### **REVIEW ARTICLE**

### Defining the concept of 'tick repellency' in veterinary medicine

L. HALOS<sup>1</sup>\*, G. BANETH<sup>2</sup>, F. BEUGNET<sup>1</sup>, A. S. BOWMAN<sup>3</sup>, B. CHOMEL<sup>4</sup>, R. FARKAS<sup>5</sup>, M. FRANC<sup>6</sup>, J. GUILLOT<sup>7</sup>, H. INOKUMA<sup>8</sup>, R. KAUFMAN<sup>9</sup>, F. JONGEJAN<sup>10</sup>, A. JOACHIM<sup>11</sup>, D. OTRANTO<sup>12</sup>, K. PFISTER<sup>13</sup>, M. POLLMEIER<sup>1</sup>, A. SAINZ<sup>14</sup> and R. WALL<sup>15</sup>

<sup>1</sup> Merial, 29 Av. Tony Garnier 69007 Lyon, France

<sup>2</sup> Gad. School of Veterinary Medicine, Hebrew University, Israel

<sup>3</sup> School of Biological Sciences (Zoology), University of Aberdeen, Tillydrone Avenue, Aberdeen AB24 2TZ, UK

<sup>4</sup> Department of Population Health and Reproduction, School of Veterinary Medicine, University of California, Davis, CA 95616, USA

<sup>5</sup> Department of Parasitology and Zoology, Faculty of Veterinary Science, Szent István University, Budapest, Hungary

<sup>6</sup> Laboratoire de Parasitologie, Ecole Nationale Vétérinaire de Toulouse, BP 87 614, 31 076 Toulouse Cedex 03, France

<sup>7</sup> Unité de Parasitologie, UMR Anses, Enva, Upec BIPAR, Ecole Nationale Vétérinaire d'Alfort, 94704 Maisons-Alfort, France

<sup>8</sup>Department of Clinical Veterinary Science, Obihiro University of Agriculture & Veterinary Medicine, Inada, Obihiro 080-8555, Japan

Department of Biological Sciences, University of Alberta, Edmonton, Canada

<sup>10</sup> Utrecht Centre for Tick-Borne Diseases (UCTD), Faculty of Veterinary Medicine, Utrecht University, Yalelaan 1, 3584 CL Utrecht, The Netherlands <sup>11</sup>Institute for Parasitology, Department of Pathobiology, University of Veterinary Medicine Vienna, Veterinaerplatz 1,

A-1210 Vienna, Austria<sup>12</sup> Dipartimento di Sanità Pubblica e Zootecnia, Faculty of Veterinary Medicine, University of Bari, Valenzano, Italy

<sup>13</sup> Institut für Parasitologie Lehrstuhl für vergleichende Tropenmedizin und Parasitologie, Leopoldstrasse 5, 80802 München, Germany

<sup>14</sup>Department of Animal Medicine and Surgery, Faculty of Veterinary Medicine, Complutense University of Madrid, Madrid, Spain

<sup>15</sup> School of Biological Sciences, University of Bristol, Bristol, BS8 1UG, UK

(Received 9 September 2011; revised 8 November 2011; accepted 10 November 2011; first published online 5 January 2012)

#### SUMMARY

Although widely used, the term repellency needs to be employed with care when applied to ticks and other periodic or permanent ectoparasites. Repellency has classically been used to describe the effects of a substance that causes a flying arthropod to make oriented movements away from its source. However, for crawling arthropods such as ticks, the term commonly subsumes a range of effects that include arthropod irritation and consequent avoiding or leaving the host, failing to attach, to bite, or to feed. The objective of the present article is to highlight the need for clarity, to propose consensus descriptions and methods for the evaluation of various effects on ticks caused by chemical substances.

Key words: tick, control, companion animals, acaricides, repellency, expellency, disruption of attachment, study design.

#### TICK REPELLENCY: AN ILL-DEFINED CONCEPT

Ectoparasitic arthropods (insects and acarines) represent a group of organisms of major importance in veterinary medicine. Ticks and fleas are the main ectoparasites of pets, followed by sand flies and mosquitoes. Ectoparasites may impact animal health directly or indirectly through bites or their ability to transmit pathogens and associated diseases, some of which are zoonotic (Otranto et al. 2009).

\* Corresponding author: Merial, 29 Av. Tony Garnier 69007 Lyon, France. Tel: +33 (0) 4 72 72 34 42. E-mail: lenaig.halos@merial.com

Several potential approaches are available for controlling ectoparasites, including the use of chemical insecticides and acaricides, vaccination or biological control (Otranto and Wall, 2008). However, the use of different formulations of chemical insecticides and acaricides remains the most widespread approach. Depending on the nature of the compound used, the following effects may be obtained, alone or in combination: (i) disruption of contact between the arthropod parasite and the host; (ii) prevention of feeding; (iii) death of the arthropod parasite, and (iv) interference with egg fertility and subsequent development of off-host life-cycle stages. Standards

Parasitology (2012), 139, 419-423. © Cambridge University Press 2012. The online version of this article is published within an Open Access environment subject to the conditions of the Creative Commons Attribution-NonCommercial-ShareAlike licence <a href="http://creative">http://creative</a> commons. org/licenses/by-nc-sa/2.5/>. The written permission of Cambridge University Press must be obtained for commercial re-use. doi:10.1017/S0031182011002228

Table 1. Definition proposal for a glossary for tickcontrol terminology

Term	Definition	
Tick repellency sensu stricto	Characterized by an irritant effect, causing the tick to move away from the treated animal leading it to fall off soon after contact with its hair coat	
Disruption of attachment	Interference with the natural process of tick fixation including inhibition of attachment, withdrawal of mouthparts by ticks in the attaching phase, or detachment of already attached ticks	
Tick expellency	Causing ticks to fall off the host animal by disrupting the mechanisms of attachment: i.e. either by causing detachment of already attached ticks or by preventing attachment of new infesting ticks	
Anti-feeding effect	Interference with the natural process of tick feeding, avoiding any bloodmeal	
Killing effect (lethal action)	Its ability to induce death of the arthropod and its acaricidal effect <i>sensu stricto</i>	

for the evaluation of the efficacy of drugs have been determined by pharmaceutical regulation worldwide (for example by the European Medicine Agency (EMA) or by the Environmental Protection Agency in the United States) as well as by scientific societies.

Usually, the range of effects described above is evaluated for a specific interval of time, along with speed of kill and residual activity (Marchiondo et al. 2007; EMA, 2007). An anti-tick product for dog, for example, should give >90% efficacy, characterized by a reduction in tick counts 48 h after treatment and the prevention of re-infestation (EMA, 2007). Nevertheless, the reduction in tick counts can be caused by a range of behaviours, not only simple mortality. These non-lethal effects can broadly be described as repellency, but definitions for the various behavioural components of which it is composed are not well established, thus resulting in the absence of specific standards for its evaluation (Bissinger and Roe, 2010). The present article aims to explore the definitions of various effects caused by chemical substances against ticks and to propose consensus descriptions, as well as to suggest a standardization of methods for evaluation of tick repellency in veterinary medicine, which could be eventually added to the EMA guidelines.

Arthropod repellents are defined as "chemical substances that cause arthropods to make oriented movements away from their source" (Dethier *et al.* 1960). While this definition of repellency is easily applied to mosquitoes and other flying insects such as sand flies, this is not the case for crawling arthropods such as fleas and ticks. Flying insects are usually highly mobile, transitory ectoparasites with only rapid, short-lasting contact with their host. Repellency in

this case is characterized simply as the effect of the chemical causing the arthropod to avoid the treated host or substrate. For crawling arthropods and especially for those with a long-lasting host-parasite association, such as fleas, ticks lice or mites, where contact with the host may last several days or even be life long, the term repellency becomes more difficult to define. In addition, the effect of a substance on a pre-existing infestation of a long-lasting parasitic infestation is not taken into account by the conventional definition. Hence, there is a need to better understand the definition of repellency. It should be specifically defined for each type of ectoparasite and, if not clearly applicable, other terminology should be proposed. Here we are particularly concerned with proposing a working definition of repellency for ticks.

## THE VARIOUS EFFECTS OF CHEMICAL PRODUCTS ON TICKS

Ticks have a specific and complex mode of parasitism. They wait for their host in the environment and are able to detect through their sensorial organs several stimuli emitted by the vertebrate host (Sonenshine *et al.* 2002). Once on their host, ticks crawl for several hours to find a suitable attachment site. The attachment process involves skin penetration, causing a feeding lesion and secretion of cement. Ticks then begin their bloodmeal within 24 h (Sonenshine *et al.* 2002). Tick-borne pathogens may be transmitted during this attachment and bloodmeal intake phase and any feature of ticks, from a molecular to a behavioural one, may affect their role as vector (Randolph, 2009).

Due to the complex host location, feedingbehaviour and vector role of ticks, the biological activity of a chemical cannot be adequately described by a single efficacy value (Marchiondo *et al.* 2007). It seems useful, therefore, to describe the behavioural effects as they disrupt the various steps leading to tick infestation (Dethier *et al.* 1960); including tick mobility (climbing), tick attachment and blood feeding. A distinction must also be made between the effect expected on pre-existing and new infestations.

The first potential influence is an irritant effect (also called 'hot foot' effect), which causes the tick to move away from the treated animal, falling off soon after the contact with its coat. This effect could therefore be considered as repellency *sensu stricto*. The effect is short-lived and variable depending on tick stage and species (Bissinger and Roe, 2010). Furthermore, it is not possible to evaluate the irritant action on a pre-existing infestation, because in such a case, the ticks are already firmly attached. The second effect is that of anti-attachment or disruption of attachment (Table 1). A contact with the substance causes disruption of attachment, the withdrawal of

mouthparts by ticks in the attaching phase, or even detachment of already attached ticks; Dryden *et al.* 2006*b*; Ian and Bryan, 1981). Notably, it is difficult to clearly distinguish the results of an irritant effect from those of a disruption of attachment effect on the behaviour of a tick. Other effects of a chemical compound include the prevention of ingestion of the bloodmeal, or an anti-feeding effect, whose evaluation remains difficult, and finally a true acaricidal toxic effect which results in tick mortality. The ability of a product to prevent the transmission of tick-borne diseases should also be considered.

The use of the term repellency in relation to tick behaviour, therefore, subsumes several effects. The use of the term 'expellency' may be more appropriate to define the combined effect of disruption of attachment and falling off the host. This term has been used previously in the pest control literature to define the effect of a product, which results in the 'expulsion' of crawling arthropods from a particular location (wood, host, hole, etc. depending on the targeted species) (Ian and Bryan, 1981; Wege *et al.* 2002).

Among the compounds which affect ticks, some substances have only an irritant effect, such as icaridin or DEET (Bissinger and Roe, 2010; Brown and Hebert, 1997). Other products may have both irritant and toxic effects, such as permethrin (Brown and Hebert, 1997). Substances such as amitraz present an effect which includes both an expellent effect, along with acaricidal properties (Folz et al. 1986). Finally, some substances, such as fipronil, have little impact on the tick's behaviour but have a strong killing potential (Narahashi et al. 2010). According to the molecule or family of molecules, efficacy against tick infestation is usually a combination of 3 types of effects: repellency sensu stricto, expellency and toxicity. Unfortunately, a standardized methodology to separately assess each type of effect is still lacking.

# TOWARDS ASSESSING TICK REPELLENCY AND RELATED EFFECTS IN COMPANION ANIMALS

*In vitro* tests (i.e. conducted in the absence of the host) to assess repellency *sensu stricto* are relatively easy to standardize. For example, in simple comparison tests in Petri dishes, repellency is determined by the number of ticks found on treated versus untreated surfaces (Bissinger and Roe, 2010). Climbing bioassays can also be used, where vertical supports are treated at some level above the base of the vertical climb with a chemical substance. Ticks that cross the treated region are considered not repelled whereas those that back down or fall from the treated surface are repelled (Bissinger and Roe, 2010).

The design of *in vivo* tests addressing separately '*sensu stricto*' repellency, expellency and killing effect is much more difficult. According to the EMA

guidelines, a repellent effect against ticks means that no ticks will attach to the animal and ticks already on the animal (attached or not) will leave the host soon after treatment. In general, no ticks should be detectable on the animal 24 h following administration of the product. However, this definition does not distinguish between ticks that are strictly repelled, or ticks that came into contact with the host and then left the host, or ticks that initially attached and then detached during the first 24 h. Distinctions between those features would particularly matter in the evaluation of the prevention for tick-borne disease transmission.

Furthermore, to date, tests used to demonstrate repellent effects lack standardized protocols. For example, the protocol used by Endris et al. (2000, 2002) exposed dogs for 2 h in individual cages to 50 unfed adult ticks (ratio male/female 1/1) at 7, 14, 21, 28 and 35 days after treatment with permethrin and other acaricidal products. After the 2-h exposure period, the numbers of live and dead ticks were counted on each dog by combing the fur and by collecting ticks from the cage. In this test, repelled ticks include those present in the cage (live and dead) and the unattached ticks on the dog 2 h after exposure. In another study (Dryden et al. 2006b), 25 adult ticks were placed on dogs for 10 min on days 3, 7, 14, 21 and 28 days post-treatment with permethrin or fipronil. During such a short time, not all ticks moved immediately into the hair coat, either on treated or untreated dogs; some dropped or crawled off the dogs. These ticks were picked up and placed back on the dogs during the 10-min infestation period. Ticks off the dog after the infestation period were considered as repelled. In a third approach (Folz et al. 1986), 50 unfed adult ticks were placed on untreated control dogs or dogs treated with amitraz on days 0, 7, 14 and 21 post-treatment. The numbers of dead and live ticks still present on the dogs 1 h after infestation were counted. In the same study, the effect of treatment on pre-existing infestation was also evaluated by counting ticks attached on dogs at 24 and 48 h after treatment.

The definition of repellency was clearly different for each of these studies. Nevertheless, a few constant features can be extracted. To evaluate repellency *sensu lato*, or expellency, it is necessary to measure the absence of ticks on a treated animal at a given time post-infestation, compared to a non-treated control. The infestation also needs to be performed in an environment, such as a crate, where ticks falling off the host can be collected at several time-points. The selection of the time-points is of major importance and should be defined according to the tick species and life stage.

The following experimental design (Table 2, Fig. 1), based on the period of time chosen for the tick count, is suggested as an example for the evaluation of tick repellency and related effects.

(The effect of any acaricidal product is evaluated according to the tick status and the time frame post-infestation, either in the crate or on dogs. Dogs are treated 24 h before the first infestation. In any case, the presence of a control group is required to be sure that the infestation process is normal.)

	Effect measured	Early expellency and repellency sensu stricto	Disruption of attachment	Acaricidal 'killing' effect
	Time frame post- infestation Tick status	0–4 hours	4–24 hours	0–48 hours
On the dog In the crate	Alive in the crate	Repelled/ Expelled	Not relevant	Ticks can be collected and placed in an insectarium. A survival evaluation can be then performed on the collected ticks after 24/48 h
	Dead in the crate	Repelled/ Expelled	Not relevant	Not relevant
	Alive free on the animal	Not Repelled/ Expelled	Anti-attachment considered	Acaricidal effect not demonstrated
	Dead free on the animal	Not Repelled/ Expelled	Anti-attachment considered	Acaricidal effect demonstrated
	Dead attached on the dog	Not relevant	Anti-attachment not considered	Acaricidal effect demonstrated
	Alive attached on the dog	Not relevant	Anti-attachment not considered	Acaricidal effect not demonstrated



Fig. 1. Flow diagram of the proposed *in vivo* test for the evaluation of tick repellency and associated effects on new infestation of an acaricidal product in dogs. The diagram gives time-line, location in crate and/or on dogs and the effect being measured. \*Tick count at 2 h is not mandatory as combing may disturb the process of infestation.

In addition to the classical 24 and 48 h counts recommended by the guidelines (EMA, 2007) earlier time-points are considered.

#### From 0 to 4 h post-infestation

This first period should be dedicated to the evaluation of repellency *sensu stricto* plus expellency. Ticks will not have enough time to attach either on treated or untreated dogs and an evaluation every hour post-infestation by counting the ticks in the crate and on dogs is suggested. A total count of ticks off-host will indicate the repellency/expellency efficacy of the compound.

#### From 4 to 24 h post-infestation

This second period should be dedicated to the evaluation of the anti-attachment effect. After 4 h, tick counts in the environment can no longer be conducted because of welfare and technical reasons. Dogs cannot be maintained in small cages for such a long time, the wastes from the dog prevent reliable tick counts and ticks that are not on the dog may also be ingested. Thus, the count of attached or unattached ticks has to be performed on the dog, and compared to untreated control dogs.

#### Between 0 and 48 h post-infestation

The lethal effect of the product can be evaluated, either on the expelled ticks collected in crates, for which survival is evaluated in an insectarium, or by counting ticks on the host, as described in classical acaricidal efficacy studies (EMA, 2007).

In addition to the evaluation of preventive efficacy described above, the measure of the effect on a preexisting infestation is also recommended to assess the curative efficacy profile of a chemical product. In that case, infestation of dogs is performed between 2 and 1 day before treatment to let the ticks attach to their host. Tick counts are performed on dogs from 0 to 48 h post-treatment as previously described. This kind of study allows measurement of expellency, through its effect on detachment, as well as acaricidal properties of the product.

During experimental studies, evaluation of tick expellency at several time-points should be performed for an accurate evaluation of tick control products. In addition, field trials can also be conducted by comparing the number of questing ticks collected on treated and untreated dogs after walking in natural tick habitats (Dryden *et al.* 2006*a*). Tick counts additional to the 24 h or 48 h counts should be performed. The same time-points as described for the laboratory studies can be chosen as they take into account the several hours during which ticks may crawl about on their host in search of a suitable feeding site.

#### CONCLUDING REMARKS

The impact of ticks on animal health is linked to irritation at the attachment site, to blood loss and to the risk of tick-borne pathogen transmission. Hence, the ultimate objective of tick control is to prevent both attachment and blood feeding. Precise definition of the effects of chemical compounds on these behaviours is therefore essential and, even if widely used, the term repellency does not seem to be fully appropriate for ticks without careful qualification. Distinction should be clearly made between several potential behavioural effects. Similar comments also apply to other long-term and permanent ectoparasites. The concept of expellency includes both the rate at which ectoparasites fall off the host and disruption of attachment and seems therefore more appropriate when used to assess the impact on tick control.

#### ACKNOWLEDGEMENTS

This paper is based on discussions carried out during a Tick Control Workshop organized by Merial in Seville, Spain, in May 2011.

#### REFERENCES

Bissinger, B. W. and Roe, R. M. (2010). Tick repellents: past, present, and future. *Pesticide Biochemistry and Physiology* **96**, 63–79.

Brown, M. and Hebert, A.A. (1997). Insect repellents: an overview. *Journal of the American Academy of Dermatology* **36**, 243–249.

Dethier, V. G., Browne, L. B. and Smith, C. N. (1960). The designation of chemicals in terms of the responses they elicit from insects. *Journal of Economic Entomology* **53**, 134–136.

Dryden, M. W., Payne, P. A., Smith, V. and Hostetler, J. (2006a). Efficacy of imidacloprid (8.8% w/w) plus permethrin (44% w/w) spot-on topical solution against *Amblyomma americanum* infesting dogs using a natural tick exposure model. *Veterinary Therapeutics* **7**, 99–106.

**Dryden, M. W., Payne, P. A., Smith, V. and Hostetler, J.** (2006b). Evaluation of an imidacloprid (8.8%w/w)-permethrin (44.0% w/w) topical spot-on and a fipronil (9.8% w/w)-(S)-methoprene (8.8% w/w) topical spot-on to repel, prevent attachment, and kill adult *Rhipicephalus sanguineus* and *Dermacentor variabilis* ticks on dogs. *Veterinary Therapeutics* **7**, 187–198.

Endris, R. G., Cooke, D., Amodie, D., Sweeney, D. L. and Katz, T. L. (2002). Repellency and efficacy of 65% permethrin and selamectin spot-on formulations against *Ixodes ricinus* ticks on dogs. *Veterinary Therapeutics* **3**, 64–71.

**Endris, R.G., Matthewson, M.D., Cooke, M.D. and Amodie, D.** (2000). Repellency and efficacy of 65% permethrin and 9.7% fipronil against *Ixodes ricinus. Veterinary Therapeutics* **1**, 159–168.

European Medicine Agency (2007). Guidelines for the Testing and Evaluation of the Efficacy of Antiparasitic Substances for the Treatment and Prevention of Tick and Flea Infestation in Dogs and Cats. EMA Committee for Medicinal Products for Veterinary Use Guideline No. EMEA/CVMP/ EWP/005/2000-Rev2-2007. European Medicine Agency, London, UK.

Folz, S. D., Ash, K. A., Conder, G. A. and Rector, D. L. (1986). Amitraz: a tick and flea repellent and tick detachment drug. *Journal of Veterinary Pharmacology and Therapeutics* 9, 150–156.

Ian, R.H. and Bryan, H.P. (1981). Further studies on amitraz as a veterinary acaricide. *Pesticide Science* 12, 467–474.

Marchiondo, A. A., Holdsworth, P. A., Green, P., Blagburn, B. L. and Jacobs, D. E. (2007). World Association for the Advancement of Veterinary Parasitology (W.A.A.V.P.) guidelines for evaluating the efficacy of parasiticides for the treatment, prevention and control of flea and tick infestation on dogs and cats. *Veterinary Parasitology* **145**, 332–344.

Narahashi, T., Zhao, X., Ikeda, T., Salgado, V. L. and Yeh, J. Z. (2010). Glutamate-activated chloride channels: unique fipronil targets present in insects but not in mammals. *Pesticide Biochemistry and Physiology* **97**, 149–152.

Otranto, D., Dantas-Torres, F. and Breitschwerdt, E.B. (2009). Managing canine vector-borne diseases of zoonotic concern: part one. *Trends in Parasitology* 25, 157–163.

Otranto, D. and Wall, R. (2008). New strategies for the control of arthropod vectors of disease in dogs and cats. *Medical and Veterinary Entomology* 22, 291–302.

Randolph, S.E. (2009). Tick-borne disease systems emerge from the shadows: the beauty lies in molecular detail, the message in epidemiology. *Parasitology* **136**, 1403–1413.

Sonenshine, D.E., Lane, R.S. and Nicholson, W.L. (2002). Ticks (Ixodida). In *Medical and Veterinary Entomology* (eds. Mullen, G. and Durden, L.). pp. 517–558. Academic Press, San Diego, CA, USA.

Wege, P. J., Bywater, A. F., Le Patourel, G. N. J. and Hoppé, M. (2002). Acquisition and transfer of a lambda-cyhalothrin microcapsule formulation by *Blattella germanica*. In *Proceedings of the 4th Conference on Urban Pests, Charleston, Virginia, USA* (eds. Jones, S. C., Zai, J. and Robinson, Wm.H.). pp. 135–146.