a solution of phosphomolybdic acid, followed by charring on a hot plate. Silica gel for flash chromatography was Merck type 60 (230-400 mesh).

Dry solvents were prepared under an inert atmosphere and transferred by oven-dried syringes. Dry THF was distilled from Na and benzophenone ketyl, dry PhH was distilled from Na, and dry CH₂Cl₂ was distilled from CaH₂.

The symbols s', d', t', and q' used for 13 C NMR spectra indicate 0, 1, 2, or 3 attached protons.

Reaction of 1,3-cyclopentadiene monoepoxide (87) with

PhMe₂SiLi. (No CuCN).



1,3-Cyclopentadiene monoepoxide (**87**) (79 mg, 0.97 mmol) in THF (1.5 mL plus 2 x 1 mL as a rinse) was added dropwise to a stirred and cooled (-78 °C) solution of PhMe₂SiLi²⁹ (0.33 M in THF, 3.0 mL, 0.99 mmol). Stirring was continued for 3 h, saturated aqueous NaHCO₃ (1 mL) was added, the cold-bath was removed, and the mixture allowed to attain room temperature (*ca.* 20 min). The mixture was diluted with EtOAc (20 mL), washed with saturated aqueous NaHCO₃ (2 x 15 mL), water (1 x 20 mL) and brine (1 x 20 mL), dried (MgSO₄), and evaporated. Flash chromatography of the residue over silica gel (5 x 25 cm), using 1:9 EtOAc-hexane and then 2:8 EtOAchexane, gave **105** (53 mg, 27%) (see **105** data after).

We did not detect (TLC) the 1,4-cis or trans isomers in the reaction mixture.

Reaction of 1,3-cyclopentadiene monoepoxide (87) with PhMe₂SiLi and CuCN (1:1 molar ratio of PhMe₂SiLi and CuCN).



Copper(I) cyanide (Aldrich, 105 mg, 1.18 mmol) was suspended in THF (1.5 mL) at -23 °C. PhMe₂SiLi (0.3 M in THF, 4 mL, 1.19 mmol) was added dropwise with stirring and, after 30 min, the mixture was cooled to -78 °C. After 10 min cyclopentene oxide (87) (104 mg, 0.99 mmol) in THF (0.5 mL plus 2 x 0.5 mL as a rinse) was added dropwise (over 2 min). Stirring was continued for ca. 3 h and then saturated aqueous NaHCO3 was added (2 mL). The cold-bath was removed and the mixture was allowed to attain room temperature. The mixture was diluted with EtOAc (20 mL) and washed with saturated aqueous NaHCO₃ (2 x 15 mL), water (1 x 20 mL) and brine (1 x 20 mL). The organic extract was dried (MgSO₄) and evaporated. Flash chromatography of residue over silica gel (5 x 25 cm), using first 1:9 EtOAc-hexane and then 2:8 EtOAchexane, gave two the isomers (186 mg, 72%) 104 (less polar) and 103 (more polar) in a 15:85 ratio, respectively, as judged by ¹H NMR (300 MHz). Pure (¹H NMR, 300 MHz) compound **104** had: FTIR (CHCl₃, cast) 3300 cm⁻¹; ¹H NMR (300 MHz, C₆D₆) δ 0.15 (s, 6 H), 0.6-0.8 (br s, 1 H), 1.5-1.6 (m, 1 H), 1.85-1.95 (m, 1 H), 2.05-2.1 (m, 1 H), 4.65-4.75 (br s, 1 H),5.55-5.65 (m, 1 H), 5.65-5.75 (m, 1 H), 7.15-7.30 (m, 3 H),

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7.35-7.45 (m, 2 H); ¹³C NMR (75.5 MHz, C_6D_6) δ -4.68, -4.39, 34.26, 35.18, 77.66, 129.50, 131.75, 133.44, 134.37, 135.92, 137.68; exact mass *m/z* calcd for $C_{13}H_{18}OSi$ 218.1127, found 218.1106. Pure (¹H NMR, 300 MHz) compound **103** had: FTIR (CHCl₃, cast) 3320 cm⁻¹; ¹H NMR (300 MHz, C_6D_6) δ 0.1 (s, 6 H), 1.7-1.9 (m, 1 H), 2.0-2.15 (m, 2 H), 2.15-2.3 (m, 1 H), 4.64-4.7 (br s, 1 H), 5.6-5.65 (m, 1 H), 5.7-5.75 (m, 1 H), 7.2-7.3 (m, 3 H), 7.35-7.45 (m, 2 H); ¹³C NMR (75.5 MHz, C_6D_6) δ -4.82, -4.67, 33.29, 36.13, 77.66, 129.99, 131.93, 134.09, 135.54, 137.96 (two overlapping signals); exact mass *m/z* calcd for $C_{13}H_{16}Si$ 200.1021, found 200.1023.

Reaction of 1,3-cyclopentadiene monoepoxide (87) and PhMe₂SiLi and CuCN (2:1 molar ratio of PhMe₂SiLi and CuCN).



The procedure for the 1:1 ratio of reagents was followed, using copper(I) cyanide (126 mg, 1.41 mmol) in THF (1.5 mL), PhMe₂SiLi (0.33 M in THF, 8.5 mL, 2.81 mmol), and 1,3-cyclopentadiene monoepoxide (87) (78 mol% in CH_2Cl_2 , 143 mg, 1.36 mmol) in THF (0.5 mL plus 2 x 1.5 mL as a rinse). A mixture (238 mg, 79%) of 105 and 103 in a ratio of 46:54, respectively, was obtained. Pure (¹H NMR, 300 MHz) compound **105** (the less polar isomer) had: FTIR (CHCl₃, cast) 3320 cm⁻¹; ¹H NMR (300 MHz, C₆D₆) δ 0.1 (s, 6 H), 2.1-2.15 (br s, 1 H), 2.15.2.25 (m, 3 H), 4.2-4.3 (br s, 1 H), 5.4-5.6 (m, 1 H), 5.6-5.7 (m, 1 H), 7.2-7.3 (m, 3 H), 7.4-7.5 (m, 2 H); ¹³C NMR (75.5 MHz, C₆D₆) δ -4.47, 43.87, 46.67, 73.48, 125.10, 128.05, 129.51, 130.20, 135.91, 137.89. See above for characterization data for **103**.

Reaction of 1,3-cyclopentadiene monoepoxide (87) with PhMe₂SiLi and CuCN (3:1 molar ratio of PhMe₂SiLi and CuCN).



The procedure for the 1:1 ratio of reagents was followed, using copper(I) cyanide (97 mg, 1.08 mmol) in THF 1.5 mL, PhMe₂SiLi (0.33 M in THF, 9.8 mL, 3.23 mmol) and cyclopentadiene oxide (87) (78 mol% in CH_2Cl_2 , 83 mg, 1.01 mmol) (1 mL plus 2 x 1 mL as rinse). A mixture (202 mg, 70%) of 105 (less polar) and 103 (more polar) in a ratio of 60:40, respectively, was obtained.

Characterization of 1,2-addition product from 1,3-

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cyclopentadiene monoepoxide (87).



Compound 105 (132 mg, 0.61 mmol) in CH₂Cl₂ (1 mL plus 2 x 1 mL as rinses) and then dry Et_3N (0.2 mL, 1.4 mmol) were added dropwise to a stirred and cooled (ice-bath) suspension of p-nitrobenzoyl chloride (241 mg, 1.30 mmol) in CH₂Cl₂ (2 mL). The ice-bath was left in place, but not recharged, and stirring was continued for 10 h. The mixture was then evaporated, and flash chromatography of the residue over silica gel (5 x 25 cm), using 1:9 EtOH-hexane and then 2:8 EtOAc-hexane, gave pure (¹H NMR, 300 MHz) **106** (133 mg, 60%): FTIR (CH₂Cl₂ cast) 1720 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 0.4 (s, 6 H), 2.4-2.5 (m, 3 H), 5.55 (d, J = 5 Hz, 1 H), 5.75-5.80 (m, 1 H), 5.75-5.86 (m, 1 H), 7.3-7.4 (m, 3 H), 7.5-7.6 $(m, 2 H), 8.1-8.2 (m, 2 H), 8.2-8.3 (m, 2 H); {}^{13}C NMR (75.5)$ MHz, CDCl₃) δ -4.72, -4.08, 40.44, 43.57, 78.87, 123.44, 124.91, 127.84, 129.44, 130.44, 130.68, 133.82, 136.23, 136.65; mass (CI) m/z calcd for $C_{20}H_{25}N_2O_4Si$ 367, found 385 (M $+ NH_4$).

Reaction of 1,3-cyclohexadiene monoepoxide (56) with PhMe₂SiLi. (No CuCN).



1,3-Cyclohexadiene monoepoxide (56) (122 mg, 1.27 mmol) in THF (0.5 mL plus 2 x 0.5 mL as rinse) was added dropwise to a stirred and cooled (-78 °C) solution of PhMe₂SiLi (0.36 M in THF, 4.0 mL, 1.44 mmol). Stirring was continued for 3 A mixture (1 mL) of 1:9 saturated aqueous NH4Cl and h. concentrated aqueous ammonia solution was added, the coldbath was removed, and the mixture allowed to attain room temperature (ca. 20 min). The mixture was diluted with EtOAc (20 mL) and washed with a mixture $(2 \times 15 \text{ mL})$ of 1:9 saturated aqueous NH4Cl and concentrated aqueous ammonia solution, and brine $(1 \times 20 \text{ mL})$. The organic extract was dried (MgSO₄) and evaporated. Flash chromatography of the residue over silica gel (2.5 x 25 cm), using first 1:9 EtOAchexane and then 2:8 EtOAc-hexane, gave pure $(^{1}H NMR, 300 MHz)$ **109** (93 mg, 40%): FTIR (CHCl₃, cast) 3330 cm⁻¹; ¹H NMR (300 MHz, C₆D₆) δ 0.35 (s, 6 H), 0.95-1.10 (br s, 1 H), 1.45-1.55 (m, 2 H), 1.75-1.85 (m, 2 H), 1.95-2.10 (m, 1 H), 3.75-3.85 (q, J = 6 Hz, 1 H), 5.50-5.60 (m, 2 H), 7.2-7.25 (m, 3 H),7.40-7.51 (m, 2 H); ¹³C NMR (75.5 MHz, C₆D₆) δ -4.10 (q'), -3.62 (q'), 22.44 (t'), 30.68 (t'), 36.42 (d'), 67.60 (d'), 125.00 (d'), 125.68 (d'), 128.11 (d'), 129.36 (d'), 134.28 (d'), 137.98 (s'); exact mass m/z calcd for $C_{14}H_{20}OSi$

Reaction of 1,3-cyclohexadiene monoepoxide (56) with PhNe₂SiLi and CuCN (1:1 molar ratio of PhNe₂SiLi and CuCN).



Copper(I) cyanide (145 mg, 1.62 mmol) was suspended in THF (1.5 mL) at -30 °C. PhMe₂SiLi (0.30 M in THF, 5.2 mL, 1.56 mmol) was added dropwise with stirring and, after 30 min, the mixture was cooled to -78 °C. After 10 min, cyclohexene oxide (**56**) (118 mg, 1.22 mmol) in THF (0.5 mL plus 2 x 0.5 mL as a rinse) was added dropwise (over 20 min). Stirring was continued for ca. 3 h, and then a mixture of 1:9 saturated aqueous NH₄Cl and concentrated aqueous ammonia solution (1 mL) was added. The cold-bath was removed and the mixture was allowed to attain room temperature. The mixture was diluted with EtoAc (20 mL) and washed with a mixture of 1:9 saturated aqueous NH₄Cl and concentrated aqueous ammonia solution (2 x 15 mL), water (1 x 20 mL) and brine (1 x 20 mL). The organic extract was dried (MgSO₄) and evaporated. Flash chromatography of residue over silica gel (2.5 x 25 cm), using first 1:9 EtOAc-hexane and then 2:8 EtOAc-hexane, gave pure (¹H NMR, 300 MHz) **107** (196 mg, 70%): FTIR (CHCl₃, cast) 3400 cm⁻¹; ¹H NMR (300 MHz, C₆D₆) δ 0.15 (s, 6 H), 1.30-1.45 (m, 1 H), 1.45-1.55 (m, 1 H), 1.55-1.75 (m, 2 H), 1.70-1.90 (m, 1 H), 2.9-3.05 (br s, 1 H), 4.0-4.15 (br s, 1 H), 5.6-5.7 (m, 1 H), 5.75-5.85 (m, 1 H), 7.18-7.25 (m, 3 H), 7.35-7.45 (m, 2 H); ¹³C (75.5 MHz, C₆D₆) δ -4.79 (q'), -4.71 (q'), 20.95 (t'), 26.12 (d'), 32.4 (t'), 65.98 (d'), 128.06 (d'), 129.32 (d'), 129.62 (d'), 130.46 (d'), 134.17 (d'), 137.79 (s'); exact mass *m/z* calcd for C₁₄H₂₀OSi 232.1377, found 232.1282.

Reaction of 1,3-cyclohexadiene monoepoxide (56) with PhNe₂SiLi and CuCN (2:1 molar ratio of PhNe₂SiLi and CuCN).



The procedure for the 1:1 ratio of reagents with 1,3cyclohexadiene monoepoxide (56) was followed, using copper(I) cyanide (119 mg, 1.33 mmol) in THF (1.5 mL), PhMe₂SiLi (0.32 M in THF, 8.2 mL, 2.62 mmol), and cyclohexene oxide (98 mg, 1.02 mmol) in THF (0.5 mL plus 2 x 0.5 mL as a rinse). A mixture (220 mg, 94%) of 109 (less polar) and 107 (more polar) was obtained, in a ratio of 17:83, respectively.

Reaction of 1,3-cyclohexadiene monoepoxide (56) with PhNe₂SiLi and CuCN (3:1 molar ratio of PhNe₂SiLi and CuCN).



The procedure for the 1:1 ratio of reagents with 1,3cyclohexadiene monoepoxide (**56**) was followed, using copper(I) cyanide (101 mg, 1.13 mmol), PhMe₂SiLi (0.32 in THF, 10.5 mL, 3.36 mmol) and 1,3-cyclohexadiene monoepoxide (**56**) (98 mg, 1.02 mmol) in THF (0.5 mL plus 2 x 0.5 mL as rinse). A mixture (212 mg, 90%) of **109** (less polar) and **107** (more polar) was obtained in a ratio of 40:60, respectively.

Hydrogenation of 1,2-addition product 109 from 1,3cyclohexadiene monoepoxide (56).



109

Pd/C (10%w/w, 50 mg) was added to a solution of 109 (150 mg, 0.64 mmol) in EtOAc (2 mL) and the mixture was stirred vigorously under hydrogen (balloon). After 2 h the mixture was filtered and evaporated. Flash chromatography of the residue over silica gel (1 x 25 cm), using 1:4 EtOAc-hexane, gave pure (¹H NMR, 300 MHz) **110** (39 mg, 26%): FTIR (CHCl₃, cast) 3400 cm⁻¹; ¹H NMR (300 MHz, C₆D₆) δ 0.4 (two singlets, 6 H), 0.60-0.70 (m, 1 H), 0.8-1.1 (m, 5 H), 1.35-1.45 (m, 1 H), 1.50-1.60 (m, 2 H), 1.65-1.75 (m, 1 H), 3.10-3.25 (m, 1 H), 7.15-7.35 (m, 3 H), 7.55-7.65 (m, 2 H); ^{13}C NMR (75.5 MHz, C_6D_6) δ -3.24 (q'), -2.78 (q'), 25.41 (t'), 27.04 (t', two overlapping signals), 34.80 (d'), 38.34 (t'), 72.86 (d'), 127.93 (d'), 128.99 (d'), 134.52 (d'), 139.93 (s'); exact mass m/z calcd for $C_{14}H_{20}Si$ (M - H₂O) 216.1334, found 216.1344; mass (CI) m/z calcd for 234, found 252 ($C_{14}H_{22}OSi +$ NH_4).

Hydrogenation of 1,2-addition product 109 from 1,3cyclohexadiene monoepoxide (56). Use of Wilkinson's catalyst.



109

A solution of **109** (150 mg, 0.64 mmol) in dry PhH (1 mL plus 1 mL as a rinse) was injected into a stirred solution of Wilkinson's catalyst (184 mg, 0.20 mmol) that had been presaturated with hydrogen (40 psi) for 1 h. The mixture was stirred at room temperature for 12 h under hydrogen (40 psi), the solvent was evaporated and the mixture was filtered through a small pad of Florisil (1 x 5 cm), using 10% EtOAchexane. Evaporation of the solvent and flash chromatography of the residue over silica gel (1 x 25 cm), using 20% EtOAchexane, gave **110** (101 mg, 67%).

Hydrogenation of trans 1,4-addition product 107 from 1,3-cyclohexadiene monoepoxide (56).



107

111

Pd/C (10% w/w, 30 mg) was added to a solution of 107 (85 mg, 0.36 mmol) in EtOAc (2 mL) and the mixture was stirred vigorously under hydrogen (balloon) for 10 h. The mixture was filtered and evaporated. Flash chromatography of the residue over silica gel (1 x 25 cm), using 20% EtOAc-hexane gave pure (¹H NMR, 300 MHz) 111 (46 mg, 52%): FTIR (CHCl₃ cast) 3400 cm⁻¹; ¹H NMR (300 MHz, C₆D₆) δ 0.1 (s, 6 H), 0.4-

0.5 (m, 1 H), 0.9-1.1 (m, 4 H), 1.1-1.2 (br s, 1 H), 1.55-1.65 (d, J = 12, 1 H), 1.95-2.05 (br, 1 H), 3.2 (tt, J =10.75, 3.8 Hz, 1 H), 7.15-7.25 (m, 3 H), 7.35-7.45 (m, 2 H); ¹³C NMR (75.5 MHz, C₆D₆) δ -5.02 (q'), 24.67 (d'), 26.22 (t'), 37.53 (t'), 70.70 (d'), 128.00 (d'), 129.11 (d'), 134.14 (d'), 138.28 (s'); exact mass m/z calcd for C₁₄H₂₂OSi 234.1488, found 234.1432.

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(I) Introduction

Angiotensin¹

The crude saline extracts of kidney contain a pressor principle called renin. It produces persistent hypertension (elevated blood pressure) in humans and animals by constricting the renal arteries. Renin is an enzyme that acts on a plasma protein substrate (called angiotensinogen) to catalyze the formation of the actual pressor material, a peptide called angiotensin.

In the 1950's, Skeggs and Peart determined the amino acid composition and sequence of angiotensin. Two forms were recognized. The first is a decapeptide (angiotensin I) and the second an octapeptide (angiotensin II). The latter is formed from angiotensin I by enzymatic cleavage by another enzyme, termed angiotensin-converting enzyme. The octapeptide, angiotensin II, was shown to be the more active species, and its synthesis in 1957 by Schwyzer and by Bumps² made the material available for intensive study.

The synthesis and degradation of the angiotensins are complex processes that are outlined in **figure 1**. Briefly, the process is initiated when the enzyme renin acts on angiotensinogen (the renin substrate) to release the decapeptide angiotensin I. This decapeptide has limited pharmacological activity, but it is cleaved by angiotensinconverting enzyme (ACE) to yield the highly active

octapeptide angiotensin II. This, in turn, undergoes hydrolysis by an aminopeptidase to yield the heptapeptide angiotensin III, which is also pharmacologically active. Further cleavage yields peptides with little activity. In an alternative (minor) path, angiotensin-converting enzyme and aminopeptidase act in the opposite sequence such that the decapeptide, angiotensin I, is hydrolyzed first to [des-asp^I] angiotensin I which, like the parent compound, has limited pharmacological activity. The [des-asp^I] angiotensin I is then cleaved by angiotensin-converting enzyme to form the active angiotensin III.^{2,3}



Formation and destruction of anxiotensins.

figure 1

There are many fatal diseases which are related to excessive amounts of angiotensin II in blood. The compound causes vasoconstriction and often has an indirect effect on the heart, such that cardiac output is lowered. Angiotensin is the most potent pressor agent known; on a molar basis, it is about 40 times more powerful than norepinephrine. It also causes kidney disease due to its indirect effects on renal tubular function, mediated by aldosterone. Angiotensin also influences urine formation through hemodynamic and intrarenal actions that interact in a complex way.

The pharmacological properties so far described are those of angiotensin II. Angiotensin I has less than 1% of the intrinsic activity of angiotensin II. However, angiotensin III retains most of the activity of angiotensin II, although in most instances it is somewhat weaker.

Angiotensin II Antagonists and inhibitors of ACE



The renin-angiotensin system plays an important role in the control of renal function and blood pressure and in the pathogenesis of some forms of hypertension. Consequently, much work has been focused on developing agents that block the renin-angiotensin system so as to maintain blood pressure in the proper range.

In the 1970's, two distinct classes of effective inhibitors of the renin-angiotensin system were identified: angiotensin II antagonists, which block receptors for the peptide, and converting enzyme inhibitors, which slow the rate of formation of angiotensin II from its inactive precursor.



Losartan, DUP 753



Telmisartan, BIBR 277

The useful antagonists of angiotensin II are slightly modified congeners in which agonist activity is profoundly attenuated by replacement of phenylalanine in position 8 with some other amino acid. The substances $[Sar^{I}, Val^{5}, Ala^{8}]$ angiotensin (1-8) octapeptide, known as the Saralasin series, were introduced by Pals in 1971. Although angiotensin II analogues are highly specific antagonists, they also retain some agonist activity, which complicates interpretation of some of their effects.⁴ However, direct blockade of the angiotensin II receptor (AT₁ receptor) has recently become feasible by the development of a series of orally active nonpeptide antagonists for the AT₁ receptor. DUP 753 (Losartan, NK954) and BIBR 277 (phase III, Boehringer Ingelheim), shown above, have been found to be very powerful AT₁ antagonists.^{5,6,7} These nonpeptide antagonists are highly selective for the AT₁ receptor and, in contrast to the peptide-based antagonists of angiotensin analogues, they do not show any partial agonism.

Angiotensin-converting enzyme (ACE) inhibitors are also commonly used for inhibiting the renin-angiotensin system (RAS).¹ The essential effect of these agents on the RAS is to inhibit conversion of the relatively inactive angiotensin I into the active angiotensin II (or the conversion of [des-Asp^I] angiotensin I to angiotensin III). In this way they attenuate or abolish responses to angiotensin I. The converting enzyme inhibitors are highly specific drugs. They do not interact directly with other components of the reninangiotensin system, including the receptor for the peptide. Also, research has revealed that ACE inhibitors cause a greater fall in blood pressure than does angiotensin II antagonism.



Captopril,⁸ the first orally active ACE inhibitor, has been marketed for the treatment of severe or drug-resistant hypertension since the 1980's. Unfortunately, it may cause some potentially hazardous side effects, perhaps related to its sulfhydryl moiety.⁹ Enalapril, a nonsulfhydrylcontaining ACE inhibitor, is also used for treatment of diseases caused by angiotensin. Enalapril is a prodrug that is not itself highly active; it must be hydrolyzed to its active parent dicarboxylic acid, enalaprilate. Enalapril has been found to be more potent than Captopril and to inhibit ACE for longer periods.^{8,9,10}

A58365A and A58365B, Inhibitors of ACE

Culture A58365, NRRL 15098, identified as a new strain of Streptomyces chromofuscus, was isolated from a soil sample collected in Brazil. It was found to produce two novel angiotensin-converting enzyme inhibitors, A58365A and A58365B. They are homologous nitrogen-containing bicyclic structures of molecular formulae $C_{12}H_{13}NO_6$ for A58365A and $C_{13}H_{15}NO_6$ for A58365B.^{11,12}







A58365B 2



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The structure elucidation of the two new ACE inhibitors has been described in detail.¹³ A58365A was determined to be 3-carboxy-1,2,3,5-tetrahydro-8-hydroxy-5-oxo-6-indolizinepropanoic acid (1); A58365B was the homologous 4-carboxy-1,3,4,6-tetrahydro-9-hydroxy-6-oxo-2*H*-quinolizine-7-propanoic acid (2). A58365A is, in fact, a naturally occurring conformationally restricted analog of α -methylglutaryl-Lproline (3), which was a part of the structure activity relationship studies leading to captopril.

Nakatsukasa and his assistants proposed a biosynthetic pathway for A58365 by Streptomyces chromofuscus NRRL 15098.¹⁴ Fermentation studies afforded an increase in the amount of the ACE inhibitor from less than 1 μ g/mL to 20 μ g/mL. Proline was the obligatory supplement for ACE inhibitor
biosynthesis. Without proline, less than $1 \mu g/mL$ of both A58365A and A58365B was synthesized. D-proline or Lhydroxyproline could be substituted in place of L-proline, but were not superior to L-proline. Greater amounts of A58365A were synthesized when L-tyrosine was added to the medium. However, this regulation was evident only in combination with proline but not with tyrosine alone. The addition of lysine to the proline-supplemented medium resulted in regulating the fermentation to produce greater amounts of A58365B. Like the regulation of A58365A by proline and tyrosine, stimulation of the synthesis of A58365B was evident with proline and lysine but not lysine alone, suggesting that A58365B synthesis is closely linked to that of A58365A. The studies also show that A58365A is synthesized before A58365B.

Total Synthesis A58365A by Danishefsky and Fang¹⁵

Enantiomerically homogeneous A58365A in the natural *S*configuration was synthesized for the first time by Danishefsky and Fang in 1989. The synthesis was done in 11 steps, but the authors pointed out that there were clearly some significant yield problems in this first generation synthesis.

Their synthesis was accomplished by starting with Lpyroglutamate. The four-step conversion of commercially available L-pyroglutamic acid (**Scheme 1**) led to vinylogous 51

urethane 6, which was the key intermediate. The annulation of 6 with α -methyleneglutaric anhydride (7) formed the hexahydroindolizidine 8. Esterification, followed by DDO



Scheme 1

oxidization, then gave pyridone 10. Monocarboxylic acid 11 was obtained after debenzylation of compound 10. Carboxyinversion of 11 was done by successive treatment with DCC and MCPBA, to produce A58365A-dimethyl ester (12) and A58365Adimethyl ester meta-chlorobenzoate (13). Compounds 12 and 13 were converted into dibenzyl ester 14 by using Otera's catalyst, the esterification being done to facilitate purification. Catalytic hydrogenation (Pd/C) of pure 14 then afforded the pure natural product A58365A 1 (Scheme 2).



Scheme 2

Total Synthesis of (-)- λ 58365A and (±)- λ 58365B by Moeller and Wong¹⁶

An anodic amide oxidation has been used to synthesize (-)-A58365A and (\pm)-A58365B by Moeller and Wong.¹⁶ Both syntheses take advantage of the ability of electrochemical methods to selectively oxidize an amide in the presence of a disubstituted acetylene nucleophile (see below).

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(-)-**A**58365**A**

In the synthesis of (-)-A58365A, the key intermediate 17 for electrolysis was synthesized in 4 steps from S-(+)-2- pyrrolidinemethanol (Scheme 3).





The oxidation of key intermediate **17** proceeded (**Scheme 4**) in an undivided cell (81.5 mA, 5.2 F/mol) to afford the methoxylated amide **18**, which was treated with TiCl₄ to give the bicyclic vinyl chloride **19**. Crude **19** was immediately ozonolyzed, followed by workup with Zn/AcOH, and keto-amide **20** was obtained in this two step process.



Scheme 4

The keto-amide 20 was treated with triisopropylsilyl triflate and triethylamine to afford enol ether 21 (Scheme 5). Treatment of 21 with DDQ led to the six-membered ring aromatic compound 22, which was then stirred with a solution of 0.01 N HCl in order to cleave the silyl ether protecting group and converge on the synthesis of Danishefsky and Fang. The optical purity of the final product, A58365A, was ca. 90% (Scheme 5).

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Scheme 5

(±)-**A**58365B

For the synthesis of (\pm) -A58365B, a nearly identical procedure was used in order to convert the electrolysis substrate 23, derived from pipecolic acid, into the required 1-aza-2,5-dioxobicyclo[4.4.0]decane ring skeleton.

Key intermediate 23 for the anodic amide oxidation was prepared using the same chemistry as described above for the preparation of substrate 17. Then anodic amide oxidation, cyclization induced by titanium tetrachloride, and ozonolysis led to compound **25** (**Scheme 6**).



The ketone 25 was treated with lithium 2,2,6,6tetramethylpiperidide as the base, and the resulting enolate was quenched with benzeneselenenyl bromide; the selenated product 26 was obtained. Treatment of 26 with MCPBA led to formation of product 27. This over-oxidized product 27 could be reduced to the desired compound 28 by using triethylsilane and TFA. The synthesis was completed by treatment of 28 with 9 N HBr, so as to obtain the natural product (\pm) -A58365B (Scheme 7).



Scheme 7

Part 1 Attempted Synthetic Approaches to (\pm) -A58365B. Synthetic Plan



(±)-A58365B 2

The structure of (\pm) -A58365B (compound 2) does not appear to be especially complicated, but synthetic work, both our own and that done earlier,^{15,16} revealed a number of unexpected difficulties. In particular, generation of the C(7)-C(8) double bond of 2 is not at all straightforward because ring A is also susceptible to desaturation. Therefore, routes that call for introduction of that double bond late in the synthesis do not seem to be highly promising. After appreciable exploratory work, we were led to the use of a radical process to construct ring B, in a sequence by which the C(7)-C(8) unsaturation is introduced early - although in the disguised form of a spirolactone. Considering all of these factors, we decided to build the molecule 29.

We hoped that the acetylenic lactone 29, representing the basic features of the target, including the propionic side chain, could be cyclized by a radical process to produce the exocyclic olefin 30. Ozonolysis of olefin 30, followed by opening of lactone 31 and hydrolysis of the mono-methyl ester 32 should then lead to the natural product (\pm) -A58365B (Scheme 8). Our plan was to synthesize the key intermediate 29 from acetylene amine 33 and the known acid 34 (which we call the "Swiss acid")^{17,18} (Scheme 9).



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Scheme 9

First Approach

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Preparation of acetylenic amine 33 proceeded without incident. We started with commercial pyridine 2,6dicarboxylic acid (35). When this was treated with a mixture of methanol and water in the presence of sulfuric acid, monomethyl ester 36 and the corresponding bis-methyl ester were formed in 36% and 30% yield, respectively. Saturation of the aromatic ring of compound 36 was done by hydrogenation in the presence of PtO_2 as catalyst, and the amino acid **37** was obtained in almost quantitative yield. Refluxing the amino acid **37** with di-*tert*-butyl dicarbonate (BOC₂O) in THF led to formation of the protected amino acid **38**.

Some difficulty was encountered in reducing acid 38 to alcohol **39**. Following a literature¹⁹ procedure, we used borane-tetrahydrofuran - a standard method for reducing an acid to the corresponding alcohol - but we did not isolate any product in our case. Then borane-dimethyl sulfide²⁰ was tried, but only a 30% yield of the desired compound 39 was obtained. Unlike many other alcohols, compound 39 was difficult to detect by TLC, and so it was difficult to monitor the reaction. Many experiments were done in an effort to improve the yield, and it was eventually found that the reaction is best done in the presence of excess boranedimethyl sulfide at -15 °C to room temperature. At least 2 mole of reducing reagent are required in order to compensate for complexation of one equivalent of the reagent by the nitrogen. When done under optimum conditions, it is possible to isolate **39** in 80% yield. The reaction is also very sensitive to the quality of the reagent, and more than 2 mol of reagent are required if the reagent is not fresh (Scheme 10).



Scheme 11

Alcohol 39 was oxidized by the Swern method and two isomers of aldehyde 40 were obtained (81%) in a ratio of 15:1. The chromatographically faster-running isomer was the major product, and only this material was used in further synthetic work. Next, acetylene 41 was made from aldehyde 40 (major isomer) following a general methodology reported in the literature 21, 22 According to this method, the reagent dimethyl diazomethylphosphonate was prepared and purified by distillation under vacuum. We found that the reagent tended to explode during distillation (fortunately done on a small scale!). For safety reasons, the crude reagent was used in our work, and acetylene **41** was obtained in 75% yield. The reaction product 41 and the starting material 39 had the same R_f on TLC when 3:7-EtOAc-hexane was used. Finally, the acetylenic amine 33 was obtained in good yield after

deprotection of the nitrogen (Scheme 11).



Scheme 12

In coupling amine 33 and the Swiss acid, we first tried DCC as the coupling agent, but did not isolate any of the desired product. Usually, primary amines are used in DCC coupling reactions, but in our case, the amine 33 is secondary and, therefore, less reactive. In addition, Swiss acid 34 is very hindered, and this would make the coupling reaction more difficult. Coupling of the two subunits was eventually achieved by treating amine 33 with the acid chloride derived from the Swiss acid, and the desired product 29 was obtained (61%) as a mixture of two isomers, which could not be separated by chromatography (Scheme 12).

Unfortunately, radical cyclization of 29 did not proceed as we had expected. Under a variety of different conditions (Scheme 13), none of the cyclized compound 30 was observed, and only the reduced compound 42 was isolated.



Radical conditions	
Ph ₃ SnH, AIBN	benzene (reflux, slow addition)
Ph ₃ SnH, AIBN	toluene (reflux, slow addition)
Bu ₃ SnH, AIBN	benzene (reflux, slow addition)
Bu ₃ SnH, AIBN	toluene (reflux, slow addition)
Ph ₃ SnH, Et ₃ B	benzene (r. t.)

Scheme 13

Formation of the reduction product might be due to the conformation of 29. Based on the experimental result, we assume that the acetylene group is in the axial conformation. When radical 43 is generated, it abstracts hydrogen from triphenyltin hydride, or gives radical 44 by intramolecular hydrogen-transfer - a process which is often a competitive side reaction in six-exo cyclizations.²³ Radical 44 would be stabilized by two resonance structures involving the acetylene and nitrogen. Both pathways (see a and b in Scheme 14) lead to formation of compound 42, and so we

decided to embark on another approach to our target molecule.



Scheme 14

Second Approach

Cobalt-mediated radical cyclization has been reported to be a powerful synthetic method to form C-C bonds.²⁴⁻³⁰ These reactions proceed via electron transfer from a cobalt(I) species to the carbon-halogen bond in the substrate (45), followed by (i) intramolecular cyclization, (ii) in situ trapping of the product radical center 46 with Co(II), and (iii) dehydrocobaltation (1,2-elimination) (Scheme 15).





In our second approach, we decided to try this method for construction of ring B in A58365B (2). This synthetic strategy would require building the lactone olefin 49. Intramolecular radical cyclization, brought about by use of Co(I), followed by trapping of the radical intermediate, would form the organocobalt compound 51. Dehydrocobaltation of 51 by either irradiation or heating could generate the exocyclic olefin 30, from which the natural product 2 should be accessible, after ozonolysis and treatment with base (Scheme 16).





As can be seen, compound **49** can be disconnected into two simple precursors, Swiss acid **34** and amine **52** (**Scheme 17**).



Scheme 17

Amine 52 was synthesized by a two-step procedure.

Aldehyde **40** (major isomer) was converted into olefin **53** by Wittig reaction (84%), and then the BOC group was removed by the action of TFA (75%). Coupling of amine **52** and Swiss acid chloride led to formation of **49** (85%) as two fractions. These can be separated by chromatography (**Scheme 18**).



Scheme 18

The individual isomers of **49** were treated with Co(I), generated by reaction of Co(II) salophen **54**³¹ and reducing reagents, such as NaBH₄³² or sodium amalgam.³³ After a reaction period of 16 hours in the dark, the intermediate was either heated or irradiated, but only complex mixtures were obtained from this experiment (**Scheme 19**).



Scheme 19

A different, but related, procedure was explored to form ring B. For this strategy, aldehyde **55** was required. Sm(II)-mediated³⁴⁻³⁷ cyclization of aldehyde **55**, followed by oxidation, could led to ketone **31**, which should be convertible into A58365B (**2**) in two steps (**Scheme 20**).



The two isomers of aldehyde **55** were obtained from **49** by ozonolysis (80%), the isomer ratio being 1:3 (**Scheme 21**). The chromatographically slower-running isomer (major product) of **55** was a mixture of rotamers and very unstable. The chromatographically faster-running isomer was just a single compound, and could be crystallized from 1:1 EtOAc-hexane.



Scheme 21

X-ray analysis of this material (the faster-running

isomer of **55**) revealed that both ester and aldehyde groups were *cis* to each other and also that both of them occupied an axial conformation.





Scheme 22

The individual isomers of **55** were treated with SmI_2 , but only complex mixtures were obtained (**Scheme 22**), and we realized, of course, that this approach was not promising.

Third Approach

As discussed above, all attempts to make B ring in compound 2 were unsuccessful, and we next considered the possibility of radical cyclization, by using substrate 57. As shown in Scheme 23, 6-exo radical cyclization of 57, followed by 1,2-elimination, would lead to olefin 30, which could subsequently be elaborated into A58365B (2).



Scheme 23

From a retrosynthetic standpoint, compound **57** could be built by joining amine **59** and Swiss acid **34** (**Scheme 24**).



Amine **59** appears to have a very simple structure, but our attempts to make it were not successful.



Scheme 25

We tried to prepare compound **60a** by the Horner-Emmons method, 38 but less than 10% of the desired compound was

obtained. We also attempted an alternative pathway, by generating compound **61** first, and then converting it into **60**. Unfortunately, the oxidation step did not appear to give us the desired product (the sulfone) because of double bond migration (**Scheme 25**).

Another approach to form ring B is related to the chemistry of **Scheme 23**, and is summarized in **Scheme 26**.



(±)-A58365B 2

Scheme 26

We hoped that radical cyclization of precursor 63, followed by deprotection of the benzyl group, would lead to carboxylic acid 64. Conversion of 64 into 65 would then be done by the Hunsdiecker method, and 1,2-elimination of 65 could then provide us with the desired olefin 30 (Scheme 26). It seemed that radical precursor 63 could be assembled from two components, amine 66 and Swiss acid 34 (Scheme 27).



Scheme 27

Scheme 28 shows the route leading to 63. Aldehyde 40 (major isomer) was treated with dry BnOOCCH=PPh₃, prepared from a 1 N solution of sodium hydroxide and BnOOCCH₂PPh₃Br, followed by azeotropic drying with benzene. The conjugated benzyl ester 67 was generated in 94%.^{39,40}

Removal of the BOC group of **67** was accomplished by treatment with dry TFA (93%). After that, coupling of amine **66** and Swiss acid chloride led to formation of the desired product **63** as two chromatographically separable isomers in a ratio of 4:6 (total yield 83%) (**Scheme 28**). The chromatographically slower-running material was the major isomer. Based on the ¹H and ¹³C NMR spectra, the fasterrunning isomer was a single compound, and the other one was a mixture of rotamers.



Scheme 28



However, when the individual fractions of **63** were treated with triphenyltin hydride and AIBN under standard thermal conditions, the ring closure product **68b** was obtained only from the slower-running material **63b** (in 75% yield), and the faster-running isomer produced only the reduction product **68a** (**Scheme 29**).

Based on the experimental results and the X-ray structure of aldehyde 55, we believe that the relative stereochemistry of the conjugated benzyl ester and methyl ester in compound 63 is as shown in Scheme 30, for the two isomers.





On this basis, if the conjugated benzyl ester in the faster-running isomer is in the axial conformation, then intramolecular hydrogen transfer occurs and leads to the reduced product **68a.** However, in the slower-running isomer, when this group is in an equatorial conformation, intramolecular hydrogen transfer is impossible as judged by inspection of Dreiding models, and radical cyclization occurs to form the desired product **68b** (Scheme 30). As compound **66** is a single substance, isomerization must have occurred in the coupling step **66** \rightarrow **63**, see **Scheme 28**).



Scheme 31

Debenzylation of **68b** under standard conditions (H_2/Pd) afforded the carboxylic acid **64**, but all efforts to make the olefin **30** by Kochi's method^{41,42} or to make **65** by Hunsdiecker reaction⁴³ were fruitless, and only complex mixtures were obtained (**Scheme 31**). This route was therefore also abandoned, and we embarked on an alternative which, in the event, led nicely to the desired racemic natural product. Part II - The Successful Route to (\pm) -A58365B.

Synthetic Plan

As indicated before, the structure of (\pm) -A58365B 2 does not appear to be especially complicated, but all our own extensive efforts to make it — those described above, as well as some others — had not been successful. We were eventually led to an approach in which the C(7)-C(8) unsaturation was introduced in the disguised form of a spirolactone, and ring B was formed by a process based on enyne radical cyclization (Scheme 32).



Scheme 32

Retrosynthetic analysis for the required intermediate

69, itself assembled from two pieces - lactone acid 71 and ε hydroxynorleucine methyl ester 72 - is shown in Scheme 33.



Scheme 33

Compound 71 represents all the carbons that make up the C(6)-C(9) segment of the natural product, including the propionic side chain. The ring A portion of (\pm) -A58365B was constructed from ϵ -hydroxynorleucine methyl ester⁴⁴ which had all the required atoms.

Synthesis of Lactone Carboxylic Acid 71

In our attempt to make the lactone carboxylic acid 71, we planned to synthesize tertiary alcohol 74, which, we hoped could be elaborated into 71 by lactonization and hydrolysis (Scheme 34).



Converting 73 into 74 proved to be a very difficult task. Four methods were tried for the synthesis of alcohol 74. The first one involved adding lithio-1-(triisopropylsilyl)propyne⁴⁵ to dimethyl α -ketoglutarate 73, itself prepared from methanol and α -ketoglutaric acid in the presence of a catalytic amount of acid. Surprisingly lactone 76 (Scheme 35) was obtained directly, but the reaction was not useful for us due to the low yield (10%). In our second attempt, the cerium salt of 3-(triisopropylsilyl)propyne was generated following a literature procedure.⁴⁶ We hoped the acetylide would add to the carbonyl group of **73** and generate **77**, but only a complex mixture was obtained from this reaction (**Scheme 35**).



scheme 35

Use of a Grignard reagent was considered next.⁴⁷ Unfortunately, the starting material simply dimerized to form compound **78** (**Scheme 36**).



scheme 37

To our delight, we then found that the three-carbon side chain could be introduced by addition of propargylaluminum sesquibromide⁴⁸ solution to **73** in dry THF at -78 °C. Compound **74** was obtained in good yield (95%). Lactonization

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did not pose any problem, but hydrolysis of methyl ester 75 was not successful. Many methods were tried to hydrolyze 75 into 71 [LiOH,⁴⁹ (Bu₃Sn)₂O,⁵⁰ Me₃SiCl/NaI⁵¹ and Ba(OH)₂⁵²], but all of them gave complex mixtures (**Scheme 37**).

However, we eventually found that ester hydrolysis, liberation of the free hydroxy diacid by ion-exchange chromatography using Amberlite IR-120 (acid form), followed by evaporation of the resulting aqueous solution at 65 °C, directly converted the hydroxy diester 74 into lactone acid 71 in good yield (95%) (Scheme 38).



In order to confirm that the reaction in **Scheme 38** gave us lactone acid **71** and not the di-acid, we prepared the benzylamide derivative. In this reaction, the standard coupling method was followed, and **79** was obtained in 85% yield (**Scheme 39**). Now we were confident that we had the required lactone acid **71** in hand.


Scheme 39

Coupling of Lactone Acid **71** and *E*-Hydroxynorleucine Methyl Ester **72**

Three strategies were tried for coupling the lactone acid **71** and ε -hydroxynorleucine methyl ester hydrochloride salt **72**, itself prepared by treating ε -hydroxynorleucine with excess Me₃SiCl in dry methanol.

In our first attempt, 72 was silylated in order to protect the free hydroxyl group, but the desired compound 80 was obtained in less than 38% yield along with the byproduct 81 (Scheme 40).



Scheme 40



Coupling of **71** and **80** under mild conditions (DCC, HOBT) gave a complex mixture, and, at this point, we decided to try more vigorous conditions. Reaction of **80** with lactone acid chloride **83** produced the coupled compound **82** as mixture of two isomers (88%), **82a** (major) and **82b** (minor) in a ratio of 7:3. Exposure of the single isomer **82** to TBAF/AcOH/THF gave the expected compound **84** (Scheme 41).

Two other methods were also explored in order to increase the yield of alcohol 84. Without protecting the free OH group, treatment of 72 with acid chloride 83 in the presence of pyridine or triethylamine gave 84 in 39% yield. However, coupling of **71** and **72** proceeded most efficiently under another set of standard conditions (*N*-methylmorpholine, HOBT, EDCI, DMF),⁵³ a mixture (6:4) of diastereoisomeric amides **84a** (major) and **84b** (minor) being obtained in 65% yield (based on ε -hydroxynorleucine).



Scheme 42

Synthesis of Intermediate 69

All that remained to reach the key intermediate 69 was to oxidize alcohol 84 and then cyclize the resulting aldehyde 70 (Scheme 43). Our initial plan called for Swern oxidation of 84, but only 20% of 70m and 30% of 70m were obtained from the individual isomers of 84, respectively. Fortunately, isomers 84m and 84m could be individually oxidized by PCC to the corresponding aldehydes, (84m \rightarrow 70m, 66%; 84m \rightarrow 70m, 87%), and these were cyclized⁴⁴ [70m \rightarrow 69m, 82%; 70m \rightarrow **69b**, 65%, after correction for recovered starting material (14%)] by exposure to freshly distilled TFA in the presence of 4Å molecular sieves, the reaction taking 4 h in the case of **70a**, and 1 h with **70b**.



Scheme 43

Synthesis of Olefin 30 from Radical Cyclization Products

Now, the stage was set for closure of ring B by stannane addition to the alkyne, followed by cyclization of the resulting vinyl radical onto the proximal terminus of the enamine double bond.⁵⁴⁻⁵⁷ Individual toluene solutions of AIBN and Ph₃SnH were added over 2 minutes to a refluxing solution of **69a** in the same solvent. The desired product **85a** (79%) was obtained from this experiment, while cyclization of **69b** under similar conditions, but using Bu₃SnH, gave **85b** (89%). Use of Bu₃SnH with **69a** was much less efficient in either refluxing toluene or benzene. It may be significant that **69a** exists as two rotamers at room temperature, while **69b**, which closes efficiently (Bu₃SnH, refluxing, toluene), does not. The ¹H NMR spectrum of **69a** shows broad signals at 85 °C, but coalesced signals at 100 °C.



Scheme 44

Thorough purification of **85a** and **85b** was difficult because of persistent tin species, but protodestannylation gave **30a** (TFA, 12 h at room temperature, 2 h at 60 °C; 62% 101

from **69a**) and **30b** (TFA, 5 h at room temperature; 89% from **69b**). Both cyclization-destannylation products **30a** and **30b** were single compounds although, of course, of different stereochemistry. However, both served equally well for the last steps of the synthesis (**Scheme 45**).

Completion of the Synthesis of (\pm) -A58365B (2)



Scheme 45

Ozonolysis ($30a \rightarrow 31a$; $30b \rightarrow 31b$) gave the corresponding ketones and, without isolation, exposure to triethylamine (THF, $60^{\circ}C$, 2 h) served both to introduce the

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critical C(7)-C(8) double bond and release the propionic acid side chain in 63% overall yield from either **30a** or **30b**. Finally, ester hydrolysis, using $(Bu_3Sn)_2O^{58}$ in refluxing benzene for 3 days, gave crystalline (±)-A58365B **2** in 67% (mp 209-213 °C, from water) (**Scheme 45**).

Conclusion

 (\pm) -A58365B 2, an inhibitor of angiotensin-converting enzyme, was synthesized⁵⁹ from two subunits, spiro lactone 71, which represents all the carbons that make up the C(6)-C(9) segment of the natural product, including the propionic side chain, and ε -hydroxynorleucine methyl ester 72, which not only has all of the required atoms of ring A, but also could be used without prior hydroxyl protection. Ring B was constructed by a process based on enyne radical cyclization (69ab \rightarrow 30ab). Our synthesis of (\pm)-A58365B 2 (7.8% over 10 steps) is summarized in Scheme 46 and Scheme 47.



Scheme 46

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Scheme 47

Synthesis of (-)-A58365B

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Our initial plan was based on the fact that racemic A58365B could be synthesized from (\pm) - ϵ -hydroxynorleucine. Consequently, optically pure material should be obtained

following an identical route, but using $(L) - \varepsilon$ -hydroxylnorleucine as the starting material.

In order to test this hypothesis we repeated the synthetic route developed for the racemic form, but using the optical pure amino acid (L)- ϵ -hydroxynorleucine, which was obtained by enzymatic resolution, according to a literature procedure⁶⁰ (Scheme 48).



Scheme 48

The synthesis was completed in a fashion identical to the one described earlier for the racemic material (see **Schemes 46** and **47**). The final compound had $[\alpha]_D{}^{21} = -114 \circ$ (c 0.16, H₂O); the literature¹¹ value is $[\alpha]_D{}^{25} = -141 \circ$ (c 0.16, H₂O). The difference between the observed and literature values might suggest that the stereogenic center had undergone a small amount of epimerization during the synthesis. However, this difference could also be attributed to the different experimental conditions under which the optical rotations were measured. In order to determine the exact ee% of the final product, several experiments were carried out, as follows.

Our first attempt was to make the amide derivative 86 from 2 by using chiral amine $(S) - \alpha$ -methylbenzylamine (Scheme 49). Unfortunately, compound 2 decomposed during attempts (use of DCC) to form the amide.



Scheme 49

Our next attempt was to prepare the trimethyl derivative of 2, by following the literature procedure¹³ (Scheme 50), and then we could apply an ¹H NMR chiral shift reagent $[Eu(hfc)_3]$ to determine the ee%. This experiment was informative and the ¹H spectra are shown below. In the racemic form of 2 (spectrum b), the three methyl groups were shifted into the six peaks, but for optically active material only three sharp signals, corresponding to these methoxy groups were observed (spectrum c). We can conclude that optically active natural product 2 was obtained in high ee. However, we are unable to exactly quantify the ee, and we plan to examine our material by HPLC, using a chiral column

.



without chiral shift reagent



(III) Experimental Section

General experimental procedures

Argon was purified by passage through a column $(3.5 \times 42 \text{ cm})$ of BASF R-311 catalyst⁶¹ and then through a similar column of Drierite. Glassware was dried in an oven for at least 3 h before use $(120 \ ^{\circ}\text{C})$ and either cooled in a desiccator over Drierite, or assembled quickly, sealed with rubber septa, and allowed to cool under a slight static pressure of argon. Reaction mixtures were stirred by Teflon-coated magnetic stirring bars.

Solvents for chromatography or extractions were distilled before use.

Products were isolated from solution by evaporation under water pump vacuum at, or below, 30 °C, using a rotary evaporator.

Melting points were determined on a Kofler block melting point apparatus.

Commercial thin layer chromatography (TLC) plates (silica gel, Merck 60F-254) were used. Spots were detected by examination under UV light or by spraying the plate with a solution of phosphomolybdic acid, followed by charring on a hot plate. Silica gel for flash chromatography was Merck type 60 (230-400 mesh).

C-18 Reverse phase silica gel (60 mesh, 10% C-18 capped with TMS, Lot 1-AB-76) (Catalogue # S4525) was obtained from Toronto Research Chemicals Inc., 4483 Chesswood Dr., Downsview, Ontario M3J 2C3 (telephone: 416-638-9696). Dry solvents were prepared under an inert atmosphere and transferred by oven-dried syringes. Dry THF was distilled from Na and benzophenone ketyl. Dry PhH was distilled from Na. Dry CH_2Cl_2 , MeOH, pyridine, DMF, and were distilled from CaH_2 , the last two solvents being distilled under water pump vacuum. Commercial (Aldrich) solutions of *n*-BuLi (in hexanes) were assumed to have the stated molarity.

The symbols s', d', t', and q' used for 13 C NMR spectra indicate 0, 1, 2, or 3 attached protons.

1-(t-Butoxycarbonyl)hexahydro-6-(methoxycarbonyl)-2pyridinecarboxylic acid (38).



BOC-anhydride (5.4 g, 24.71 mmol) was added to a suspension of amino acid **37** (3.9 g, 20.9 mmol) in dry THF (60 mL). The resulting mixture was refluxed for 48 h under Ar and then cooled to room temperature. Evaporation of the solvent and flash chromatography of the residue over silica gel (4 x 20 cm), using 1:9 MeOH-CHCl₃, gave pure (¹H NMR 400 MHz) **38** as a colorless oil (5 g, 83% yield): FTIR (CH₂Cl₂ cast) 1700, 1741, 1801 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 1.4-1.6 (br signal, 10 H), 1.6-1.7 (br, 1 H), 1.7-2.1 (br, 2 H), 2.1-

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2.4 (br, 2 H), 3.8-4 (br, 3 H), 4.6-4.9 (br, 2 H) (the carboxyl H was not observed; possibly it was an very broad signal); ¹³C NMR (CDCl₃, 50.3 MHz) δ 16.65 (t'), 25.8 (t'), 26.5 (t'), 27.9 (q'), 52.9 (d'), 53.65 (d'), 56.03 (q'), 82.5 (s'), 154.9 (s'), 174.5 (s'), 178.6 (s'); exact mass m/z calcd for C_{12H20}NO₄ (M - COOH) 242.1391, found 242.1389.

Methyl 1-(t-Butoxycarbonyl)hexahydro-6-(hydroxymethyl)-2-pyridinecarboxylate (39).



A mixture of acid **38** (2.9 g, 10.10 mmol) and dry THF (20 mL) was cooled in an ice-salt bath (-14 °C), and $BH_3 \cdot Me_2S$ (2 M in THF, 11 mL, 22.01 mmol) was added over a period of 15 min (evolution of H_2). Stirring was continued for 16 h, the ice-salt bath being allowed to reach room temperature. The reaction mixture was hydrolyzed by dropwise addition of water (10 mL) at 0 °C (stirring). No hydrogen evolution was observed, indicating complete utilization of the $BH_3 \cdot Me_2S$. The aqueous phase was treated with K_2CO_3 (6 g) and the THF phase was separated. The aqueous phase was extracted with Et_2O (3 x 50 mL). The combined ether extracts were washed with brine (3 x 20 mL) and dried (MgSO₄). Evaporation of solvent and flash chromatography of the residue over silica

gel (3 x 20 cm), using 3:7 EtOAc-hexane, gave pure (¹H NMR, 200 MHz) **39** (2.2 g, 80%): FTIR (CH₂Cl₂ cast) 1693, 1734, 1750, 3471 cm⁻¹; ¹H NMR (CDCl₃, 200 MHz) δ 1.4-1.6 (br, 11 H), 1.6-1.8 (m, 3 H), 2-2.2 (br, 1 H), 3.1 (t, J = 7 Hz, 1 H), 3.5 (t, J = 7 Hz, 2 H), 3.8 (s, 3 H), 4.3-4.4 (m, 1 H), 4.7-5 (br, 1 H); ¹³C NMR (CD₂Cl₂, 50.3 MHz) δ 17.1 (t'), 25.0 (t'), 26.4 (t'), 28.4 (q'), 51.9 (d'), 52.7 (d'), 53.6 (q'), 63.6 (t'), 80.5 (s'), 156.2 (s'), 175.5 (s'), exact mass *m/z* calcd for C₁₃H₂₃NO₅ 273.1572, found 273.1571.

Methyl 1-(t-Butoxycarbonyl)-6-formylhexahydro-2pyridinecarboxylate (40).



Freshly distilled DMSO (1.9 mL, 26.76 mmol) was added slowly (Ar atmosphere) to a cooled (-78 °C) and stirred solution of (COCl)₂ (1.2 mL, 13.76 mmol) in dry CH_2Cl_2 (15 mL). There was immediate gas evolution. Stirring was continued and, after 0.5 h, the alcohol **39** (1.76 g, 6.45 mmol) in dry CH_2Cl_2 (5 mL) was added dropwise. Stirring was continued for 2 h at -78 °C, and then Et_3N (3.5 mL, 25.11 mmol) was added dropwise. The cooling bath was removed, stirring was continued for 3 h, and the mixture was then quenched with water (30 mL), and extracted with $CHCl_3$ (3 x 20 mL). The combined organic extracts were washed with water (3 x 20 mL) and brine (2 x 20 mL), and dried (MgSO₄). Evaporation of solvent and flash chromatography of the residue over silica gel (3 x 20 cm), using 1:4 EtOAc-hexane, gave two isomers in ratio 15:1 [faster-running isomer (40a) on TLC, 1.32 g; slower-running isomer (40b) on TLC, 87 mg; total yield 81%].

The pure (¹H NMR, 200 MHz) faster-running isomer (**40a**) had: FTIR 1699, 1729, 1743, 2868 cm⁻¹; ¹H NMR (CD₂Cl₂, 200 MHz) δ 1.1-1.3 (br, 1 H), 1.3-1.6 (br, 11 H), 1.6-1.8 (m, 1 H), 1.9-2.1 (br, 1 H), 2.2-2.4 (br, 1 H), 3.7 (s, 3 H), 4.4-4.6 (br, 1 H), 4.7-4.9 (br, 1 H), 9.7-9.8 (br, 1 H); ¹³C NMR (CD₂Cl₂, 50.3 MHz) δ 17.4 (t'), 22.7 (t'), 26.8 (t'), 28.3 (q'), 52.4 (q'), 58.9 (d'), 58.9 (d'), 81.4 (s'), 155.7 (s'), 173.7 (s'), 201.96 (d'); exact mass *m/z* calcd for C₁₂H₂₀NO₄ (M - CHO) 242.13924, found 242.13254.

The slower-running isomer (**40b**) had: ¹H NMR (CD₂Cl₂, 200 MHz) δ 1.1-1.5 (br, 11 H), 1.6-1.8 (br, 3 H), 2.1-2.2 (br d, 1 H), 3.6 (s, 3 H), 3.8-4.0 (br, 1 H), 4.6-4.7 (br d, 1 H), 9.2-9.4 (m, 1 H); ¹³C NMR (CD₂Cl₂, 50.3 MHz) (mixture of rotamers) δ 18.6 (t'), 25.7 (t'), 26.8 (t'), 28.2 (q'), 52.38 (q'), 54.43 (d'), 55.97 (d'), 61.42 (d'), 61.7 (d'), 82.00 (s'), 157.7 (s'), 173.1 (s'), 196.9 (d'), 197.9 (d'); exact mass *m/z* calcd for C₁₂H₂₀NO₄ (M - CHO) 242.13924, found 242.13912. Methyl 1-(t-Butoxycarbonyl)-6-ethynylhexahydro-2-

pyridinecarboxylate (41).



A stirred slurry of t-BuOK (300 mg, 2.67 mmol) in dry THF (3 mL) was cooled to -78 °C under Ar. A solution of dimethyl (diazomethyl)phosphonate (383 mg, 2.54 mmol) in THF (1 mL) was added dropwise over 2 min and the mixture was stirred for 5 min. During this time, the color of the mixture changed from pale yellow to brown. A solution of aldehyde 40 (518 mg, 1.91 mmol) in THF (2 mL) was added over 1 min. N_2 was evolved immediately. The resulting solution was stirred at -78 °C for 12 h, the cooling bath was removed, and stirring was continued for an additional 4 h. The mixture was then quenched with water (50 mL) and extracted with Et_2O (3 x 20 mL). The combined organic extracts were washed with water $(3 \times 10 \text{ mL})$ and brine $(2 \times 3 \text{ mL})$, and dried (MgSO₄). Evaporation of solvent and flash chromatography of the residue over silica gel (2 x 20 cm), using 3:17 EtOAchexane, gave **41** as a pure (¹H NMR, 200 MHz), white powder (380 mg, 75%): FTIR (CH₂Cl₂ cast) 1701, 1738, 3265 cm⁻¹; ¹H NMR (CD₂Cl₂, 200 MHz) δ 1.4 (s, 10 H), 1.5-1.9 (m, 5 H), 2.2 (d, J = 2 Hz, 1 H), 3.7 (s, 3 H), 4.7-4.85 (br, 1 H), 4.95.05 (br, 1 H); ¹³C NMR (CD₂Cl₂, 50.3 MHz) (mixture of rotamers) δ 16.9 (t', two peaks), 25.3 (t'), 28.3 (q'), 31.0 (t'), 42.6 (d', two peaks), 51.5 (q', two peaks), 55.5 (d'), 71.4 (s'), 80.7 (d'), 81.3 (s', two peaks), 155.0 (s', two peaks), 172.2 (s', two peaks); exact mass *m/z* calcd for C₁₄H₂₁NO₄ 267.14706, found 267.14520.





TFA (2.5 mL, 32.45 mmol) was added to a cooled (0 °C, ice bath) and stirred solution of the acetylene **41** (207 mg, 0.77 mmol) in CH₂Cl₂ (3 mL). The cold bath was removed, and stirring was continued for 4 h. The solvent was evaporated, the residue was dissolved in EtOAc (20 mL), and the solution was washed with saturated aqueous NaHCO₃ (3 x 10 mL), water (2 x 10 mL) and brine (1 x 20 mL), and dried (MgSO₄). Evaporation of solvent and flash chromatography of the residue over silica gel (1.5 x 20 cm), using 3:7 EtOAchexane, gave the desired product **33** (120 mg, 92%) as a colorless oil: FTIR (CHCl₃, cast) 1696, 1739, 3290 cm⁻¹; ¹H NMR (CD₂Cl₂, 200 MHz) δ 1.4-1.6 (m, 3 H), 1.8-2.0 (m, 3 H), 2.25-2.34 (br d, J = 2 Hz, 2 H), 3.23-3.30 (m, 1 H), 3.453.52 (m, 1 H), 3.7 (s, 3 H); ¹³C NMR (CD_2Cl_2 , 50.3 MHz) δ 24.3 (t'), 28.7 (t'), 32.9 (t'), 48.0 (d'), 52.1 (d'), 58.9 (q'), 70.5 (d'), 173.2 (s') (one acetylenic signal is not observed under the conditions used to measure the spectrum); exact mass m/z calcd for $C_{9H_{13}NO_2}$ 167.09464, found 167.09383.

Methyl 1-[[2-(Bromomethyl)tetrahydro-5-oxo-2-furanyl]carbonyl]-6-ethynylhexahydro-2-pyridinecarboxylate (29).



SOCl₂ (1.8 mL, 24.66 mmol) was added to a suspension of Swiss acid 34 (562 mg, 2.52 mmol) in dry PhH (4 mL) and the mixture was refluxed with stirring for 2 h (Ar atmosphere), cooled and evaporated to dryness, with protection from moisture. The resulting freshly prepared acid chloride in CH_2Cl_2 (2 mL) was added to a stirred and cooled (0 °C) mixture of amine 33 (374 mg, 2.25 mmol), pyridine (0.7 mL, 8.65 mmol) and CH_2Cl_2 (4 mL) (Ar atmosphere). The cold bath was left in place and stirring was continued overnight. The solution was then evaporated. The residue was dissolved in EtOAc (30 mL), washed with 1N HCl solution (4 x 10 mL), water (3 x 10 mL) and brine (2 x 20 mL), dried (MgSO₄), and evaporated. Flash chromatography of the residue over silica gel (2 x 20 cm), using 7:3 hexane-ethyl acetate, gave **29** as a mixture of isomers (500 mg, 61%): exact mass m/z calcd for C₁₅H₁₈⁸¹BrNO₅ 373.03479, found 373.03591; calcd for C₁₅H₁₈⁷⁹BrNO₅ 371.03683, found 371.03749. Because the material is a mixture of isomers, we can not get good ¹³C and ¹H NMR spectra.

Methyl 1-(t-Butoxycarbonyl)-6-ethenylhexahydro-2pyridinecarboxylate (53).



For this experiment, t-BuOK was weighed in a glove bag filled with Ar. Ph₃PCH₃Br was dried under good oil pump vacuum at 60 °C (oil bath), and dioxane was dried by refluxing over Na.

t-BuOK (333 mg, 2.96 mmol) was suspended in dry dioxane (10 mL), and Ph₃PCH₃Br (1.05 g, 2.93 mmol) was added in small portions from a side-arm addition tube (Ar atmosphere). The reaction mixture was stirred for 1 h at 50 °C (oil bath), then cooled to room temperature. A solid was deposited. The liquid above the solid was added by cannula to a solution of

aldehyde 40 (600 mg, 2.22 mmol) in dry THF (10 mL), and the resulting solution was stirred for 24 h at room temperature (Ar atmosphere). The mixture was diluted with EtOAc (20 mL), washed with water $(3 \times 10 \text{ mL})$ and brine $(2 \times 20 \text{ mL})$, dried (MgSO₄) and evaporated. The residue was suspended in distilled hexane (20 mL) and stored overnight in a refrigerator. The resulting white solid (Ph₃PO) was filtered off and the filtrate was evaporated. Flash chromatography of the residue over silica gel (2 x 20 cm), using 3:17 EtOAchexane, gave 53 as a pure (¹H NMR, 300 MHz), colorless oil (497 mg, 84%): FTIR (CDCl₃, cast) 1695, 1737, 2881, 2949, 2974 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 1.4 (s, 9 H), 1.5-1.7 (m, 4 H), 1.8-1.9 (m, 1 H), 2.2-2.3 (m, 1 H), 3.7 (s, 3 H), 4.56-4.62 (br, 1 H), 4.78-4.84 (m, 1 H), 5.0-5.2 (m, 2 H), 6.0-6.1 (m, 1 H); ¹³C NMR (CD₂Cl₂, 50.3 MHz) δ 16.6 (t'), 26.2 (t'), 28.4 (t'), 28.5 (q'), 51.9 (d'), 52.7 (q'), 80.2 (s'), 114.6 (t'), 139.1 (d'), 155.7 (s'), 173.5 (s'); exact mass m/zcalcd for C14H23NO4 269.16272, found 269.16215.

Methyl 6-Ethenylhexahydro-2-pyridinecarboxylate (52).



TFA (2 mL, 25.96 mmol) was added to a stirred and cooled

(0 °C) solution of olefin 53 (1.3 g, 4.83 mmol) in dry CH_2Cl_2 (2 mL). The cold bath was left in place, and stirring was continued for 5 h. The solvent was then evaporated. The residue was dissolved in EtOAc (40 mL), washed with saturated aqueous NaHCO₃ (3 x 20 mL), water (2 x 20 mL) and brine (2 x 30 mL), dried (MgSO₄), and evaporated. Flash chromatography of the residue over silica gel (3 x 20 cm), using 3:7 EtOAchexane, gave pure **52** (¹H NMR, 300 MHz) (610 mg, 75%): ¹H NMR (CDCl₃, 300 MHz) δ 1.2-1.3 (m, 1 H), 1.4-1.6 (m, 2 H), 1.7-1.8 (m, 1 H), 1.9-2.1 (m, 3 H), 3.1-3.2 (m, 1 H), 3.4-3.44 (dd, J)= 13.5, 1.8 Hz, 1 H), 3.75-3.8 (m, 3 H), 5.0-5.05 (dd, J =13.5, 3.6 Hz, 1 H), 5.2-5.25 (dd, J = 18, 3.6 Hz, 1 H), 5.9-5.95 (m, 1 H); ¹³C NMR (CD₂Cl₂, 75.5 MHz) δ 24.7 (t'), 29.1 (t'), 32.3 (t'), 51.9 (d'), 59.3 (q'), 59.4 (d'), 113.9 (t'), 142.0 (d'), 173.8 (s'); exact mass m/z calcd for C₉H₁₅NO₂ 169.11028, found 169.11033. We did not get the FTIR.

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Methyl 1-[[2-(Bromomethyl)tetrahydro-5-oxo-2-furanyl]carbonyl]-6-ethenylhexahydro-2-pyridinecarboxylate (49).



SOCl₂ (2.3 mL, 25.13 mmol) was added to a suspension of Swiss acid **34** (740 mg, 3.32 mmol) in dry PhH (15 mL), and the mixture was stirred and refluxed for 2 h (Ar atmosphere). The solution was then cooled to room temperature and evaporated to dryness. Dry pyridine (0.54 mL, 6.61 mmol) and the freshly prepared acid chloride in PhH (5 mL) were added sequentially to a stirred and cooled (ice bath) solution of amine **52** (510 mg, 3.01 mmol) in PhH (10 mL) (Ar atmosphere). The ice bath was left in place, and stirring was continued overnight. The mixture was diluted with EtOAc (50 mL), washed with 1 N HCl (4 x 20 mL), water (3 x 20 mL) and brine (2 x 30 mL), and dried (MgSO₄). Evaporation of solvent and flash chromatography of the residue over silica gel (3 x 20 cm), using 7:3 hexane-EtOAc, gave **49** as two separable fractions (total yield 1.01 g, 85%). The faster-running fraction (TLC) (minor reaction product) was a mixture of rotamers and/or isomers (394 mg): FTIR (CHCl₃ cast) 1130, 1685, 1737, 1794, 2950 cm⁻¹; exact mass m/z calcd for $C_{15}H_{20}^{79}BrNO_5$ 373.05249, found 373.05225. Because the material is a mixture of rotamers and/or isomers, satisfactory NMR data could not be obtained.

The slower-running fraction (TLC) (major reaction product) was also a mixture of rotamers and/or isomers (620 mg): FTIR (CHCl₃ cast) 1631, 1739, 1794, 2921, 2953; exact mass m/z calcd for $C_{15}H_{20}^{81}BrNO_5$ 375.05045, found 375.05042; calcd for $C_{15}H_{20}^{79}BrNO_5$ 373.05249, found 373.05259. Because the material is a mixture of rotamers and/or isomers, satisfactory NMR dates could not be obtained.

Methyl 1-[[2-(Bromomethyl)tetrahydro-5-oxo-2-furanyl]carbonyl]-6-formylhexahydro-2-pyridine-carboxylate (55).



Ozonized oxygen was passed into a stirred and cooled (-78 °C) solution of spirolactone **49** (mixture of two isomers, 90 mg, 0.24 mmol) in CH_2Cl_2 (5 mL) contained in a three-necked flask equipped with a reflux condenser (not connected to a water supply) closed by a drying tube packed with Drierite, an inlet tube (for ozone), and a glass stopper. When the solution became pale blue, ozonization was stopped, and the excess of ozone was removed with a stream of oxygen. Ph₃P (130 mg, 0.49 mmol) was tipped into the solution, the cooling bath was removed, and stirring was continued for 2 h. Evaporation of solvent and flash chromatography of the residue over silica gel (1.5 x 20 cm), using 3:7 EtOAchexane, gave pure (¹H NMR, 300 MHz) **55** (68 mg, 75%) as a mixture of two isomers.

The chromatographically faster-running isomer (17 mg) crystallized from EtOAc and had: mp 149-151 °C; FTIR (CHCl₃, cast) 1637, 1731, 1796, 2867, 2954; ¹H NMR (CDCl₃, 300 MHz) δ 1.4-1.7 (m, 3 H), 1.8-1.9 (m, 1 H), 2.3-2.5 (m, 3 H), 2.6-2.7 (m, 2 H), 3.1-3.2 (m, 1 H), 3.6 (d, 1 H), 3.7 (s, 3 H), 3.9-4.0 (dd, 1 H), 5.1-5.2 (m, 1 H), 5.4-5.5 (m, 1 H), 9.8 (s, 1 H); ¹³C NMR (CDCl₃, 100.6 MHz) (mixture of rotamers or isomers) δ 16.9 (t'), 21.9 (t'), 26.5 (t'), 27.2 (t'), 27.4 (t'), 32.4 (t'), 33.4 (t'), 37.4 (t'), 49.5 (t'), 53.0 (d'), 56.1 (q'), 58.9 (d'), 87.9 (s'), 88.5 (s'), 170.5 (s'), 173.1 (s'), 173.9 (s'), 199.1 (d'); exact mass *m/z* calcd for C_{13H17}⁸¹BrNO₅ (M - CHO) 348.02698, found 348.02628; calcd for C_{13H17}⁷⁹BrNO₅ 346.02902, found 346.02894.

The chromatographically slower-running isomer (51 mg, mixture of rotamers) was a colorless oil and had: FTIR (CHCl₃, cast) 1642, 1732, 1796, 2869, 2954 cm⁻¹; ¹H NMR (CDCl₃, 360 MHz) δ 1.3-1.8 (m, 3 H), 2.15-2.26 (m, 2 H), 2.34-

2.44 (m, 2 H), 2.5-2.8 (m, 2 H), 3.0-3.15 (m, 1 H), 3.6-4 (m, 5 H), 4.9-5.2 (br d, 1 H), 5.3-5.6 (br d, 1 H), 9.6-9.8 (d, 1 H); ¹³C NMR (CDCl₃, 50.3 MHz) (mixture of rotamers) δ 16.7 (t'), 16.6 (t'), 21.8 (t'), 23.2 (t'), 25.9 (t'), 26.5 (t'), 27.3 (t'), 27.5 (t'), 27.9 (t'), 30.4 (t'), 31.3 (t'), 31.8 (t'), 32.7 (t'), 36.9 (t'), 39.196 (t'), 52.5 (q'), 52.9 (q'), 53.2 (d'), 55.5 (d'), 59.7 (d'), 60.7 (d'), 87.1 (s'), 87.5 (s'), 88.1 (s'), 169.9 (s'), 170.2 (s'), 170.4 (s'), 171.7 (s'), 172.4 (s'), 174.0 (s'), 174.5 (s'), 199.1 (d'), 199.7 (d'); exact mass *m/z* calcd for C₁₃H₁₇⁸¹BrNO₅ (M - CHO) 348.02698, found 348.02731; calcd for C₁₃H₁₇⁷⁹BrNO₅ 346.02902, found 346.02896.

The structure of the fast-running isomer was confirmed by X-ray analysis (see appendix for details of the X-ray determination).

Methyl 1-(t-Butoxycarbonyl)hexahydro-6-[2-(phenylsulfonyl)ethenyl]-2-pyridinecarboxylate (60a).



BuLi (2.5 M in hexane, 1.1 mL, 2.75 mmol) was added slowly (Ar atmosphere) to a stirred and cooled (-78 °C)

solution of $(EtO)_2POCH_2SO_2Ph$ (773 mg, 2.64 mmol) in dry THF (15 mL). This mixture was stirred for 1 h, and then aldehyde **40** (670 mg, 2.47 mmol) in dry THF (5 mL) was added dropwise. Stirring was continued for another 2.5 h, the cold bath was removed and, when the mixture had attained room temperature, saturated aqueous NH₄Cl (30 mL)was added. The aqueous solution was extracted with Et₂O (3 x 20 mL) and the combined organic extracts were washed with water (2 x 10 mL) and brine (2 x 20 mL), dried (MgSO₄), and evaporated. Flash chromatography of the residue over silica gel (3 x 20 cm), using 1:5 EtOAc-hexane, gave three pure compounds (¹H NMR, 200 MHz): **60a** (fastest-running on TLC) 38 mg; **60c** (slowest-running on TLC) 230 mg; **60b** (unidentified structure) 410 mg; total yield 65% [**60b** has the same mass (mass spectrum) as **60a**].

Isomer **60a** had: ¹H NMR (CD₂Cl₂, 200 MHz) δ 1.30-1.40 (br, s, 9 H), 1.4-1.6 (m, 2 H), 1.8-2.0 (m, 3 H), 2.1-2.2 (m, 1 H), 3.7 (s, 3 H), 4.7-4.75 (dd, J = 8, 2 Hz, 1 H), 5.7-5.9 (m, 1 H), 6.1-6.15 (dd, J = 12, 2 Hz, 1 H), 7.1-7.15 (dd, J =12, 8 Hz, 1 H), 7.5-7.6 (m, 3 H), 8.0-8.2 (m, 2 H); ¹³C NMR (CD₂Cl₂, 75.5 MHz) δ 17.17 (t'), 26.6 (t'), 28.4 (q'), 31.5 (t'), 48.7 (d'), 52.6 (q'), 54.5 (d'), 80.9 (s'), 127.6 (d'), 127.9 (d'), 129.5 (d'), 133.8 (d'), 141.6 (s'), 149.6 (d'), 155.7 (s'), 174.6 (s'); exact mass m/z calcd for C₁₆H₁₉NO₆S (M - C₄H₈) 352.09332, found 353.09382.

Isomer 60c had: ¹H NMR (CD₂Cl₂, 200 MHz) δ 1.1-1.2 (m, 2 H), 1.4 (s, 9 H), 1.6-1.9 (m, 3 H), 2.0-2.1 (m, 1 H), 3.7 (s, 3 H), 3.8-3.85 (dd, J = 8, 4 Hz, 2 H), 4.7-4.8 (m, 1 H), 5.5 (t', J = 8 Hz, 1 H), 7.5-7.7 (m, 3 H), 7.9-8.0 (m, 2 H); ¹³C NMR (CD₂Cl₂, 75.5 MHz) δ 20.8 (t'), 26.2 (t'), 26.2 (t'), 28.3 (q'), 52.3 (q'), 55.9 (t'), 57.3 (d'), 81.3 (s'), 110.6 (d'), 129.0 (d'), 129.4 (d'), 133.9 (d'), 139.2 (s'), 141.9 (s'), 153.9 (s'), 172.2 (s'); exact mass m/z calcd for $C_{15H_{19}NO_{4}S$ (M - C₄H₉CO₂) 309.10349, found 309.10299.

Methyl 1-(t-Butoxycarbonyl)hexahydro-6-[2-(phenylthio)ethenyl]-2-pyridinecarboxylate (61).



BuLi (2.5 M in hexane, 0.16 mL, 0.41 mmol) was added slowly to a stirred and cooled (-78 °C) solution of PhSCH₂PPh₃Cl (170 mg, 0.41 mmol) in dry THF (1 mL). Stirring was continued for 1 h (Ar atmosphere) at -78 °C, and then aldehyde **40** (106 mg, 0.39 mmol) in THF (1 mL) was added dropwise. The cold bath was removed, stirring was continued for 4 h, and the mixture was diluted with Et₂O (20 mL), washed with saturated aqueous NH₄Cl (3 x 10 mL), water (3 x 10 mL) and brine (2 x 20 mL), and dried (MgSO₄). Evaporation of solvent and flash chromatography of the residue over silica gel (1.5 x 20 cm), using 1:9 EtOAc-harane, gave three isomers: isomer 61a (fastest-running on in TLC) (11 mg); isomer 61b (23 mg); isomer 61c (slowest-running on TLC) (36 mg); total yield 48%.

Isomer **61a** had: ¹H NMR (CD₂Cl₂, 200 MHz) δ 1.4 (s, 9 H), 1.5-1.9 (m, 5 H), 2.23-2.33 (m, 1 H), 3.7 (s, 3 H), 4.78-4.82 (br s, 2 H), 6.0-6.2 (dd, J = 14, 8 Hz, 1 H), 6.3-6.5 (dd, J = 14, 0.8 Hz, 1 H), 7.2-7.4 (m, 5 H); ¹³C NMR (CD₂Cl₂, 75.5 MHz) δ 16.6 (t'), 26.2 (t'), 28.5 (q'), 28.5 (t'), 52.2 (q'), 52.4 (d'), 53.1 (d'), 80.4 (s'), 123.5 (d'), 126.7 (d'), 129.1 (d'), 129.4 (d'), 135.4 (d'), 136.6 (s'), 155.5 (s'), 173.4 (s'); exact mass m/z calcd for C₂₀H₂₇NO₄S 377.16608, found 377.16538.

Isomer **61b** had: ¹H NMR (CD₂Cl₂, 200 MHz) δ 1.4 (s, 9 H), 1.5-1.6 (m, 2 H), 2.7-2.9 (m, 3 H), 2.2-2.3 (m, 1 H), 3.7 (s, 3 H), 4.8 (dd, J = 6, 2 Hz, 1 H), 5.3-5.4 (m, 1 H), 6.2-6.4 (m, 2 H), 7.2-7.4 (m, 5 H); ¹³C NMR (CD₂Cl₂, 75.5 MHz) δ 17.1 (t'), 26.5 (t'), 28.5 (q'), 30.5 (t'), 50.7 (d'), 52.3 (d'), 53.1 (q'), 80.4 (s'), 122.5 (d'), 176.7 (d'), 129.3 (d'), 129.4 (d'), 134.4 (d'), 136.7 (s'), 155.7 (s'), 174.1 (s', two signals); exact mass *m/z* calcd for C₂₀H₂₇NO₄S 377.16608, found 377.16589.

Isomer 61c [a mixture of rotamers (¹H NMR)] had: ¹H NMR (CD₂Cl₂, 200 MHz) δ 1.3-1.5 (br s, 9 H), 1.5-1.8 (m, 4 H), 1.8-2.8 (m, 2 H), 3.7 (s, 3 H), 4.2-4.4 (m, 1 H), 4.7-4.9 (m, 1 H), 5.8-6.0 (m, 1 H), 6.2-6.3 (m, 1 H), 7.2-7.4 (m, 5 H); ¹³C NMR (CD₂Cl₂, 75.5 MHz) δ 16.3 (t'), 17.3 (t'), 28.4 (q'), 127

28.6 (t'), 52.1 (q'), 55.1 (d'), 55.5 (d'), 80.7 (s'), 122.2 (d'), 122.7 (d'), 126.6 (d'), 129.3 (d'), 129.4 (d'), 134.3 (d'), 136.6 (s'), 173.6 (s'); exact mass m/z calcd for $C_{20H_27}NO_4S$ 377.16608, found 377.17008.

Methyl 6-[2-(Benzyloxycarbonyl)ethenyl]-1-(t-butoxycarbonyl)hexahydro-2-pyridinecarboxylate (67).



Aqueous NaOH (2 N, 40 mL) was added to a solution of BnOOCCH₂PPh₃Br (986 mg, 2.00 mmol) and CHCl₃ (40 mL) in a separatory funnel, and the mixture was shaken gently for 5 min. The organic solution was dried (MgSO₄) and evaporated. The residue (BnOOCCH=PPh₃) was kept under oil pump vacuum for 5 min and then dissolved in dioxane (10 mL). Aldehyde **40** (444 mg, 1.638 mmol) in dioxane (4 mL) was added to the above solution of BnOOCCH=PPh₃, and the mixture was stirred and heated in an oil bath set 90 °C. Stirring was continued for 4 h and the resulting solution was then cooled to room temperature. Evaporation of the solvent gave a white residue which was suspended in distilled hexane (20 mL) and kept overnight in a refrigerator. The resulting solid (Ph₃PO) was filtered off and the filtrate was evaporated. Flash chromatography of the residue over silica gel (3 x 20 cm), using 1:9 EtOAc-hexane, gave pure 67 (¹H NMR, 400 MHz) (620 mg, 94%): FTIR (CH₂Cl₂, cast) 1653, 1696, 1722, 2950, 2973 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 1.4 (s, 9 H), 1.45-1.53 (m, 2 H), 1.67-1.72 (m, 2 H), 1.84 (d, J = 12 Hz, 1 H), 2.24 (d, J= 12 Hz, 1 H), 3.6 (s, 3 H), 4.78-4.85 (br s, 2 H), 5.2 (s, 2 H), 5.89 (d, J = 12 Hz, 1 H), 7.01-7.15 (dd, J = 12, 6 Hz, 1 H), 7.38-7.43 (m, 5 H); ¹³C NMR (CDCl₃, 75.5 MHz) δ 16.2 (t'), 25.7 (t'), 27.6 (t'), 28.2 (q'), 51.2 (d'), 51.9 (q'), 52.8 (d'), 65.9 (t'), 80.7 (s'), 120.9 (d'), 128.2 (d'), 128.5 (d'), 136.1 (s'), 148.4 (d'), 155.2 (s'), 166.2 (s'), 172.8 (s'); exact mass m/z calcd for C₁₈H₂₁NO₆ (M - C₄H₈) 347.13690, found 347.13648.

Methyl 6-[2-(Benzyloxycarbonyl)ethenyl]hexahydro-2pyridinecarboxylate (66).



TFA (1 mL) was added to a stirred and cooled (0 °C) solution of 67 (134 mg, 0.33 mmol) in dry CH_2Cl_2 (1 mL). The ice bath was removed, stirring was continued for 5 h, and the

solvent was then evaporated. The residue was dissolved in EtOAc (20 mL) and the organic solution was washed with saturated aqueous NaHCO₃ (3 x 10 mL), water (2 x 20 mL) and brine $(2 \times 20 \text{ mL})$, and dried $(MgSO_4)$. Evaporation of solvent and flash chromatography of the residue over silica gel (1.5 x 20 cm), using 3:7 EtOAc-hexane, gave pure (¹H NMR, 400 MHz) 66 (90 mg, 93%) as a colorless oil: FTIR (CH_2Cl_2 , cast) 1656, 1621, 1737, 2855, 2948, 3350 cm⁻¹; ¹H NMR (CDC1₃, 400) MHz) δ 1.2-1.3 (m, 1 H), 1.4-1.6 (m, 2 H), 1.7 (d, J = 12 Hz, 1 H), 1.95-2.0 (m, 2 H), 2.0 (d, J = 12 Hz, 1 H), 3.3-3.4 (m, 1 H), 3.4 (dd, J = 12, 3.6 Hz, 1 H), 3.7 (s, 3 H), 5.2 (s, 2 H), 6.1 (d, J = 14 Hz, 1 H), 7.0 (dd, J = 14, 6 Hz, 1 H), 7.4-7.7 (m, 5 H); ¹³C NMR (CDCl₃, 75.5 MHz) δ 24.1 (t'), 28.4 (t'), 31.2 (t'), 51.9 (d'), 51.1 (d'), 58.6 (q'), 66.1 (t'), 120.0 (d'), 128.0 (d'), 128.1 (d'), 128.5 (d'), 135.9 (s'), 150.2 (d'), 166.1 (s'), 173.0 (s'); exact mass m/z calcd for $C_{17}H_{20}NO_4$ (M - 1) 302.13922, found 302.13909; calcd for C₁₅O₁₈NO₂ (M - COOMe) 244.13376, found 244.13385.

Methyl 6-[2-(Benzyloxycarbonyl)ethenyl]hexahydro-1-[[2-(bromomethyl)tetrahydro-5-oxo-2-furanyl]carbonyl]-2-pyridinecarboxylate (63).



SOCl₂ (1 mL, 13.71 mmol) was added to a stirred suspension of acid **34** (423 mg, 1.89 mmol) in dry PhH (10 mL). The mixture was refluxed and stirred for 2 h (Ar atmosphere), and then cooled to room temperature and evaporated to dryness. The freshly prepared acid chloride in PhH (5 mL) was added at room temperature (Ar atmosphere) to a solution of amine **66** (330 mg, 1.089), pyridine (0.4 mL, 4.91 mmol), and DMAP (10 mg, 0.08 mmol) in PhH (10 mL). The mixture was stirred overnight at room temperature, diluted with EtOAc (50 mL), and then washed with 1 N HCl (4 x 20 mL), water (3 x 20 mL) and brine (2 x 30 mL), and dried (MgSO₄). Evaporation of solvent and flash chromatography of the residue over silica gel (2 x 20 cm), using 7:3 hexane-EtOAc, gave the product as two isomers: isomer **63a** (faster-running on TLC) (240 mg) and isomer 63b (slower-running on TLC) (220 mg) (total yield 83%).

Isomer 63a (faster-running on TLC) was a pure (¹H NMR, 400 MHz) single compound: FTIR (CH₂Cl₂, cast) 1636, 1720, 1737, 2872, 2951, 3007, 3032 cm^-1; ¹H NMR (CDCl₃, 400 MHz) δ 1.56-1.61 (m, 2 H), 1.78-1.81 (m, 2 H), 2.05 (d, J = 12 Hz, 1 H), 2.34-2.4 (m, 1 H), 2.45 (d, J = 12 Hz, 1 H), 2.55-2.6 (m, 2 H), 3.0-3.1 (m, 1 H), 3.6 (s, 3 H), 3.7-4.1 (m, 2 H), 5.2 (s, 2 H), 5.34-5.42 (br, 1 H), 5.45-5.53 (m, 1 H), 6.0 (d, J = 12 Hz, 1 H), 7.1-7.2 (dd, J = 12, 6 Hz, 1 H), 7.34-7.42 (m, 5 H); ¹³C NMR (CDCl₃, 75.5 MHz) δ 16.0 (t'), 26.1 (t'), 27.2 (t'), 27.6 (t'), 33.1 (t'), 38.2 (t'), 50.9 (d'), 52.7 (d'), 54.7 (q'), 66.3 (t'), 88.2 (s'), 121.9 (d'), 128.3 (d'),128.6 (d'), 135.9 (s'), 146.8 (d'), 165.9 (s'), 169.6 (s'), 172.1 (s'), 174.2 (s'); exact mass m/z calcd for C₂₃H₂₆⁸¹BrNO₇ 509.08722, found 509.08761; calcd for $C_{23}H_{26}^{79}BrNO_7$ 507.08926, found 507.08986; $C_{21}H_{23}^{81}BrNO_5$ (M - COOMe) 450.07391, found 450.07476; calcd for $C_{21}H_{23}^{79}BrNO_5$ 448.07596, found 448.07673.

Isomer **63b** (slower-running on TLC) was a mixture of rotamers and had: FTIR (CH₂Cl₂ cast) 1640, 1684, 1721, 1738, 1796, 2951, 3008 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 1.6-1.7 (m, 1 H), 1.7-1.9 (m, 3 H), 2.0-2.1 (m, 1 H), 2.3-2.8 (m, 4 H), 3.0-3.1 (m, 1 H), 3.50 and 3.60 (two s, 3 H), 3.7-4.1 (m, 2 H), 5.15-5.22 (br, 3 H), 5.34-5.44 (br 1 H), 6.0-6.2 (dd, J =18, 6 Hz, 1 H), 6.9-7.1 (m, 1 H), 7.36-7.44 (m, 5 H); ¹³C NMR (CDCl₃, 75.5 MHz) δ 15.8 (t'), 15.9 (t'), 24.5 (t'), 25.5 (t'), 26.6 (t'), 27.7 (t'), 27.9 (t'), 28.0 (t'), 31.7 (t'),
32.9 (t'), 37.3 (t'), 51.7 (d'), 52.36 (q'), 52.6 (q'), 53.5 (d'), 54.6 (d'), 66.4 (t'), 87.5 (s'), 88.1 (s'), 122.6 (d'), 128.2 (d'), 128.5 (d'), 135.9 (s'), 145.8 (d'), 147.5 (d'), 165.6 (s'), 169.7 (s'), 171.3 (s'), 171.5 (s'), 174.1 (s'); exact mass m/z calcd for $C_{23}H_{26}^{81}BrNO_7$ 509.08722, found 509.08811; calcd for $C_{23}H_{26}^{79}BrNO_7$ 507.08926, found 507.08936; calcd for $C_{21}H_{23}^{81}BrNO_5$ (M - COOMe) 450.07391, found 450.07459; calcd for $C_{21}H_{23}^{79}BrNO_5$ (M - COOMe) 448.07596, found 448.07637.

Methyl 1'-[(Benzyloxycarbonyl)methyl]octahydro-4',5dioxospiro[furan-2(3H), 3'(4'H)-[2H]quinolizine]-6'-



Ph₃SnH (225 mg, 0.67 mmol) in PhH (10 mL) and AIBN (6 mg, 0.04 mmol) in PhH (10 mL) were added simultaneously over 5 h, by syringe pump, to a stirred and refluxing solution of **63b** (220 mg, 0.43 mmol) in PhH (5 mL). After the addition, refluxing was continued for another 10 h and the mixture was then cooled and evaporated. Flash chromatography (done twice) over silica gel (2 x 20 cm), using 1:1 EtOAc-hexane, gave **68b** (139 mg, 75%) as a mixture of isomers: exact mass

m/z calcd for $C_{23}H_{27}NO_7$ 429.17874, found 429.17786; calcd for $C_{21}H_{24}NO_5$ (M - CCOMe) 370.16544, found 370.16515; calcd for $C_{14}H_{18}NO_5$ (M - CH₂COOBn) 280.11850, found 280.11790. We can not get ¹H and ¹³C due to the mixture of isomers.

Methyl 1'-[Carboxymethyl]octahydro-4', 5-dioxospiro-[furan-2(3H), 3'(4'H)-[2H]quinolizine]-6'-carboxylate (64).



Benzyl ester **68b** (34 mg, 0.08 mmol) was hydrogenated (60 psi) overnight in EtOAc over Pd/C (10 mg). The catalyst was filtered off and the solvent was evaporated to leave **64** as a white foam (22 mg, 85%): FTIR (CDCl₃, cast) 1699, 1717, 1735, 1782, 3500, 2990 (br) cm⁻¹; ¹H NMR (CDCl₃, 200 MHz) 1.5-1.8 (m, 7 H), 1.9-2.1 (m, 3 H), 2.2-3.9 (m, 6 H), 3.1 (d, 6 Hz, 1 H), 3.7 (s, 3 H); exact mass m/z calcd for C₁₆H₂₁NO₇ 339.13181, found 339.13136; calcd for C₁₄H₁₈NO₅ (M - COOMe) 280.11850, found 280.11841. An informative ¹³C NMR spectrum could not be obtained because of the presence of several isomers.

Dimethyl α -Ketoglutarate (73).



A round bottomed flask equipped with a Dean-Stark apparatus was charged with α -ketoglutaric acid (5 g, 34 mmol), TsOH.H2O (580 mg, 3.31 mmol), dry MeOH (5 mL, distilled from Mg) and dry PhMe (10 mL). The solution was refluxed for 5 h, cooled and poured into saturated aqueous $NaHCO_3$ (250 mL). The aqueous solution was extracted with EtOAc $(3 \times 50 \text{ mL})$, and the combined organic extracts were washed with saturated aqueous NaHCO₃ (3 x 50 mL), water (3 x 50 mL) and brine $(2 \times 50 \text{ mL})$, and dried $(MgSO_4)$. Evaporation of solvent gave pure (400 ¹H NMR) 73 as colorless oil (3.5 g, 59%): FTIR (CH₂Cl₂ cast) 1735, 2850, 2957 cm⁻¹; ¹H NMR $(CDCl_3, 400 \text{ MHz}) \delta 2.65 (t, J = 6 \text{ Hz}, 2 \text{ H}), 3.15 (t, J = 6 \text{ Hz}, 2 \text{ H})$ 2 H), 3.65 (s, 3 H), 3.85 (s, 3 H); ${}^{13}C$ NMR (CDCl₃, 75.5 MHz) δ 27.3 (t'), 34.1 (t'), 51.8 (q'), 52.9 (q'), 160.8 (s'), 172.3 (s'), 192.2 (s'); exact mass m/z calcd for $C_7H_{1,0}O_5$ 174.05283, found 174.05249.

Methyl Tetrahydro-5-oxo-2-[(3-triisopropylsilyl)-2propynyl)]furancarboxylate (76).



BuLi (1.6 M in hexane, 0.4 mL, 0.64 mmol) was added to a stirred and cooled (-20 °C) solution of 1-(triisopropylsilyl)propyne (82 mg, 0.42 mmol) and THF (1 mL). The mixture was stirred 20 min at -20 °C and then cooled to -78 °C (Ar atmosphere). Dimethyl α -ketoglutarate (106 mg, 0.611 mmol) in THF (1 mL) was added dropwise, and stirring was continued for 6 h at -78 °C (Ar atmosphere). The cooling bath was removed, the solution was quenched with saturated aqueous NH₄Cl (10 mL) and extracted with Et_2O (3 x 10 mL). The combined organic extracts were washed with water $(3 \times 5 \text{ mL})$ and brine $(2 \times 10 \text{ mL})$, and dried (MgSO₄). Evaporation of solvent and flash chromatography of the residue over silica gel (1 x 15 cm), using 2:8 EtOAc-hexane, gave pure (¹H NMR 300 MHz) **76** as a colorless oil (15 mg, 10%): ¹H NMR (CDCl₃, 300 MHz) δ 0.9-1.1 (br. 21 H), 2.45-2.50 (m, 2 H), 2.56-2.60 $(m, 2 H), 2.95 (d, J = 2.7 Hz, 2 H), 3.85 (s, 3 H); {}^{13}C NMR$ (CDCl₃, 75.5 MHz) δ 11.2 (d'), 18.6 (q'), 28.3 (t'), 28.7 (t'), 29.6 (t'), 53.2 (q'), 84.338 (s'), 85.4 (s'), 100.6 (s'), 170.8 (s'), 175.3 (s'); exact mass m/z calcd for

C₁₅H₂₃O₄Si (M - CHMe₂) 295.13657, found 295.13658.

Dimethyl 2-Hydroxy-2-[2-(propynyl)]pentanedioate (74).



A mixture of Al powder (84 mg, 3.11 mmol) and HgCl₂ (5 mg, 0.02 mmol) in dry THF (3 mL) was stirred vigorously for 1 h (Ar atmosphere) in a three-necked flask fitted with a reflux condenser, and closed by septa. Most of the solvent was then withdrawn by syringe from the resulting shiny Al, and fresh THF (3 mL) was injected. The mixture was warmed in an oil bath set at 40 °C, and propargyl bromide (363 mg, 3.05 mmol) in THF (1 mL) was then added slowly with vigorous stirring and at such a rate that the THF did not boil. The addition took ca. 10 min. After the addition, stirring at 40 °C was continued until a dark gray solution was obtained (ca. 1 h). The solution of propargyl aluminum sesquibromide was cooled to room temperature, and added by cannula at a fast dropwise rate to a stirred and cooled (-78 °C) solution of dimethyl 2-ketoglutarate (158 mg, 0.90 mmol) in THF (5 mL). Stirring at -78 °C was continued for 4 h, and the mixture was then poured into ice-water (100 mL), and extracted with Et_2O $(4 \times 30 \text{ mL})$. The combined organic extracts were washed with brine (50 mL), dried (MgSO₄), and evaporated. Flash

chromatography of the residue over silica gel (2 x 16 cm), using 3:7 EtOAc-hexane, gave pure (¹H NMR, 300 MHz) **74** (185 mg, 95%) as a colorless oil: FTIR (KBr) 3502, 3286, 1739 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 2.05-2.10 (m, 3 H), 2.15-2.26 (m, 1 H), 2.4-2.5 (m, 1 H), 2.5-2.63 (ABX, J_{AB} = 16.9, J_{AX} = 2.7, J_{BX} = 2.7 Hz, 2 H), 3.52 (s, 1 H), 3.65 (s, 3 H), 3.78 (s, 3 H); ¹³C NMR (CDCl₃, 75.5 MHz) 28.6 (t'), 30.2 (t'), 32.8 (t'), 51.8 (q'), 53.2 (q'), 71.6 (s'), 75.8 (d'), 78.4 (s'), 173.3 (s'), 174.8 (s'); exact mass m/z calcd for C₉H₁₁O₄ (M -OCH₃) 183.06573 found 183.06573.

Methyl Tetrahydro-5-oxo-2-[2-(propynyl)]furancarboxylate (75).



A three-necked 100-mL round bottomed flask equipped with a Dean-Stark apparatus and condenser was charged with 74 (1.610 g, 7.51 mmol), TsOH.H₂O (181 mg, 0.95 mmol) and dry PhMe (70 mL). The solution was refluxed for 3 h, cooled to room temperature, and poured into saturated aqueous NaHCO₃ (100 mL). The mixture was extracted with EtOAc (3 x 50 mL) and the combined organic extracts were washed with saturated aqueous NaHCO₃ (3 x 10 mL), water (3 x 20 mL) and brine (2 x 30 mL), and dried (MgSO₄). Evaporation of solvent and flash chromatography of the residue over silica gel (2.5 x 20 cm), using 3:7 EtOAc-hexane, gave pure (¹H NMR, 300 MHz) **75** as colorless oil (900 mg, 66%): FTIR (CHCl₃, cast) 1743, 1787, 3283 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 2.1 (t, J = 2.3 Hz, 1 H), 2.5-2.6 (m, 2 H), 2.7-2.8 (m, 2 H), 2.9 (narrow multiplet of ABX spectrum, 2 H), 3.8 (s, 3 H); ¹³C NMR (CDCl₃, 75.5 MHz) δ 27.3 (t'), 28.2 (t'), 29.7 (t'), 53.3 (q'), 72.3 (d'), 84.0 (s'), 170.7 (s'), 175.3 (s'); exact mass *m/z* calcd for C_{7H7O2} (M - COOMe) 123.04460, found 123.04468.

Tetrahydro-5-oxo-2-[2-(propyny1)]-2-furancarboxylic acid (71).



Note that in the following experiment we do not know the order in which hydrolysis and lactonization occur.

LiOH (509 mg, 10.37 mmol) in water (2 mL) was added to a solution of diester 74 (1.00 g, 4.67 mmol) in THF (20 mL), and the mixture was stirred overnight at room temperature. Evaporation of the solvent gave what we assumed to be the dilithium salt, as a solid.

A column packed with Amberlite IR-120 ion-exchange resin

(20-50Å mesh, 2.5 x 16 cm) was washed with water until the eluent was colorless. The column was then washed successively with 2 N aqueous NaOH (4 bed volumes), water (until the eluent was neutral to pH paper), 2 N HCl (4 bed volumes), and finally with water (until the eluent was neutral to pH paper). The dilithium salt was dissolved in water (ca. 2-3 mL) and the solution was passed down the column, using water. The eluent was monitored with pH paper or by TLC (silica, 1:9 MeOH-CHCl₃). Evaporation of the combined acidic fractions (water pump, rotary evaporator, bath temperature 65-70 °C) gave lactone acid 71 (746 mg, 95%) as a pure (¹H NMR, 300 MHz), light yellow powder: FTIR (KBr) 1720 1770, 2500-3500 (br) cm⁻¹; ¹H NMR (D₂O, 300 MHz) δ 2.45-2.70 (m, 3 H), 2.75-2.85 (m, 2 H), 2.9-3.1 (ABX, $J_{AB} = 16.9$, $J_{\text{AX}} = 2.9$, $J_{\text{BX}} = 2.9$ Hz, 2 H); ¹³C NMR (D₂O, 75.5 MHz) δ 27.8 (t'), 29.0 (t'), 30.7 (t'), 73.3 (d'), 79.1 (s'), 86.9 (s'), 174.8 (s'), 180.5 (s'); exact mass m/z calcd for C₅H₅O₄ (M -CH₂C≡CH) 129.01878, found 129.01892; mass (FAB) m/z calcd for C₈H₈O₄ 168.15056, found (FAB) 168.9.

N-Benzyl Tetrahydro-5-oxo-2-[2-(propynyl)]furancarboxamide (79).



Lactone acid 71 (84 mg, 0.51 mmol) was dissolved in a mixture of dry DMF (1 mL) and dry CH₂Cl₂ (1 mL). HOBT (64 mg, 0.47 mmol) and benzylamine (49 mg, 0.46 mmol) were added. followed by DCC (97 mg, 0.47 mmol). The mixture was stirred overnight (Ar atmosphere) at room temperature, and the resulting solid was filtered off. The filtrate was diluted with EtOAc (20 mL) and washed with water (5 x 10 mL) and brine (2 x 20 mL), and dried (MgSO₄). Evaporation of the solvent and flash chromatography of the residue over silica gel (1.5 x 20 cm), using 3:7 EtOAc-hexane, gave pure (¹H NMR, 300 MHz) 79 as a yellow oil (112 mg, 93%): FTIR (CHCl₃, cast) 1605, 1672, 1787, 3290, 3350 cm⁻¹; ¹H NMR (CDCl₃, 200 MHz) δ 2.1-2.15 (m, 1 H), 2.45-2.5 (m, 2 H), 2.67-2.7 (m, 2 H), 2.9-2.95 (m, 2 H), 4.45-4.5 (m, 2 H), 7.00-7.1 (br s, 1 H), 7.3-7.33 (m, 5 H); ¹³C NMR (CDCl₃, 75.5 MHz) δ 28.3 (t'), 28.6 (t'), 29.8 (t'), 43.5 (t'), 72.3 (d'), 85.6 (s'), 127.7 (d'), 127.8 (d'), 128.7 (d'), 137.3 (s'), 170.5 (s'), 175.3 (s'); exact mass m/z calcd for C₁₅H₁₅NO₃ 257.10519, found 257.10513.





6-Hydroxynorleucine (1.06 g, 7.21 mmol) and then Me₃SiCl

(4.6 mL, 36.24 mmol) were added to dry MeOH (20 mL, freshly distilled from Mg), and the solution was stirred for 42 h at room temperature (protection from moisture by a tube filled with Drierite). The solvent was then evaporated. PhMe (20 mL) was added and evaporation was repeated. PhMe (20 mL) was added twice more and, after each addition, the solvent was evaporated. Finally, the residue was kept under good oil pump vacuum for 3 days to remove the last traces of solvent, and to afford the pure (¹H NMR, 300 MHz) **72** (1.42, 100%) as a colorless oil: ¹H NMR (D₂O, 300 MHz) δ 1.5-1.6 (m, 2 H), 1.6-1.7 (m, 2 H), 2.0-2.1 (m, 2 H), 3.6 (t, J = 6.8 Hz, 2 H), 3.9 (s, 3 H), 4.2 (t, J = 6.8 Hz, 2 H); ¹³C NMR (D₂O, 100.6 MHz) δ 21.6 (t'), 30.3 (t'), 31.5 (t'), 53.6 (q'), 54.3 (d'), 61.9 (t'), 171.6 (s').

6-[(t-Butyldimethylsilyl)oxy]norleucine Methyl Ester (80) and N-Formy1-6-[(t-butyldimethylsilyl)oxy]norleucine Methyl Ester (81).



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TBDMSCl (241 mg, 1.59 mmol) and imidazole (216 mg, 3.14 mmol) were added to a solution of 72 (310 mg, 1.57 mmol) in dry DMF (1 mL). Stirring was continued overnight (Ar atmosphere) and the mixture was diluted with Et_2O (30 mL), washed with water (5 x 15 mL) and brine (3 x 10 mL), and dried (MgSO₄). Evaporation of solvent and flash chromatography of the residue over silica gel (2.5 x 20 cm), using 1:1 EtOH:hexane, gave two compounds **80** (slower-running on TLC) (159 mg, 37%) and **81** (faster-running on TLC) (250 mg, 52%).

Compound **80** was a pure (¹H NMR, 300 MHz), colorless oil (¹H NMR, 300 MHz) and had: FTIR (CHCl₃, cast) 1698, 1702, 3384 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 0.05 (s, 6 H), 0.9 (s, 9 H), 1.4-1.5 (m, 2 H), 1.5-1.6 (m, 5 H), 1.8-1.7 (m, 1 H), 3.4 (dd, J = 8.5, 7 Hz, 1 H), 3.6 (t, J = 6.8 Hz, 2 H), 3.7 (s, 3 H); ¹³C NMR (CDCl₃, 75.5 MHz) δ -5.4 (q'), 18.2 (s'), 21.8 (t'), 25.8 (q'), 32.4 (t'), 34.7 (t'), 51.7 (d'), 54.3 (q'), 62.7 (t'), 176.4 (s'); exact mass *m/z* calcd for C_{13H29}NO₃Si 275.19168, found 275.19198.

Compound **81** was a pure, colorless oil (¹H 400 MHz) and had: FTIR (CHCl₃ cast) 1665, 1747, 3295 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 0.5 (s, 6 H), 0.9 (s, 9 H), 1.4-1.45 (m, 2 H), 1.5-1.53 (m, 2 H), 1.7-1.73 (m, 1 H), 1.9-1.93 (m, 1 H), 3.6-3.63 (m, 2 H), 3.7 (s, 3 H), 4.7-4.73 (m, 1 H), 6.2-6.23 (br, 1 H), 8.2 (s, 1 H); ¹³C (CDCl₃, 75.5 MHz) δ -5.42 (q'), 18.2 (s'), 21.5 (t'), 25.8 (q'), 32.03 (t'), 32.05 (t'), 50.7 (d'), 52.3 (q'), 62.5 (t'), 160.8 (d'), 172.5 (s'); exact mass m/z calcd for $C_{13}H_{26}NO_4Si$ (M - Me) 288.16312, found 288.16284; calcd for $C_{13}H_{26}NO_3Si$ (M - OCH₃) 272.16818, found 272.16788.

6-[(t-Butyldimethylsilyl)oxy]-N-[[tetrahydro-5-oxo-2-(2-propynyl)-2-furanyl]carbonyl]norleucine Methyl Ester (82).



SOCl₂ (0.5 mL, excess) was added to a stirred mixture of **71** (82 mg, 0.48 mmol) in dry PhH (5 mL) (protection from moisture by a drying tube filled with Drierite). The mixture was refluxed and Ar was bubbled through the solution to expel HCl. After 1 h at reflux, the solvent was evaporated (protection from moisture). The resulting freshly prepared acid chloride in CH_2Cl_2 (2 mL) was added to a solution of amine **80** (138 mg, 0.51 mmol) and pyridine (0.2 mL, excess) in dry CH_2Cl_2 (3 mL). The resulting mixture was stirred at room temperature under Ar for 4 h and then evaporated. Flash chromatography of the residue over silica gel (2 x 20 cm), using 3:7 EtoAc-hexane, gave two pure isomers **82a** (fasterrunning on TLC) (131 mg) and **82b** (slower-running on TLC) (30 mg) (total yield 76%).

The pure (¹H NMR, 300 MHz) isomer **82a** had: FTIR (CHCl₃, cast) 1678, 1756, 1791, 2953, 3311, 3349 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 0 (s, 6 H), 0.8 (s, 9 H), 1.3-1.6 (m, 4 H), 1.7-1.8 (m, 1 H), 1.9-2.0 (m, 1 H), 2.1 (t, J = 2.9 Hz, 1 H), 2.5-2.8 (m, 4 H), 2.9 ABX, J = 2.9 1.26 Hz, 2 H), 3.6 (t, J =5.9 Hz, 2 H), 3.7 (s, 3 H), 4.5-4.6 (m, 1 H), 6.9 (d, J = 8.7Hz, 1 H); ¹³C NMR (CDCl₃, 75.5 MHz) δ -5.4 (q'), 18.3 (s'), 22. (t'), 25.9 (q'), 28.4 (t'), 28.5 (t'), 29.5 (t'), 31.9 (t'), 32.1 (t'), 52.3 (d'), 52.4 (q'), 62.6 (t'), 72.4 (d'), 77.2 (s'), 85.3 (s'), 170.8 (s'), 171.8 (s'), 174.9 (s'); exact mass m/z calcd for C₂₁H₃₅NO₆Si 425.22336, found 425.22400.

The pure (¹H NMR, 360 MHz) isomer **82b** had: FTIR (CHCl₃, cast) 1678, 1745, 1792, 3312, 3350 cm⁻¹; ¹H NMR (CDCl₃, 360 MHz) δ 0.0 (s, 6 H), 0.9 (s, 9 H), 1.4-1.5 (m, 2 H), 1.5-1.6 (m, 2 H), 1.7-1.8 (m, 1 H), 1.9-2.0 (m, 1 H), 2.1 (t, J = 3.2 Hz, 1 H), 2.4-2.5 (m, 1 H), 2.6-2.7 (m, 2 H), 2.7-2.8 (m, 1 H), 2.9 (d, J = 2.8 Hz, 2 H), 3.6 (t, J = 6.8 Hz, 2 H), 3.9 (s, 3 H), 4.5-4.6 (m, 1 H), 6.8 (d, J = 8.3 Hz, 1 H); ¹³C NMR (CDCl₃, 75.5 MHz) δ -5.3 (q'), 18.3 (s'), 21.9 (t'), 25.9 (q'), 28.3 (t'), 28.6 (t'), 29.9 (t'), 31.7 (t'), 32.1 (t'), 52.38 (q'), 52.4 (d'), 62.6 (t'), 72.4 (d'), 77.3 (s'), 85.4 (s'), 170.7 (s'), 172.0 (s'), 175.1 (s'); exact mass m/z calcd for C₂₀H₃₂NO₆Si (M - CH₃) 410.19989, found 410.19955.

6-Hydroxy-N-[[tetrahydro-5-oxo-2-(2-propyny1)-2-

furanyl]carbonyl]norleucine Methyl Ester (84).



TBAF (1 M in THF, 1.4 mL, 1.41 mmol) and acetic acid (0.12 mL, 3.41 mmol) were added to a solution of **82a** (chromatographically faster-running isomer of **82** (410 mg, 0.97 mmol) in CH_2Cl_2 (3 mL) at 0 °C. The cold bath was removed and stirring was continued for 4 h. The solution was then diluted with EtOAc (40 mL), washed with water (4 x 10 mL) and brine (2 x 20 mL), and dried (MgSO₄). Evaporation of solvent and flash chromatography of the residue over silica gel (2 x 20 cm), using 7:3 EtOAc-hexane, gave pure (¹H NMR, 300 MHz) **84a** (279 mg, 93%) as an oil. This material is the same (¹H NMR) as the chromatographically slower-running product obtained from the EDCI coupling reaction of the unprotected alcohol (see later).

The above procedure was followed, using TBAF (1 M, 0.9 mL, 0.91 mmol), acetic acid (0.08 mL, 1.39 mmol), **82b** (chromatographically slower-running isomer of **82**, (240 mg, 0.56 mmol). Flash chromatography of the crude product over silica gel (1.5 x 20 cm), using 7:3 EtOAc-hexane, gave **84b**

(149 mg, 85%) as an oil. The material is the same as the chromatographically faster-running product obtained from the EDCI coupling reaction of the unprotected alcohol (see later).

6-Hydroxy-N-[[tetrahydro-5-oxo-2-(2-propyny1)-2furany1]carbony1]norleucine Methyl Ester (84). (Alternative way to make 84).



SOCl₂ (0.5 mL, 6.91 mmol) was added to a suspension of the acid **71** (173 mg, 1.02 mmol) in dry PhH (5 mL), and the mixture was refluxed with stirring for 2 h (protection from moisture by a drying tube packed with Drierite), cooled and evaporated to dryness, with protection from moisture. The resulting freshly prepared acid chloride in CH_2Cl_2 (3 mL) was added to a stirred and cooled (0 °C) mixture of amine salt **72** (257 mg, 1.29 mmol), pyridine (0.6 mL, 7.31 mmol) and Et₃N (0.3 mL, 2.15 mmol) in CH_2Cl_2 (5 mL) (Ar atmosphere). Stirring was continued for 2 h and the mixture was quenched with water (30 mL) and extracted with $CHCl_3$ (3 x 10 mL). The combined organic extracts were washed with water (3 x 10 mL) and brine $(2 \times 20 \text{ mL})$, and dried (MgSO₄). Evaporation of the solvent and flash chromatography of the residue over silica gel (1.5 x 20 cm), using 7:3 EtOAc-hexane, gave **84** as an oil in three fractions, two containing the individual pure isomers, and the third containing both isomers (total weight 123 mg, total yield 39%).

The chromatographically slower-running isomer is identical to the chromatographically slower-running isomer prepared by EDCI coupling of the unprotected alcohol (see later). The chromatographically faster-running isomer is identical to the chromatographically faster-running isomer prepared by EDCI coupling of the unprotected alcohol.

6-Oxo-N-[[tetrahydro-5-oxo-2-(2-propyny1)-2-furany1]carbony1]norleucine Methyl Ester (70).



Freshly distilled DMSO (0.175 mL, 2.51 mmol) was added slowly (Ar atmosphere) to a stirred and cooled (-78 °C) solution of oxalyl chloride (0.14 mL, 1.61 mmol) in dry CH_2Cl_2 (2 mL). There was immediate gas evolution. After 0.5 h, 84a (chromatographically slower-running isomer of 84 (150 mg, 0.48 mmol) in dry CH_2Cl_2 (2 mL) was added dropwise. The mixture was stirred for 1 h at -78 °C, and then Et_3N (0.5 mL, 3.58 mmol) was added, and the cooling bath was removed. The mixture was stirred for another 2 h, quenched with water (20 mL), and extracted with CHCl₃ (3 x 10 mL). The combined extracts were washed with water (3 x 10 mL) and brine (2 x 10 mL), and dried (MgSO₄). Evaporation of solvent and flash chromatography of the residue over silica gel (1 x 20 cm), using 1:1 EtOAc-hexane, gave pure (¹H NMR, 300 MHz) **70a** (53 mg, 36%). The material was identical to that prepared by PCC oxidation (see later).

The above procedure was followed, using oxalyl chloride (0.078 mL, 0.89 mmol), DMSO (0.126 mL, 1.71 mmol), **84b** (chromatographically faster-running isomer of **84**, 125 mg, 0.40 mmol) and Et₃N (0.246 mL, 1.76 mmol). Flash chromatography of the crude product over silica gel (1 x 20 cm), using 1:1 EtOAc-hexane, gave pure (¹H NMR, 300 MHz) **70b** (37 mg, 30%). The material was identical to that obtained by PCC oxidation (see later).

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6-Hydroxy-N-[[tetrahydro-5-oxo-2-(2-propyny1)-2furanyl]carbonyl]norleucine Methyl Ester (84). (Alternative way to make 84).



The methyl ester hydrochloride salt was prepared from Ehydroxynorleucine (666 mg, 4.53 mmol) by treatment with saturated methanolic HCl, exactly as described in the literature.⁴⁴ Evaporation of the solvent gave the crude ester hydrochloride, which was used directly after being kept overnight under oil pump vacuum. The material was covered with dry DMF (2 mL) and CH₂Cl₂ (8 mL), and the mixture was stirred with a glass rod for ca. 1 h, by which stage all of the salt had dissolved [use of a magnetic stirring bar was unsuccessful because of the sticky nature of the material]. *N*-Methylmorpholine (4 mL, 36.38 mmol) was then added, and the mixture was stirred (magnetic stirring bar) (Ar atmosphere). After 1 h, the solid lactone acid (760 mg, 4.52 mmol) and 1hydroxybenzotriazole (608 mg, 4.50 mmol) were tipped into the solution, followed by 1-(3-dimethylaminopropyl)-3ethylcarbodiimide (872 mg, 4.55 mmol). Stirring at room temperature was continued overnight (Ar atmosphere), and then water (100 mL) was added. The mixture was extracted with CHCl₃ (4 x 70 mL), and the combined organic extracts were washed with water (3 x 50 mL) and brine (2 x 70 mL), dried (MgSO₄), and evaporated. The crude product was kept overnight under oil pump vacuum to remove traces of *N*methylmorpholine. Flash chromatography of the material over silica gel (2.5 x 20 cm), using 3:1 EtoAc-hexane, gave **84** as an oil in three fractions, two containing the individual pure isomers, and the third fraction containing both isomers.

The chromatographically slower-running material (isomer 84a) (320 mg) had: FTIR (CHCl₃ cast) 1672, 1741, 1788, 3286, 3355 cm⁻¹; ¹H NMR (CD₂Cl₂, 400 MHz) δ 1.3-1.42 (m, 2 H), 1.5-1.6 (m, 2 H), 1.7-1.8 (m, 2 H), 1.85-1.95 (m, 1 H), 2.18 (t, J = 2.4 Hz, 1 H), 2.4-2.65 (m, 3 H), 2.7-2.75 (m, 1 H), 2.8-2.83 (ABX, J = 2.8, 0.8 Hz, 2 H) 3.6 (t, J = 6.4, 2 H), 3.75 (s, 3 H), 4.45-4.5 (m, 1 H), 6.95 (d, J = 8.0 Hz, 1 H); ¹³C (CDCl₃, 100.6 MHz) δ 21.8 (t'), 28.4 (t'), 28.4 (t'), 29.5 (t'), 31.5 (t'), 31.6 (t'), 52.2 (q'), 52.5 (d'), 62.1 (t'), 72.4 (s'), 77.2 (d'), 85.4 (s'), 170.9 (s'), 171.8 (s'), 175.3 (s'); exact mass m/z calcd for C₁₅H₂₁NO₆ 311.13690, found 311.13648.

The chromatographically faster-running material (isomer **84b**) (250 mg) had: FTIR (CHCl₃ cast) 1674, 1742, 1790, 3286, 3341 cm⁻¹; ¹H NMR (CD₂Cl₂, 400 MHz) δ 1.4-1.5 (m, 2 H), 1.5-1.6 (m, 3 H), 1.7-1.75 (m, 1 H), 1.9-1.95 (m, 1 H), 2.16 (t, J = 2.4 Hz, 1 H, 2.35-2.45 (m, 1 H), 2.50-2.65 (m, 2 H), $2.65-2.75 \text{ (m, 1 H)}, 2.75-2.90 \text{ (ABX, } J_{AB} = 17.6, J_{AX} = 2.5, J_{BX}$ = 2.5 Hz, 2 H, 3.60-3.70 (dd, J = 10.9, 5.4 Hz, 2 H), 3.70 (s, 3 H), 4.45-4.50 (m, 1 H), 6.9-7.00 (d, J = 7.6 Hz, 1 H); $^{13}\text{C} \text{ (CDCl}_3, 100.6 \text{ MHz}) \delta 21.9 \text{ (t')}, 28.3 \text{ (t')}, 28.6 \text{ (t')}, 30.0 \text{ (t')}, 31.5 \text{ (t')}, 31.8 \text{ (t')}, 52.4 \text{ (q')}, 52.5 \text{ (d')}, 62.3 \text{ (t')},$ $^{72.4} \text{ (s')}, 85.5 \text{ (s')}, 170.8 \text{ (s')}, 172.0 \text{ (s')}, 175.1 \text{ (s')} \text{ (two signals overlap)}; exact mass <math>m/z$ calcd for $\text{C}_{15}\text{H}_{21}\text{NO}_6$ $^{311.133690}$, found $^{311.13723}$.

The mixed fraction (both isomers) weighed 343 mg. The total yield amounted to 65% in all.

Oxidation of slower-running isomer 84a to 6-Oxo-N-[[tetrahydro-5-oxo-2-(2-propynyl)-2-furanyl]carbonyl]norleucine Methyl Ester (70a).



PCC (1.20 g, 5.45 mmol) was added to a stirred mixture of the chromatographically slower-running alcohol **84a** (300 mg, 0.96 mmol) and crushed 4Å molecular sieves (1.20 g) in dry CH_2Cl_2 (10 mL). Stirring was continued for 1.5 h (Ar atmosphere), and the mixture was then filtered. The solids were washed with a little CH_2Cl_2 , and the combined filtrates were evaporated. Flash chromatography of the residue over silica gel (2 x 22 cm), using 1:1 EtOAc-hexane, gave aldehyde **70a** as a pure (¹H NMR, 300 MHz), colorless oil (190 mg, 66%): FTIR (CHCl₃ cast) 1676, 1724, 1741, 1789, 2732, 3281, 3353 cm⁻¹; ¹H NMR (CD₂Cl₂, 300 MHz) δ 1.6-1.85 (m, 3 H), 1.85-1.95 (m, 1 H), 2.2 (t, J = 2.9 Hz, 1 H), 2.4-2.5 (m, 3 H), 2.5-2.8 (m, 3 H), 2.85 (ABX, 2 H) 3.75 (s, 3 H), 4.45-4.55 (m, 1 H), 6.9-7.0 (br, 1 H), 9.7 (t, J = 1.2 Hz, 1 H); ¹³C NMR (CDCl₃, 75.5 Hz) 17.7 (t'), 28.4 (t'), 28.6 (t'), 29.7 (t'), 31.4 (t'), 42.9 (t'), 52.0 (q'), 52.6 (d'), 72.1 (d'), 77.2 (s') 85.3 (s'), 171.0 (s'), 171.4 (s'), 175.1 (s'), 201.2 (d'); exact mass m/z calcd for C_{15H19}NO₆ 309.12125, found 309.12152.

Oxidation of faster-running isomer 84b to 6-Oxo-N-[[tetrahydro-5-oxo-2-(2-propynyl)-2-furanyl]carbonyl]norleucine Methyl Ester (70b).



The above procedure was followed, using PCC (880 mg, 4.01 mmol), the chromatographically faster-running alcohol 84b (200 mg, 0.64 mmol), crushed 4Å molecular sieves (880 mg) and dry CH₂Cl₂ (10 mL). Flash chromatography of the crude product over silica gel (2 x 22 cm), using 1:1 EtOAc-hexane, gave aldehyde **70b** as a pure (¹H NMR, 300 MHz), colorless oil (173 mg, 87%): FTIR (CHCl₃ cast) 1676, 1723, 1741, 1789, 2731, 3283, 3357 cm⁻¹; ¹H NMR (CD₂Cl₂, 300 MHz) δ 1.65-1.8 (m, 3 H), 1.85-1.90 (m, 1 H), 2.15 (t, J = 2.7 Hz, 1 H), 2.3-2.4 (m, 1 H), 2.45-2.75 (m, 5 H), 2.80-2.95 (ABX, $J_{AB} = 17.4$, J_{AX} = 2.7, $J_{BX} = 2.7$ Hz, 2 H), 3.75 (s, 3 H), 4.5-4.6 (m, 1 H), 6.9-7.0 (br, 1 H), 9.75 (t, J = 1.2 Hz, 1 H); ¹³C NMR (CD₂Cl₂, 50.3 Hz) δ 18.0 (t'), 28.1 (t'), 28.6 (t'), 30.1 (t'), 30.9 (t'), 42.9 (t'), 52.0 (q'), 52.4 (d'), 72.0 (d'), 77.7 (s'), 85.5 (s'), 170.6 (s[']), 171.7 (s'), 175.1 (s'), 201.7 (d'); exact mass m/z calcd for C₁₅H₁₉NO₆ 309.12125, found 309.12229; calcd. for C₁₄H₁₈O₅N (M - CHO) 280.11850, found 280.11862.

Note that the two aldehydes (from the two alcohols) have the same $R_{\rm f}.$

Methyl 1,2,3,4-Tetrahydro-1-[[tetrahydro-5-oxo-2-(2propynyl)-2-furanyl]carbonyl]-2-pyridinecarboxylate (69a).



For this reaction the CF3COOH must be freshly distilled from P_2O_5 , but can be stored for 1 week in a tightly stoppered flask kept in a small desiccator containing Drierite. TFA (0.40 mL, 5.19 mmol) was added to a stirred mixture of aldehyde 70a (150 mg, 0.49 mmol) from the chromatographically slower-running alcohol and crushed 4Å molecular sieves (150 mg) in dry CH₂Cl₂ (5 mL) (Ar atmosphere). Stirring at room temperature was continued for 3-4 h, the course of the reaction being closely followed by TLC (silica, 3:7 EtOAchexane). As soon as the reaction was complete, the mixture was filtered, and the solids were washed with a little CH_2Cl_2 . Evaporation of the combined filtrate and flash chromatography of the residue over silica gel (1.5 x 20 cm), using 3:7 EtOAc-hexane, gave pure (¹H NMR, 300 MHz) 69a (122 mg, 82%) as a colorless oil. At room temperature the material consisted of a mixture of two rotamers (variable temperature ¹H NMR): FTIR (CHCl₃, cast) 1643, 1743, 1794, 3270 cm⁻¹; ¹H NMR (CD₂Cl₂, 300 MHz) δ 1.9-2.17 (m, 3 H), 2.18-2.25 (m, 1 H) 2.25-2.60 (m, 3 H), 2.60-2.85 (m, 2 H), 2.90-3.05 (m, 2 H), 3.75 (s, 3 H), 5.05-5.50 (br, 2 H), 7.10-7.20 (br, 1 H); ^{13}C NMR (CD₂Cl₂, 100.6 MHz) (mixture of rotamers) δ 19.7 (t'), 19.2 (t'), 23.7 (t'), 24.3 (t'), 27.8 (t'), 28.2 (t'), 28.8 (t'), 30.3 (t'), 30.7 (t'), 33.1 (t'), 52.7 (d'), 53.0 (d'), 54.1 (q'), 56.5 (q'), 72.5 (d'), 72.9 (d'), 77.1 (s'), 77.8 (s'), 86.7 (s'), 87.6 (s'), 108.6 (d'), 110.8 (d'), 124.5 (d'), 125.2 (d'), 167.8 (s'), 168.1 (s'), 170.8 (s'), 171.6 (s'), 174.9 (s'), 175.4 (s'); exact mass *m/z* calcd for

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C₁₅H₁₇NO₅ 291.11069, found 291.11123.

Methyl 1,2,3,4-Tetrahydro-1-[[tetrahydro-5-oxo-2-(2propynyl)-2-furanyl]carbonyl]-2-pyridinecarboxylate (69b).



The above procedure was followed, using TFA (0.50 mL, 6.49 mmol), the chromatographically faster-running alcohol 70b (222 mg, 0.72 mmol), crushed 4Å molecular sieves (222 mg) and dry CH_2Cl_2 (5 mL). Stirring at room temperature was continued for 3-4 h, the course of the reaction being closely followed by TLC (silica, 3:7 EtOAc-hexane). Flash chromatography of the crude product over silica gel (1.5 x 20 cm), using 3:7 EtOAc-hexane, gave pure (¹H NMR, 300 MHz) **69b** (120 mg, 65% after correction for recovered starting material) as a colorless oil, and recovered starting material (31 mg, 14%). An increase in reaction time gave a lower yield. At room temperature the product is a single isomer [sharp signals in the ¹H MHz NMR spectrum (300 MHz)]: FTIR (CHCl₃ cast) 1643, 1667, 1743, 1793, 3268 cm⁻¹; ¹H NMR (CD₂Cl₂, 300 MHz) δ 1.85-2.05 (m, 3 H) 2.15 (t, J = 2.7 Hz, 1 H), 2.30-2.40 (m, 2 H), 2.50-2.75 (m, 2 H), 2.85-2.95 (ABX, 2 H), 2.98-3.05 (m, 1 H), 3.75 (s, 3 H), 5.00-5.10 (br, 1 H), 5.12-5.22 (br, 1 H), 7.1 (d, J = 8.7 Hz, 1 H); ¹³C NMR (CD₂Cl₂, 100.6 MHz) δ 19.3 (t'), 23.9 (t'), 28.2 (t'), 29.0 (t'), 30.8 (t'), 52.8 (d'), 54.1 (q'), 72.9 (d'), 77.2 (s'), 86.7 (s'), 109.2 (d'), 124.9 (d'), 167.2 (s'), 170.9 (s'), 175.3 (s'); exact mass m/z calcd for C₁₅H₁₇NO₅ 291.11069, found 291.11014.

Methyl Octahydro-4',5-dioxo-1'-[(triphenylstannyl)methylene]-spiro[furan-2(3H),3'(4H')-[2H]quinolizine]6'-carboxylate (85a).



A solution of AIBN (7 mg, 0.04 mmol) and Ph₃SnH (148 mg, 0.42 mmol) in PhMe (1 mL) was injected over 2 min into a stirred and refluxing solution of acetylene **69a** (derived from the chromatographically slower-running alcohol) (46 mg, 0.16 mmol) in PhMe (2 mL) (Ar atmosphere). Refluxing was continued for 4 h, and the mixture was then cooled to room temperature and evaporated. Flash chromatography of the

residue over silica gel $(1 \times 20 \text{ cm})$, using 3:7 EtOAc-hexane, gave **85a** (72 mg) as a white powder contaminated with triphenyltin residues (¹H NMR). The crude material was directly subjected to protodestannylation.

Note that with Bu₃SnH, slow addition led to recovery of considerable starting material; hence, the stannane was added quickly this time. Fast addition of Bu₃SnH also led to another product, which was not identified.

Methyl Octahydro-4',5-dioxo-1'-[(triphenylstannyl)methylene]-spiro[furan-2(3H),3'(4H')-[2H]quinolizine]6'-carboxylate (85b).



A solution of AIBN (10 mg, 0.05 mmol) and Bu₃SnH (210 mg, 0.72 mmol) in PhMe (2 mL) was injected over 2 min into a stirred and refluxing solution of acetylene **69b** (derived from the chromatographically faster-running alcohol) (100 mg, 0.34 mmol) in PhMe (2 mL) (Ar atmosphere). Refluxing was continued for 2 h, and the mixture was then cooled to room temperature and evaporated. Flash chromatography of the

residue over silica gel $(1.5 \times 20 \text{ cm})$, using 3:7 EtOAchexane, gave **85b** (170 mg) as a white powder contaminated with tributyltin residues (¹H NMR). The crude material was directly subjected to protodestannylation.

Methyl Octahydro-1'-methylene-4',5-dioxospiro[furan-2(3H),3'(4'H')-[2H]quinolizine]-6'-carboxylate (30a).



TFA (0.5 mL, 6.51 mmol) was added to a stirred solution of vinyl stannane **85a** (derived from the slower-running alcohol) (72 mg) in THF (2 mL). Stirring was continued overnight at room temperature and then at 60 °C for 2 h. The solution was cooled to room temperature and evaporated. Flash chromatography of the residue over silica gel (1 x 20 cm), using 1:1 EtOAc-hexane, gave pure **30a** (29 mg, 62% over two steps) as a pure (¹H NMR, 360 MHz), white powder: FTIR (CHCl₃ cast) 1655, 1741, 1785 cm⁻¹; ¹H NMR (CDCl₃, 360 MHz) δ 1.7-1.8 (m, 3 H), 1.9-2.2 (m, 4 H), 2.4-2.5 (m, 2 H), 2.7-3 (m, 3 H), 3.75 (s, 3 H), 4.10-4.15 (br. 1 H), 4.3 (t, J = 5.7 Hz, 1 H), 5.05 (d, J = 7.9 Hz, 2 H); ¹³C NMR (CDCl₃, 75.5 MHz) δ 18.2 (t'), 24.2 (t'), 27.6 (t'), 28.5 (t'), 32.4 (t'), 41.6 (t'), 52.2 (d'), 56.2 (d'), 57.0 (q'), 81.8 (s'), 113.8 (t'), 138.5 (s'), 169.6 (s'), 171.6 (s'), 176.4 (s'); exact mass m/z calcd for C₁₅H₁₉NO₅ 293.12631, found 293.12580.

Methyl Octahydro-1'-methylene-4',5-dioxospiro[furan-2(3H),3'(4'H')-[2H]quinolizine]-6'-carboxylate (30b).



TFA (3 drops) was added to a stirred solution of vinyl stannane **85b** (derived from the faster-running alcohol) (170 mg) in THF (5 mL). Stirring was continued at room temperature for 5 h, and the solution was then evaporated. Flash chromatography of the residue over silica gel (1.5 x 20 cm), using 1:1 EtOAc-hexane, gave **30b** (86 mg, 89% over two steps) as a pure (¹H NMR, 300 MHz), white powder: FTIR (CHCl₃ cast) 1659, 1723.35, 1772.74 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 1.4-1.5 (m, 2 H), 1.55-1.65 (m, 1 H), 1.75-1.85 (m, 1 H), 1.9-2.0 (m, 2 H), 2.25-2.35 (m, 2 H), 2.45-2.55 (m, 1 H), 2.65 (d, J = 13.5, 1 H), 2.75-2.85 (m, 1 H), 2.95 (d, J = 13.5, 1 H), 3.75 (s, 3 H), 4.10 (d, J = 10.2 Hz, 1 H), 5.15 (s, 2 H), 5.35 (d, J = 5.4 Hz, 1 H); ¹³C NMR (CDCl₃, 100.6 MHz) δ 20.6 (t'), 26.2 (t'), 28.4 (t'), 30.3 (t'), 33.0 (t'),

41.4 (t'), 52.4 (d'), 52.5 (d'), 57.7 (q') 81.9 (s'), 113.9 (t'), 138.4 (s'), 169.6 (s'), 170.9 (s'), 176.3 (s'); exact mass m/z calcd for C_{15H19}NO₅ 293.12631, found 293.12936.





Ozonized oxygen was passed into a stirred and cooled (-78 °C) solution of olefin **30a** (derived from the slowerrunning alcohol) (30 mg, 0.10 mmol) in CH₂Cl₂ (4 mL) contained in a three-necked flask equipped with a reflux condenser (not connected to a water supply) and closed by a drying tube packed with Drierite, an inlet tube (for ozone), and a glass stopper. When the solution became pale blue, the ozonization was stopped, and the excess of ozone was removed with a stream of oxygen. Ph₃P (55 mg, 0.21 mmol) was tipped into the solution, the cooling bath was removed, and stirring was continued for 1.5 h.

The solvent was evaporated and Et_3N (0.10 mL, 0.72 mmol) in dry THF (3 mL) was added. The mixture was stirred at 60 °C for 2 h, cooled to room temperature, diluted with EtOAc (10 mL), and extracted with water (4 x 5 mL). The combined aqueous extracts were washed with $CHCl_3$ (4 x 5 mL). Evaporation of the aqueous phase gave a solid residue.

A column packed with Amberlite IR-120 ion-exchange resin $(20-50\text{\AA mesh}, 1 \times 10 \text{ cm})$ was washed with water until the eluent was colorless. The column was then washed successively with 2 N aqueous NaOH (4 bed volumes), water (until the eluent was neutral to pH paper), 2 N HCl (4 bed volumes), and finally with water (until the eluent was neutral to pH paper). The above solid residue was dissolved in water (ca. 0.5 mL) and the solution was passed down the column, using water. The eluent was monitored by TLC [silica, 2:4:4 AcOH-hexane-EtOAc-water (1 drop water to 10 mL of the organic phase), visualization under uv light]. Evaporation of the combined uv-active fractions (water pump, rotary evaporator, bath temperature below 60 °C) gave ester 32 (19 mg, 63%) as a pure (¹H NMR, 400 MHz), yellow foam: FTIR (microscope) 1654, 1717, 1740, 2800-3400 (br) cm⁻¹; ¹H NMR (D₂O, 400 MHz) δ 1.58-1.60 (m, 1 H), 1.80-1.88 (m, 1 H), 2.10-2.20 (m, 1 H), 2.25-2.35 (m, 1 H), 2.6-2.70 (m, 2 H), 2.7-2.8 (m, 3 H), 1.90-1.95 (m, 1 H), 3.75 (s, 3 H), 5.10-5.15 (dd, J = 6.8, 4.4 Hz, 1 H), 7.35 (s, 1 H); ¹³C NMR (D₂O, 100.6 MHz) δ 16.2 (t'), 23.8 (t'), 26.1 (t'), 26.5 (t'), 33.4 (t'), 54.3 (d'), 57.7 (q'), 126.9 (s'), 132.9 (s'), 133.7 (d'), 137.8 (s'), 161.9 (s'), 175.0 (s'), 178.6 (s'); exact mass m/z calcd for C₁₄H₁₇NO₆ 295.10559, found 295.10566.

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1,3,4,6-Tetrahydro-9-hydroxy-4-(methoxycarbonyl)-6-

oxo-2H-quinolizine-7-propanoic acid (32) from 30b.



The above procedure was followed exactly, using exocyclic olefin **30b** (derived from the faster-running alcohol) (58 mg, 0.20 mmol) in CH_2Cl_2 (5 mL), Ph_3P (107 mg, 0.41 mmol), and Et_3N (0.20 mL, 1.43 mmol) in THF (3 mL). Ester **32** (37 mg, 63%) was obtained as a pure (¹H NMR, 400 MHz), yellow foam, spectroscopically identical to material obtained in the previous experiment.

4-Carboxy-1,3,4,6-tetrahydro-9-hydroxy-6-oxo-2Hquinolizine-7-propanoic acid (2).



(Bu₃Sn)₂O (0.5 mL, 0.98 mmol) was added to a solution of

methyl ester 32 (91 mg, 0.31 mmol) in dry PhH (2 mL), and the mixture was refluxed for 3 days (Ar atmosphere) [TLC control, silica, 2:4:4 AcOH-hexane-EtOAc-water (1 drop water to 10 mL of the organic phase)]. The deep red solution was cooled to room temperature and diluted with hexane (10 mL). The resulting solution was extracted with 1% AcOH-water (4 x 5 mL), and the combined aqueous extracts were washed with CHCl3 (4 x 10 mL). Evaporation of the aqueous extracts (rotary evaporator, water pump, below 60 °C) and flash chromatography of the residue over C-18 reverse phase silica gel (60 mesh, 10% C-18 capped with TMS, 1 x 20 cm), using 9:1 water-MeCN, gave the crude product. This was dissolved in the minimum amount of water at room temperature. The solution was stored overnight in a refrigerator to obtain pure $(^{1}H NMR, 400 MHz)$ (\pm) -A58365B¹⁶ (2) (60 mg, 67%) as a yellow crystalline solid: mp 209-213 °C; FTIR (microscope) 1651, 1709, 2600-3200 (br) cm⁻¹; UV (MeOH) λ_{max} : 340, 238 nm; ¹H NMR (D₂O, 300 MHz) δ 1.65-1.70 (m, 1 H), 1.85-1.90 (m, 1 H), 2.15-2.25 (m, 1 H), 2.3-2.35 (m, 1 H), 2.65-2.85 (m, 5 H), 2.95-3.00 (m, 1 H), 5.10-5.15 (dd, J = 6.4, 3.6 Hz, 1 H), 7.35 (s, 1 H); ¹³C NMR $(D_2O, 100.6 \text{ MHz}) \delta 16.1 (t'), 23.7 (t'), 26.1 (t'), 26.5 (t'),$ 33.4 (t'), 57.5 (d'), 126.8 (s'), 132.7 (s'), 133.5 (d'), 137.6 (s'), 161.8 (s'), 176.2 (s'), 178.6 (s'); Exact mass m/z calcd for C₁₃H₁₅NO₆ 281.08994, found 281.08743.

(+)-6-Hydroxy-N-[[tetrahydro-5-oxo-2-(2-propyny1)-2furany1]carbony1]norleucine Methyl Ester [(+)-84].



The methyl ester hydrochloride salt was prepared from (L)-E-hydroxynorleucine (603 mg, 3.05 mmol) by treatment with saturated methanolic HCl, exactly as described in the literature.⁴⁴ Evaporation of the solvent gave the crude ester hydrochloride, which was used directly after being kept overnight under oil pump vacuum. The material was covered with dry DMF (4 mL) and CH_2Cl_2 (15 mL), and the mixture was stirred with a glass rod for ca. 1 h, by which stage all of the salt had dissolved [use of a magnetic stirring bar was unsuccessful because of the sticky nature of the material]. N-Methylmorpholine (4 mL, 36.38 mmol) was then added, and the mixture was stirred (magnetic stirring bar) (Ar atmosphere). After 1 h, the solid lactone acid (640 mg, 3.81 mmol) and 1hydroxybenzotriazole (717 mg, 4.68 mmol) were tipped into the solution, followed by 1-(3-dimethylaminopropyl)-3ethylcarbodiimide (805 mg, 4.11 mmol). Stirring at room

temperature was continued overnight (Ar atmosphere), and then water (100 mL) was added. The mixture was extracted with CHCl₃ (4 x 70 mL), and the combined organic extracts were washed with water (3 x 50 mL) and brine (2 x 70 mL), dried (MgSO₄), and evaporated. The crude product was kept overnight under oil pump vacuum to remove traces of *N*methylmorpholine. Flash chromatography of the material over silica gel (2.5 x 20 cm), using 3:1 EtOAc-hexane, gave (+)-84 as an oil in three fractions, two containing the individual pure isomers, and the third fraction containing both isomers.

The chromatographically slower-running material [isomer (+)-84a] (303 mg) had: $[\alpha]_D^{21} = +3.57$ ° (c 1.485 CHCl₃); FTIR (CHCl₃ cast) 1672, 1741, 1788, 3286, 3355 cm⁻¹; ¹H NMR (CDCl₃, 360 MHz) δ 1.35-1.42 (m, 2 H), 1.5-1.6 (m, 2 H), 1.7-1.85 (m, 2 H), 1.85-1.95 (m, 1 H), 2.18 (t, J = 2.9 Hz, 1 H), 2.4-2.65 (m, 3 H), 2.7-2.75 (m, 1 H), 2.8 (d, J = 2.52 Hz, 2 H) 3.65 (t, J = 6.12, 2 H), 3.78 (s, 3 H), 4.5-4.6 (m, 1 H), 7.0 (d, J = 7.9 Hz, 1 H); ¹³C (CDCl₃, 100.6 MHz) δ 21.8 (t'), 28.4 (t'), 29.5 (t'), 31.5 (t'), 31.6 (t'), 52.2 (q'), 52.5 (d'), 62.1 (t'), 72.4 (s'), 77.2 (d'), 85.4 (s'), 170.9 (s'), 171.8 (s'), 175.3 (s'); exact mass m/z calcd for C₁₅H₂₂NO₆ (M⁺ + 1) 312.14471, found 312.14228; Calcd for C₁₃H₁₈NO₄ (M⁺ -COOMe) 252.12358, found 252.12339.

The chromatographically faster-running material [isomer (+)-84b] (160 mg) had: $[\alpha]_D^{21} = +9.43$ ° (c 1.1126 CHCl₃); FTIR (CHCl₃ cast) 1674, 1742, 1790, 3286, 3341 cm⁻¹; ¹H NMR (CDCl₃, 360 MHz) δ 1.4-1.5 (m, 2 H), 1.5-1.6 (m, 3 H), 1.7166

1.75 (m, 1 H), 1.9-1.95 (m, 1 H), 2.15 (t, J = 2.88 Hz, 1 H), 2.35-2.45 (m, 1 H), 2.50-2.65 (m, 2 H), 2.65-2.75 (m, 1 H), 2.8-2.85 (ABX, 2 H), 3.65 (t, J = 6.12, 2 H), 3.80 (s, 3 H), 4.45-4.50 (m, 1 H), 6.9-7.00 (d, J = 7.6 Hz, 1 H); ¹³C (CDC1₃, 100.6 MHz) δ 21.9 (t'), 28.3 (t'), 28.6 (t'), 30.0 (t'), 31.5 (t'), 31.8 (t'), 52.4 (q'), 52.5 (d'), 62.3 (t'), 77.4 (s'), 85.5 (s'), 170.8 (s'), 172.0 (s'), 175.1 (s'); exact mass m/zcalcd for C₁₅H₂₁NO₆ 311.13690, found 311.13712.

The mixed fraction (both isomers) weighed 551 mg. The total yield amounted to 83% in all.

Oxidation of slower-running isomer (+)-84a to (+)-6-Oxo-N-[[tetrahydro-5-oxo-2-(2-propyny1)-2-furany1]carbony1]-norleucine Methyl Ester [(+)-70a].



PCC (310 mg g, 1.41 mmol) was added to a stirred mixture of the chromatographically slower-running alcohol (+)-84a (110 mg, 0.35 mmol) and crushed 4Å molecular sieves (500 mg) in dry CH_2Cl_2 (10 mL). Stirring was continued for 1.5 h (Ar atmosphere), and the mixture was then filtered. The solids were washed with a little CH_2Cl_2 , and the combined filtrates were evaporated. Flash chromatography of the residue over silica gel (2 x 22 cm), using 1:1 EtOAc-hexane, gave aldehyde (+)-70a as a pure (¹H NMR, 300 MHz), colorless oil (78 mg, 76%): $[\alpha]_D^{21} = +6.22$ ° (c 0.724, CHCl₃); FTIR (CHCl₃ cast) 1676, 1724, 1741, 1789, 3281, 3353 cm⁻¹; ¹H NMR (CDCl₃, 360 MHz) δ 1.6-1.85 (m, 3 H), 1.85-1.95 (m, 1 H), 2.15 (t, J =2.63 Hz, 1 H), 2.4-2.5 (m, 3 H), 2.5-2.8 (m, 3 H), 2.85 (ABX, 2 H) 3.75 (s, 3 H), 4.45-4.55 (m, 1 H), 6.9-7.0 (br, 1 H), 9.7 (t, J = 1.08 Hz, 1 H); ¹³C NMR (CDCl₃, 100.6 Hz) 17.7 (t'), 28.4 (t'), 28.6 (t'), 29.7 (t'), 31.4 (t'), 42.9 (t'), 52.0 (q'), 52.6 (d'), 72.1 (d'), 77.2 (s') 85.3 (s'), 171.0 (s'), 171.4 (s'), 175.1 (s'), 201.2 (d'); exact mass m/z calcd for C_{15H18}NO₆ (M⁺ - 1) 308.11340, found 308.11295.

Oxidation of faster-running isomer (+)-84b to (+)-6-Oxo-N-[[tetrahydro-5-oxo-2-(2-propynyl)-2-furanyl]carbonyl]-norleucine Methyl Ester [(+)70b].



The above procedure was followed, using PCC (468 mg, 2.12 mmol), the chromatographically faster-running alcohol (+)-84b (468 mg, 2.12 mmol), crushed 4Å molecular sieves (480
mg) and dry CH₂Cl₂ (10 mL). Flash chromatography of the crude product over silica gel (2 x 22 cm), using 1:1 EtOAc-hexane, gave aldehyde (+)-70b as a pure (¹H NMR, 300 MHz), colorless oil (140 mg, 64%): $[\alpha]_D^{21} = +11.5$ ° (c 1.139, CHCl₃); FTIR (CHCl₃ cast) 1676, 1723, 1741, 1789, 2731, 3283, 3357 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 1.65-1.8 (m, 3 H), 1.85-1.90 (m, 1 H), 2.15 (t, J = 2.52 Hz, 1 H), 2.3-2.4 (m, 1 H), 2.45-2.75 (m, 5 H), 2.80-2.95 (ABX, 2 H), 3.75 (s, 3 H), 4.5-4.6 (m, 1 H), 6.9-7.0 (br, 1 H), 9.75 (t, J = 1.08 Hz, 1 H); ¹³C NMR (CDCl₃, 100.6 Hz) δ 18.0 (t'), 28.1 (t'), 28.6 (t'), 30.1 (t'), 30.9 (t'), 42.9 (t'), 52.0 (q'), 52.4 (d'), 72.0 (d'), 77.7 (s'), 85.5 (s'), 170.6 (s'), 171.7 (s'), 175.1 (s'), 201.7 (d'); exact mass m/z calcd for C₁₅H₁₉NO₆ 309.12125, found 309.12268; calcd. for C₁₄H_{18N}O₅ (M - CHO) 280.11850, found 280.11707.

Note that the two aldehydes (from the two alcohols) have the same $R_{\rm f}.$

(-)-Methyl 1,2,3,4-Tetrahydro-1-[[tetrahydro-5-oxo-2-(2-propynyl)-2-furanyl]carbonyl]-2-pyridinecarboxylate [(-)-69a].



For this reaction the CF3COOH must be freshly distilled from P_2O_5 , but can be stored for 1 week in a tightly stoppered flask kept in a small desiccator containing Drierite. TFA (0.40 mL, 5.19 mmol) was added to a stirred mixture of aldehyde (+)-70a (315 mg, 1.02 mmol) from the chromatographically slower-running alcohol and crushed 4Å molecular sieves (300 mg) in dry CH₂Cl₂ (5 mL) (Ar atmosphere). Stirring at room temperature was continued for 3-4 h, the course of the reaction being closely followed by TLC (silica, 3:7 EtOAc-hexane). As soon as the reaction was completed, the mixture was filtered, and the solids were washed with a little CH_2Cl_2 . Evaporation of the combined filtrate and flash chromatography of the residue over silica gel (1.5 x 20 cm), using 3:7 EtOAc-hexane, gave pure (¹H NMR, 300 MHz) (-)-69a (151 mg, 96% after correction for recovered starting material 150 mg)) as a colorless oil. At room temperature the material consisted of a mixture of two rotamers (variable temperature ¹H NMR): $[\alpha]_D^{21} = -62.7^\circ$ (c 1.01, CHCl₃); FTIR (CHCl₃, cast) 1643, 1743, 1794, 3270 cm⁻¹; ¹H NMR (CDCl₃, 360 MHz) δ 1.9-2.17 (m, 3 H), 2.18 (t, J = 2.7 Hz 1 H), 2.25-2.60 (m, 3 H), 2.60-2.85 (m, 2 H), 2.90-3.05 (m, 2 H), 3.75 (d, 3 H), 5.05-5.50 (br, 2 H), 7.10-7.20 (br, 2 H)1 H); ¹³C NMR (CDCl₃, 100.6 MHz) (mixture of rotamers) δ 18.7 (t'), 18.8 (t'), 23.2 (t'), 23.9 (t'), 27.4 (t'), 27.8 (t'), 28.4 (t'), 30.3 (t'), 30.7 (t'), 32.8 (t'), 52.5 (d'), 52.8 (d'), 53.7 (q'), 56.1 (q'), 72.4 (d'), 72.9 (d'), 77.1 (s'), 77.8 (s'), 86.4 (s'), 87.4 (s'), 108.5 (d'), 110.6 (d'),

124.3 (d'), 124.9 (d'), 167.5 (s'), 167.8 (s'), 170.5 (s'), 171.3 (s'), 174.6 (s'), 175.2 (s'); exact mass m/z calcd for $C_{15H_{17}NO_5}$ 291.11069, found 291.11042.

(-)-Methyl 1,2,3,4-Tetrahydro-1-[[tetrahydro-5-oxo-2-(2-propynyl)-2-furanyl]carbonyl]-2-pyridinecarboxylate [(-)69b].



The above procedure was followed, using TFA (0.50 mL, 6.49 mmol), the chromatographically faster-running alcohol (+)-70b (470 mg, 1.52 mmol), crushed 4Å molecular sieves (422 mg) and dry CH_2Cl_2 (5 mL). Stirring at room temperature was continued for 3-4 h, the course of the reaction being closely followed by TLC (silica, 3:7 EtOAc-hexane). Flash chromatography of the crude product over silica gel (1.5 x 20 cm), using 3:7 EtOAc-hexane, gave pure (¹H NMR, 300 MHz) (-)-69b (159 mg, 73% after correction for recovered starting material 240 mg) as a colorless oil. An increase in reaction time gave a lower yield. At room temperature the product is a single isomer [sharp signals in the ¹H MHz NMR spectrum (300 MHz): $[\alpha]_D^{21} = -114.9$ ° (c 0.684, CHCl₃); FTIR (CHCl₃ cast) 1643, 1667, 1743, 1793, 3268 cm⁻¹; ¹H NMR (CDCl₃, 360 MHz) δ 1.85-2.05 (m, 3 H) 2.15 (t, J = 2.54 Hz, 1 H), 2.30-2.40 (m, 2 H), 2.50-2.75 (m, 2 H), 2.85-2.95 (ABX, 2 H), 2.98-3.05 (m, 1 H), 3.75 (s, 3 H), 5.00-5.10 (br, 1 H), 5.12-5.22 (br, 1 H), 7.05-7.12 (d, J = 8.7 Hz, 1 H); ¹³C NMR (CDCl₃, 75.5 MHz) δ 19.3 (t'), 23.9 (t'), 28.2 (t'), 29.0 (t'), 30.8 (t'), 52.8 (d'), 54.1 (q'), 72.9 (d'), 77.2 (s'), 86.7 (s'), 109.2 (d'), 124.9 (d'), 167.2 (s'), 170.9 (s'), 175.3 (s'); exact mass m/z calcd for C_{15H17}NO₅ 291.11069, found 291.11049.

(-)-Methyl Octahydro-4', 5-dioxo-1'-[(triphenylstannyl)methylene]-spiro[furan-2(3H), 3'(4H')-[2H]quinolizine]-6'-carboxylate [(-)-85a].



A solution of AIBN (14 mg, 0.08 mmol) and Ph₃SnH (363 mg, 1.03 mmol) in PhMe (1 mL) was injected over 2 min into a stirred and refluxing solution of acetylene (-)-69a (derived from the chromatographically slower-running alcohol) (127 mg, 0.44 mmol) in PhMe (6 mL) (Ar atmosphere). Refluxing was

continued for 4 h, and the mixture was then cooled to room temperature and evaporated. Flash chromatography of the residue over silica gel (1 x 20 cm), using 3:7 EtOAc-hexane, gave (-)-85a (228 mg) as a white powder contaminated with triphenyltin residues (¹H NMR). The crude material was directly subjected to protodestannylation.

(-)-Methyl Octahydro-4', 5-dioxo-1'-[(triphenylstannyl)methylene]-spiro[furan-2(3H), 3'(4H')-[2H]quinolizine]-6'-carboxylate [(-)-85b].



A solution of AIBN (10 mg, 0.05 mmol) and Bu_3SnH (367 mg, 1.26 mmol) in PhMe (2 mL) was injected over 2 min into a stirred and refluxing solution of acetylene (-)-69b (derived from the chromatographically faster-running alcohol) (150 mg, 0.58 mmol) in PhMe (2 mL) (Ar atmosphere). Refluxing was continued for 2 h, and the mixture was then cooled to room temperature and evaporated. Flash chromatography of the residue over silica gel (1.5 x 20 cm), using 3:7 EtOAc-hexane, gave (-)-85b (220 mg) as a white powder contaminated with tributyltin residues (¹H NMR). The crude material was

directly subjected to protodestannylation.

(-)-Methyl Octahydro-1'-methylene-4',5-dioxospiro-[furan-2(3H),3'(4'H')-[2H]quinolizine]-6'-carboxylate [(-)30a].



TFA (0.5 mL, 6.51 mmol) was added to a stirred solution of vinyl stannane (-)-85a (derived from the slower-running alcohol) (228 mg) in THF (2 mL). Stirring was continued overnight at room temperature and then at 60 °C for 2 h. The solution was cooled to room temperature and evaporated. Flash chromatography of the residue over silica gel (1×20) cm), using 1:1 EtOAc-hexane, gave pure (-)-30a (85 mg, 65% over two steps) as a pure (¹H NMR, 360 MHz), white powder: $[\alpha]_D^{21} = -21.58$ ° (c 0.811, CHCl₃); FTIR (CHCl₃ cast) 1655, 1741, 1785 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 1.7-1.8 (m, 3 H), 1.9-2.2 (m, 4 H), 2.4-2.5 (m, 2 H), 2.7-3 (m, 3 H), 3.75 (s, 3 H), 4.10-4.15 (br. 1 H), 4.3 (t, J = 5.7 Hz, 1 H), 5.05-5.10 (d, J = 7.9 Hz, 2 H); ¹³C NMR (CDCl₃, 100.6 MHz) δ 18.2 (t'), 24.2 (t'), 27.6 (t'), 28.5 (t'), 32.4 (t'), 41.6 (t'), 52.2 (d'), 56.2 (d'), 57.0 (q'), 81.8 (s'), 113.8 (t'), 138.5

(s'), 169.6 (s'), 171.6 (s'), 176.4 (s'); exact mass m/z calcd for C₁₅H₁₉NO₅ 293.12631, found 293.12662.

(-)-Methyl Octahydro-1'-methylene-4',5-dioxospiro-[furan-2(3H),3'(4'H')-[2H]quinolizine]-6'-carboxylate [(-)-30b].



TFA (3 drops) was added to a stirred solution of vinyl stannane (-)-**85b** (derived from the faster-running alcohol) (220 mg) in THF (5 mL). Stirring was continued at room temperature for 5 h, and the solution was then evaporated. Flash chromatography of the residue over silica gel (1.5 x 20 cm), using 1:1 EtOAc-hexane, gave (-)-**30b** (97 mg, 68% over two steps) as a pure (¹H NMR, 300 MHz), white powder: $[\alpha]_D^{21} = -101.86^{\circ}$ (c 0.91, CHCl₃); FTIR (CHCl₃ cast) 1659, 1723.35, 1772.74 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 1.4-1.5 (m, 2 H), 1.55-1.65 (m, 1 H), 1.75-1.85 (m, 1 H), 1.9-2.0 (m, 2 H), 2.25-2.35 (m, 2 H), 2.45-2.55 (m, 1 H), 2.6 (d, J = 13.5, 1 H), 2.75-2.85 (m, 1 H), 2.95 (d, J = 13.5 Hz, 1 H), 3.75 (s, 3 H), 4.1 (d, J = 10.2 Hz, 1 H), 5.1 (s, 2 H), 5.35 (d, J = 5.4 Hz, 1 H); ¹³C NMR (CDCl₃, 75.5 MHz) δ 20.6 (t'), 26.2

(t'), 28.4 (t'), 30.3 (t'), 33.0 (t'), 41.4 (t'), 52.4 (d'), 52.5 (d'), 57.7 (q') 81.9 (s'), 113.9 (t'), 138.4 (s'), 169.6 (s'), 170.9 (s'), 176.3 (s'); exact mass m/z calcd for C_{15H19}NO₅ 293.12631, found 293.12611.

(-)-1,3,4,6-Tetrahydro-9-hydroxy-4-(methoxycarbony1)-6-oxo-2H-quinolizine-7-propanoic acid [(-)-32] from [(-)-30a].



Ozonized oxygen was passed into a stirred and cooled (-78 °C) solution of olefin (-)-**30a** (derived from the slowerrunning alcohol) (170 mg, 0.58 mmol) in CH₂Cl₂ (4 mL) contained in a three-necked flask equipped with a reflux condenser (not connected to a water supply) and closed by a drying tube packed with Drierite, an inlet tube (for ozone), and a glass stopper. When the solution became pale blue, the ozonization was stopped, and the excess of ozone was removed with a stream of oxygen. Ph₃P (250 mg, 0.95 mmol) was tipped into the solution, the cooling bath was removed, and stirring was continued for 1.5 h.

The solvent was evaporated and Et₃N (0.10 mL, 0.72 mmol)

in dry THF (3 mL) was added. The mixture was stirred at 60 °C for 2 h, cooled to room temperature, diluted with EtOAc (10 mL), and extracted with water (4 x 5 mL). The combined aqueous extracts were washed with $CHCl_3$ (4 x 5 mL). Evaporation of the aqueous phase gave a solid residue.

A column packed with Amberlite IR-120 ion-exchange resin $(20-50\text{\AA} \text{ mesh}, 1 \times 10 \text{ cm})$ was washed with water until the eluent was colorless. The column was then washed successively with 2 N aqueous NaOH (4 bed volumes), water (until the eluent was neutral to pH paper), 2 N HCl (4 bed volumes), and finally with water (until the eluent was neutral to pH paper). The above solid residue was dissolved in water (ca. 0.5 mL) and the solution was passed down the column, using water. The eluent was monitored by TLC [silica, 2:4:4 AcOH-hexane-EtOAc-water (1 drop water to 10 mL of the organic phase), visualization under uv light]. Evaporation of the combined uv-active fractions (water pump, rotary evaporator, bath temperature below 60 °C) gave ester (-)-32 (110 mg, 63%) as a pure (¹H NMR, 400 MHz), yellow foam: $[\alpha]_D^{21} = -132$ ° (c 1.073, H₂O); FTIR (microscope) 1654, 1717, 1740, 2800-3400 (br) cm⁻¹; ¹H NMR (D₂O, 360 MHz) δ 1.58-1.60 (m, 1 H), 1.80-1.88 (m, 1 H), 2.10-2.20 (m, 1 H), 2.25-2.35 (m, 1 H), 2.6-2.70 (m, 2 H), 2.7-2.8 (m, 3 H), 1.90-1.95 (m, 1 H), 3.75 (s, 3 H), 5.10-5.15 (dd, J = 6.8, 4.4 Hz, 1H), 7.35(s, 1 H); ¹³C NMR (D₂O, 100.6 MHz) δ 16.2 (t'), 23.8 (t'), 26.1 (t'), 26.5 (t'), 33.4 (t'), 54.3 (d'), 57.7 (q'), 126.9 (s'), 132.9 (s'), 133.7 (d'), 137.8 (s'), 161.9 (s'),

175.0 (s'), 178.6 (s'); exact mass m/z calcd for $C_{14}H_{17}NO_6$ 295.10559, found 295.10599.

(-)-1,3,4,6-Tetrahydro-9-hydroxy-4-(methoxycarbonyl)-6-oxo-2H-quinolizine-7-propanoic acid [(-)-32] from [(-)30b].



The above procedure was followed exactly, using exocyclic olefin (-)**30b** (derived from the faster-running alcohol) (58 mg, 0.20 mmol) in CH_2Cl_2 (5 mL), Ph_3P (107 mg, 0.41 mmol), and Et_3N (0.20 mL, 1.43 mmol) in THF (3 mL). Ester **32** (37 mg, 63%) was obtained as a pure (¹H NMR, 400 MHz), yellow foam, spectroscopically identical to material obtained in the previous experiment. (-)-4-Carboxy-1,3,4,6-tetrahydro-9-hydroxy-6-oxo-2Hquinolizine-7-propanoic acid (-)-A58365B.



(Bu₃Sn)₂O (0.5 mL, 0.98 mmol) was added to a solution of methyl ester 32 (42 mg, 0.14 mmol) in dry PhH (2 mL), and the mixture was refluxed for 3 days (Ar atmosphere) [TLC control, silica, 2:4:4 AcOH-hexane-EtOAc-water (1 drop water to 10 mL of the organic phase)]. The deep red solution was cooled to room temperature and diluted with hexane (10 mL). The resulting solution was extracted with 1% AcOH-water (4 \times 5 mL), and the combined aqueous extracts were washed with CHCl3 $(4 \times 10 \text{ mL})$. Evaporation of the aqueous extracts (rotary evaporator, water pump, below 60 °C) and flash chromatography of the residue over C-18 reverse phase silica gel (60 mesh, 10% C-18 capped with TMS, 1 x 20 cm), using 9:1 water-MeCN, gave the crude product. This was dissolved in the minimum amount of water at room temperature. The solution was stored overnight in a refrigerator to obtain pure $(^{1}H NMR, 400 MHz)$ (-)-A58365B (2) (27 mg, 70%) as a yellow crystalline solid: mp 209-213 °C; $[\alpha]_D^{21} = -114$ ° (c 0.16, H₂O); literature¹¹ value: $[\alpha]_D^{25} = -141^{\circ}$ (c 0.16, H₂O); FTIR (microscope) 1651,

1709, 2600-3200 (br) cm⁻¹; UV (MeOH) λ_{max} : 340, 238 nm; ¹H NMR (D₂O, 360 MHz) δ 1.65-1.70 (m, 1 H), 1.85-1.90 (m, 1 H), 2.15-2.25 (m, 1 H), 2.3-2.35 (m, 1 H), 2.65-2.85 (m, 5 H), 2.95-3.00 (m, 1 H), 5.10-5.15 (dd, J = 6.4, 3.6 Hz, 1 H), 7.35 (s, 1 H); ¹³C NMR (D₂O, 75.5 MHz) δ 16.1 (t'), 23.7 (t'), 26.1 (t'), 26.5 (t'), 33.4 (t'), 57.5 (d'), 126.8 (s'), 132.7 (s'), 133.5 (d'), 137.6 (s'), 161.8 (s'), 176.2 (s'), 178.6 (s'); exact mass *m/z* calcd for C₁₃H₁₅NO₆ 281.08984, found 281.08753.

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