

1 **Nutrient availability, the microbiome and intestinal**
2 **transport during pregnancy**

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30

31 **Abstract**

32 Adequate adaptation of the gastrointestinal (GI) tract is important during
33 pregnancy to ensure that the increased metabolic demands by the developing
34 fetus are met. These include changes in surface area mediated by villus
35 hypertrophy and enhanced functional capacity of individual nutrient receptors
36 including those transporting glucose, fructose, leucine, and calcium. These
37 processes are regulated either by the enhanced nutrient demand or are
38 facilitated by changes in the secretion of pregnancy hormones. Our review also
39 covers recent research into the microbiome, and how pregnancy could lead to
40 microbial adaptations, which are beneficial to the mother, yet are also similar to
41 those seen in the metabolic syndrome. The potential role of diet in modulating
42 the microbiome during pregnancy, as well as the potential for the intestinal
43 microbiota to induce pregnancy complications are examined.

44
45 Gaps in the current literature are highlighted including those where only
46 historical evidence is available, and we suggest areas that should be a priority for
47 further research. In summary, although a significant degree of adaptation has
48 been described, there are both well-established processes, and more recent
49 discoveries such as changes within the maternal microbiome that pose new
50 questions as to how the GI tract effectively adapts to pregnancy, especially in
51 conjunction with maternal obesity.

52

53 **Keywords:** gastrointestinal adaptation, nutrition, pregnancy, microbiome,
54 nutrient transport, maternal health.

55

56 **Introduction**

57 The perinatal period is associated with widespread adaptations in a majority of
58 maternal organ systems in order to ensure that nutrient supply to both the
59 mother and developing fetus can be maintained. These temporary changes,
60 collectively known as homeorhesis (Bauman and Currie 1980), are necessary to
61 optimise health during pregnancy, and to enable the mother to meet the
62 additional energetic and nutrient demands that accompany lactation. Many of
63 these maternal physiological adaptations have been previously reviewed. This
64 includes articles summarising the changes in energy metabolism (Herrera 2000,
65 Prentice and Goldberg 2000), and circulation (Hunter and Robson 1992), in both
66 women of normal (King 2000) and excess body mass index (BMI) (King 2006).
67 There is also a substantial body of literature covering adaptation of the placenta
68 to different nutrient intakes (Jones et al. 2007, Lager and Powell 2012), but there
69 is comparatively little work exploring whether the gut and bacterial inhabitants
70 of the gut are subject to similar changes.

71

72 The aim of this review is to discuss the variety of changes that have been
73 demonstrated to occur in the small and large intestine in pregnancy, ranging
74 from the anatomical to the molecular. It will also highlight the need for further
75 research, as much of the evidence discussed is now dated, and therefore has not
76 been considered in light of the significant increases in average pre-pregnancy
77 BMI of women of child-bearing age over the last two decades. Depending on the
78 cut-off used, the incidence of obesity in pregnant women in the USA ranged
79 between 18.5 and 38.3% (Galtier-Dereure et al. 2000, Catalano and Ehrenberg
80 2006, Guelinckx et al. 2008), and there has been a marked increase in the

81 prevalence of obesity in women of childbearing age of ~33% between 1988 and
82 2000 (Kim et al. 2007). Among Canadian women aged 18-79, the proportion with
83 a BMI classified as underweight, normal weight overweight or obese is 2.2%,
84 44.7%, 29.5% and 23.6% respectively, and the prevalence of obesity among
85 women of child-bearing age (18-44) ranges between 5.5 and 19.4%
86 (Government of Canada 2011).

87

88 **Changes in gut physiology during pregnancy and lactation**

89 Gross changes of the alimentary tract using wet weight as an indicator of overall
90 size have been assessed in pregnant and lactating rats in studies dating back to
91 the 1930s (Lew et al. 1939), with more detailed documentation focusing on the
92 intestine beginning in the 1960s. Relative to the non-pregnant state, increases in
93 weight of the stomach and the small intestine of rats have been recorded
94 consistently during lactation but not pregnancy (Souders and Morgan 1957, Fell
95 et al. 1963, Campbell and Fell 1964). One study in sheep reported a 45% increase
96 in small intestine weight ($p < 0.05$ vs age matched non-pregnant controls) in the
97 third trimester (Fell et al. 1972). Although intestinal weight does not appear to
98 change in pregnant rats, surface area increases throughout pregnancy. Villus
99 height increases in the duodenum by mid-pregnancy (Cripps and Williams 1975)
100 and this is accompanied by intestinal dilatation commencing at the beginning of
101 the final week of gestation; these observations have also been reported in
102 pregnant sheep (Fell et al. 1964). These changes persist through lactation (Fell et
103 al. 1963, Boyne et al. 1966) as evidenced by increased serosal circumference and
104 villus height in the jejunum and ileum at the end of lactation in rats, and lactating
105 mice (Campbell and Fell 1964). More recent research in rats shows villus heights

106 in the jejunum significantly increasing by gestational day 21, with no change in
107 the ileum or duodenum (Sarvestani et al. 2015). Intestinal length has been
108 reported to be unchanged during pregnancy in rats, but will increase by almost
109 25% during lactation (Craft 1970, Cripps and Williams 1975).

110

111 The mechanisms proposed to mediate these gross anatomical changes include
112 changing caloric intake, so called “work hypertrophy” (Fell et al. 1963). This has
113 been widely debated with two studies in non-pregnant rats showing significant
114 increases in stomach and colon weight with raised nutrient intake but no change
115 in the small intestine (Addis 1932, Dowling et al. 1967). Both an increase in
116 plasma thyroxine and insulin resistance may also play a role, both are known to
117 occur in pregnancy (Branch 1992), and both have been independently associated
118 with intestinal hypertrophy (Middleton 1971, Fujita et al. 1998). However this is
119 currently speculation as both effects have only been shown in non-pregnant
120 animal models.

121

122 When relating findings from rodent models to humans, substantial differences in
123 energy expenditure must be taken into account. The higher metabolic rate,
124 shorter gestation and larger litter sizes of rodents compared with human and
125 sheep pregnancies could contribute to more pronounced intestinal adaptations
126 than are observed in humans (Hammond 1997). Detailed information about
127 specific anatomical adaptations in pregnant is limited by the fact that collection
128 of GI tissue from healthy pregnant women throughout pregnancy is not ethically
129 feasible.

130

131 *Changes in gut motility and transit time*

132 An increased occurrence of heartburn, bloating and constipation during

133 pregnancy has been well documented in humans (Feeney 1982) suggesting that

134 intestinal motility and transit time may both be increased. This was initially

135 thought to be caused by the expanding uterus placing pressure on the GI tract

136 (Byrne 1972) but additional studies suggest hormonally driven changes may add

137 to mechanical influences that slow these processes. Exposure to high

138 concentrations of progesterone reduces GI motility, with in-vitro treatment of rat

139 GI sections with progesterone leading to reduced contractile activity in

140 oesophageal, antral and colonic tissue (Bruce and Behsudi 1979). Studies of the

141 effect of progesterone on motilin, a hormone which stimulates GI motility in the

142 stomach, showed significant inverse correlations between motilin and plasma

143 progesterone both during fasting and after a glucose load in humans

144 (Christofides et al. 1982, Holst et al. 1992). This suggests that progesterone has

145 a direct effect on GI tissue motility as well as an inhibitory effect on the action of

146 other hormones. Studies in humans using the lactulose hydrogen breath test to

147 measure oro-caecal transit report no significant changes in transit time in the

148 first trimester, despite women displaying dyspeptic symptoms such as heartburn

149 and bloating. Gut transit time increases during the third trimester (Chiloiro et al.

150 2001) when these symptoms can disappear.

151

152 **Changes in intestinal absorption and permeability**

153 The increased intestinal surface area and transit time observed in pregnancy

154 have the potential to affect fluid and electrolyte balance as well as nutrient

155 absorption. Absorption of sodium and water increases by almost 50%, between

156 12 to 20 weeks gestation in humans. Such changes could be mediated, in part, by
157 a raised plasma aldosterone concentration (Brown et al. 1992) but may also be
158 affected by hormonal adaptations to pregnancy. Changing concentrations of
159 oestrogen could influence sodium absorption by effects on the adrenal gland and
160 the renin-angiotensin system (RAS). Angiotensin II promotes fluid absorption in
161 the small intestine (Fändriks 2011), and plasma concentrations of several RAS
162 hormones including angiotensin II are elevated during pregnancy (Irani and Xia
163 2008). Enhanced absorption could reflect the raised transit time in late gestation
164 (Chiloiro et al. 2001), whereas chlorine (Cl) secretion in the colon may be
165 inhibited during pregnancy due to raised oestrogen (Condliffe et al. 2001). In
166 rats this response could be mediated by activating protein kinase-C delta
167 (O'Mahony et al. 2007), whilst blocking its action prevents the decrease in Cl
168 secretion on administration of 17 β -oestradiol (Doolan et al. 2000). This is
169 consistent with the water retention observed towards the end of pregnancy,
170 when oestrogen levels are at their highest (Atherton et al. 1982, Schrier et al.
171 2001) .

172

173 Gene expression analyses using microarray and qPCR has shown that several ion
174 transporters associated with sodium, calcium and magnesium are all
175 upregulated in the rat duodenum during pregnancy and lactation
176 (Teerapornpuntakit et al. 2014). This has been suggested to be in part due to the
177 hypertrophy observed in the small intestine, however, as discussed above the
178 absorption of specific nutrients are affected by stage of pregnancy rather than
179 reflecting an increase in absorptive surface.

180

181

182 **Absorption of Major Dietary Constituents**183 *Carbohydrates*

184 Carbohydrates are the main substrate of fetal and placental metabolism, and
185 adequate intake, absorption and distribution are required for healthy pregnancy
186 outcome (Battaglia 1989, Hay 1991). Pregnancy is an insulin resistant state, with
187 both hyperinsulinaemia and insulin resistance peaking in the third trimester
188 (Cousins et al. 1980, Buchanan et al. 1990, Catalano et al. 1993). Fructose intake
189 in pregnancy is also becoming increasingly relevant given its increased role in
190 the Western diet since the 1980s, and its potential for inducing lipogenesis much
191 more readily than glucose (Regnault et al. 2013).

192

193 The relationship between pregnancy and glucose homeostasis is therefore of
194 clinical relevance, particularly with regard to how carbohydrates are absorbed
195 across the gut. An inhibitory effect of oestrogen on glucose absorption in the
196 small intestine has been demonstrated. Glucose uptake from the small intestine
197 in rats is increased by a third following ovariectomy and is reduced back to the
198 same level as sham-operated controls with 17 β -estradiol and progesterone
199 replacement (Singh et al. 1985). This is at odds with more recent research
200 showing an upregulation in both glucose transporter 5 (GLUT5) and sodium-
201 linked glucose transporter 1 (SGLT1) expression in late pregnancy compared to
202 age-matched non-pregnant controls (Teerapornpuntakit et al. 2014). Thus, we
203 speculate that upregulation in transport protein expression may offset
204 reductions in glucose absorption as a result of oestrogen-related changes. In the
205 case of fructose, to our knowledge no research has been conducted focusing on

206 the transport of fructose during pregnancy. Rat studies have demonstrated that
207 exposure to luminal fructose in the fetus during critical stages of development
208 will program the offspring intestine by increasing expression of the fructose
209 transporter GLUT5 (Jiang and Ferraris 2001, Suzuki et al. 2011), suggesting that
210 fructose intake and intestinal uptake during pregnancy is of interest.

211

212 Future pregnancy based studies could utilise Ussing chambers in conjunction
213 with inhibiting SGLT1 or other carbohydrate transporters, techniques that have
214 already been employed when studying transport of other nutrients across the
215 small intestine (Wolffram et al. 2002). Focus on GLUT2 would also be warranted
216 as this has been shown to play an equally important role as SGLT1 in intestinal
217 glucose transport (Kellett and Brot-Laroche 2005, Kellett et al. 2008)

218

219 *Amino acids*

220 Protein intake, absorption and distribution is also vital during pregnancy, with
221 the fetus, uterus, placenta, and amniotic fluid together accounting for ~925g of
222 protein accreted over pregnancy in humans. This is to be obtained from a
223 recommended dietary intake of 1.1g per kg body weight per day (an average
224 increase of 25g/day compared to the non-pregnant state), and a macronutrient
225 distribution whereby protein contributes between 10 and 35% of total calories
226 (United States National Research Council 2005). The effect of protein restriction
227 during pregnancy on the offspring has been well defined in animal models using
228 <9% of calories from protein (Lakshmy 2013), leading to growth restriction in
229 the offspring and an increase in incidence of the metabolic syndrome in later life.
230 The precise role of a low protein intake in mediating such responses is difficult

231 to determine as all low protein diets contain additional carbohydrate in order to
232 ensure they are isocaloric compared to the control diet (Symonds et al. 2006). In
233 addition protein requirements are much higher in rodent pregnancies compared
234 to humans (Symonds et al. 2006).

235

236 Protein turnover increases linearly throughout pregnancy, shown indirectly
237 from measuring the fate of radio-labelled leucine (Thompson and Halliday 1992)
238 and glycine (de Benoist et al. 1985) in women. A lack of studies covering
239 potential mechanisms for the increase in amino acid absorption has been
240 highlighted in earlier reviews (Karasov and Diamond 1983), and this does not
241 appear to have changed. Therefore research into expression of the variety of
242 different transport proteins present in the small intestine (Bröer 2008) and how
243 expression changes throughout pregnancy is of interest.

244 *Fatty acids*

245 Maternal dietary fat intake is important throughout pregnancy not only as a
246 maternal energy source but also to provide the developing fetus with essential
247 fatty acids for optimal development (Budge and Symonds 2006). It has been
248 suggested that total fat requirements as a proportion of energy intake during
249 pregnancy should not be increased compared to non-pregnant women, as they
250 can be met by small increases in consumption of a balanced diet (United States
251 National Research Council 2005). There is debate as to whether the pre-formed
252 long-chain polyunsaturated fatty acids (LCPUFA) should be considered as
253 conditionally essential in pregnancy, and whether the normal dietary supply of
254 essential fatty acids, specifically linoleic and α -linolenic acid are sufficient for
255 optimal fetal development and long term health (Haggarty 2014). Current

256 literature has examined the lack of maternal dietary docosahexaenoic acid
257 (DHA), particularly during late pregnancy (Haggarty 2004), and the mobilisation
258 of maternal adipose tissue, but not the intestinal transport of fatty acids. The US
259 Dietary Reference Intakes (United States National Research Council 2005)
260 highlight a lack of evidence available to determine optimal intakes of n-3 and n-6
261 fatty acids during pregnancy, in part because of difficulties defining a range of
262 intakes in Western populations that would lead to a deficiency. This potentially
263 explains the lack of research interest shown in the uptake of fatty acids during
264 pregnancy.

265

266 To summarise, although it is well established that glucose and fructose transport
267 across the intestine is a dynamic process modulated by the expression of
268 transport proteins, very little work has been done to ascertain how pregnancy
269 affects the expression and function of these transporters. Transport of both
270 amino and fatty acids across the intestinal epithelium during pregnancy is even
271 less well studied, despite the fact that in the case of amino acids, increased
272 absorption in pregnancy has been demonstrated.

273

274 *Absorption of Micronutrients*

275 Micronutrients can have an important role in preventing adverse events such as
276 premature birth and low birth weight (Ramakrishnan et al. 1999, United States
277 National Research Council 2001, 2011). The intestinal absorption of calcium,
278 vitamin D and iron absorption in pregnancy are well described but many other
279 micronutrients remain unstudied. This includes magnesium and zinc, which
280 when deficient are associated with adverse outcomes such as pre-eclampsia and

281 preterm delivery (Black 2001). There is promising preliminary research into the
282 transport of some of these micronutrients, with microarrays showing
283 upregulation in duodenal transporters of both zinc and calcium during
284 pregnancy in rats; but no associated mechanisms have been identified
285 (Teerapornpuntakit et al. 2014).

286

287 *Calcium and vitamin D absorption*

288 The human fetus requires the transfer of ~30 g of calcium from maternal stores
289 between conception and birth. Both calcium uptake, excretion (Kent et al. 1991,
290 Ritchie et al. 1998) and regulation of vitamin D metabolism adapt to pregnancy
291 (Phillips et al. 2000, Prentice 2000). In rats, intestinal calcium transport
292 increases throughout pregnancy to peak at day 14 of lactation coincident with
293 changes in plasma 1,25 dihydroxyvitamin D, a hormonally active metabolite of
294 vitamin D. However an increase in absorption was also observed in a cohort of
295 vitamin D deficient rats following the same study design, suggesting the
296 adaptations in calcium uptake may be independent of vitamin D status (Halloran
297 and DeLuca 1980). Further evidence for this is provided from pregnant mice
298 lacking the vitamin D receptor (VDR) which display osteomalacia at baseline
299 (before mating), and an increase in bone mineral content throughout pregnancy,
300 that is comparable to pregnant wild-type controls. (Fudge and Kovacs 2010).
301 Taken together these findings suggest other mechanisms of calcium transport
302 may be active during pregnancy.

303

304 *Iron absorption*

305 Maternal iron requirements in humans increase during pregnancy in order to
306 meet the needs of the raised erythrocyte mass, formation of the placenta, and
307 ~1g of stored iron accumulated by the fetus at term. These are met through a
308 combination of the maternal mobilisation of stored iron, reduced iron loss, and
309 potentially increased absorption from the maternal diet. Iron deficiency in
310 pregnancy, however, remains a problem, leading to an increased risk of
311 preeclampsia, intrauterine growth restriction and low birth weight (Cetin et al.
312 2011).

313

314 Upregulation of iron absorption has been shown during the second and third
315 trimester of pregnancy irrespective of iron status using labelled ^{54}Fe in a small
316 study of 12 pregnant women (Barrett et al. 1994). Animal studies have proposed
317 potential mechanisms for these changes. For example, duodenal gene expression
318 of Dcytb, an enzyme responsible for converting dietary Fe^{3+} to Fe^{2+} for
319 metabolism, and divalent metal transporter 1 (DMT1), a major intestinal iron
320 transporter, have been shown to increase through pregnancy and decline within
321 48 hours after birth (Millard et al. 2004). The positive association between the
322 timing of changes in oestrogen concentrations and Dcytb and DMT1 protein
323 expression suggests that oestrogen may regulate iron absorption, but firm
324 evidence for this remains to be established.

325

326 **Microbial changes**

327 Over the last decade the microbiome has been shown to be an important
328 mediator of human health, and the composition of different bacterial species
329 making up the microbiome and potential mechanisms by which they can be

330 altered is now a well-established field of research (Cho and Blaser 2012). There
331 is growing appreciation that diet (Tilg and Kaser 2011, David et al. 2014, Walter
332 2015) and pregnancy (Koren et al. 2012, Jost et al. 2014) modulate the
333 microbiome, as well as speculation that the microbiome can influence offspring
334 development (Ma et al. 2014, Aagaard et al. 2014).

335

336 The influence of the microbial environment of the intestine on host physiology is
337 now well established (Hooper and Gordon 2001), and has led to a number of
338 studies examining how diet, and more recently pregnancy, affect it. The maternal
339 GI microbiome undergoes profound changes during pregnancy, some of which
340 are not dissimilar to those characteristics found in obesity (Koren et al. 2012),
341 i.e. a decrease in microbial diversity (Turnbaugh et al. 2009, Qin et al. 2010,
342 Greenblum et al. 2012). An “obese” type microbiome has been defined as one
343 with an increased capacity for energy utilisation, primarily due to the greater
344 abundance of bacterial species capable of fermenting and therefore increasing
345 availability of otherwise indigestible sugars. The most significant example of this
346 is the increased abundance of *Firmicutes* species relative to *Bacteroidetes*, with
347 *Firmicutes* being more efficient sugar fermenters. Analysis of the colonic
348 contents of ob/ob mice also shows an elevated concentration of the fermentation
349 products butyrate and acetate relative to lean controls (Turnbaugh et al. 2006).
350 A shift in *Bacteroidetes* and *Firmicutes* was not observed in the only maternal
351 microbiome sequencing study to date conducted in normal weight participants
352 (Koren et al. 2012), however a greater representation of the lactic acid
353 fermenters *Lactobacillus*, *Streptococcus* and *Enterococcus* was reported in the
354 third trimester. This has been suggested to be an evolutionary adaptation, and

355 the concomitant transfer of these microbes to the newborn could enable it to
356 take advantage of the lactose in its mother's milk (Cox and Blaser 2013).

357

358 Similarities between the pregnant and obese microbiome are of interest since
359 both conditions are associated with increased fat mass, although the relative
360 distribution of this additional fat is different between pregnant and obese
361 individuals (Straughen et al. 2013). There are a number of potential explanations
362 for the development of an "obese type" microbiota during pregnancy. Species of
363 *Bacteroidetes* contain a number of glycoside hydrolases which are able to
364 ferment sugars which would otherwise pass through the large intestine and
365 remain undigested (Bäckhed et al. 2005). Increased availability of
366 polysaccharides is regarded as a hallmark of an obese microbiota (Turnbaugh et
367 al. 2006), but would also result in increased energy availability which may be
368 beneficial in pregnancy. This could help in meeting maternal energy demands
369 and supplying the additional demands of the fetus, and suggest that during
370 pregnancy the microbiome could be modulated to increase energy available for
371 absorption. However, it is currently unclear whether these adaptations are due
372 to the changing microbial environment, or are modulated by the other intestinal
373 adaptations outlined earlier in this review.

374

375 It is not only changes in the relative proportion of individual species that occur,
376 as the diversity of the microbiome also adapts, as defined by the relative number
377 and abundance of different types of organism. For example, both obesity and
378 inflammatory bowel disease have been linked with a low diversity of gut
379 microbes (Turnbaugh et al. 2009, Qin et al. 2010). Diversity is measured within

380 an individual (alpha diversity, for example, the number of different types
381 organisms within an individual's intestinal microbiome) and between
382 individuals (beta diversity). Patterns of alpha and beta diversity differ
383 significantly when making comparisons from the same habitat. For example, a
384 high alpha diversity indicates a diverse microbiome but this can be concomitant
385 with a low beta diversity, with members of a defined population all sharing
386 similar organisms. Currently a functional explanation for changes in diversity
387 has not been identified, with a need for studies taking into account responses to
388 short and long term dietary modifications, diurnal rhythms as well as mode of
389 delivery at birth and host genetics (The Human Microbiome Project Consortium
390 2012).

391

392 The current evidence base for understanding the changing gut microbiome has
393 compared the effect of different diets and physiological states including
394 pregnancy. Although this field is very new, it appears that the gut microbiome
395 does change with pregnancy (Koren et al. 2012), and it has been hypothesised
396 that inappropriate adaptation of the gut microbiome, such as that seen in a
397 number of inflammatory bowel disorders (Kamada et al. 2013) may contribute
398 to the development of pregnancy complications including pre-eclampsia,
399 intrauterine growth restriction and miscarriage (Zhang et al. 2015). Pregnancy is
400 associated with a significant reduction in alpha diversity and an increase in beta
401 diversity by the third trimester (Koren et al. 2012). At the species level, there is a
402 significant increase in abundance of *Proteobacteria* in the third trimester, which
403 has previously been observed in obesity (Turnbaugh et al. 2009) and chronic
404 inflammatory states (Mukhopadhyaya et al. 2012).

405
406 Changes in intestinal permeability to larger molecules and bacteria are of
407 interest especially given the above changes in the gut microbiota during
408 pregnancy, and the recent description of a unique microbiome in the placenta
409 (Aagaard et al. 2014). The effect of pregnancy on epithelial tight junctions, which
410 mediate intestinal permeability (Turner 2009), has not been studied. There is
411 overlap here with research into the role of the gut microbiota in pregnancy, with
412 lipopolysaccharide (LPS) present on the membranes of Gram-negative bacteria
413 passing into the systemic circulation and leading to the low-level adipose tissue
414 inflammation characteristic of obesity (Lam et al. 2011). Catecholamines
415 released as part of the maternal stress response have been hypothesised to lead
416 to gut barrier failure (Friebe and Arck 2008), and the subsequent release of LPS
417 linked to an increased risk of spontaneous abortion (Friebe et al. 2011).

418
419 Small studies in non-human primates (n=2-4 animals/group) have
420 demonstrated that a high fat diet in pregnancy has potentially adverse effects on
421 the offspring microbiome (Ma et al. 2014). An upregulation in amino acid,
422 carbohydrate and lipid metabolic pathways was observed in 1-year-old offspring
423 exposed to a high-fat diet during gestation and lactation. These changes were
424 partly reversed in offspring switched from a high-fat diet containing 36% fat to a
425 control diet containing 13% fat at weaning suggesting modest flexibility of the
426 offspring microbiome. The authors suggest that these results show the influence
427 of the maternal diet on establishment of the microbiome, rather than an
428 “obesity-causing” microbiome as indicated in the above research. However the
429 small numbers of animals studied together with the lack of an adverse

430 phenotype in the offspring (McCurdy et al. 2009) suggest that these findings
431 need following-up.

432

433 **Conclusion**

434 Maternal adaptations occur throughout pregnancy and lactation, ranging from
435 changes in gross gut anatomy to changes in the expression of specific nutrient
436 transporters as summarised in Figure 1. The evidence here demonstrates that
437 adaptation of the intestine to pregnancy is equally important as adaptation of
438 other organs. However much of the material presented here is based on dated
439 research, and there is a clear need for an expanded body of evidence using
440 contemporary techniques in animal models and humans where possible. This
441 would enable the addition of mechanistic data especially in pregnancies at risk
442 from complications. The increasing prevalence of obesity in the Western world
443 makes the study of obesogenic or diabetogenic diets during pregnancy critical to
444 furthering understanding of important adaptations and processes in the gut. The
445 microbiota is now considered a metabolically active organ, and the evidence
446 presented suggests that its adaptation in pregnancy is a part of increasing energy
447 extracted from the diet to provide for the fetus. Understanding these
448 adaptations, and how they may be modulated by diet could therefore be
449 beneficial in treating overweight and obesity during pregnancy.

450

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452

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457

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811 **Figure caption**

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813 *Figure 1: Summary of the primary known gastrointestinal adaptations and*
814 *nutrient transporters affected during pregnancy. All processes shown are*
815 *upregulated, those with blue arrows have mechanisms suggested in the literature,*
816 *those without have been demonstrated but currently lack a well-defined*
817 *mechanism.*

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