Pre and postnatal maternal distress and infant gut immunity in the Canadian Healthy Infant Longitudinal Development (CHILD) birth cohort

by

Liane Joo Kang

A thesis submitted in partial fulfillment of the requirements for the degree of

Master of Science

in

Medical Sciences - Paediatrics

University of Alberta

© Liane Joo Kang, 2017

Abstract

Introduction: Secretory immunoglobulin A (sIgA) plays a critical role in gut mucosal immunity and is a marker of immune maturation in early life. Delayed IgA production is associated with increased risk of allergic diseases. Animal models of stressful events before birth and during infancy show changes in the maternal vaginal microbiome and in intestinal microbial composition of offspring. A human study found infants born to mothers with greater prenatal stress more likely to have gut dysbiosis, imbalance in microbial composition, but there is a gap in literature on stress-microbiome-immunity pathways in humans. This study investigated differences in infant fecal sIgA levels according to the depression and stress status of the mother pre and postnatally.

Methods: Data were obtained from a sub-sample of 403 term infants from the Vancouver, Edmonton and Winnipeg sites of the general cohort in the Canadian Healthy Infant Longitudinal Development (CHILD) Study. Mothers of the infants were enrolled during pregnancy and were asked to report stress and depressive symptoms through scored-scales administered to the general CHILD cohort at several time points throughout pregnancy and postpartum. Center for Epidemiologic Studies Depression (CES-D) Scale ascertained depressive symptoms, and Perceived Stress Scale (PSS) determined perceived stress. Infant stool samples were collected at a mean age of 3.8 months, and fecal sIgA levels were measured using the Immundiagnostik sIgA ELISA kit. Using IBM SPSS version 24, Mann-Whitney U-tests detected median differences in IgA levels according to maternal distress status, and logistic regression models estimated the odds of lowest and highest quartile, lowest tertile and below median sIgA in infants.

ii

Results: About 12% of women had prenatal depressive symptoms, 9% had symptoms postpartum and 9% had symptoms both pre and postnatally. With perceived stress scores in the highest quartile, 16.7% of mothers fell in this category at prenatal only, 10.2% for postnatal only and 18.7% for both pre and postnatal. Mothers with any depressive symptoms had infants with significantly lower sIgA compared to mothers without symptoms (p=0.004). Median sIgA levels are 6.3 (IQR=3.6 - 12.4) mg/g feces, 5.2 (IQR = 2.0 - 9.8) mg/g feces, 5.7 (IQR = 2.6 - 8.6) mg/g feces, 4.4 (IQR = 2.4 - 8.0) mg/g feces for exposure to less symptoms (below cut-off), prenatal, postnatal, both pre and postnatal symptoms respectively, which are mutually exclusive categories. Median sIgA levels with higher stress were not significantly different compared to the reference group, but after stratification for the presence of other children and pets at home, significant differences were seen when the mother had both pre and postnatal higher stress. The odds of low fecal sIgA was 3.07 times higher when the mother had both pre and postnatal symptoms (95% CI: 1.25 - 7.55) compared to less symptoms (reference group), after controlling for infant age, antibiotics exposure, maternal asthma or allergy, prenatal SSRI use, pets, breastfeeding and gravida. In the same model, the odds ratio for prenatal only depressive symptoms was 2.44 (95% CI: 1.07 – 5.57). No significant odds ratios were found with higher perceived stress.

Conclusion: Infants born to mothers with higher levels of depressive symptoms and perceived stress appear to have lower fecal sIgA levels independent of various study covariates. Due to the association with lower sIgA and possibly changes in the infant gut microbiome, maternal distress may put infants at higher risk of later development of allergic diseases.

iii

Preface

This thesis is an original work by Liane Joo Kang. The thesis was written in a traditional thesis format according to the guidelines of the Faculty of Graduate Studies and Research at the University of Alberta. Another version of this thesis is in preparation for submission to the Journal of Allergy and Clinical Immunology titled as "Maternal perceived stress and depressive symptoms is associated with reduced secretory immunoglobulin A levels in Canadian infants".

Chapter 1 is the introduction that consists of a literature review on maternal distress, infant gut immunity and potential covariates between the association of distress and gut immunity, study objectives and hypotheses and sample size calculations.

In Chapter 2, the relevant methods of the CHILD study and the information on variables used for the study on distress and gut immunity are outlined. Chapter 3 includes the results of this quantitative study. The final chapter (Chapter 4) contains the discussion and conclusions. This chapter contains the main findings and interpretation of the findings from Chapter 3, the strengths and limitations of the study, the clinical significance and implications for future research.

Dedication

Dedicated to my family, friends, mentors and my heavenly Father

Acknowledgments

I would like to thank my mentor and supervisor, Dr. Anita Kozyrskyj, for accepting me as her student and for supporting and teaching me in every step during my Master's program. Also, I would like to thank my supervisory committee members, Dr. Eytan Wine and Dr. Gerald Giesbrecht for guiding me and providing their knowledge and expertise.

I would like to give many thanks to the previous and current members of the SyMBIOTA (Synergy in Microbiota Research) Research Group for the many helpful suggestions and assistance with the data. Petya, Sarah, Mon, Meghan, Tedd, Manjeet, Hein, Usha and Brittany, thank you for being part of the journey to complete this thesis project.

Thank you to the families and staff of the CHILD (Canadian Healthy Infant Longitudinal Development) study for giving me the opportunity to work with their data. Special thanks to Petya Koleva and Dr. Catherine Field's lab for analyzing the sIgA from the stool samples. This thesis would not have been possible without their support.

Finally, I would like to thank my beloved parents for sacrificing everything they had in Korea to come to Canada so that I would be able to pursue higher education and have better opportunities. Thank you for your love, patience, guidance and prayers.

Financial support for this study was provided by AllerGen NCE, University of Alberta Faculty of Medicine & Dentistry and Canadian Institutes of Health Research (CIHR).

Table of Contents

Chapter 1: Introduction		
1.1 Distress, Asthma and Allergic Diseases	1	
1.2 Maternal Distress	1	
1.2.1 Prenatal distress	1	
1.2.2 Prenatal distress impact on fetal development	2	
1.2.3 Postnatal distress	3	
1.2.4 Postnatal distress and mother-infant interactions	3	
1.2.5 Long-term outcomes to mother and child	4	
1.3 Gut Microbiome Development in Early Life	5	
1.4 Infant Gut Secretory IgA	6	
1.4.1 Role in gut immunity	6	
1.4.2 Breastmilk sIgA	8	
1.4.3 sIgA Production in Infants	8	
1.4.4 Importance of gut immunity at early life	8	
1.5 Potential Covariates	10	
1.5.1 Maternal asthma and allergies	10	
1.5.2 Pregnancy overweight	10	
1.5.3 Delivery mode	12	
1.5.4 Gravidity	13	

1.5.5 Small-for-gestational-age (SGA)	14
1.5.6 Antibiotics exposure	14
1.5.7 Selective serotonin reuptake inhibitors (SSRI)	15
1.5.8 Breastfeeding	16
1.5.9 Smoke exposure	17
1.5.10 Home environment	18
1.5.11 Infant allergic eczema	19
1.6 Distress and Infant Gut Development	20
1.6.1 Distress and infant gut development	20
1.6.2 Distress and gut immunity	21
1.6.3 Summary of gaps in literature	22
1.7 Hypothesis and Objectives	23
1.8 Sample Size Calculation	23
Chapter 2: Methods and Materials	
2.1 Methods overview	25
2.2 Study population	25
2.3 Exposures	26
2.4 Definition of potential covariates	27
2.5 Stool collection and storage	29
2.6 Extraction and analysis of sIgA	29

2.7 Statistical analyses	30
Chapter 3: Results	
3.1 Sub-sample Characteristics	31
3.2 Maternal Distress and Infant sIgA	35
3.2.1 Infant sIgA with exposure to both pre and postnatal distress (primary question)	37
3.2.2 Infant sIgA with exposure to only prenatal distress	40
3.2.3 Infant sIgA with exposure to only postnatal distress	42
3.2.4 Simple and multiple logistic regression analyses with lowest quartile sIgA	44
Chapter 4: Discussion and Conclusions	
4.1 Summary and interpretation of the findings	48
4.1.1 General overview	48
4.1.2 Time-specific distress effects	49
4.1.3 Pre and postnatal distress effects on infant sIgA levels	50
4.1.4 Prenatal only distress effects on infant sIgA levels	52
4.1.5 Postnatal only distress effects on infant sIgA	54
4.1.6 Birth characteristics and home environment impacting on association of interest	55
4.2 Strengths of study	56
4.3 Limitations of study	57
4.4 Clinical relevance	60
4.5 Implications for future research	61

	4.6 Conclusion	61
R	References	62
A	Appendices	
	Appendix A: Preliminary analyses with 304 sample and pregnancy overweight	84
	Appendix B: Preliminary analyses with perceived stress cut-off at mean score of CHI	LD
	cohort (12.96)	90
	Appendix C: Logistic regression analyses on below median sIgA, highest quartile and	
	lowest tertile sIgA as outcome variables (yes/no)	95
	Appendix D: Sensitivity analyses for home births, age, gestational age and SSRI use	100
	Appendix E: Maternal distress and sIgA median comparisons and logistic regression	
	models	106

List of Tables and Figures

 Table 1 – Proportion of depressive symptoms, higher perceived stress and potential covariates in

 403 sub-sample and CHILD cohort

Table 2 – Median infant fecal sIgA levels and percentage distribution of maternal depressive symptoms and lowest quartile sIgA in relation to potential covariates (n=403)

Table 3 – Median infant fecal sIgA levels and percentage distribution of maternal higher

 perceived stress and lowest quartile sIgA in relation to potential covariates (n=403)

Table 4 – Crude and adjusted odds ratios (OR) of low sIgA for exposure to maternal depressive symptoms

 Table 5 – Crude and adjusted odds ratios (OR) of low sIgA for exposure to maternal higher

 perceived stress

Figure 1 – Infant fecal sIgA levels depending on distress status of mother

Figure 2 – sIgA levels among breastfed and formula-fed infants at time of stool collection for different depressive symptoms groups

Figure 3 – sIgA levels among depressive symptoms groups after stratification for potential covariates

Figure 4 – Infant sIgA levels among higher perceived stress groups after stratification for other children and pets living at home

xi

Figure 5 – Infant sIgA levels based on maternal depressive symptoms groups after stratification for pet ownership

Figure 6 – Infant sIgA levels based on maternal depressive symptoms groups after stratification for maternal asthma or allergy during pregnancy and child sex

Chapter 1: Introduction

1.1 Distress, Asthma and Allergic Diseases

The prevalence of asthma and allergies is increasing worldwide, and these conditions remain as a public health concern in both children and adults (1-3). Although various factors may lead to the development of these diseases, many studies show a relationship between distress (stress, depression and anxiety) and allergic diseases (4-7). A meta-analysis of 10 studies on prenatal maternal stress (higher perceived stress, stress exposure or difficult life circumstances) and child asthma and wheezing, where four studies were on asthma, reports a significant OR of 1.56 for respiratory morbidity with the exposure of prenatal stress, supporting the negative impact of maternal psychological stress on the respiratory health of the child (4). And ersson's systematic review of 16 studies, primarily prospective cohort studies, found support for the association between maternal stress during pregnancy and atopic diseases in children, but there were diverse measures for the exposure and outcome (5). Many questions still need to be answered about the mechanistic links between distress and allergy and asthma development. One of the possible links is through the development of the infant gut microbiome, as early changes in the infant microbiome have been linked to asthma and allergies. This thesis will observe the associations between pre and postnatal distress (stress and depression) in mothers and gut immunity in infants.

1.2 Maternal Distress

1.2.1 Prenatal distress

Major depression during pregnancy affects approximately 10% of women based on a systematic review of 28 studies from developed countries (8). However, the prevalence of

psychosocial stress during pregnancy is not known but has been reported to be common in an ethnically diverse sample of pregnant women (9). A small minority of pregnant women receives care for when they have depression (10), and stress is not routinely measured in obstetric care (9). Distress may be occurring despite the fact that both pre and postnatal distress are detrimental to the cognitive, socio-emotional and psychomotor development in the infant (11). Valid psychometric measures of stress and anxiety are needed for clinical care; however, Bann et al. (12) was able to identify the Perceived Stress Scale, Pregnancy Experience Scale and State Trait Anxiety Inventory – Trait-Anxiety Subscale as reliable and valid measures of higher levels of stress and anxiety in nulliparous pregnant women.

1.2.2 Prenatal distress impact on fetal development

According to the DSM-5, the symptoms during a depressive episode include depressed mood for the majority of the day almost every day, clinically significant weight loss or gain, inability to sleep or oversleeping, loss of energy, feelings of worthlessness or inappropriate guilt, diminished ability to think or concentrate and lower interest in activities (13). These symptoms, occurring at clinically significant levels for a pregnant woman, may pose health risks on both the pregnant mother and the developing fetus. Lack of nutritional foods in the mother's diet could lead to poor transfer of nutrients from placenta to fetus, affecting fetal development, and lead to iron deficiency, oxidative stress, chronic inflammation and smaller birth size in the children (14). Increased stress may elevate glucocorticoid transfer between mother and fetus via the placenta due to maternal distress having the potential to downregulate enzymes that prevent transfer (14, 15). Increased fetal exposure to glucocorticoids may impact the responsivity of stress neuroendocrine and immunological systems in the child later by affecting density of receptor sites and epigenetic programming (14, 15).

1.2.3 Postnatal distress

Overall prevalence of postpartum depression is 19%, but prevalence differs from country to country (8, 16). Like prenatal depression, majority of affected women do not seek treatment for their symptoms of sadness and anxiety, reduced energy leading to less activity, loss of enjoyment and interest, changes in appetite and sleeping patterns, and feelings of inadequacy when caring for their infant (10, 17, 18). Mothers with prenatal distress or with a history of mental disorders are more likely to develop postpartum depression (19). However, the impact of distress differs depending on the time period. New mothers may be at higher risk because of the stress that comes with the initiation of parenting (20). A study found that there is a common tendency for depressive symptoms in the postpartum period to peak after the first month of the infant's life (20). It should also be acknowledged that maternal depression during the perinatal period is rather heterogeneous as several different symptom profiles and trajectories have been found with different risk factors and child outcomes, but there is limited information as to which specific symptoms and combinations contribute to the heterogeneity (21).

1.2.4 Postnatal distress and mother-infant interactions

Mothers who are depressed are less likely to interact with their infants most likely due to the symptoms of low self-esteem and loss of concentration (22). Maternal depression has been associated with greater instances of abuse and neglect on the child (23). This lack of interaction with the child affects the mother-child bonding and attachment and is associated with both shortterm and long-term child health outcomes (24). In 2-4 month-old infants raised by mothers with depressive symptoms, there were reduced odds of parental behaviours that are important for child development, such as playing and conversing with the child, and there was lower likelihood and reduction of breastfeeding, the gold standard for infant feeding (22, 25). Maternal distress has been shown to affect the stress reactivity of the infant through affected maternal behaviours potentially leading to epigenetic changes in the infant (26).

Postpartum maternal distress may also affect the environment in which the child develops. Santos et al. (27) reported significant differences in the quality of the home environment between mothers with high and lower levels of distress based on social-emotional and stimulation characteristics, but this study was focused on preterm infants. Stressful environments during the infants first year of life may impact the HPA axis and immune development of the infants (28, 29).

1.2.5 Long-term outcomes to mother and child

Some studies have found that toddlers did not appear to be affected behaviourally or developmentally by maternal depression (30). However, child developmental delays in language and cognitive function and poor child mental health have been seen with maternal depression (23, 31). A recent systematic review of 16 studies on adolescent outcomes found that adolescents exposed to postnatal depression at infancy had higher rates of attentional difficulties and conduct problems but found inconsistent reports for psychosocial outcomes (23). The review also included two studies from a research group that found higher depression incidence at age 16 and

cortisol levels at age 13 with postnatal depression, mediated by postnatal depression and insecure attachment during infancy.

Common co-morbidities with depression, asthma and allergic diseases, have been found in children that were born to or raised by distressed mothers (5, 32). Caregiver stress is a major predictor and mediating risk factor of wheeze in children (33, 34). A recent publication on the association between postpartum distress and asthma in a nationally representative sample of Canadian infants (7), found the association diminished with child age, which may suggest for less concern for mothers that are worried about the effects of their distress on their child's health. The directionality and causality of the association between poor mother-infant relationships and asthma have yet to be determined due to the lack of empirical studies (28).

1.3 Gut Microbiome Development in Early Life

The early gut microbiota composition in infants, rather than later composition, is considered important to the development of the immune system (35). The *in-utero* fetal gut microbiome development may allow for the development of the gut immune system in the fetus during pregnancy (36). Microbial seeding in the infant gut is currently thought to start *in-utero* through sources from the placental and maternal gut microbiome that can be transferred through the bloodstream and swallowed by the infant in amniotic fluid (36), as shared microbial composition features are found between the placental and amniotic samples with infant meconium (37).

The newborn gut is first colonized by aerobes and facultative anaerobes that are tolerant to the aerobic environment in the newborn gut, and these colonizers create an anaerobic environment for other microbes, obligate anaerobes, to thrive in the gut (36). Initially, the infant

gut has lower microbial diversity and greater variability between individuals than the adult gut. Eventually at around one year of age, the infant gut microbiome shows some similarities with the adult gut (36). Differences in infant gut microbiome composition in the first few months of life have been shown between vaginal and C-section births, breastfed and not breastfed and with environmental factors. Infants born from vaginal births more likely have maternal vaginal and fecal bacteria colonized in their gut; whereas, those born via C-sections have maternal skin and hospital bacteria colonized in the gut (36). Breastmilk also contributes to the establishment of gut microbiome in infants, as breastfeeding is linked to reduced gut microbial diversity in 3-month infants (38), due to the human milk oligosaccharides (HMOs) that favour higher abundance of microbes from the genera *Bifidobacteria* and *Lactobacillus* (39). Formula-fed infants have higher gut microbial diversity and have microbes from the genera *Bacteriodes, Veillonella* and *Clostridium* (36). Environmental factors, such as the presence of older siblings and pets, have been shown to affect the infant gut microbial composition (40).

1.4 Infant Gut Secretory IgA

1.4.1 Role in gut immunity

Secretory immunoglobulin A (sIgA) is an antibody that is produced by plasma B cells in the Peyer's patches, lamina propria and cecal patches in the gut, and IgA response against antigens are thought to involve both T cell-dependent and independent mechanisms (41). Secretory IgA is also present in other secretions, such as breastmilk, tears and mucus. The structure of sIgA differs from serum IgA, and plays a more decisive role in mucosal immune defense (42). As the first line of defense in the gut, sIgA plays a critical role in the gut mucosal immune system in early life because it prevents pathogens from entering the gut serum through

immune exclusion and allows for the induction of oral tolerance to commensal gut bacteria at early life (42). Secretory IgA can change the expression of surface molecules in commensal bacteria to reduce inflammation, but the exact mechanisms have yet to be established (43). On the other hand, sIgA may allow for transcytosis of pathogenic microbial species via Microfold cells (M cells) so that they can be degraded by other immune cells (43). Animal studies show that supplementing sIgA in mice that are deficient in sIgA can adequately restore the microbiota composition, which suggests a possibility of the development of immunoglobulin-based therapies that can restore optimal microbiota composition (44, 45). Therefore, sIgA seems to play a key role in shaping the gut microbiota composition in early life, and because of its significant role, sIgA is considered a marker of immune maturation in infants (46).

The initial infant gut microbiota composition is also thought to be important to the development of the immune system (35). Germ free mice, mice that do not have any microbes, are lacking in sIgA-producing B cells (47). However, when there is colonization of microbes in these mice, sIgA plasma cells are seen again. Certain microbial species can also stimulate sIgA production in the gut, and thus factors affecting gut microbial composition may also affect the production of sIgA. This suggests interplay between gut microbiota composition and gut immunity. Mice provided with lactic acid bacteria, such as *Lactobacillus* and *Bifidobacterium* species, had increased both serum and secretory IgA production as well as increased the number of Peyer's patches (48). Proteobacteria, for instance pathogenic *Escherichia coli* and *Salmonella enteritidis*, have been reported to promote sIgA production in the intestine (49, 50).

1.4.2 Breastmilk sIgA

Newborns have limited production of sIgA in the first few weeks of life (42, 51); therefore, breastmilk provides sIgA to protect the child from pathogens during this vulnerable period. Breastfeeding has had significant associations with IgA levels in later months of infancy (52). Six-month old infants when breastfed secreted higher levels of salivary sIgA than formulafed infants, which suggests breastfeeding may help stimulate the production of sIgA in infants (53). In addition, after the cessation of breastfeeding, changes in microbiota composition in 12month infants indicate the importance of sIgA in breastmilk in shaping and driving the maturation of the infant gut microbiome (54). An experimental study on sIgA-deficient dams found changes in gut microbiota composition in the pups that lasted to adulthood as well as a compromised gut epithelial barrier at early life that may prevent sensitization to microbe antigens from an immature immune system (55). This study suggests that sIgA from breastmilk given to the infant may influence the development of the gut microbiome and immune system that could influence later health.

1.4.3 sIgA Production in Infants

Peyer's patches and B cells develop at 11 and 12-16 weeks gestation respectively, and IgA H chain transcripts have been found in cord blood at 27 weeks gestation (47). Although endogenous IgA is thought to start producing several months after birth in humans (56), fecal total IgA have been found in exclusively formula-fed infants as early as 3 months of age (52). Endogenous IgA production slowly increases over the first few years after birth (56).

1.4.4 Importance of gut immunity at early life

According to data from the World Health Organization (WHO), about 1.1 million newborns in the world die from early infections every year (57). The exposure to a myriad of pathogenic microbes in their new environment after birth necessitates newborn infants to transition from *in-utero* protection via their mother's immune system to dependence on their own immunity, which is relatively immature at birth. The reason as to why some newborns are able to survive this transition is still not known. Since infants are not able to produce sIgA until as early as one month after birth, they rely on sIgA derived from maternal breastmilk to obtain passive immunity during early life. There are several reports of breastfeeding and its protective role against infections (47).

Delayed immune maturation and production of serum and intestinal IgA in early life is associated with increased risk of allergic diseases in later childhood (46, 58, 59). Higher total infant fecal IgA at 3 months has been associated with reduced *Clostridium difficile* colonization, and early colonization of this microbial species in infants have been associated with increased risk of allergic diseases (60). Another study on secretory IgA found that total fecal sIgA measured at 1 month and 12 months of age were not significantly different between healthy and allergic/asthmatic groups of children at age 7, but the sample sizes for both groups were under 30 (61). However, the microbe-sIgA interactions at 1 year of age were significantly different in the allergic and asthmatic groups compared to the control group, notably lower sIgA binding to *Bacteriodes* genus and *Escherichia* species in allergic and asthmatic children. Different IgA responses to gut microbiota are preceding the development of allergic conditions; therefore, screening sIgA responses may provide more insight about allergic diseases. Secretory IgA has many links to asthma and allergies and may potentially reveal more about pathway links to maternal distress.

1.5 Potential Covariates

1.5.1 Maternal asthma and allergies

Higher rates of depression and stress are present in children and adults with asthma and allergies, and animal and human research studies indicate chronic stress disrupts physiological systems that increase risk of allergies and asthma (6, 62). Stress in the mother also exacerbates allergen responses in infants (63). Corticosterone, a glucocorticoid that is increased during pregnancy and can pass through the placenta, seems to enhance Th2 cytokine responses, and therefore, it seems to be a mediator that is transferred from mother to fetus and increases allergy responsiveness in the child (63). In addition, studies seem to suggest that maternal transfer of allergy risk to offspring may occur prenatally (63). Th2 responses in mothers with allergies, such as antibody depletion of maternal T cells, seem to modulate the allergen responsiveness development in the infants. Differences in allergen responsiveness appear to be induced by maternal alterations in the dendritic cells of offspring in the murine model (63), and these cells are important for sIgA production (41).

1.5.2 Pregnancy overweight

Women who are depressed have a higher risk of becoming overweight and obese (64). Depression is directly related to overweight during pregnancy, but this association is multidirectional as the concern of overweight can increase depressive symptoms and depression prior to pregnancy can increase risk of excessive gestational weight gain, while the relationship seems to stronger for the latter (64, 65). Alternatively, maternal stress is not associated with overweight during pregnancy. Maternal obesity during pregnancy has several health risks on both the mother and infant, such as gestational diabetes and higher likelihood of having a Csection in the mothers and increased infant birth weight and likelihood of fetal defects (66).

In addition, overweight in mothers after birth may impact the mother-infant relationship as according to Bergmann et al. (67), obese mothers score significantly lower in emotional availability, in terms of sensitivity to child's signals, appropriate guidance of child activity, nonhostility and the child's response and involvement in the interaction with the mother. Lower emotional availability has been previously reported to predict childhood obesity. Interestingly, depressive symptoms in the Bergmann study did not have strong associations with emotional availability perhaps due to depressive symptoms not reaching clinical significance in the sample.

Pregnancy overweight is linked to changes in infant gut microbiome. One seminal study based on a prospective birth cohort looked at maternal pre-pregnancy and pregnancy weight and infant fecal microbiota composition. This study found lower abundance of *Bacteriodes* and *Bifidobacterium* at one month and higher abundance of *Clostridium histolyticum* at six months with higher pre-pregnancy weight and excessive weight gain during pregnancy (68). Some of these microbiota composition changes may influence IgA production in the gut. Whether maternal overweight directly affects IgA production in the infant gut is yet to be determined, but lower salivary sIgA have been found in obese children (69). A recent review suggests that maternal obesity is associated with immune dysregulation in the mother, which may affect the fetal immune development and postnatal immunity (39). One study compared sIgA levels in colostrum and found significantly higher levels in the colostrum of overweight and obese mothers (70). Although immunological changes were observed, the group did not find support for the negative impact of obesity on immune protection through breastfeeding. One point to add is that obese mothers are less likely to initiate and have shorter duration of breastfeeding (66),

which may be one explanation to the reduced *Bifidobacterium* abundance in infants who are born to overweight mothers. This may suggest infants of overweight women may be provided with less immune protection in the early months of life due to reduced breastfeeding.

1.5.3 Delivery mode

Studies on the delivery mode and pre and postnatal distress have conflicting results as to whether the association is significant possibly due to different tools used to measure distress and demographic differences. A longitudinal cohort study of over 55 000 women found no association between postpartum emotional distress and mode of delivery after adjusting for confounding variables, but this study used a distress measure that is not commonly used for diagnosing depression due to low specificity (71). However, this study suggests that there is a strong association between prenatal emotional distress and the occurrence of scheduled/elective C-sections. While a much smaller Canadian cohort used the Edinburgh Postnatal Depression Scale (EPDS), they found no significant associations suggesting that the place of birth and other confounding factors may explain the co-occurrence of delivery mode and depression (72).

The studies that did find significant associations postpartum distress and mode of delivery found higher incidence of depression with C-sections (73, 74) as well as primary C-sections or first C-section for the mother (75). Delivery mode preference of pregnant women prior to delivery may be a factor in the stress response of women to the actual delivery mode. Garthus-Niegel et al. (76) found increased post-traumatic stress symptoms in women who preferred delivery by C-section but delivered vaginally compared to women who both preferred and delivered vaginally with depression and anxiety being significant covariates. The same effect

was not seen in mothers who preferred vaginal delivery but delivered via C-section. The psychological factors in mothers may be more relevant to the performance of scheduled C-sections rather than vaginal and emergency C-sections.

A population-based study in Denmark found caesarean section associated with diseases involving immune function and mucosal surfaces in the infants: asthma, gastroenteritis, ulcerative colitis, celiac disease, and differences were seen between elective and acute caesarean sections (77). An older study found 3-, 6-, 12-month total fecal IgA was lower in the vaginallydelivered babies compared to the babies delivered through caesarean section, but this study did not look at elective and emergent caesarean section separately that have different circumstances (78). Early colonization of microbiota in infant gut is widely known to be affected by the mode of delivery (36, 40), and therefore mode of delivery may influence the immune response and development in the infant gut.

1.5.4 Gravidity

A preliminary study on a pilot cohort of the CHILD study by Bridgman et al. (52) found associations between parity and total fecal IgA levels in 3-4-month-old infants. First-borns had higher total fecal IgA than those with older siblings, and this may be because first-borns were more likely to be breastfed in this study. A study on breastfeeding durations among women with postpartum depressive symptoms found greater prevalence of clinically significant depressive symptoms when the mother was a new parent (79). These new mothers were less likely to breastfeed up to the recommended period. Bascom et al. (79) reports a common reason for early cessation of breastfeeding in women with depressive symptoms was the burden of too many

household duties. This reason along with sleep problems and fatigue that are symptoms of depression may affect the mother-infant interactions and reduce breastfeeding.

1.5.5 Small-for-gestational-age (SGA)

Prenatal stress and depression increases risk of small-for-gestational-age (SGA) during the second trimester, and this risk appears to be higher in male infants (80). The use of antidepressants during pregnancy also appears to increase SGA risk (81), which may be part of explanation to the association between distress and birth weight. Epigenetic evidence in placenta supports the link between distress and SGA, which shows lower methylation levels for the transcription site of a leptin receptor in SGA births and when the mother had active depression that was not treated (82). Leptin is considered to have an important regulatory role in fetal nutritional intake and growth. Low birth weight and SGA are associated with greater risks of neonatal hyperbilirubinemia, which is linked to three to four times greater risk of asthma and allergic rhinitis (39). Whether SGA affects IgA production has not be studied thus far. SGA infants may have more IgA as higher IgA content has been found in the breastmilk of mothers that gave birth to a low birth weight and small-for-date infant (83).

1.5.6 Antibiotics exposure

Antibiotics have a profound effect on gut microbiota. Although there is no conclusive evidence on the impact of prenatal antibiotics use on maternal microbial transmission to the infant gut (84), antibiotics usage during the delivery and the first few months of life appear to cause major perturbations in the infant gut microbial composition. Distressed women during pregnancy have higher usage of antibiotics during pregnancy possibly attributable to the

increased susceptibility of infections (85), which may affect the composition of the maternal vaginal microbiome. Infants born to depressed mothers have more infections in the early weeks of postpartum (86), and thus they may have increased use of antibiotics that can alter the gut microbiota establishment. Antibiotics taken by the mother have been found in breastmilk in small concentrations, but even the small amounts can change the gut microbiota in the infant (87). Intrapartum antibiotics use for vaginal delivery and C-sections have been studied in the CHILD study, and it has been reported that with intrapartum antibiotics exposure there is a lowering of microbiota richness and *Bacteriodetes* abundance in 3-month infant gut (38).

1.5.7 Selective serotonin reuptake inhibitors (SSRI)

Pregnant women may take antidepressants, selective serotonin reuptake inhibitors (SSRIs) being the first choice, if they do not respond to psychological therapies alone when treating for moderate to severe depression (18). These medications sometimes have negative effects on the developing fetus and cause immunomodulatory effects (18, 88, 89). In addition, short-lived neonatal adaptation syndrome occurs with SSRI use (18). Antidepressants are taken to prevent the risk of relapse of depression, which counterbalances the pharmacological risks on the fetus (90). A recent review on antidepressant use during pregnancy, discusses the favouring of preventing depressive symptoms over risks of antidepressant use due to currently published studies that reveal statistically significant but minor clinical outcomes. However, there are no clinical trials that confirm the safety of antidepressants that are used during this critical developmental period (91).

1.5.8 Breastfeeding

Breastfeeding seems to be the main covariate between distress and infant gut IgA levels. According to some studies (79, 92), mothers are less likely to interact and continue breastfeeding their infants when depressed starting at early postpartum, but the directionality of this association is yet to be determined as some evidence show difficulty breastfeeding can precede the development of postpartum depression. Reasons for breastfeeding cessation in depressed mothers are lack of social support, the need to go back to work, self-consciousness about breastfeeding in public and other medical reasons (30, 79). Infants cared by depressed mothers may be provided with less sIgA from breastmilk. Lower breastfeeding could reduce the amount of exposure to antidepressants taken by the mother during lactation, but usually the levels of SSRIs are low and undetectable in breastmilk and few adverse effects have been reported in infants (81). Increased levels of breastfeeding have been associated with increased levels of total fecal IgA (52). Whether breastfeeding induces sIgA production is still not known because of the inability to differentiate between maternal-derived and infant-produced sIgA of breastfeed infants.

Breastmilk favours greater abundance of *Bifidobacteria* and *Lactobacillus* because of the presence of bioactive ingredients like the human oligosaccharides (36, 39). Infants supplemented with probiotics that include microbial species belonging to the two genera are likely to have higher fecal IgA than infants without supplementation. This altering of colonization may impact the secretion of IgA, since certain microbial species may be stimulatory of sIgA production. However, evidence is still not clear as to whether breastfeeding has protective abilities against later development of allergy and asthma (39, 93).

1.5.9 Smoke exposure

As distress in the perinatal time period is linked with higher levels of smoking (94-96), smoke exposure to the infant pre and postnatally may have impact on the development of infant gut immune system. Maternal smoking may disturb the establishment of the gut microbiota in early life as smoking cessation has been linked to major changes in proportions of *Firmicutes* and *Bacteriodetes* in the adult human gut (97). Mothers who are smokers or were exposed to environmental tobacco smoke (ETS) gave birth to infants with tobacco smoke metabolites in the meconium, the first stool of a newborn (98). Another study on 20 human meconium samples reported maternal smoking during the entire pregnancy had no meconium samples with predominantly lactic acid bacteria (99), which are stimulatory of IgA production (46).

Conflicting results have been shown for the association between maternal smoking and infant salivary IgA levels, but these differences may be due to difference in age and other covariates. Four to six year old children who live with a smoker tended to have lower salivary sIgA (100). Total salivary IgA in 12-month infants was higher when born to mothers who smoked during pregnancy and during the first year of life, which may be due to the increased vulnerability to infections from smoking since IgA was significantly associated with chest infections (101). The same study did not find differences in common respiratory pathogenspecific IgA with maternal smoking status. This study also found that mothers who smoked were more often stopping breastfeeding earlier, and other studies have reported the same association along with lower rates of breastfeeding initiation (102-104). Breastmilk from mothers who smoked during lactation tended to have lower sIgA levels, but this result was not significant (105). Currently, there are no studies on tobacco smoke and gut sIgA.

1.5.10 Home environment

Interactions with animals appear to have beneficial effects on psychological stress in humans. A review by Beetz et al. (106) summarized that human-animal interactions were seen with an improvement of social attention, interpersonal interaction and mood as well as a reduction of self-reported anxiety. However, there is limited evidence on the association between humananimal interactions and reduction of stress hormones, epinephrine and norepinephrine. The review proposes that the interactions with animals may work through the oxytocin system and reduce psychological stress, as increased oxytocin levels are seen with pet ownership (106). Animal-assisted therapy has been used in conjunction with psychotherapies for various psychological disorders (107). Although patients report effectiveness of this therapy, evidencebased support is still needed. The absence or presence of siblings at home in relation to maternal distress may be similar to the relationship between gravida and distress as discussed in Section 1.5.4.

The presence of pets and other children at home are considered because of reported changes in gut microbiome. Martin et al. (40) reported significantly lower *Lactobacillus reuteri* species in the 6-month old infant gut with pet ownership and if there were older siblings. Based on a systematic review of 6 studies that looked at the gut microbial profile of children with eczema, increased *Lactobacilli* and *Bacteriodes* have been found with increased number of siblings, while a lack of older siblings was associated to a rapid colonization of *Clostridium* (108). It is generally assumed that pets and older siblings increase exposure to environmental microbes in early life, which may help the development of the infant gut microbiome and immune system according to the "hygiene hypothesis" (40, 109). Two preliminary studies on the CHILD cohort found increased microbiota richness and diversity when the infant lived with pets, greater

representation of *Clostridiaceae* and *Veillonella* and lower representation of *Bifidobacteria* when living with pets and pet exposure modifying the association between total fecal IgA and breastfeeding (52, 110).

1.5.11 Infant allergic eczema

Infant allergic eczema was considered due to the possibility of reverse causation of postpartum maternal stress. Caregiving an infant with eczema may increase stress on the parents. There is very little literature on the parental experience of caregiving an infant with eczema. One report by Faught et al. (111) was able to find increased stress levels in mothers who were caring for a child with eczema of higher severity at less than six years of age. A more recent study in the Korean population showed lower family quality of life and more negative emotions in the parents associated with increased severity of atopic dermatitis (112). Possible reasons for the increased stress are the child's behaviours and demands when having eczema that increase pressure on parenting, as well as the financial burden for caring moderate to severe eczema (113). Due to the small sample size, large variations of age and possibilities of selection bias in the sample in the two studies, this association requires further investigation.

Infants with allergic conditions have different levels of immune factors. Tendency of lower concentrations of 6-month fecal IgA and other inflammation markers was found in infants with IgE-associated eczema at two years of age, but the difference was not statistically significant (46). A recent study on IgA interactions with gut microbiota explored the differences between allergic or asthmatic children and healthy children, and they found significantly more IgA-bound fecal bacteria in the healthy children at 12 months of age (61).

1.6 Distress and Infant Gut Development

1.6.1 Distress and infant gut development

An animal study by Martinez-Carrillo et al. (114) found reduced plasma B cell counts in Peyer's patches with the administration of dexamethasone, a corticosteroid, which suggests that distress may affect the development of the fetal gut immune system if greater than normal levels of glucocorticoids are passed through the placenta. Maternal glucocorticoids are tightly regulated from passing the placental barrier by the 11β-hydroxysteroid dehydrogenase enzyme. However, in the presence of higher maternal stress levels, some maternal corticosterone can cross the placenta and affect fetal corticosterone levels (115). Mice models of chronic stress reveal alterations of the vaginal microbiome in pregnant mice (116). Lactobacillus abundance was decreased in the vagina of pregnant mice after exposure to chronic stress, and this vaginal abundance was positively correlated to the gut *Lactobacillus* abundance in the offspring. This lowering of abundance Lactobacillus was also seen in pregnant rhesus monkeys (117). A possible reason for this change is that stress during pregnancy increases susceptibility to vaginal infections due to the immunosuppression (118). These infections can lead to responses that deplete the Lactobacillus in the vagina, and potentially affect the early colonization of microbiota in infants when vaginally delivered. However, studies on the impact of prenatal distress on vaginal microbiome in humans are still lacking. Increased stress in the mother may increase infection susceptibility and increase antibiotic use during pregnancy, which may affect microbiome seeding in the infant gut before and during birth (119). Whether prenatal antibiotics use has a sole major impact on seeding is yet to be determined since many metabolic and immune changes occur during pregnancy (84).

1.6.2 Distress and gut immunity

Infant immunity differences have been found when the mother was distressed during pregnancy (120). Therefore, maternal stress and depression may have an impact on the development of the infant gut immune system at early life. Zijlmans et al. (121) was the first to report, based on longitudinal data of 56 infants, human infants born to mothers with greater prenatal anxiety and stress more likely to have gut dysbiosis, an imbalance in gut microbiome composition. Altered infant gut microbiome composition has been associated with the later development of asthma and allergies (119). Although this first report on humans found that stress predisposes allergic symptoms in early life, this study did not have enough power to investigate the influence of breastfeeding, an important covariate, and did not look at immune factors that may lead to changes in gut microbiome composition (121). Since distress has been shown to affect the gut microbiome, maternal distress has potential to affect the gut immunity and IgA levels in infants.

Jarillo-Luna et al. (122) reported lower secretory intestinal IgA concentrations in chronically stressed mice when compared to the control group. Adrenalectomy was performed to determine the role of adrenal glands on IgA levels, and it confirmed adrenal stress hormones, catecholamines and glucocorticoids, play a role in reducing intestinal IgA levels when chronically stressed. A later study from the same group on repeated stress found lower sIgAproducing B cells in the Peyer's patches (114). This suggests that distress may have an influence on the production and release of immune factors in the gut. Currently, there is a paucity of literature on connections between distress and gut immunity in humans.

1.6.3 Summary of gaps in literature

Few studies have investigated the association between distress and gut immunity. Gut microbiome studies in animal models and humans show various links in changes in early life gut composition with prenatal distress that could possibly lead to alterations in infant immunity (116,121). However, the animal models are not representative for the depression and stress that humans experience. A commonly used animal model for depression is the olfactory bulbectomized rat that is used to test antidepressants and other treatments for depression-like symptoms (123), but this model has not been used in the animal studies that investigated gut immunity and microbiome differences with distress. Instead, the animal studies used restraint stress when testing for differences in intestinal immunity (114, 122), which may not be applicable to humans. The animal studies also looked at effects of direct stress and not maternal stress effects on the infant gut immunity.

No studies have investigated the impact of postnatal distress on infant gut microbiome and immunity in humans. Although Zijlmans et al. (121) controlled for postnatal distress in their analyses for prenatal distress and infant gut microbiome composition, the impact of postnatal distress on the infant gut was not tested. This study also did not measure maternal depression, but focused on prenatal stress and anxiety. A recently published study on rat pups by Moussaoui et al. (124) reports differences in plasma corticosterone levels, intestinal permeability and microbial diversity and composition in pups that were exposed to postnatal chronic stress (limited nesting room and brief maternal separation), suggesting that stressful environments postnatally may affect the stress response and gut microbiome development in offspring. There is a gap in literature on the potential effects of maternal pre and postnatal distress specifically on human infant gut immunity, which is an important contributor of gut microbiome development that may

assist the prevention of allergies and asthma. Therefore, this thesis will provide some insight on the impact of pre and postnatal maternal distress on infant gut immunity in the first few months of life.

1.7 Hypothesis and Objectives

The hypothesis for this study is that infants born from mothers that self-report having stress or depressive symptoms during and after pregnancy will have lower fecal secretory IgA than infants born from mothers that experience lower levels of stress or depression. Having longer exposure to distress (during both pre and postnatal periods) may greatly affect sIgA levels. Secondary hypotheses are that distress in the prenatal only period or postnatal only period will also affect gut immunity by lowering sIgA levels, and differences in effects may be observed between pre and postnatal distress exposure.

The objective of this study is to determine whether infants born from mothers that selfreport stress and depressive symptoms during and after pregnancy have lower fecal sIgA than infants born from non-distressed mothers. We are testing this association while controlling for the effects of breastfeeding status, antidepressant use, delivery mode, antibiotics exposure, gravidity, maternal asthma/allergy and overweight status, depression history, tobacco smoke exposure, infant allergic eczema and number of pets and children at home.

1.8 Sample Size Calculation

In an animal study by Jarillo-Luna et al. (122), the stressed group had intestinal IgA concentrations of 1μ g/ml; whereas, the control group had 3μ g/ml of IgA.

Percent decrease:

$$\overline{X}_1 = 3\mu g/m I \, \overline{X}_2 = 1\mu g/m I$$

 $[(3-1)\mu g/ml / 3\mu g/ml] \times 100\% = 66.67\%$

Mean IgA of infants from non-depressed mothers in human sample = 10.74mg/g of protein

Estimated IgA of infants from depressed mothers in human sample = $10.74 - (10.74 \text{mg/g} \times 0.6667) = 3.5809 \text{mg/g}$ of protein

Pooled SD =
$$\sqrt{\frac{(n_1-1)s_1^2 + (n_2-1)s_2^2}{n_1+n_2-2}} = \sqrt{\frac{(88-1)(7.0991)^2 + (25-1)(5.7116)^2}{88+25-2}} = 6.8230$$
mg/g of protein

To determine the sample size with a 2-sided α of 0.05 and β of 0.20 (power=80%), the Power Index (PI) will be: 1.96 + 0.84=2.80

N per group = $[PI * SD/clinical difference]^2$

$$= [2.80*\frac{6.8230mg/g}{(10.74-3.5809)mg/g}]^2 = 7$$

The minimum required number of participants per group is seven.

Chapter 2: Methods and Materials

2.1 Methods overview

This chapter describes the methods and materials used in this thesis study. This study is a quantitative study based on data from sub-sample of the Canadian Healthy Infant Longitudinal Development (CHILD) study, a nationally representative prospective longitudinal birth cohort study.

2.2 Study population

The CHILD study has 3623 enrolled pregnant mothers and full-term infants recruited between 2008 and 2012 from four Canadian cities: Vancouver, Edmonton, Manitoba and Toronto (125). Mothers were followed throughout pregnancy and children are clinically assessed at birth, at three months, and at 1-, 3-, and 5-year visits. Moraes et al. (125) documented the inclusion and exclusion criteria for this cohort study. Pregnant women who are 18 years and older, who lives in residence in reasonable proximity to the delivery hospital and plans to give birth at a designated recruitment center participating hospital, who is able to read, write and speak English and who provides informed consent to the study and cord blood donation were recruited. Their infants were born at or after 35 weeks and the families were able to provide name, address and telephone numbers of two alternate contact individuals. The exclusion criteria for this cohort are children born with major congenital abnormalities or respiratory distress syndrome, expectation of moving away within one year, children of multiple births, children resulting from in vitro fertilization, children who will not spend at least 80% of nights in the home and children born before 35 weeks.

The subsample of 403 infants for this study used data from three of the four study sites (excluding Toronto), and these infants were selected based on availability of the stool samples. Although home births were originally meant to be excluded from the study, some infants born from home births were initially included in our sample. Sensitivity analyses show that including these infants had a major effect on our results, and therefore, these infants (n=12) were excluded from our sample (Appendix D; Table D1). Late preterm infants (35-37 weeks gestation) were also meant to be excluded from the CHILD study, but they were included in our sample because of the low impact on our results (Table D3).

2.3 Exposures

Maternal distress data collected at recruitment (mean 27 weeks) and 36 weeks gestation, and at 6 months and 12 months of child age were used for this study. Mothers with scores above the cut-offs only at 27 weeks or 36 weeks were grouped in the Prenatal group, mothers with higher scores only at 6 months or 12 months of child age were grouped into the Postnatal group and mothers with higher scores in both time periods were group into the Both group. The groups were mutually exclusive.

Perceived Stress

Maternal stress was assessed using the 10-item version of the Perceived Stress Scale (PSS) (126). The items measured how frequent mothers had found their lives stressful, unmanageable and overwhelming in the past month, and the items were scored on a scale ranging from zero (never) to four (very often). The scores for the items are summed, and the higher sums indicate greater stress. Since there are no established criteria when evaluating perceived stress, we used the highest quartile of perceived stress scores to indicate higher stress

and the cut-off was 17. A preliminary analysis used the PSS mean in the entire CHILD cohort (12.96) as the cut-off for higher level of perceived stress, which is close to the mean score of females in the general population of USA (127). However, associations with infant sIgA were not significant with this cut-off (see Appendix B).

<u>Depressive Symptoms</u>

Pre and postnatal depressive symptoms were determined using the 20-item Center for Epidemiologic Studies Depression (CES-D) Scale (128). Mothers reported the frequency of experiencing various depression behaviors, cognitions and affect during the past one week using scores ranging from zero (none of the time or less than one day) to three (most or all the time or five to seven days). The scores were summed, and sums of 16 and greater reflect clinically significant levels of depressive symptoms.

2.4 Definition of potential covariates

Covariates investigated in this study were infant age at stool collection, breastfeeding status at time of stool collection, antidepressant (SSRI) use, tobacco smoke exposure, antibiotic exposure to infant from birth to three months, delivery mode, gravidity, number of children and pets at home, small-for-gestational-age (SGA), pregnancy overweight, maternal asthma or allergy status, depression history and infant allergic eczema. These variables were created from data obtained from the CHILD general cohort questionnaires.

Depression history was determined from mother reports of having depression before pregnancy. *Pregnancy overweight* indicated whether the mother was overweight or obese based on maternal BMI during pregnancy. This information was available for only 304 mothers in our

sample, and so separate analyses on the 304 mother-infant dyads were conducted with pregnancy overweight (Appendix A). *Maternal asthma or allergy status* was based on mother reports of having asthma or allergies during pregnancy. *Antidepressant (SSRI) use* indicated if mothers took selective serotonin reuptake inhibitors (SSRIs) based on prenatal and postnatal maternal medication questionnaires.

Antibiotic exposure from birth to three months determined whether infant was exposed to antibiotics at any time from birth to 3 months of age based on questionnaire data (caesarean section, antibiotics during vaginal delivery, newborn given antibiotics, antibiotics used before the 3-month visit). *Delivery mode* information came from birth chart questionnaires and had three categories: emergency caesarean section (C-section), scheduled caesarean section and vaginal delivery. *Gravida* indicated whether the mothers were primigravida or not. *Small-for-gestational-age (SGA)* was defined as born with birthweight below 10th percentile for corresponding gestational age and gender based on Canadian birthweight for gestational age chart.

Breastfeeding status at time of stool collection indicated whether the infant was exclusively breastfed (exclusively breastfed at time of collection), partially breastfed (breastfed and formula-fed) or had zero breastfeeding (formula-fed). *Infant age at stool collection* was the infant age in weeks during the time the stool samples were collected for the CHILD study 3month visits. *Prenatal smoke exposure* was determined from recruitment questionnaires on maternal health and home environment, and was defined as mothers who smoked during pregnancy or had a household member smoking inside the house. *Postnatal smoke exposure* was defined as infants living with a household member that smokes inside the house based on the 3month home environment questionnaire. *Number of children and pets at home* living with the

infant before the 3-month visit was determined from home environment questionnaires. *Infant allergic eczema* was determined if there was rash in more than one area excluding diaper rash (face, inside elbow, ankle, back of knee, wrist/hand, scalp, other) or if there was a physician diagnosis of atopic dermatitis as reported by the mother.

2.5 Stool collection and storage

Infant stool was collected during 3-month CHILD study visits. Stool samples (5-10g) were collected from a freshly soiled diaper using a sterile spatula, divided into aliquots and stored at -80°C. Freezing of stool only caused marginal decreases in sIgA levels as reported by Forrest et al. (129), and so freezing does not have a significant impact on the quantification of sIgA.

2.6 Extraction and analysis of sIgA

The sIgA ELISA kit from Immundiagnostik AG is used to measure the amounts of fecal sIgA in mg per gram of protein. Immundiagnostik AG is a diagnostics company operating globally to develop and produce immunoassays and other analytical detection methods for medical research (130). After thawing the stool samples, sIgA is extracted with IDK Extract extraction buffer. The stool samples are diluted 1:125 in wash buffer. The diluted patient samples, controls, and 100µl standards are put into a microtiter plate, washed and incubated at 15-30°C. After the wells are aspirated and washed, the microtiter plate is tapped on absorbent paper. 100µl of conjugate is added, and then the samples are incubated at 15-30°C and shaken on a horizontal mixer. After the final washing step and adding TMB substrate, the samples are incubated in a dark room for 10-20 minutes at 15-30°C.

450nm against 620nm as a reference. The results of the microplate reader were multiplied by the dilution factor of 12500. Standards with known concentrations that were provided in the kit were used to create standard curves and determine sIgA concentrations.

2.7 Statistical analyses

The sIgA concentrations were highly skewed, and so nonparametric tests, Mann Whitney U-tests and Kruskal-Wallis tests, were used to detect differences in sIgA medians according to the distress status of the mother. Bonferroni sequential correction was used for critical p-values for multiple comparisons when comparing distressed groups with the reference group. Mothers with both pre and postnatal distress was compared to a critical p-value of 0.05 as determined a priori since our primary hypothesis is to see if distress at both time periods affect immunity (131). Prenatal only and postnatal only comparisons with the reference group were adjusted for two pairs of interest using Bonferroni sequential correction (132). Since the prenatal only group infants had exposure to distress before the time of stool collection whereas the postnatal only group had exposure measure during or after stool collection, it was considered the next hypothesis to be tested and was compared to an adjusted critical p-value of 0.05/2 = 0.025. The postnatal only group comparisons with the reference group had p-values compared to the adjusted critical p-value of 0.05/1 = 0.05. Logistic regression models used sIgA binary outcome variables (yes/no) of lowest quartile, highest quartile, lowest tertile and below median. Models were created using purposeful selection of covariates. Statistical analyses were conducted using IBM SPSS 24 and figures were created using GraphPad Prism 6.

Chapter 3: Results

3.1 Sub-sample Characteristics

The 403 sub-sample did not differ significantly for the majority of potential covariates than all the infants in the three sites of the CHILD cohort (Table 1). Breastfeeding at 3 months was slightly higher in the CHILD cohort compared to the sub-sample; however, 38 infants in the CHILD cohort were missing information for breastfeeding status at 3 months, which may have contributed to the difference in proportions. About 12.2% of women in our sample had clinically significant prenatal only depressive symptoms, 8.7% had symptoms postpartum only and 9.2% had symptoms both pre and postnatally (Table 1). For PSS scores higher than the CHILD mean, 19.5% of all mothers in the sample had stress scores above the mean for prenatal only, 14.7% had postnatal only and 37.4% had both pre and postnatal stress scores above the mean (Table B2). With PSS scores in the highest quartile, 16.7% of mothers fell in this category at prenatal only, 10.2% for postnatal only and 18.7% for both pre and postnatal (Table 1).

Although the stool samples were meant to be collected at three months of infant age, some infants had their stool collected earlier or much later than the proposed time point. The mean of stool collection time was 3.8 months, while the range was 2-8 months. Thirty infants had their stool collected at greater than six months of age, but sensitivity analyses indicate including these infants did not affect our results (Appendix D; Table D2), and thus these infants were included in our sample. Instead of excluding infants, we controlled for age at stool collection in the logistic regression models as shown in Section 3.2.4.

 Table 1. Proportion of depressive symptoms, higher perceived stress and potential covariates in 403 sub-sample and CHILD cohort

	sIgA Sample % (95% CI) N=403	CHILD Cohort (Three Sites*) % N=2373
Depressive Symptoms	1 403	IN 2575
Less (below cut-off)	69.8 (65.3 - 74.3)	66.6
Prenatal Only	12.2(9.0-15.4)	13.8
Postnatal Only	8.7 (5.9 – 11.5)	8.6
Both Pre and Postnatal	9.2 (6.4 – 12.0)	10.9
Higher Perceived Stress (highest quartile)		
Less (below cut-off)	54.4 (49.5 - 59.3)	51.5
Prenatal Only	16.7(13.0-20.4)	18.8
Postnatal Only	10.2(7.2 - 13.2)	12.1
Both Pre and Postnatal	18.7 (14.9 – 22.5)	17.6
Depression History		
Yes	20.0 (16.1 – 23.9)	19.8
No	80.0 (76.1 - 83.9)	80.2
Maternal Asthma/Allergy	``````````````````````````````````````	
Yes	63.9 (59.2 - 68.6)	65.4
No	36.1 (31.4 - 40.8)	34.6
Child Sex		
Male	55.8 (51.0 - 60.6)	52.3
Female	44.2 (39.4 - 49.0)	47.7
Gravida		
Primigravida	38.1 (33.3 - 42.9)	35.8
Multigravida	61.9 (57.1 – 66.7)	64.2
Birth Mode		
Caesarean Section	28.8 (24.4 - 33.2)	24.0
Vaginal	71.2 (66.8 - 75.6)	76.0
Antibiotics Exposure		
Yes	56.4 (51.5 - 61.3)	51.3
No	43.6 (38.7 - 48.5)	48.7
Breastfeeding at 3 Months		
Yes	78.6 (74.6 – 82.6)	84.8**
No	21.4 (17.4 – 25.4)	15.2**
Prenatal Smoke Exposure		
Yes	6.1 (3.7 – 8.5)	7.1
No	93.9 (91.5 - 96.3)	92.9
Postnatal Smoke Exposure		
Yes	16.0 (12.4 – 19.6)	15.7
No	84.0 (80.4 - 87.6)	84.3
Pets at Home		
Yes	46.4 (41.5 – 51.3)	45.2
No	53.6 (48.7 - 58.5)	54.8
Infant Allergic Eczema		
Yes	10.8 (7.7 – 13.9)	13.4
No Since the sub-sample came from three site	89.2 (86.1 – 92.3)	86.6

*Since the sub-sample came from three sites, only infants from the three sites of the entire CHILD cohort were used to compare with the sub-sample.

**Significant difference since percentage is not within the 95% CI calculated for the sub-sample proportions

Table 2. Median infant fecal sIgA levels and percentage distribution of maternal depressive symptoms and lowest quartilesIgA in relation to potential covariates (n=403)

	Less symptoms (69.8% overall)	Prenatal only (12.2% overall)	Postnatal only (8.7% overall)	Both (9.2% overall)		Fecal sIgA		Lowest qua (% yes)	rtile sIgA
	N (%)	N (%)	N (%)	N (%)	p-value (χ^2)	median (IQR)	p-value	N (%)	p-value (χ^2)
Child sex									
Male	159 (71)	25 (11.2)	20 (8.9)	20 (8.9)	0.89	6.1 (3.3 – 12.0)	0.58	54 (24)	0.67
Female	121 (68.4)	24 (13.6)	15 (8.5)	17 (9.6)		5.6 (3.1 - 10.4)		46 (25.8)	
Depression History	40 (50 ()	10 (22 0)	10 (12 7)	11 (12 0)	-0.001	(2(22) 11.4)	0.04	25 (21.0)	0.12
Yes No	40 (50.6) 236 (74.9)	18 (22.8) 29 (9.2)	10 (12.7) 25 (7.9)	11 (13.9) 25 (7.9)	<0.001	6.2(3.3 - 11.4) 5.1(2.3 - 9.3)	0.04	25 (31.6) 74 (23.4)	0.13
Maternal asthma/allergy during pregnancy	250 (74.9)	29 (9.2)	25 (7.9)	23 (7.9)		5.1 (2.5 - 9.5)		74 (23.4)	
Yes	175 (69.2)	33 (13)	18 (7.1)	27 (10.7)	0.19	6.0 (3.3 - 11.0)	0.61	61 (24.1)	0.59
No	101 (71.1)	15 (10.6)	17 (12)	9 (6.3)	0.17	5.7(3.0 - 11.1)	0.01	38 (26.6)	0.57
Delivery mode	101 (/1.1)	15 (10.0)	17 (12)) (0.5)		5.7 (5.0 11.1)		50 (20.0)	
Vaginal	198 (70.2)	30 (10.6)	30 (10.6)	24 (8.5)	0.16	6.1 (3.2 – 11.2)	0.50	70 (24.6)	0.56
Scheduled C-section	47 (69.1)	8 (17)	4 (8.5)	3 (6.4)		5.6(3.5-10.9)		19 (27.9)	
Emergency C-section	32 (68.1)	11 (16.2)	1 (1.5)	9 (13.2)		5.4(3.0 - 10.5)		9 (19.1)	
Antibiotics exposure up to 3 months									
Yes	155 (70.5)	31 (14.1)	16 (7.3)	18 (8.2)	0.28	5.6 (3.1 – 11.2)	0.28	58 (26.2)	0.43
No	120 (70.6)	15 (8.8)	18 (10.6)	17 (10)		6.1 (3.3 – 12.0)		39 (22.8)	
Gravida									
Primigravida	112 (73.7)	19 (12.5)	12 (7.9)	9 (5.9)	0.32	5.1 (2.9 - 10.0)	0.077	43 (28.3)	0.18
Multigravida	165 (67.3)	30 (12.2)	23 (9.4)	27 (11)		6.4 (3.5 – 12.1)		55 (22.3)	
Small-for-gestational-age	20 (74.1)	4 (14.0)	2 (7 1)	1 (2 5)	0.55	75 (15 160)	0.073		0.00
Yes	20 (74.1)	4 (14.8)	2 (7.4)	1 (3.7)	0.77	7.5(4.5 - 16.0)	0.063	3 (11.1)	0.09
No Prenatal SSRI use	256 (69.6)	45 (12.2)	33 (9)	34 (9.2)		5.7 (3.1 - 10.8)		95 (25.7)	
Yes	5 (27.8)	8 (44.4)	0 (0)	5 (27.8)	<0.001	3.6(2.5 - 7.0)	0.12	6 (33)	0.42
No	269 (71.9)	40 (10.7)	34 (9.1)	31 (8.3)	<0.001	6.0(3.2 - 11.4)	0.12	93 (24.8)	0.42
Postnatal SSRI use	207 (71.7)	40 (10.7)	J+ ().1)	51 (0.5)		0.0 (5.2 11.4)		<i>))</i> (24.0)	
Yes	10 (43.5)	8 (34.8)	1 (4.3)	4 (17.4)	0.002	4.6 (3.3 - 7.4)	0.18	6 (26.1)	0.89
No	266 (71.9)	40 (10.8)	31 (8.4)	33 (8.9)		6.0(3.2 - 11.5)		92 (24.8)	
Prenatal smoke exposure		. ()	- ()						
Yes	10 (62.5)	3 (18.8)	2 (12.5)	1 (6.3)	0.78	5.1 (1.8 - 12.3)	0.39	6 (37.5)	0.24
No	203 (72)	32 (11.3)	26 (9.2)	21 (7.4)		6.0 (3.2 – 11.1)		69 (24.4)	
Postnatal smoke exposure									
Yes	31 (67.4)	7 (15.2)	2 (4.3)	6 (13)	0.26	4.9 (2.8 – 9.6)	0.25	14 (30.4)	0.37
No	184 (72.2)	29 (11.4)	25 (9.8)	17 (6.7)		6.1 (3.2 – 11.5)		62 (24.4)	
Pets at home	105 ((0, 0)	22 (11 0)	12 (7)	22 (12 1)	0.00	5.2 (2.5 10.4)		55 (20 5)	
Yes	127 (68.6)	22 (11.9)	13 (7)	23 (12.4)	0.20	5.3(2.7-10.4)	0.031	55 (29.7)	0.045
No Children et herre	151 (70.9)	27 (12.7)	21 (9.9)	14 (6.6)		6.4 (3.7 – 12.0)		45 (21)	
Children at home Yes	135 (71.4)	20 (10.6)	17 (9)	17 (9)	0.79	6.5 (3.6 - 11.9)	0.11	43 (22.6)	0.34
No	133 (71.4) 144 (68.9)	29 (13.9)	17 (9)	19 (9.1)	0.79	5.3(3.1-10.7)	0.11	43 (22.8) 56 (26.8)	0.34
Breastfeeding at stool collection	144 (00.2)	29 (13.9)	1/(0.1)	19 (9.1)		5.5 (5.1 - 10.7)		50 (20.0)	
Zero	64 (64)	15 (15)	11 (11)	10 (10)	0.30	3.4(2.0-5.9)	< 0.001	47 (47)	<0.001
Partial	108 (72)	13 (8.7)	16 (10.7)	13 (8.7)	5.50	5.5(3.2 - 10.0)	0.001	39 (26)	0.001
Exclusive	99 (75.6)	15 (11.5)	6 (4.6)	11 (8.4)		10.1 (5.6 - 16.2)		11 (8.4)	
Infant allergic eczema	,								
Yes	28 (65.1)	4 (9.3)	4 (9.3)	7 (16.3)	0.38	5.9 (3.4 - 11.2)	0.84	9 (20.9)	0.55
No	251 (71.1)	44 (12.5)	28 (7.9)	30 (8.5)		5.8 (3.1 - 11.0)		89 (25.1)	

Table 3. Median infant fecal sIgA levels and percentage distribution of maternal higher perceived stress and lowest quartile sIgA in relation to potential covariates (n=403)

	Less stress (54.4% overall)	Prenatal only (16.7% overall)	Postnatal only (10.2% overall)	Both (18.7% overall)		Fecal sIgA		Lowest quartile sIgA (% yes)	
	N (%)	N (%)	N (%)	N (%)	p-value (χ^2)	median (IQR)	p-value	N (%)	p-value (χ^2)
Child sex									
Male	125 (55.8)	33 (14.7)	22 (9.8)	44 (19.6)	0.64	6.1 (3.3 – 11.7)	0.58	54 (24)	0.67
Female	93 (52.5)	34 (19.2)	19 (10.7)	31 (17.5)		5.6 (3.2 - 10.4)		46 (25.8)	
Depression History									
Yes	31 (39.2)	17 (21.5)	8 (10.1)	23 (29.1)	0.009	6.1 (3.3 – 11.4)	0.04	25 (31.6)	0.13
No	184 (58.4)	49 (15.6)	32 (10.2)	50 (15.9)		5.1 (2.3 – 9.0)		74 (23.4)	
Maternal asthma/allergy during pregnancy									
Yes	136 (53.8)	44 (17.4)	24 (9.5)	49 (19.4)	0.88	6.0 (3.2 - 12.0)	0.49	66 (24.7)	0.99
No	79 (55.6)	22 (15.5)	16 (11.3)	25 (17.6)		5.7 (3.2 - 9.8)		32 (24.8)	
Delivery mode									
Vaginal	157 (55.7)	41 (14.5)	31 (11)	53 (18.8)	0.26	5.6 (3.6 - 10.4)	0.50	70 (24.6)	0.56
Scheduled C-section	26 (55.3)	7 (14.9)	4 (8.5)	10 (21.3)		5.3 (2.9 - 10.4)		19 (27.9)	
Emergency C-section	33 (48.5)	19 (27.9)	5 (7.4)	11 (16.2)		6.2 (3.2 – 11.4)		9 (19.1)	
Antibiotics exposure up to 3 months		. , ,	. ,						
Yes	123 (55.9)	41 (18.6)	16 (7.3)	40 (18.2)	0.25	5.6 (3.1 - 11.0)	0.28	58 (26.2)	0.43
No	92 (54.1)	25 (14.7)	22 (12.9)	31 (18.2)		6.1(3.4 - 12.0)		39 (22.8)	
Gravida	<u> </u>	(··· /	· · · /	()		(
Primigravida	85 (55.9)	27 (17.8)	16 (10.5)	24 (15.8)	0.72	5.2(3.0 - 10.0)	0.077	43 (28.3)	0.18
Multigravida	131 (53.5)	40 (16.3)	24 (9.8)	50 (20.4)		6.4(3.5-12.0)		55 (22.3)	
Small-for-gestational-age	101 (00.0)	10 (10.0)	2. (3.0)	20 (20.1)		0.1 (0.0 12.0)		00 (22.0)	
Yes	14 (51.9)	7 (25.9)	3 (11.1)	3 (11.1)	0.52	7.5 (4.7 – 15.7)	0.063	3 (11.1)	0.09
No	201 (54.6)	60 (16.3)	37 (10.1)	70 (19)	0.02	6.0(3.2 - 10.9)	0.005	95 (25.7)	0.07
Prenatal SSRI use	201 (51.0)	00 (10.5)	57 (10.1)	/0(1))		0.0 (5.2 10.7)		<i>y</i> (20.1)	
Yes	4 (22.2)	6 (33.3)	1 (5.6)	7 (38.9)	0.012	3.8(2.7-6.7)	0.12	6 (33)	0.42
No	208 (55.6)	61 (16.3)	39 (10.4)	66 (17.6)	0.012	6.0(3.2 - 11.0)	0.12	93 (24.8)	0.12
Postnatal SSRI use	200 (00.0)	01 (10.5)	55 (10.1)	00 (17.0)		0.0 (0.2 11.0)		<i>y</i> (21.0)	
Yes	7 (30.4)	7 (30.4)	2 (8.7)	7 (30.4)	0.062	4.7 (3.2 – 7.1)	0.18	6 (26.1)	0.89
No	208 (56.2)	59 (15.9)	38 (10.3)	65 (17.6)	0.002	6.1 (3.2 – 11.4)	0.10	92 (24.8)	0.07
Prenatal smoke exposure	200 (30.2)	57 (15.7)	50 (10.5)	05 (17.0)		0.1 (5.2 11.1)		<i>y</i> ² (21.0)	
Yes	7 (29.2)	7 (29.2)	3 (12.5)	7 (29.2)	0.072	5.1 (1.8 - 12.3)	0.39	6 (37.5)	0.24
No	208 (56.1)	59 (15.9)	37 (10)	67 (18.1)	0.072	6.0(3.2 - 11.1)	0.57	69 (24.4)	0.24
Postnatal smoke exposure	200 (00.1)	57 (15.7)	57 (10)	07 (10.1)		0.0 (0.2 11.1)		(27.7)	
Yes	30 (46.9)	11 (17.2)	5 (7.8)	18 (28.1)	0.17	4.9 (2.8 - 9.6)	0.25	14 (30.4)	0.37
No	188 (56)	56 (16.7)	36 (10.7)	56 (16.7)	0.17	4.9(2.8 - 9.0) 6.1(3.2 - 11.5)	0.25	62 (24.4)	0.57
Pets at home	100 (50)	50 (10.7)	50 (10.7)	50 (10.7)		5.1 (5.2 = 11.5)		02 (24.4)	
Yes	96 (51.9)	31 (16.8)	18 (9.7)	40 (21.6)	0.46	5.3 (2.7 - 10.4)	0.031	55 (29.7)	0.045
No	121 (56.8)	36 (16.9)	23 (10.8)	33 (15.5)	0.40	6.4(3.7-12.0)	0.031	45 (21)	0.045
Children at home	121 (30.0)	50 (10.9)	25 (10.0)	55 (15.5)		0.4(5.7 - 12.0)		45 (21)	
Yes	105 (55.6)	28 (14.8)	21 (11.1)	35 (18.5)	0.76	6.5 (3.6 - 11.9)	0.11	56 (26.8)	0.34
No	112 (53.6)	39 (18.7)	20 (9.6)	38 (18.2)	0.70	5.3(3.1 - 10.7)	0.11	43 (22.6)	0.34
Breastfeeding at stool collection	112 (33.0)	37 (10.7)	20 (9.0)	50 (10.2)		5.5 (5.1 - 10.7)		43 (22.0)	
Zero	49 (49)	17 (17)	13 (13)	21 (21)	0.67	3.3 (2.0 - 6.0)	< 0.001	47 (47)	<0.001
Zero Partial	49 (49) 88 (58.7)	17 (17) 19 (12.7)	13 (13) 14 (9.3)		0.07	5.3(2.0-0.0) 5.4(3.2-9.8)	~0.001		~0.001
				29 (19.3)				39 (26)	
Exclusive	76 (58)	22 (16.8)	12 (9.2)	21 (16)		10.0 (5.6 - 16.1)		11 (8.4)	
Infant allergic eczema	20 (46 5)	0 (20 0)	4 (0.2)	10 (22.2)	0.0	57 (24 11 4)	0.04	0 (20 0)	0.55
Yes	20 (46.5)	9 (20.9)	4 (9.3)	10 (23.3)	0.62	5.7(3.4 - 11.4)	0.84	9 (20.9)	0.55
No	196 (55.5)	57 (16.1)	37 (10.5)	63 (17.8)		6.0 (3.2 – 11.2)		89 (25.1)	

Tables 2 and 3 report the frequency distribution of the exposure, maternal distress, and the outcome of infant fecal sIgA across putative confounding factors. Depressive symptoms and high perceived stress were significantly more common in women with a history of for depression and among those treated with SSRIs during or after pregnancy. Four percent of women with minimal depressive symptoms took SSRIs, while this was 18.8% in the prenatal group, 3.2% in the postnatal group and 16.7% in the pre and postnatal group (Appendix E; Table E1). Significantly lower median fecal sIgA levels were observed in infants born to mothers having no depression history compared to those with history of depression (p=0.04), living with pets compared to no breastfeeding at stool collection time (p<0.001; Table 2 and 3). Borderline significant differences were observed in median sIgA for gravida and small-for-gestational-age (SGA), where infants born to primigravidas and not SGA infants had lower sIgA concentrations. Percentage of those in the lowest quartile for sIgA was significantly different with the pet ownership and breastfeeding status at stool collection time.

3.2 Maternal Distress and Infant sIgA

Due to the skewness of the sIgA data, comparisons between the distress groups and the reference group were conducted for median sIgA levels. Infant sIgA levels were significantly lower in the infants exposed to both pre and postnatal depressive symptoms compared to the reference group (p=0.033; Figure 1). Median sIgA levels were 6.3 (IQR=3.6 - 12.4) mg/g feces for exposure to less depressive symptoms (reference group) and 4.4 (IQR = 2.4 - 8.0) mg/g feces for exposure to both pre and postnatal symptoms (see Appendix E). Infants in the prenatal only group had median sIgA levels of 5.19 (IQR=2.02 - 9.78) mg/g feces, and those in the postnatal only group had median levels of 5.69 (IQR=2.62 - 8.56) mg/g feces; however, statistically

significant differences in these levels were not detected in relation to the reference group. There were no significant differences in sIgA levels among the perceived stress groups.

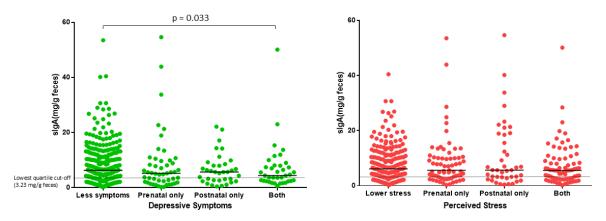


Figure 1. Infant fecal sIgA levels depending on distress status of mother. Secretory IgA levels were significantly lower in the group exposed to both pre and postnatal depressive symptoms (green) compared to the group exposed to lower symptoms (not clinically significant). No significant differences were seen in sIgA with maternal higher perceived stress status (red). Lowest quartile cut-off for sIgA used in logistic regression models was at 3.23 mg/g feces (dotted line).

Sections 3.2.1 to 3.2.3 contain the median comparisons for sIgA with exposure to both pre and postnatal (primary question), prenatal only and postnatal only maternal distress respectively after stratifying for covariates. The final section shows the simple and multiple logistic regression analyses using lowest quartile sIgA (yes or no) as the outcome (Table 4 and 5). Logistic regression analyses using highest quartile, lowest tertile and below median sIgA cut-offs are listed in Appendix C. 3.2.1 Infant sIgA with exposure to both pre and postnatal distress (primary question)

After stratifying for breastfeeding status at stool collection time, significantly lower sIgA levels were seen in formula-fed infants with exposure to both pre and postnatal depressive symptoms compared to the reference group (p=0.033; Figure 2). No significant differences were seen in the breastfed infants between the depressive symptoms groups and the reference group. Breastfed infants not exposed to clinically significant depressive symptoms had higher sIgA than formula-fed infants born to mothers experiencing both pre and postnatal depressive symptoms (p<0.001; Figure 2). Median sIgA was not significantly different between perceived stress groups after breastfeeding stratification, but similar trends were seen (Table E4).

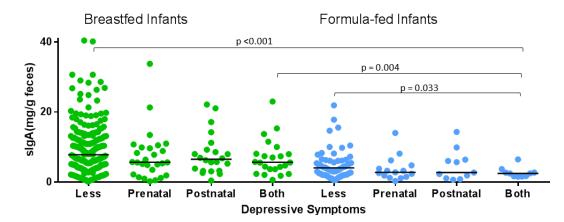


Figure 2. sIgA levels among breastfed and formula-fed infants at time of stool collection for different depressive symptoms groups. Breastfed infants (green) did not have significant differences in sIgA levels based on distress levels, while formula-fed infants (blue) had significantly lower sIgA when exposed to both pre and postnatal depressive symptoms. Formula-fed infants had significantly lower sIgA levels than breastfed infants for all depressive symptoms groups.

After stratifying by other covariates, there were many instances of significantly lower infant sIgA levels when the mother experienced both pre and postnatal distress. This was the case when the infant was exposed to tobacco smoke after birth (p=0.005), exposed to antibiotics (p=0.027), born to a mother with no depression history (p=0.021), born to a multigravida (p=0.022) and living in a home with other children (p=0.038; Figure 3). Figures 5 and 6 in Section 3.2.2 and 3.2.3 also show statistically significant differences between the both group and the reference group. The presence of pet ownership (p=0.022; Figure 5) and maternal asthma or allergy during pregnancy (p=0.032; Figure 6) showed the same lowering of infant sIgA in the presence of both pre and postnatal depressive symptoms. Interactions for the previously mentioned covariates were significant except for antibiotics and maternal asthma or allergy (Table E1). Subsequent stratifications are listed in Appendix E.

Lower median sIgA levels were found with the presence of higher perceived stress in mothers when the dyad was living with other children (p=0.004) and pets at home (p=0.041; Figure 4). Interactions were significant for both covariates in the association between sIgA and higher stress (Table E4). With subsequent stratifications listed in Table E4, lower sIgA was seen with the exposure of both pre and postnatal stress in breastfed infants that live with other children (p=0.025) and in infants exposed to antibiotics that live with pets (p=0.006). With or without antibiotics exposure, having other children in the household had infants with significantly lower sIgA levels after exposure to both pre and postnatal stress than the reference group (Table E4). In the absence of pets, infants living with other children had lower sIgA levels (p=0.037), while infants living without other children had higher sIgA levels when their mom experienced higher pre and postnatal stress (p=0.015; Table E4).

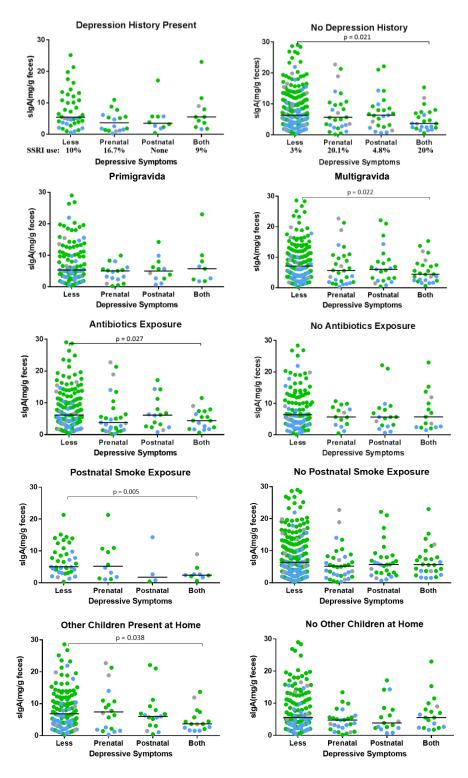


Figure 3. sIgA levels among depressive symptoms groups after stratification for potential covariates. Breastfed infants are in green and exclusively formula-fed infants are in blue. 2-10 points were omitted in each graph for having an sIgA level greater than the scale.

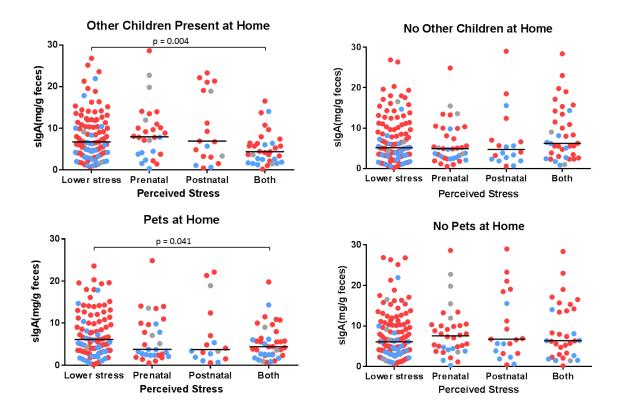


Figure 4. Infant sIgA levels among higher perceived stress groups after stratification for other children and pets living at home. Statistically significant differences were seen between infants exposed to both pre and postnatal higher maternal stress compared to the lower stress group (reference group). Breastfed infants are in red, exclusively formula-fed infants are in blue and infants with unknown breastfeeding status at time of stool collection are in gray. 4-7 points were omitted in each graph for having sIgA greater than the scale.

3.2.2 Infant sIgA with exposure to only prenatal distress

For the group of mothers who experienced clinically significant depressive symptoms during the prenatal period only, a significantly lower median value of sIgA than the reference group was seen with pet ownership, and significance remained in this group of infants when exposed to antibiotics and born to a primigravida (Figure 2, Table E1). Significant interactions were found with pet ownership (Table E1). In infants born to mothers that reported perceived stress in the highest quartile, no significant differences in sIgA levels were seen compared to the reference group (Table E4).

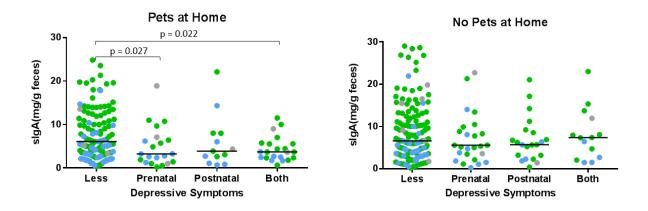


Figure 5. Infant sIgA levels based on maternal depressive symptoms groups after stratification for pet ownership. With pet ownership, prenatal only and both pre and postnatal groups had significantly lower sIgA levels than the reference group. No differences were seen among the depressive symptom categories with the absence of pets. Green data points indicate breastfed infants during the time of stool collection, blue indicates exclusively formula-fed infants and gray indicates unknown breastfeeding status. 2-10 points were omitted for having sIgA greater than the scale.

3.2.3 Infant sIgA with exposure to only postnatal distress

Sex differences were seen with the exposure to only postnatal depressive symptoms. Figure 6 shows significantly lower infant sIgA levels with exposure to postnatal depressive symptoms compared to the reference group in male infants (median (IQR)=4.56(2.00 - 7.99)) mg/g feces versus 6.62 (3.53 - 13.15) mg/g feces), but this difference was not observed in female infants. After stratifying by antibiotics exposure up to 3 months of age, the male infants with no antibiotics exposure still showed this significant difference (p=0.037; Table E1); however, the number of males in this category was small. Additionally, when the mother had asthma or allergy during pregnancy, a statistically significant difference was seen in infant sIgA levels with lower levels after exposure to postnatal depressive symptoms (p=0.041; Figure 6). Infants born to mothers with asthma or allergy continued to have significantly lower sIgA levels with exposure to postnatal depressive symptoms when they had antibiotics exposure (p=0.02) and were living with pets (p=0.014; Table E1). Interestingly, the differences with depressive symptoms were not seen with higher perceived stress (Table E4). Higher sIgA concentrations were seen with exposure to postnatal higher stress when the infant was breastfed and had antibiotics, when exposed to antibiotics and pets and when the mother was primigravida and living with pets (Table E4). Since the counts were small in these stratification groups, these results should be taken with caution.

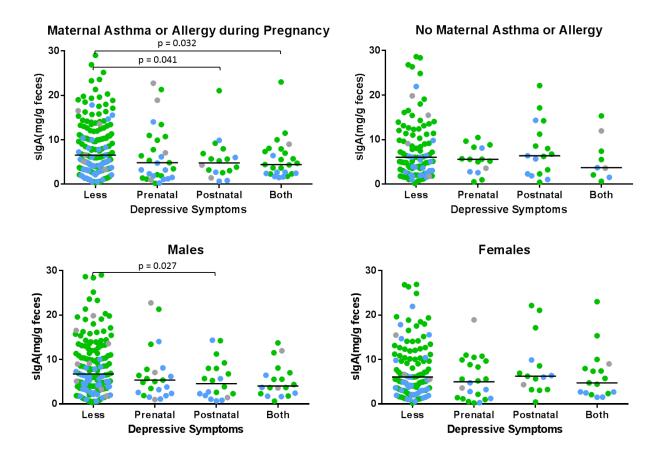


Figure 6. Infant sIgA levels based on maternal depressive symptoms groups after stratification for maternal asthma or allergy during pregnancy and child sex. In infants born to mothers with asthma or allergy and in male infants, the postnatal only group had significantly lower sIgA levels than the reference group. No differences were seen among the depressive symptom categories in the absence of maternal asthma or allergy and in female infants. Green data points indicate breastfed infants during the time of stool collection, blue indicates exclusively formula-fed infants and gray indicates unknown breastfeeding status. 2-10 points were omitted for having sIgA greater than the scale.

3.2.4 Simple and multiple logistic regression analyses with lowest quartile sIgA

A preliminary logistic regression analysis, shown in Appendix C, looking at the odds of being below median for sIgA levels showed significant odds ratios with only prenatal depressive symptoms when adjusting for antibiotics exposure (1.96, 95% CI (1.03 - 3.73)) and collectively adjusting for antibiotics, breastfeeding, pets and gravida (2.57, 95% CI (1.17 - 5.65); Table C1). Highest quartile IgA was also used as the outcome variable in, and the only significant OR was found after controlling for breastfeeding status at time of stool collection with the exposure of prenatal depressive symptoms (Table C2). With depressive symptoms during the prenatal period only, there was a 68% reduction in odds of having infant sIgA levels in the highest quartile (Table C2). Although a first look at the sIgA data suggested more differences detected between higher distress and lower distress when categorizing sIgA into tertiles (Table B1), the final regression models using lowest tertile did not detect many significant odds ratios (Table C4).

Instead of using lowest tertile of sIgA for the outcome, we used lowest quartile for the main regression analyses, which looks at the odds of the infant having sIgA levels in the lowest quartile depending on the maternal distress status. The crude OR for both pre and postnatal depressive symptoms is 2.12 (95% CI: 1.02 - 4.42; Table 4), which means with the exposure of maternal depressive symptoms, the likelihood of the infant having sIgA levels in the lowest quartile is 2.12 times greater than when there are less symptoms (below cut-off). The OR lost significance after single adjustment for the pet ownership, postnatal smoke exposure and prenatal SSRI use. After including covariates of confounding influence and statistically significance at p<0.20 in various combinations in the multiple logistic regression models, the final models were created. Final regression model for low sIgA showed a 3.07 times increase in the odds of having fecal sIgA in the lowest quartile with the exposure of higher pre and postnatal depressive

symptoms than less symptoms after multiple adjustment for infant age, antibiotics exposure,

maternal asthma or allergy, prenatal SSRI use, pets, breastfeeding and gravida (95% CI: 1.25 -

7.55; Table 4).

Table 4. Crude and adjusted odds ratios (OR) of low sIgA for exposure to maternal
depressive symptoms

(Ref: less depressive symptoms)	OR (95% CI) for pre and postnatal	OR (95% CI) for prenatal only	OR (95% CI) for postnatal only	
Crude	2.12 (1.02 – 4.42)	2.08 (1.08 - 4.01)	2.31 (1.10 – 4.87)	
Adjusted for age (quartiles)	2.35 (1.11 - 4.97)	1.88 (0.96 - 3.67)	2.12 (0.99 - 4.55)	
Adjusted for maternal asthma or allergy during pregnancy	2.27 (1.08 – 4.76)	2.18 (1.12 – 4.21)	2.28 (1.08 - 4.82)	
Adjusted for antibiotics exposure	2.39 (1.13 - 5.05)	2.30 (1.18 - 4.49)	2.22 (1.03 – 4.78)	
Adjusted for gravida (primigravida or not)	2.44 (1.15 - 5.16)	2.17 (1.12 – 4.20)	2.46 (1.16 - 5.21)	
Adjusted for birth characteristics (gravida, antibiotics)	2.56 (1.20 - 5.45)	2.36 (1.20 - 4.62)	2.29 (1.06 - 4.94)	
Adjusted for pets at home	1.96 (0.93 – 4.12)	2.08 (1.08 - 4.03)	2.51 (1.18 - 5.35)	
Adjusted for home environment (pets and children at home)	1.81 (0.85 – 3.86)	2.05 (1.06 - 3.96)	2.50 (1.17 – 5.34)	
Adjusted for prenatal smoke exposure	2.15 (1.01 – 4.55)	1.96 (1.00 – 3.96)	2.33 (1.10 - 4.95)	
Adjusted for postnatal smoke exposure	2.00 (0.95 - 4.21)	1.96 (1.01 - 3.81)	2.48 (1.16 - 5.26)	
Adjusted for breastfeeding at stool collection	2.39 (1.06 - 5.39)	2.21 (1.05 - 4.65)	1.78 (0.79 – 4.01)	
Adjusted for prenatal SSRI	2.11 (1.00 - 4.48)	1.86 (0.94 - 3.70)	2.36 (1.12 - 5.01)	
Adjusted for postnatal SSRI	2.11 (1.01 – 4.41)	2.15 (1.10 - 4.21)	2.01 (0.92 - 4.41)	
Adjusted for postnatal SSRI and breastfeeding	2.41 (1.06 - 5.48)	2.37 (1.10 - 5.10)	1.52 (0.64 - 3.58)	
Adjusted for breastfeeding and prenatal smoke	2.42 (1.06 - 5.52)	2.26 (1.05 - 4.83)	1.81 (0.80 - 4.09)	
Adjusted for age, breastfeeding, antibiotics, postnatal SSRI, maternal asthma/allergy, pets, gravida (final model for postnatal only)	3.33 (1.36 - 8.19)	2.82 (1.23 - 6.47)	1.43 (0.57 – 3.60)	
Adjusted for age, antibiotics, maternal asthma/allergy, pets, breastfeeding, gravida, prenatal SSRI (final model for prenatal only and both)	3.07 (1.25 – 7.55)	2.44 (1.07 – 5.57)	1.93 (0.80 – 4.68)	
Final model for prenatal only and both with interaction term for antibiotics	5.04 (1.32 - 19.26)	0.70 (0.16 - 3.06)	3.13 (0.83 - 11.78)	

With the same model, the OR for prenatal depressive symptoms only was 2.44 (95% CI: 1.07 - 5.57). Adjusting for most of the study covariates individually did not affect the statistical significance of the odds ratios for prenatal depressive symptoms, but when adjusting for prenatal SSRI use and smoking in mothers and infant age, significance was lost (Table 4). The interaction term between depressive symptoms and antibiotics exposure from birth to three months of age was significant (Table E3), and so when the interaction term was added to the model, the OR for prenatal only lost significance. However, the OR for both pre and postnatal raised to 5.04 (95% CI: 1.32 - 19.26) with the addition of the interaction in the final model. With the exposure of only postnatal depressive symptoms, we found a crude odds ratio of 2.31 (95% CI: 1.10 - 4.87), which means infants have 2.3 times greater odds of having sIgA levels in the lowest quartile. The odds ratio maintained significance until we controlled for breastfeeding and postnatal SSRI use in the mothers (Table 4). The final model did not have a significant odds ratio for postnatal depressive symptoms.

The crude OR for pre and postnatal higher stress (highest quartile) in mothers was not statistically significant (OR=1.65, 95% CI: 0.92 - 2.98; Table 5). The odds ratio only became significant with the adjustment of gravida and antibiotics exposure (OR=1.87, 95% CI: 1.03 - 3.41). Therefore, after controlling for gravida and antibiotics, the odds of having low sIgA is 1.87 times greater when having higher stress during the perinatal period than having lower stress (reference group). The final multivariate model that adjusts for infant age, breastfeeding, antibiotics, postnatal SSRI and maternal asthma or allergy did not have a significant with the exposure to higher prenatal only stress and higher postnatal only stress (Table 5).

Table 5. Crude and adjusted odds ratios (OR) of low sIgA for exposure to maternal higher perceived stress

(Ref: lower stress)	OR (95% CI) for pre and postnatal	OR (95% CI) for prenatal only	OR (95% CI) for postnatal only	
Crude	1.65 (0.92 - 2.98)	1.37 (0.73 – 2.58)	1.74 (0.83 – 3.62)	
Adjusted for child age	1.44 (0.80 – 2.58)	1.27 (0.63 – 2.54)	1.28 (0.61 – 2.71)	
Adjusted for maternal asthma or allergy during pregnancy	1.63 (0.90 – 2.97)	1.42 (0.75 – 2.67)	1.82 (0.87 – 3.81)	
Adjusted for antibiotics exposure	1.82 (1.00 – 3.30)	1.40 (0.74 - 2.65)	1.59 (0.73 – 3.46)	
Adjusted for gravida (primigravida or not)	1.76 (0.97 – 3.19)	1.39 (0.87 – 2.22)	1.63 (0.77 – 3.46)	
Adjusted for birth characteristics (gravida, antibiotics)	1.87 (1.03 – 3.41)	1.40 (0.74 – 2.65)	1.59 (0.73 - 3.48)	
Adjusted for pets at home	1.64 (0.90 - 2.98)	1.36 (0.72 – 2.56)	1.74 (0.83 - 3.63)	
Adjusted for home environment (pets and children at home)	1.57 (0.86 – 2.86)	1.33 (0.70 – 2.51)	1.74 (0.83 – 3.63)	
Adjusted for prenatal smoke exposure	1.75 (0.97 – 3.17)	1.43 (0.76 – 2.71)	1.82 (0.87 – 3.81)	
Adjusted for postnatal smoke exposure	1.51 (0.82 – 2.76)	1.35 (0.72 – 2.55)	1.77 (0.84 – 3.69)	
Adjusted for breastfeeding at stool collection	1.44 (0.76 – 2.74)	1.72 (0.86 - 3.44)	1.70 (0.77 – 3.78)	
Adjusted for prenatal SSRI	1.52 (0.83 – 2.78)	1.30 (0.69 – 2.45)	1.73 (0.83 – 3.63)	
Adjusted for postnatal SSRI	1.62 (0.89 – 2.96)	1.38 (0.73 – 2.61)	1.38 (0.73 – 2.61)	
Adjusted for postnatal SSRI and breastfeeding	1.45 (0.74 – 2.81)	1.70 (0.84 - 3.44)	1.46 (0.65 - 3.31)	
Adjusted for breastfeeding and prenatal smoke	1.53 (0.80 - 2.94)	1.87 (0.93 – 3.79)	1.76 (0.79 – 3.93)	
Adjusted for breastfeeding and postnatal smoke	1.38 (0.71 – 2.67)	1.69 (0.84 - 3.41)	1.71 (0.77 – 3.79)	
Adjusted for age, breastfeeding, antibiotics, postnatal SSRI, maternal asthma/allergy (final model for perceived stress)	1.39 (0.73 – 2.65)	1.26 (0.59 – 2.69)	0.97 (0.40 - 2.32)	

Chapter 4: Discussion and Conclusions

This final chapter will summarize the findings that were mentioned in Chapter 3 and interpret the results to see how it relates to other studies. The strengths and limitations of the study design mentioned in Chapter 2, as well as clinical relevance and directions for future research are also outlined in this chapter.

4.1 Summary and interpretation of the findings

4.1.1 General overview

We tested for possible associations between pre and postnatal maternal distress and infant fecal sIgA in humans based on a sub-sample of 403 infants from a prospective birth cohort. We found that after higher exposure or clinically significant exposure to maternal perceived stress and depressive symptoms during both the pre and postnatal period, infant fecal sIgA levels in the first few months of life were significantly lower compared to the reference group. This result is consistent with the sole animal study that found that chronic restraint stress lowered total intestinal IgA in mice (122). Maternal prenatal stress affects the maternal gut and vaginal microbiome, which are speculated to affect the colonization of microbiota in the infant gut during the pregnancy and birth process (15). Although a first report on humans has shown prenatal maternal stress associated with gut dysbiosis in infants linked to the development of increased gastrointestinal symptoms and allergic reactions (121), whether the stress was also affecting gut immunity in infants has not been previously investigated. Our results show that distress may lower infant fecal sIgA levels, which is a vital component in the first-line of immune defense and development of the gut microbiota composition in early life. A possible reason for the low levels of sIgA is that distress may affect the gut immune system development

during pregnancy. Along with this change, delivery or factors that co-occur with distress in the perinatal period may affect colonization of microbes that favour sIgA production, such as an altered stress response seen in infants of distressed mothers.

4.1.2 Time-specific distress effects

Median comparisons with covariate stratifications suggest interaction effects of covariates with time and type of distress, such as pet ownership for prenatal only depressive symptoms, maternal asthma or allergy and infant sex for postnatal only depressive symptoms and breastfeeding status, depression history, gravida, antibiotics, tobacco smoke exposure, other children and pets for both pre and postnatal distress. The different results found with the time period and duration of distress emphasizes that stress and depression effects on infant gut health have time-dependent variations, and other factors related to the association that co-occur with distress during a specific period may affect the association between distress and infant gut immunity. Lee et al. (133) found dose-response relationships between maternal stress and childhood asthma for prenatal and postnatal periods respectively in a prospective birth cohort and found that higher stress in both pre and postnatal periods increased the odds of diagnosis of asthma in girls. Our results support the temporal-specific effects of distress on infant gut immunity, which suggest a need for greater understanding on the different mechanisms in different periods and how the prenatal stress combined with postnatal stress works to affect infant immunity and risk of developing allergic diseases.

4.1.3 Pre and postnatal distress effects on infant sIgA levels

In the logistic regression analyses, the odds ratio for higher exposure to both pre and postnatal depressive symptoms lost significance after controlling individually for pets, other children, postnatal smoke exposure and prenatal SSRI use, but only pets and prenatal SSRIs stayed in the final model. Breastfeeding, when combined with postnatal tobacco smoke exposure or SSRI use, made the smoke and SSRI variables have little influence on the odds ratios, suggesting that breastfeeding status may explain for the influence of tobacco smoke exposure and SSRI use on the association. Smoke and SSRIs may affect the breastmilk composition or breastfeeding status or duration due to their associations with distress. The final model collectively controlled for infant age, antibiotics, maternal asthma or allergy, breastfeeding, pets and gravida, suggests that despite controlling for the many study covariates, there appears to be a unique association between pre and postnatal depressive symptoms and low infant sIgA. For instance, although being born to a mother with asthma or allergy could cause immunological differences and allergic responsiveness in the infant as shown in Section 1.5.1, regardless of this status infants still appear to have higher risk of low sIgA when their mom has clinically significant perinatal depressive symptoms.

The odds ratio for higher perceived stress both pre and postnatally in mothers was only significant after controlling for gravida and antibiotics, suggesting that these covariates may mask the association between perinatal stress and infant IgA. In other words, if the mother-infant dyads had the same gravida status and antibiotics exposure status, a significant association between higher stress and low sIgA would be observed. No other odds ratios were found with higher perceived stress and low sIgA, even for the prenatal only and postnatal only odds ratios. This may have happened because of the higher levels of sIgA seen with higher stress in some of

our median comparisons. Perhaps more infections occurred in the infants of these groups, which may elevate the fecal IgA levels (101), but the number of infections was not determined in our study. Another explanation may be other physiological processes occurring with stress, and some animal models of acute stress have shown upregulation of immune cells (134). Restraint stress in rats have shown increased levels of IgA and increased mRNA expression of pIgR, the polymeric immunoglobulin receptor that is important for sIgA transcytosis into the gut lumen (134).

The combination of the prenatal distress affecting fetal stress response and gut immune development along with environmental circumstances co-occurring with distressed mothers during the postpartum period may have greater impact on the infant gut immunity than having only prenatal distress or only postnatal distress in the first few months of life. Maternal stress appears to affect the programming of the fetal hypothalamic–pituitary–adrenal (HPA) axis, as maternal stress seems to stimulate corticotrophin-releasing hormones that elevate fetal secretion of glucocorticoids to excess levels (135). One possible mechanism for this programming may be from methylation of fetal glucocorticoid receptors (136). This change in infant stress response may affect the infant immune response to environmental factors that occur with postpartum distress. Maternal distress usually does not end at birth and continues during postpartum (15), possibly impairing the interaction between mother and infant and the stress and immune response of offspring (22, 137). Offspring have elevated cortisol levels when the mother is distressed during the early months in both animal and human studies (135).

Depressed mothers have been known to interact less with their infant and are less likely to breastfeed their child, which may make the infant more vulnerable to infections (79). No significant differences in sIgA were observed in breastfed infants among the maternal depressive symptoms groups compared to reference but differences were seen in formula-fed infants. This

may indicate the protective effects of breastfeeding against the impact of depressive symptoms on infant gut immunity. Breastfeeding has been associated with higher levels of total fecal IgA in the infant (52), and is associated with the reduced risks of allergic disease (39, 59). Interventions encouraging breastfeeding in depressed mothers may allow for sufficient maturation of the infant immune system and reduce the risk of developing allergic diseases. However, it should be acknowledged that breastfeeding is not a simple task, and it could cause more stress and frustration on mothers. In addition, breastfeeding was controlled in the final logistic regression models for lowest quartile sIgA, which suggests that depressive symptoms still affect infant gut immunity regardless of the infant's diet in the early months.

4.1.4 Prenatal only distress effects on infant sIgA levels

For the prenatal only analyses, we found that exposure to depressive symptoms at prenatal only increased the crude odds of having low fecal sIgA in infants by 2.1 times. Prenatal SSRI use in mothers was the only covariate that made the odds ratio lose significance, suggesting that serotonin levels in mothers during pregnancy may affect the fetal development of the gut immune system. This confirms the suggested immunomodulatory effects of SSRIs as previously discussed in Section 1.5.7. Serotonin has a negative association with inflammation in depressed patients, and there are serotonin receptors in blood cells involved in immunity, B cells, T cells, and dendritic cells, which are involved in the class switching and secretion of sIgA (138). However, the multiple subtypes of serotonin receptors may cause serotonin to have various inhibitory or stimulatory effects on the gut immune system. The final logistic regression model was the same as the model for both pre and postnatal depressive symptoms, and the statistically significant odds ratio indicates 2.4 times greater likelihood of having low sIgA after higher

exposure during the prenatal period. Even after taking into account the SSRI use and other covariates, the association between higher prenatal depressive symptoms and gut sIgA levels remained significant, suggesting that the physiology of depressed mothers may have influence on the fetal gut immunity development. Murine models have shown lower plasma B cell counts after direct exposure to restraint stress due to the effects of glucocorticoids and catecholamines (114), and these hormones were responsible for the reduction of intestinal IgA (122). Although catecholamines have not been associated with maternal perceived distress (stress and anxiety) (139), one particular glucocorticoid that is associated with maternal stress, corticosterone, at elevated levels is a strong Th2 cytokine inducer and increases allergic responses in the offspring after it crosses the placenta (63).

An interesting finding was the interaction between prenatal depressive symptoms and antibiotics exposure from birth to 3 months of infant age. In another study, there was a significant reduction in the odds ratio for prenatal distress and childhood asthma at age seven when adjusting for antibiotics use during infancy (140). Once the interaction was added to our final logistic regression models, the prenatal only OR greatly reduced and lost statistical significance, while the opposite occurred for the pre and postnatal depressive symptoms OR. This result may suggest the difference in gut immunity of infants born to a mother that experiences prenatal only and both pre and postnatal depressive symptoms. Antibiotics use for those in the prenatal only group may explain for the association between prenatal distress and low sIgA, but this is not the case for the both pre and postnatal group.

4.1.5 Postnatal only distress effects on infant sIgA

The odds ratio for only postnatal depressive symptoms became non-significant after individually controlling for breastfeeding, postnatal SSRI use and tobacco smoke exposure, indicating that postnatal depressive symptoms seem to not have a unique association with fecal sIgA after the effects of the covariates are removed. The final model for postnatal depressive symptoms, which adjusted for infant age, antibiotics exposure, maternal asthma/allergy, postnatal SSRI, breastfeeding and gravida, was not statistically significant. The odds ratios for postnatal higher stress remained non-significant for both crude and adjusted.

SSRI use and living with a household member that smokes, both which are more common with maternal depression (141), may be the factors that affect gut immunity. SSRIs can be passed through the breastmilk and have been related to some neonatal health outcomes (18), and our model shows that it may be part of the explanation for the lower sIgA levels with postnatal depressive symptoms. Not only does serotonin affect the immune system, but it also works closely with the gut microbiota, since some gut microbes contribute to the serotonin system and the serotonin system can alleviate the cognitive impairment effects of antibioticinduced gut dysbiosis (142, 143). Percent SSRI use may be an explanation for the differences seen with depression history (see Figure 3). Lower percent SSRI use was seen in women with less depressive symptoms who did not have depression history compared to women with less symptoms who did have depression history. The number of women who did take SSRIs pre and postnatally was low (n < 30); therefore, median comparisons could not be made with stratification of SSRI use. The findings with SSRIs should be taken with caution, as inadequate adjustment of SSRI use may have occurred in the regression models due to the low number of SSRI use. In addition, the effect of tobacco smoke exposure was not significant when breastfeeding status was

added to the model. Women living with a smoker may be more likely to be depressed, which may affect their breastfeeding status, or tobacco smoke may change the breastmilk content and sIgA levels (105). Therefore, breastfeeding status may be the primary explanation for why there are lower fecal sIgA levels in infants that are exposed to postnatal depressive symptoms only.

4.1.6 Birth characteristics and home environment impacting on association of interest

As we hypothesized, birth characteristics and the environment appeared to affect the relationship between distress and fecal sIgA. Although new mothers have been documented to have greater depressive symptoms (20, 79), we found significantly lower sIgA after depressive symptoms exposure in multigravidas. Other birth characteristics, antibiotic exposure and non-SGA were also associated with lower sIgA levels possibly due to the gut dysbiosis caused by antibiotics (144) and increased levels of IgA secreted in breastmilk in SGA infants (145). The presence of other children in the house is related to the number of pregnancies of the mother, but we were able to see separate effects on sIgA from the presence of children in the house after exposure to maternal stress with the median comparisons. Even though pets are seen as beneficial for psychological health in adults (146), there were significant associations between depressive symptoms and lower infant sIgA levels in the presence pets. This is contrary to the "hygiene hypothesis" and studies that revealed pets and other children increasing sIgA levels in breastmilk and exposure to pets inducing total IgA production in non-breastfed infants to similar levels of IgA as breastfed infants (52, 59, 109). This may suggest that the presence of pets and other children may not have protective effects against maternal distress affecting infant gut immunity. The increased exposure to microbes in early life may cause competition and reduce the abundance of microbes that are stimulatory of sIgA. Lower Lactobacillus reuteri and

Bifidobacteria have been found with pet ownership (40,110), which are stimulators of sIgA. However, most of these variables were controlled for in the logistic regression analyses, and we still found significant associations between maternal depressive symptoms and infant gut sIgA.

4.2 Strengths of study

Although stress and gut microbiome and immunity have been studied in animal models, this is the first study to ever report associations between maternal distress and the development of gut immunity in humans. Since animal models are beneficial to determine mechanisms but sometimes not applicable to humans, our study greatly adds suggestions to the literature on the links between maternal psychosocial health and the gut immune maturation in early life. As having a trial testing this association would violate ethics, the prospective cohort design of the CHILD study allows us to investigate the exposure-outcome relationship of our question of interest in humans.

Although causality cannot be concluded with our study design, the prospective CHILD birth cohort can support the temporality of the relationship between exposure of perinatal distress and outcome of infant sIgA levels in the first few months of life. The cohort is a nationally representative birth cohort for Canada, and our sub-sample seems to be representative of the entire cohort based on Table 1. This sufficient representation of the country makes our sample more generalizable to the Canadian population. Since the cohort study was able to recruit over 3500 mothers, we were able to investigate with a sizeable sample and conduct various stratification analyses with potential covariates. The CHILD cohort obtained multifarious

information on home environment, maternal medications, maternal and infant health, birth characteristics and breastfeeding that allowed our study to look into many covariates.

Many previous studies used negative or stressful life events as a measure of parental distress. In our study, mothers reported perceived stress and depressive symptoms, which would indicate whether life stressors did affect the mother's psychosocial health, as opposed to relying on reports of negative life events that have uncertainty as to whether there was an experience of stress. Our distress measures were both from validated questionnaires that have been used widely in the research and clinical community (126, 128, 147).

We used single and multiple logistic regression analyses to statistically control for potential confounding variables and covariates for our question of interest. Since stress and depressive symptoms may co-occur with various other factors that affect infant gut immunity, such as smoking, antidepressant use, less breastfeeding and lower quality of relationship between mother and child, controlling for covariates may strengthen the validity of the association that was studied. In the final multivariate models, we were able to still find significant adjusted odds ratios when the mother had prenatal only depressive symptoms and when the mother had both pre and postnatal depressive symptoms.

4.3 Limitations of study

One limitation is that our stool samples were analyzed based on availability from the three study sites, which may have led to selection bias in our sample. Although our sub-sample was representative of the rest of the CHILD cohort for most of the covariates that we analyzed, there is the possibility that residual confounding that was not controlled. For instance, the

pregnancy overweight variable was only available for 304 infants in the sub-sample, and so we could not control for this variable in the final model. Covariates self-reported by mothers may be subject to reporting bias, such as maternal distress, maternal asthma and allergy, medications, smoke exposure, breastfeeding, depression history and the home environment. We did not have access to information about whether the infant was experiencing stress, if there was supplementation of prebiotics and probiotics, the quality of relationship and interactions between mother and child, the cleanliness of the home, the availability of social support for the mothers and the use of psychological treatments for distressed mothers (*e.g.*, Cognitive Behavior Therapy).

Our distress measure is not without limitations. The concept of distress includes anxiety along with depression and stress, but our study only measured perceived stress and depressive symptoms. A measure for maternal anxiety was not included in the CHILD study despite the strong associations with asthma (148, 149). In addition, the CES-D scores themselves cannot confirm the diagnosis of depression in the mothers (147).

Another limitation is that there is uncertainty as to whether the ELISA is detecting both free and bound sIgA. The manufacturing company claims that the kit partially separates sIgA from the bacteria, and therefore, the antibodies detect both free and bound sIgA. One study that looked at total sIgA using ELISA did not find significant differences in 1-month and 12-month total fecal sIgA between infants that developed allergy and did not develop allergy but found less sIgA-coated bacteria in the allergy group, which was determined by flow cytometry (61). This may suggest the need for further studies on sIgA and the interaction with gut microbiota in relation to maternal distress.

Some 3-month visits of the CHILD study were conducted when the infant was 2-8 months of age due to difficulties in communication. As no set timeframe was used to exclude infants from our sample, this wide range of infant age may have increased the variation of sIgA levels in our sample, and perhaps lower sIgA was observed because of the younger and older ages (61). However, there were also many variations in sIgA levels within 3-4 months of age. In the logistic regression analyses, we controlled for infant age, but still obtained significant results.

Although stool collected greater than 6 months had significantly lower sIgA levels than stool collected less than 6 months, sensitivity analyses show that having these differences did not have a significant impact on our results. Having these infants in our sample may help with the comparisons for postnatal depressive symptoms since the depression information was collected at 6 months of child age, which leads to another limitation about the postnatal measure for distress being based on time points that come after the proposed time of stool collection. One study on the EDEN mother-child cohort reported depressive symptom trajectories with measures at 4 months and 8 months postpartum that had very small changes in symptom scores between the two time points for both prenatal and postnatal trajectories (150). McCall-Hosenfield et al. (151) also reported postpartum depressive symptom trajectories with time points at 1 month and 6 months, and no major changes were observed.

Lastly, we detected few significant associations between higher perceived stress and low sIgA, and sometimes with the presence of stress, sIgA levels were higher than the reference group. Although we used a validated questionnaire for perceived stress, we did not have a recommended cut-off for very high levels of stress but instead used the highest quartile cut-off for all CHILD mothers, which may have affected our detection of an association between maternal stress and infant fecal sIgA. In addition, immigrant families were included in this study,

and so the questions in the maternal distress surveys may not capture the variations of the concept of stress depending on the culture.

4.4 Clinical relevance

Asthma and allergic diseases are the most common chronic diseases among children worldwide, and the prevalence is increasing (3, 152). This study will greatly contribute to the scholarly literature on the impact of maternal stress and depression on the immune system maturation in human infants. In addition, determining the existence of gut immunity changes in infants of mothers with pre and postnatal distress will provide insight on potential infant biomarkers to predict development of common chronic conditions in young individuals. The findings may encourage interventions in reducing distress during pregnancy and after birth to become more valuable to mothers and their families, as they have the potential to reduce the risk of future chronic diseases in newborns.

Encouraging breastfeeding without inflicting more stress on distressed mothers or having prebiotic or probiotic treatments for mother and infant may improve the infant gut health. Our results show that breastfeeding may have protective effects against distress lowering infant fecal sIgA levels. Several studies have found supplementation of prebiotics and probiotics containing microbes that are stimulatory of sIgA can increase sIgA levels in the gut (46, 48, 153). Prebiotics and probiotics also show benefits for reducing stress, depressive and anxiety-like symptoms (154-156). These may be future interventions to allow for sufficient establishment of the infant gut microbiota and prevent many chronic illnesses that are related to alterations in early gut microbiome composition.

4.5 Implications for future research

More studies on the mechanisms of maternal distress impacting infant gut immunity are needed. Still it is not known whether stress and depression have similar mechanisms. The compounds in tobacco smoke and the use of SSRIs in depressed women may be some of the factors that affect infant gut health, but this could not be directly tested with our study design. It would be beneficial to know which sIgA-microbe interactions distress affects as well as the optimal level of sIgA that is needed for infants to have healthy gut microbiome compositions. These various ideas may assist with the future developments of interventions that allow for the development of healthy infant gut microbiome and immunity.

4.6 Conclusion

This thesis investigated the impact of perinatal maternal distress on infant gut immunity in the first few months of life. We found that when the mother experiences both pre and postnatal depressive symptoms, the infants are more likely to have lower sIgA in their gut, even after controlling for various covariates. With population-level findings, this study highlights the importance of psychosocial health in mothers during the pre and postnatal period as it may contribute to the health of children in the early months of life and in later life.

61

References

(1) To T, Stanojevic S, Moores G, Gershon AS, Bateman ED, Cruz AA, et al. Global asthma prevalence in adults: findings from the cross-sectional world health survey. BMC Public Health 2012 Mar 19;12:1-8.

(2) Prescott SL, Pawankar R, Allen KJ, Campbell DE, Sinn JK, Fiocchi A, et al. A global survey of changing patterns of food allergy burden in children. World Allergy Organ J 2013 Dec 4;6(1):1-12.

(3) Asher MI, Montefort S, Bjorksten B, Lai CK, Strachan DP, Weiland SK, et al. Worldwide time trends in the prevalence of symptoms of asthma, allergic rhinoconjunctivitis, and eczema in childhood: ISAAC Phases One and Three repeat multicountry cross-sectional surveys. Lancet 2006 Aug 26;368(9537):733-743.

(4) van de Loo KF, van Gelder MM, Roukema J, Roeleveld N, Merkus PJ, Verhaak CM.Prenatal maternal psychological stress and childhood asthma and wheezing: a meta-analysis. EurRespir J 2016 Jan;47(1):133-146.

(5) Andersson NW, Hansen MV, Larsen AD, Hougaard KS, Kolstad HA, Schlunssen V. Prenatal maternal stress and atopic diseases in the child: a systematic review of observational human studies. Allergy 2016 Jan;71(1):15-26.

(6) Van Lieshout RJ, Macqueen GM. Relations between asthma and psychological distress: an old idea revisited. Chem Immunol Allergy 2012;98:1-13.

(7) Kozyrskyj A, Letourneau N, Kang L, Salmani M. Associations between postpartum depressive symptoms and childhood asthma diminish with child age. Clin Exp Allergy 2016 Oct 22.

(8) Gavin NI, Gaynes BN, Lohr KN, Meltzer-Brody S, Gartlehner G, Swinson T. Perinatal depression: a systematic review of prevalence and incidence. Obstet Gynecol 2005 Nov;106(5 Pt 1):1071-1083.

(9) Woods SM, Melville JL, Guo Y, Fan MY, Gavin A. Psychosocial stress during pregnancy.Am J Obstet Gynecol 2010 Jan;202(1):61.e1-61.e7.

(10) Byatt N, Xiao RS, Dinh KH, Waring ME. Mental health care use in relation to depressive symptoms among pregnant women in the USA. Arch Womens Ment Health 2016 Feb;19(1):187-191.

(11) Kingston D, Tough S, Whitfield H. Prenatal and postpartum maternal psychological distress and infant development: a systematic review. Child Psychiatry Hum Dev 2012 Oct;43(5):683-714.

(12) Bann CM, Parker CB, Grobman WA, Willinger M, Simhan HN, Wing DA, et al.Psychometric properties of stress and anxiety measures among nulliparous women. J PsychosomObstet Gynaecol 2017 Mar;38(1):53-62.

(13) American Psychiatric Association. Diagnostic and statistical manual of mental disorders.5th ed.: American Psychiatric Publishing; 2013.

(14) Lewis AJ, Austin E, Knapp R, Vaiano T, Galbally M. Perinatal maternal mental health, fetal programming and child development. Healthcare (Basel) 2015 Nov 26;3(4):1212-1227.

(15) Beijers R, Buitelaar JK, de Weerth C. Mechanisms underlying the effects of prenatal psychosocial stress on child outcomes: beyond the HPA axis. Eur Child Adolesc Psychiatry 2014 Oct;23(10):943-956.

(16) Norhayati MN, Hazlina NH, Asrenee AR, Emilin WM. Magnitude and risk factors for postpartum symptoms: a literature review. J Affect Disord 2015 Apr 1;175:34-52.

(17) McGarry J, Kim H, Sheng X, Egger M, Baksh L. Postpartum depression and help-seeking behavior. J Midwifery Womens Health 2009 Jan-Feb;54(1):50-56.

(18) Vigod SN, Wilson CA, Howard LM. Depression in pregnancy. BMJ 2016 Mar 24;352:i1547.

(19) Batmaz G, Dane B, Sarioglu A, Kayaoglu Z, Dane C. Can we predict postpartum depression in pregnant women? Clin Exp Obstet Gynecol 2015;42(5):605-609.

(20) Iwata H, Mori E, Sakajo A, Aoki K, Maehara K, Tamakoshi K. Prevalence of postpartum depressive symptoms during the first 6 months postpartum: association with maternal age and parity. J Affect Disord 2016 Oct;203:227-232.

(21) Santos H,Jr, Tan X, Salomon R. Heterogeneity in perinatal depression: how far have we come? A systematic review. Arch Womens Ment Health 2017 Feb;20(1):11-23.

(22) McLearn KT, Minkovitz CS, Strobino DM, Marks E, Hou W. Maternal depressive symptoms at 2 to 4 months post partum and early parenting practices. Arch Pediatr Adolesc Med 2006 Mar;160(3):279-284.

(23) Sanger C, Iles JE, Andrew CS, Ramchandani PG. Associations between postnatal maternal depression and psychological outcomes in adolescent offspring: a systematic review. Arch Womens Ment Health 2015 Apr;18(2):147-162.

(24) Mantymaa M, Puura K, Luoma I, Salmelin R, Davis H, Tsiantis J, et al. Infant-mother interaction as a predictor of child's chronic health problems. Child Care Health Dev 2003 May;29(3):181-191.

(25) Walker A. Breast milk as the gold standard for protective nutrients. J Pediatr 2010Feb;156(2 Suppl):S3-7.

(26) Weaver IC, Cervoni N, Champagne FA, D'Alessio AC, Sharma S, Seckl JR, et al. Epigenetic programming by maternal behavior. Nat Neurosci 2004 Aug;7(8):847-854.

(27) Santos H, Jr, Yang Q, Docherty SL, White-Traut R, Holditch-Davis D. Relationship of maternal psychological distress classes to later mother-infant interaction, home environment, and infant development in preterm infants. Res Nurs Health 2016 Jun;39(3):175-186.

(28) Yatsenko O, Pizano J, Nikolaidis A. Revisiting maternal–infant bonding's effects on asthma:A brief history. Cogent Psychology 2016;3(1):1161267.

(29) Wright RJ. Prenatal maternal stress and early caregiving experiences: implications for childhood asthma risk. Paediatr Perinat Epidemiol 2007 Nov;21 Suppl 3:8-14.

(30) Roux G, Anderson C, Roan C. Postpartum depression, marital dysfunction, and infant outcome: a longitudinal study. J Perinat Educ 2002;11(4):25-36.

(31) Grace SL, Evindar A, Stewart DE. The effect of postpartum depression on child cognitive development and behavior: a review and critical analysis of the literature. Arch Womens Ment Health 2003 Nov;6(4):263-274.

(32) Mojtabai R. Parental psychopathology and childhood atopic disorders in the community.Psychosom Med 2005 May-Jun;67(3):448-453.

(33) Wright RJ, Cohen S, Carey V, Weiss ST, Gold DR. Parental stress as a predictor of wheezing in infancy: a prospective birth-cohort study. Am J Respir Crit Care Med 2002 Feb 1;165(3):358-365.

(34) Alton ME, Zeng Y, Tough SC, Mandhane PJ, Kozyrskyj AL. Postpartum depression, a direct and mediating risk factor for preschool wheeze in girls. Pediatr Pulmonol 2015 Oct 8.

(35) Penders J, Stobberingh EE, van den Brandt PA, Thijs C. The role of the intestinal microbiota in the development of atopic disorders. Allergy 2007 Nov;62(11):1223-1236.

(36) Koleva PT, Kim JS, Scott JA, Kozyrskyj AL. Microbial programming of health and disease starts during fetal life. Birth Defects Res C Embryo Today 2015 Dec;105(4):265-277.

(37) Collado MC, Rautava S, Aakko J, Isolauri E, Salminen S. Human gut colonisation may be initiated in utero by distinct microbial communities in the placenta and amniotic fluid. Sci Rep 2016 Mar 22;6:1-13.

(38) Azad MB, Konya T, Persaud RR, Guttman DS, Chari RS, Field CJ, et al. Impact of maternal intrapartum antibiotics, method of birth and breastfeeding on gut microbiota during the first year of life: a prospective cohort study. BJOG 2016 May;123(6):983-993.

(39) Lodge CJ, Dharmage SC. Breastfeeding and perinatal exposure, and the risk of asthma and allergies. Curr Opin Allergy Clin Immunol 2016 Jun;16(3):231-236.

(40) Martin R, Makino H, Cetinyurek Yavuz A, Ben-Amor K, Roelofs M, Ishikawa E, et al. Early-life events, including mode of delivery and type of feeding, siblings and gender, shape the developing gut microbiota. PLoS One 2016 Jun 30;11(6):e0158498.

(41) Reboldi A, Arnon TI, Rodda LB, Atakilit A, Sheppard D, Cyster JG. IgA productionrequires B cell interaction with subepithelial dendritic cells in Peyer's patches. Science 2016 May 13;352(6287):aaf4822.

(42) Brandtzaeg P. Secretory IgA: designed for anti-microbial defense. Front Immunol 2013 Aug 6;4:1-17.

(43) Zagato E, Mazzini E, Rescigno M. The variegated aspects of Immunoglobulin A. Immunol Lett 2016 Oct;178:45-49.

(44) Kato LM, Kawamoto S, Maruya M, Fagarasan S. Gut TFH and IgA: key players for regulation of bacterial communities and immune homeostasis. Immunol Cell Biol 2014 Jan;92(1):49-56.

(45) Suzuki K, Meek B, Doi Y, Muramatsu M, Chiba T, Honjo T, et al. Aberrant expansion of segmented filamentous bacteria in IgA-deficient gut. Proc Natl Acad Sci U S A 2004 Feb 17;101(7):1981-1986.

(46) Kukkonen K, Kuitunen M, Haahtela T, Korpela R, Poussa T, Savilahti E. High intestinal IgA associates with reduced risk of IgE-associated allergic diseases. Pediatr Allergy Immunol 2010 Feb;21(1 Pt 1):67-73.

(47) Brugman S, Perdijk O, van Neerven RJ, Savelkoul HF. Mucosal immune development in early life: setting the stage. Arch Immunol Ther Exp (Warsz) 2015 Aug;63(4):251-268.

(48) Karamese M, Aydin H, Sengul E, Gelen V, Sevim C, Ustek D, et al. The immunostimulatory effect of lactic acid bacteria in a rat model. Iran J Immunol 2016 Sep;13(3):220-228.

(49) Karaffova V, Bobikova K, Husakova E, Levkut M, Herich R, Revajova V, et al. Interaction of TGF-beta4 and IL-17 with IgA secretion in the intestine of chickens fed with E. faecium AL41 and challenged with S. Enteritidis. Res Vet Sci 2015 Jun;100:75-79.

(50) Nagano K, Taguchi K, Tokoro S, Tatsuno I, Mori H. Adhesion of enterohemorrhagic Escherichia coli O157:H7 to the intestinal epithelia is essential for inducing secretory IgA antibody production in the intestine of mice. Biol Pharm Bull 2014;37(3):409-416.

(51) Maruyama K, Hida M, Kohgo T, Fukunaga Y. Changes in salivary and fecal secretory IgA in infants under different feeding regimens. Pediatr Int 2009 Jun;51(3):342-345.

(52) Bridgman SL, Konya T, Azad MB, Sears MR, Becker AB, Turvey SE, et al. Infant gut immunity: a preliminary study of IgA associations with breastfeeding. J Dev Orig Health Dis 2016 Feb;7(1):68-72.

(53) Piirainen L, Pesola J, Pesola I, Komulainen J, Vaarala O. Breastfeeding stimulates total and cow's milk-specific salivary IgA in infants. Pediatr Allergy Immunol 2009 May;20(3):295-298.

(54) Backhed F, Roswall J, Peng Y, Feng Q, Jia H, Kovatcheva-Datchary P, et al. Dynamics and stabilization of the human gut microbiome during the first year of life. Cell Host Microbe 2015 Jun 10;17(6):690-703.

(55) Rogier EW, Frantz AL, Bruno ME, Wedlund L, Cohen DA, Stromberg AJ, et al. Secretory antibodies in breast milk promote long-term intestinal homeostasis by regulating the gut microbiota and host gene expression. Proc Natl Acad Sci U S A 2014 Feb 25;111(8):3074-3079.

(56) Pabst O, Cerovic V, Hornef M. Secretory IgA in the coordination of establishment and maintenance of the microbiota. Trends Immunol 2016 May;37(5):287-296.

(57) Battersby AJ, Gibbons DL. The gut mucosal immune system in the neonatal period. Pediatr Allergy Immunol 2013 Aug;24(5):414-421.

(58) Sandin A, Bjorksten B, Bottcher MF, Englund E, Jenmalm MC, Braback L. High salivary secretory IgA antibody levels are associated with less late-onset wheezing in IgE-sensitized infants. Pediatr Allergy Immunol 2011 Aug;22(5):477-481.

(59) Orivuori L, Loss G, Roduit C, Dalphin JC, Depner M, Genuneit J, et al. Soluble immunoglobulin A in breast milk is inversely associated with atopic dermatitis at early age: the PASTURE cohort study. Clin Exp Allergy 2014 Jan;44(1):102-112.

(60) Bridgman SL, Konya T, Azad MB, Guttman DS, Sears MR, Becker AB, et al. High fecalIgA is associated with reduced Clostridium difficile colonization in infants. Microbes Infect2016 May 24;18(9):543-549.

(61) Dzidic M, Abrahamsson TR, Artacho A, Bjorksten B, Collado MC, Mira A, et al. Aberrant IgA responses to the gut microbiota during infancy precedes asthma and allergy development. J Allergy Clin Immunol 2016 Aug 13;139(3):1017-1025.

(62) Warren CM, Otto AK, Walkner MM, Gupta RS. Quality of life among food allergic patients and their caregivers. Curr Allergy Asthma Rep 2016 May;16(5):1-8.

(63) Cook-Mills JM. Maternal influences over offspring allergic responses. Curr Allergy Asthma Rep 2015 Feb;15(2):1-10.

(64) Mannan M, Mamun A, Doi S, Clavarino A. Is there a bi-directional relationship between depression and obesity among adult men and women? Systematic review and bias-adjusted meta analysis. Asian J Psychiatr 2016 Jun;21:51-66.

(65) Hartley E, McPhie S, Skouteris H, Fuller-Tyszkiewicz M, Hill B. Psychosocial risk factors for excessive gestational weight gain: a systematic review. Women Birth 2015 Dec;28(4):e99-e109.

(66) Marchi J, Berg M, Dencker A, Olander EK, Begley C. Risks associated with obesity in pregnancy, for the mother and baby: a systematic review of reviews. Obes Rev 2015 Aug;16(8):621-638.

(67) Bergmann S, von Klitzing K, Keitel-Korndorfer A, Wendt V, Grube M, Herpertz S, et al. Emotional availability, understanding emotions, and recognition of facial emotions in obese mothers with young children. J Psychosom Res 2016 Jan;80:44-52.

(68) Collado MC, Isolauri E, Laitinen K, Salminen S. Effect of mother's weight on infant's microbiota acquisition, composition, and activity during early infancy: a prospective follow-up study initiated in early pregnancy. Am J Clin Nutr 2010 Nov;92(5):1023-1030.

(69) Kozyrskyj AL, Kalu R, Koleva PT, Bridgman SL. Fetal programming of overweight through the microbiome: boys are disproportionately affected. J Dev Orig Health Dis 2016 Feb;7(1):25-34.

(70) Fujimori M, Franca EL, Fiorin V, Morais TC, Honorio-Franca AC, de Abreu LC. Changes in the biochemical and immunological components of serum and colostrum of overweight and obese mothers. BMC Pregnancy Childbirth 2015 Aug 12;15:1-8.

(71) Adams SS, Eberhard-Gran M, Sandvik AR, Eskild A. Mode of delivery and postpartum emotional distress: a cohort study of 55,814 women. BJOG 2012 Feb;119(3):298-305.

(72) Sword W, Landy CK, Thabane L, Watt S, Krueger P, Farine D, et al. Is mode of delivery associated with postpartum depression at 6 weeks: a prospective cohort study. BJOG 2011 Jul;118(8):966-977.

(73) Chang S, Chen K, Ho H, Lai Y, Lin M, Lee C, et al. Depressive symptoms, pain, and sexual dysfunction over the first year following vaginal or cesarean delivery: A prospective longitudinal study. Int J Nurs Stud 2015 9;52(9):1433-1444.

(74) Yang S, Shen L, Ping T, Wang Y, Chien C. The delivery mode and seasonal variation are associated with the development of postpartum depression. J Affect Disord 2011;132(1–2):158-164.

(75) Rauh C, Beetz A, Burger P, Engel A, Haberle L, Fasching PA, et al. Delivery mode and the course of pre- and postpartum depression. Arch Gynecol Obstet 2012 Dec;286(6):1407-1412.

(76) Garthus-Niegel S, von Soest T, Knoph C, Simonsen TB, Torgersen L, Eberhard-Gran M. The influence of women's preferences and actual mode of delivery on post-traumatic stress symptoms following childbirth: a population-based, longitudinal study. BMC Pregnancy and Childbirth 2014;14:1-10.

(77) Kristensen K, Henriksen L. Cesarean section and disease associated with immune function.J Allergy Clin Immunol 2016 Feb;137(2):587-590.

(78) Huurre A, Kalliomaki M, Rautava S, Rinne M, Salminen S, Isolauri E. Mode of delivery - effects on gut microbiota and humoral immunity. Neonatology 2008;93(4):236-240.

(79) Bascom EM, Napolitano MA. Breastfeeding duration and primary reasons for breastfeeding cessation among women with postpartum depressive symptoms. J Hum Lact 2015 Dec 7;32(2):282-291.

(80) Khashan AS, Everard C, McCowan LM, Dekker G, Moss-Morris R, Baker PN, et al. Second-trimester maternal distress increases the risk of small for gestational age. Psychol Med 2014 Oct;44(13):2799-2810.

(81) Becker M, Weinberger T, Chandy A, Schmukler S. Depression during pregnancy and postpartum. Curr Psychiatry Rep 2016;18(3):1-9.

(82) Ciesielski TH, Marsit CJ, Williams SM. Maternal psychiatric disease and epigeneticevidence suggest a common biology for poor fetal growth. BMC Pregnancy Childbirth 2015 Aug 25;15:1-9.

(83) Grumach AS, Carmona RC, Lazarotti D, Ribeiro MA, Rozentraub RB, Racz ML, et al. Immunological factors in milk from Brazilian mothers delivering small-for-date term neonates. Acta Paediatr 1993 Mar;82(3):284-290.

(84) Cox LM, Blaser MJ. Antibiotics in early life and obesity. Nat Rev Endocrinol 2015 Mar;11(3):182-190.

(85) Field T, Hernandez-Reif M, Diego M. Risk factors and stress variables that differentiate depressed from nondepressed pregnant women. Infant Behavior and Development 2006 4;29(2):169-174.

(86) Groer MW, Morgan K. Immune, health and endocrine characteristics of depressed postpartum mothers. Psychoneuroendocrinology 2007 2;32(2):133-139.

(87) Newton ER, Hale TW. Drugs in breast milk. Clin Obstet Gynecol 2015 Dec;58(4):868-884.

(88) Nguyen T, Kramer J, Vallejo R, Stanton G, Heidenreich BA, Benyamin R, et al. Citalopram enhances B cell numbers in a murine model of morphine-induced immunosuppression. Pain Pract 2009 May-Jun;9(3):195-205.

(89) Ho PS, Yeh YW, Huang SY, Liang CS. A shift toward T helper 2 responses and an increase in modulators of innate immunity in depressed patients treated with escitalopram. Psychoneuroendocrinology 2015 Mar;53:246-255.

(90) Breedlove G, Fryzelka D. Depression screening during pregnancy. J Midwifery Womens Health 2011 Jan-Feb;56(1):18-25.

(91) Muzik M, Hamilton SE. Use of antidepressants during pregnancy?: what to consider when weighing treatment with antidepressants against untreated depression. Matern Child Health J 2016 Nov;20(11):2268-2279.

(92) Woolhouse H, James J, Gartland D, McDonald E, Brown SJ. Maternal depressive symptoms at three months postpartum and breastfeeding rates at six months postpartum: Implications for primary care in a prospective cohort study of primiparous women in Australia. Women Birth 2016 Aug;29(4):381-387.

(93) Victora CG, Bahl R, Barros AJD, França GVA, Horton S, Krasevec J, et al. Breastfeeding in the 21st century: epidemiology, mechanisms, and lifelong effect. The Lancet 2016;387(10017):475-490. (94) Yedid Sion M, Harlev A, Weintraub AY, Sergienko R, Sheiner E. Is antenatal depression associated with adverse obstetric and perinatal outcomes? The Journal of Maternal-Fetal & Neonatal Medicine 2016;29(6):863-867.

(95) Salimi S, Terplan M, Cheng D, Chisolm MS. The relationship between postpartum depression and perinatal cigarette smoking: an analysis of PRAMS data. J Subst Abuse Treat 2015 2016;56:34-38.

(96) Lynch ME, Johnson KC, Kable JA, Carroll J, Coles CD. Smoking in pregnancy and parenting stress: maternal psychological symptoms and socioeconomic status as potential mediating variables. Nicotine Tob Res 2011 Jul;13(7):532-539.

(97) Biedermann L, Zeitz J, Mwinyi J, Sutter-Minder E, Rehman A, Ott SJ, et al. Smoking cessation induces profound changes in the composition of the intestinal microbiota in humans. PLoS One 2013;8(3):e59260.

(98) Narkowicz S, Polkowska Z, Kielbratowska B, Namiesnik J. Meconium samples used to assess infant exposure to the components of ETS during pregnancy. Int J Occup Med Environ Health 2015;28(6):955-970.

(99) Gosalbes MJ, Llop S, Valles Y, Moya A, Ballester F, Francino MP. Meconium microbiota types dominated by lactic acid or enteric bacteria are differentially associated with maternal eczema and respiratory problems in infants. Clin Exp Allergy 2013 Feb;43(2):198-211.

75

(100) Avsar A, Darka O, Bodrumlu EH, Bek Y. Evaluation of the relationship between passive smoking and salivary electrolytes, protein, secretory IgA, sialic acid and amylase in young children. Arch Oral Biol 2009 May;54(5):457-463.

(101) Noakes P, Taylor A, Hale J, Breckler L, Richmond P, Devadason SG, et al. The effects of maternal smoking on early mucosal immunity and sensitization at 12 months of age. Pediatr Allergy Immunol 2007 Mar;18(2):118-127.

(102) Goldade K, Nichter M, Nichter M, Adrian S, Tesler L, Muramoto M. Breastfeeding and smoking among low-income women: results of a longitudinal qualitative study. Birth 2008 Sep;35(3):230-240.

(103) Amir LH, Donath SM. Does maternal smoking have a negative physiological effect on breastfeeding? The epidemiological evidence. Breastfeed Rev 2003 Jul;11(2):19-29.

(104) Jedrychowski W, Perera F, Mroz E, Edwards S, Flak E, Rauh V, et al. Prenatal exposure to passive smoking and duration of breastfeeding in nonsmoking women: Krakow inner city prospective cohort study. Arch Gynecol Obstet 2008 Nov;278(5):411-417.

(105) Bachour P, Yafawi R, Jaber F, Choueiri E, Abdel-Razzak Z. Effects of smoking, mother's age, body mass index, and parity number on lipid, protein, and secretory immunoglobulin A concentrations of human milk. Breastfeed Med 2012 Jun;7(3):179-188.

(106) Beetz A, Uvnas-Moberg K, Julius H, Kotrschal K. Psychosocial and psychophysiological effects of human-animal interactions: the possible role of oxytocin. Front Psychol 2012 Jul 9;3:1-15.

(107) Amerine JL, Hubbard GB. Using animal-assisted therapy to enrich psychotherapy. Adv Mind Body Med 2016 Summer;30(3).

(108) Chan CW, Wong RS, Law PT, Wong CL, Tsui SK, Tang WP, et al. Environmental factors associated with altered gut microbiota in children with eczema: a systematic review. Int J Mol Sci 2016 Jul 16;17(7):1-14.

(109) Brooks C, Pearce N, Douwes J. The hygiene hypothesis in allergy and asthma: an update.Curr Opin Allergy Clin Immunol 2013 Feb;13(1):70-77.

(110) Azad MB, Konya T, Maughan H, Guttman DS, Field CJ, Sears MR, et al. Infant gut microbiota and the hygiene hypothesis of allergic disease: impact of household pets and siblings on microbiota composition and diversity. Allergy Asthma Clin Immunol 2013;9(1):1-9.

(111) Faught J, Bierl C, Barton B, Kemp A. Stress in mothers of young children with eczema. Arch Dis Child 2007 Aug;92(8):683-686.

(112) Jang HJ, Hwang S, Ahn Y, Lim DH, Sohn M, Kim JH. Family quality of life among families of children with atopic dermatitis. Asia Pac Allergy 2016 Oct;6(4):213-219.

(113) Filanovsky MG, Pootongkam S, Tamburro JE, Smith MC, Ganocy SJ, Nedorost ST. The financial and emotional impact of atopic dermatitis on children and their families. J Pediatr 2016 Feb;169:284-290.

(114) Martinez-Carrillo BE, Godinez-Victoria M, Jarillo-Luna A, Oros-Pantoja R, Abarca-Rojano E, Rivera-Aguilar V, et al. Repeated restraint stress reduces the number of IgAproducing cells in Peyer's patches. Neuroimmunomodulation 2011;18(3):131-141. (115) Lim R, Fedulov AV, Kobzik L. Maternal stress during pregnancy increases neonatal allergy susceptibility: role of glucocorticoids. Am J Physiol Lung Cell Mol Physiol 2014 Jul 15;307(2):L141-L148.

(116) Jasarevic E, Howerton CL, Howard CD, Bale TL. Alterations in the vaginal microbiome by maternal stress are associated with metabolic reprogramming of the offspring gut and brain. Endocrinology 2015;156:3265-3276.

(117) Bailey MT, Lubach GR, Coe CL. Prenatal stress alters bacterial colonization of the gut in infant monkeys. J Pediatr Gastroenterol Nutr 2004 Apr;38(4):414-421.

(118) Jasarevic E, Rodgers AB, Bale TL. A novel role for maternal stress and microbial transmission in early life programming and neurodevelopment. Neurobiology of Stress 2014;1:81-88.

(119) van Best N, Hornef MW, Savelkoul PH, Penders J. On the origin of species: factors shaping the establishment of infant's gut microbiota. Birth Defects Res C Embryo Today 2015 Dec;105(4):240-251.

(120) Kianbakht S, Mashhadi E, Jamillian HR, Ghazavi A. Immune phenomena in neonates of women with depression during pregnancy: a case-control study. The Journal of Maternal-Fetal & Neonatal Medicine 2013 04/01;26(6):608-610.

(121) Zijlmans MA, Korpela K, Riksen-Walraven JM, de Vos WM, de Weerth C. Maternal prenatal stress is associated with the infant intestinal microbiota. Psychoneuroendocrinology 2015 Mar;53:233-245.

(122) Jarillo-Luna A, Rivera-Aguilar V, Garfias HR, Lara-Padilla E, Kormanovsky A, Campos-Rodriguez R. Effect of repeated restraint stress on the levels of intestinal IgA in mice.Psychoneuroendocrinology 2007 Jul;32(6):681-692.

(123) Morales-Medina JC, Iannitti T, Freeman A, Caldwell HK. The olfactory bulbectomized rat as a model of depression: the hippocampal pathway. Behav Brain Res 2017;317:562-575.

(124) Moussaoui N, Jacobs JP, Larauche M, Biraud M, Million M, Mayer E, et al. Chronic earlylife stress in rat pups alters basal corticosterone, intestinal permeability, and fecal microbiota at weaning: influence of sex. J Neurogastroenterol Motil 2017 Jan 30;23(1):135-143.

(125) Moraes TJ, Lefebvre DL, Chooniedass R, Becker AB, Brook JR, Denburg J, et al. The Canadian Healthy Infant Longitudinal Development birth cohort study: biological samples and biobanking. Paediatr Perinat Epidemiol 2015 Jan;29(1):84-92.

(126) Cohen S, Kamarck T, Mermelstein R. A global measure of perceived stress. J Health Soc Behav 1983 Dec;24(4):385-396.

(127) Cohen S. Perceived stress in a probability sample of the United States. 1988.

(128) Radloff LS. The CES-D scale: a self-report depression scale for research in the general population. Applied Psychological Measurement 1977 June 01;1(3):385-401.

(129) Forrest BD. Effects of sample processing on the measurement of specific intestinal IgA immune responses. Vaccine 1992;10(11):802-805.

(130) Immundiagnostik AG. 2009; Available at:

http://www.immundiagnostik.com/en/home.html.

(131) Glickman ME, Rao SR, Schultz MR. False discovery rate control is a recommended alternative to Bonferroni-type adjustments in health studies. J Clin Epidemiol 2014;67(8):850-857.

(132) Holm S. A simple sequentially rejective multiple test procedure. Scandinavian journal of statistics 1979:65-70.

(133) Lee A, Mathilda Chiu YH, Rosa MJ, Jara C, Wright RO, Coull BA, et al. Prenatal and postnatal stress and asthma in children: temporal- and sex-specific associations. J Allergy Clin Immunol 2016 Sep;138(3):740-747.

(134) Campos-Rodríguez R, Godínez-Victoria M, Abarca-Rojano E, Pacheco-Yépez J, Reyna-Garfias H, Barbosa-Cabrera RE, et al. Stress modulates intestinal secretory immunoglobulin A. Front Integr Neurosci 2013;7:1-10.

(135) Wright RJ. Prenatal maternal stress and early caregiving experiences: implications for childhood asthma risk. Paediatr Perinat Epidemiol 2007;21(s3):8-14.

(136) Oberlander TF, Weinberg J, Papsdorf M, Grunau R, Misri S, Devlin AM. Prenatal exposure to maternal depression, neonatal methylation of human glucocorticoid receptor gene (NR3C1) and infant cortisol stress responses. Epigenetics 2008 Mar-Apr;3(2):97-106.

(137) Trapolini T, Ungerer JA, McMahon CA. Maternal depression: relations with maternal caregiving representations and emotional availability during the preschool years. Attach Hum Dev 2008 Mar;10(1):73-90.

(138) Cloez-Tayarani I, Changeux JP. Nicotine and serotonin in immune regulation and inflammatory processes: a perspective. J Leukoc Biol 2007 Mar;81(3):599-606.

(139) Wroble-Biglan MC, Dietz LJ, Pienkosky TV. Prediction of infant temperament from catecholamine and self-report measures of maternal stress during pregnancy. J Reprod Infant Psychol 2009;27(4):374-389.

(140) Kozyrskyj AL, Mai XM, McGrath P, Hayglass KT, Becker AB, Macneil B. Continued exposure to maternal distress in early life is associated with an increased risk of childhood asthma. Am J Respir Crit Care Med 2008 Jan 15;177(2):142-147.

(141) Cottrell L, Gibson M, Harris C, Rai A, Sobhan S, Berry T, et al. Examining smoking and cessation during pregnancy among an Appalachian sample: a preliminary view. Subst Abuse Treat Prev Policy 2007 May 7;2:14.

(142) Praveen V, Praveen S. Microbiome-gut-brain axis: a pathway for improving brainstem serotonin homeostasis and successful autoresuscitation in SID-A novel hypothesis. Front Pediatr 2016;4:1-11.

(143) Fröhlich EE, Farzi A, Mayerhofer R, Reichmann F, Jačan A, Wagner B, et al. Cognitiveimpairment by antibiotic-induced gut dysbiosis: analysis of gut microbiota-brain communication.Brain Behav Immun 2016 Aug;56:140-155.

(144) Kozyrskyj AL, Bahreinian S, Azad MB. Early life exposures: impact on asthma and allergic disease. Curr Opin Allergy Clin Immunol 2011 Oct;11(5):400-406.

(145) Grumach AS, Carmona RC, Lazarotti D, Ribeiro MA, Rozentraub RB, Racz ML, et al.Immunological factors in milk from Brazilian mothers delivering small-for-date term neonates.Acta Paediatr 1993 Mar;82(3):284-290.

(146) Wisdom JP, Saedi GA, Green CA. Another breed of "service" animals: STARS study findings about pet ownership and recovery from serious mental illness. Am J Orthopsychiatry 2009 Jul;79(3):430-436.

(147) Vilagut G, Forero CG, Barbaglia G, Alonso J. Screening for depression in the general population with the center for epidemiologic studies depression (CES-D): a systematic review with meta-analysis. PLoS One 2016 May 16;11(5):e0155431.

(148) Cookson H, Granell R, Joinson C, Ben-Shlomo Y, Henderson AJ. Mothers' anxiety during pregnancy is associated with asthma in their children. J Allergy Clin Immunol 2009 Apr;123(4):847-53.

(149) Guxens M, Sonnenschein-van der Voort AM, Tiemeier H, Hofman A, Sunyer J, de Jongste JC, et al. Parental psychological distress during pregnancy and wheezing in preschool children: the Generation R Study. J Allergy Clin Immunol 2014 Jan;133(1):59-67.

(150) van der Waerden J, Bernard JY, De Agostini M, Saurel-Cubizolles MJ, Peyre H, Heude B, et al. Persistent maternal depressive symptoms trajectories influence children's IQ: The EDEN mother-child cohort. Depress Anxiety 2017 Sep 7;34:105-117.

82

(151) McCall-Hosenfeld JS, Phiri K, Schaefer E, Zhu J, Kjerulff K. Trajectories of depressive symptoms throughout the peri- and postpartum period: results from the first baby study. J Womens Health (Larchmt) 2016 Jun 16;25(11):1112-1121.

(152) Pawankar R, Canonica GW, Holgate ST, Lockey RF. Allergic diseases and asthma: a major global health concern. Curr Opin Allergy Clin Immunol 2012 Feb;12(1):39-41.

(153) Radke M, Picaud JC, Loui A, Cambonie G, Faas D, Lafeber HN, et al. Starter formula enriched in prebiotics and probiotics ensures normal growth of infants and promotes gut health: a randomized clinical trial. Pediatr Res 2017 Dec 21.

(154) Liu X, Cao S, Zhang X. Modulation of gut microbiota-brain axis by probiotics, prebiotics, and diet. J Agric Food Chem 2015 Sep 16;63(36):7885-7895.

(155) Mika A, Rumian N, Loughridge AB, Fleshner M. Exercise and prebiotics produce stress resistance: converging impacts on stress-protective and butyrate-producing gut bacteria. Int Rev Neurobiol 2016;131:165-191.

(156) Messaoudi M, Violle N, Bisson JF, Desor D, Javelot H, Rougeot C. Beneficial psychological effects of a probiotic formulation (Lactobacillus helveticus R0052 and Bifidobacterium longum R0175) in healthy human volunteers. Gut Microbes 2011 Jul-Aug;2(4):256-261.

Appendices

Appendix A: Preliminary analyses with 304 sample and pregnancy overweight

Table A1. Infant fecal sIgA levels and percentage distribution of potential covariates in relation to prenatal maternal depressive symptoms and lowest quartile sIgA (n=304)

Child ex Control Contro Control <thcontrol< th=""> <th< th=""><th></th><th>Lower symptoms (71.2% overall)</th><th>Prenatal only (11.9% overall)</th><th>Postnatal only (9.3% overall)</th><th>Both (7.6% overall)</th><th></th><th>Fecal sIgA</th><th></th><th>Low sIgA</th><th></th></th<></thcontrol<>		Lower symptoms (71.2% overall)	Prenatal only (11.9% overall)	Postnatal only (9.3% overall)	Both (7.6% overall)		Fecal sIgA		Low sIgA	
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $			N (%)	N (%)	N (%)	p-value (χ^2)	median (IQR)	p-value	N (%)	p-value
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Child sex									
						0.71		0.60		0.79
$ \begin{array}{ccccccc} \mathbf{Yes} & 27(509) & 11(20.3) & 8(15.1) & 7(13.2) & 0.004 & 5.1(2.2-8.7) & 0.073 & 16(20.2) & 8.2 \\ Maternal asthmallergy during pregnatey model of the set of the$		92 (68.1)	19 (14.1)	13 (9.6)	11 (8.1)		6.1 (3.2 – 11.0)		33 (24.3)	
No62 (62)24 (68)20 (82)15 (6.1)62 (3 - 1.5)59 (24)Weat and studing brigging pregnate135 (71.8)21 (11.2)15 (8.8)17 (9)0.3757 (3.3 - 1.0)0.4444 (3.2)0.51No126 (70.9)14 (12.7)13 (11.8)54 (4.5)17 (9)0.3757 (3.3 - 1.0)0.4144 (23.9)0.51Pregnate0128 (73.6)19 (15.3)11 (8.9)0.5451 (3.0 - 1.0)0.1152 (3.2)0.20Vaginal128 (71.7)23 (10.8)14 (46.0)0.266.3 (3.2 - 11.4)0.5151 (3.8)0.51Petitize puole22 (71.9)31 (9.4)41 (2.5)21 (6.1)0.2652 (2.7 - 11.2)17 (11.5)17 (13.5) <t< td=""><td></td><td>27 (50.0)</td><td>11 (20.0)</td><td>0 (15.1)</td><td>7 (12 2)</td><td></td><td>5 1 (3 3 0 7)</td><td>0.072</td><td>16 (20.2)</td><td>0.25</td></t<>		27 (50.0)	11 (20.0)	0 (15.1)	7 (12 2)		5 1 (3 3 0 7)	0.072	16 (20.2)	0.25
$\begin{tabular}{ c c c c c c c c c c c c c c c c c c c$						0.004		0.073		0.35
Yes135 (71.8)21 (11.2)15 (8)17 (9)0.375.7 (3.3 - 11.0)0.444 (23.4)(3.3 (27)Pregnancy overwight/obse1112.7 (23 11.2)12.7 (24 11.2)12.7 (24		186 (75.9)	24 (9.8)	20 (8.2)	15 (6.1)		6.2 (3.3 - 11.5)		59 (24)	
No78 (70.9)14 (127)13 (11.8)5 (4.5)5 7 (2.8 - 11.2)31 (7.9)Pregnancy workpit/block85 (68.5)19 (15.3)11 (8.9)9 (7.3)0.545 1 (3.0 - 10.4)0.1135 (2.82)0.2No125 (71.7)23 (10.8)23 (10.8)14 (6.6)0.265 (3.3 2 - 11.4)0.515 (12.3)0.34Wainal23 (10.8)24 (12.5)26 (3.0)25 (12.7 - 10.1)0.5112 (2.3)0.340.26Multivity Cosetion20 (12.7)23 (10.8)11 (7.1)10 (6.5)0.265 (2.9 - 11.0)0.884 (2.6 0)0.43Multivity Cosetion12 (7.3)22 (14.2)11 (7.1)10 (6.5)0.515 (1.2 - 10.0)0.844 (2.6 0)0.43No98 (7.2)11 (8.1)16 (11.8)10 (6.5)0.245 (2.7 - 10.0)0.94635 (0.7)0.93Multivity Exploration age12 (2.7)0.94413 (2.2 0)0.9425 (0.1)0.9425 (0.1)0.9425 (0.1)0.94No16 (9.7)21 (14.1)20 (10.9)15 (8.2)0.9425 (8.1)0.9425 (8.1)0.9425 (8.1)0.9425 (8.1)0.9426 (8.1)0.9426 (8.1)0.9426 (8.1)0.9426 (8.1)0.9426 (8.1)0.9426 (8.1)0.9426 (8.1)0.9426 (8.1)0.9426 (8.1)0.9426 (8.1)0.9426 (8.1)0.9426 (8.1)0.9426 (8.1)0.9426 (8.1) </td <td></td> <td>125 (71.8)</td> <td>21 (11 2)</td> <td>15 (8)</td> <td>17 (0)</td> <td>0.37</td> <td>57(33 110)</td> <td>0.64</td> <td>44 (23.4)</td> <td>0.38</td>		125 (71.8)	21 (11 2)	15 (8)	17 (0)	0.37	57(33 110)	0.64	44 (23.4)	0.38
Presenters Image: second						0.57		0.04		0.58
$\begin{array}{c c c c c c c c c c c c c c c c c c c $		78 (70.5)	14 (12.7)	15 (11.6)	5 (4.5)		5.7 (2.0 - 11.2)		51 (27.7)	
No128 (7.5)17 (9.8)15 (8.614 (8)6.3 (3.5 - 12.0)40 (2.7)Delivery mode152 (71.7)23 (10.8)23 (10.8)14 (6.6)0.266.3 (3.2 - 11.4)0.5151 (2.3.8)0.3Emergency Casetion23 (10.8)10 (12.5)2 (6.3)52 (3.7 - 12.2)17 (31.5)17 (31.5)17 (31.5)Emergency Casetion37 (6.8.5)10 (18.5)11 (19.1)0 (6.1.1)5.1 (2.3 - 10.1)63 (8.2.2)17 (31.5)Emergency Casetion37 (6.8.5)20 (14.2)11 (7.1)10 (6.5)0.235.5 (2.9 - 11.0)0.384.2 (2.6.9)0.4Autibidite casetion11 (8.1)10 (6.5)0.235.5 (2.9 - 11.0)0.4233 (2.0.1)0.4335 (30.7)0.0Mattriggravida84 (73.7)15 (13.2)8 (7.7)7 (6.1)0.615.1 (2.7 - 10.0)0.4635 (30.7)0.0Multiggravida128 (9.6)21 (11.4)3 (13.0)09.437 5 (3.6 - 15.6)0.203 (13.00.7Nond to gettafonal age		85 (68 5)	19 (15.3)	11 (8 9)	9 (7.3)	0.54	51(30-104)	0.11	35 (28.2)	0.28
Deliver modeDeliver modeDeliver modeVaginal152 (71.7)23 (10.8)24 (10.8)14 (6.6)0.2053 (3.2 - 11.4)0.5117 (31.5)0.51<										
	Delivery mode	- ()			(-)					
Image of Section 37 (85) 10 (18) 1 (19) 6 (1.1) 5 (12, 8 - 10.1) 6 (18, 8) Ves 12 (7.3) 22 (14.2) 11 (7.1) 10 (6.5) 0.23 5 (5.9 - 11.0) 0.38 42 (26.9) 0.4 Gravia vertice veritite <th< td=""><td></td><td>152 (71.7)</td><td>23 (10.8)</td><td>23 (10.8)</td><td>14 (6.6)</td><td>0.26</td><td>6.3 (3.2 - 11.4)</td><td>0.51</td><td>51 (23.8)</td><td>0.36</td></th<>		152 (71.7)	23 (10.8)	23 (10.8)	14 (6.6)	0.26	6.3 (3.2 - 11.4)	0.51	51 (23.8)	0.36
Antibiotics exposure up to 3 months 1 (272.3) 22 (14.2) 11 (7.1) 10 (6.5) 23 (5 (2.9-110) 0.38 42 (2.6.9) 0.4 No 98 (72.1) 11 (8.1) 16 (11.8) 11 (8.1) 0.6 (5.3) 52 (2.9-110) 0.38 42 (2.6.9) 0.4 Printigravida 84 (73.7) 15 (13.2) 8 (7) 7 (6.1) 0.61 5.1 (2.7-10.0) 0.046 55 (3.0.7) 0.00 Small for gestational age Ves 16 (69.6) 21 (11.4) 20 (10.9) 15 (8.2) 6.4 (3.5-12.2) 0.046 35 (3.0.7) 0.0 Small for gestational age Ves 16 (69.6) 41 (7.4) 3 (13) 0 0.43 57 (3.6-15.6) 0.20 3 (13) 0.20 7 (12.5.0) 7 (1	Scheduled C-section	23 (71.9)	3 (9.4)	4 (12.5)	2 (6.3)		5.2 (3.7 – 11.2)		17 (31.5)	
Yes112 (2.3) No22 (14.2) 98 (72.1)11 (7.1) 16 (11.8)10 (6.5) (6.5)0.23 6.1 (3.4 - 12.1)0.38 6.1 (3.4 - 12.1)42 (26.9) 	Emergency C-section	37 (68.5)	10 (18.5)	1 (1.9)	6 (11.1)		5.1 (2.8 - 10.1)		6 (18.8)	
No98 (21)11 (81)16 (11.8)11 (81)6.1 (3.4 - 12.1)31 (22.6)Gravida128 (69.6)15 (13.2)8 (7)7 (6.1)0.615.1 (2.7 - 10.0)0.04635 (30.7)9.00Multigravida128 (69.6)21 (11.4)20 (10.9)15 (8.2)6.4 (3.5 - 12.2)0.04635 (30.7)9.00Small for gestational age77 (6.1)0.615.1 (2.7 - 10.0)0.04635 (30.7)9.00Small for gestational age16 (69.6)41 (74)3 (13)00.437.5 (3.6 - 15.6)0.203 (13.00.2Prematal SSRI use904 (28.6)<0.0113.9 (2.4 - 6.7)0.145 (35.7)0.30.60 (3.2 - 11.3)0.12 (4.8)0.00Postnatal SSRI use900 (10.7)25 (9.8)3 (17.6)0.0055.3 (3.0 - 8.7)0.594 (23.5)0.8Yes7 (41.2)6 (35.3)1 (5.9)3 (17.6)0.0055.3 (3.0 - 8.7)0.594 (23.5)0.8No203 (72)23 (21.3)26 (9.2)21 (6.3)0.775.1 (1.8 - 12.3)0.396 (37.5)0.20No203 (72)23 (11.8)2 (12.5)1 (6.3)0.775.1 (1.8 - 12.3)0.396 (37.5)0.20No203 (72)29 (11.4)26 (9.3)1 (76.7)0.0895.1 (2.7 - 10.0)0.0455.3 (2.7 - 11.0)0.0455.3 (2.7 - 11.0)0.0455.3 (2.7 - 11.0)0.0455.3 (2.7 - 11.0)0.0455.3 (2.7 - 11.0)0.045	Antibiotics exposure up to 3 months									
Gravital Convertiant						0.23		0.38		0.40
Primigravida Multigravida 14 (73.7) (28 (69.6) 15 (13.2) (11.4) 8 (7) (20 (10.9) 7 (6.1) (5.8.2) 0.61 5.1 (2.7 - 10.0) (4.3.5 - 12.2) 0.046 35 (30.7) (30.2) 0.046 35 (30.7) 0.046 35 (30.7) 0.046 35 (30.7) 0.046 35 (30.7) 0.046 35 (30.7) 0.01 39 (24 - 67) 0.14 5 (35.7) 0.33 0		98 (72.1)	11 (8.1)	16 (11.8)	11 (8.1)		6.1 (3.4 – 12.1)		31 (22.6)	
Multigravida 128 (69.6) 21 (11.4) 20 (10.9) 15 (8.2) 6.4 (3.5 - 12.2) 0.0 (3.5 - 12.2) 0.0 (3.5 - 12.2) Small for gestational age										
Small for gestational age Inc.						0.61		0.046		0.058
Yes16 (69, 6)4 (17, 4)3 (13)00.437,5 (3, 6 - 15, 6)0.203 (13)0.2Prenatal SSRI use22 (8)5,7 (3, 1 - 10, 9)71 (25, 6) </td <td></td> <td>128 (69.6)</td> <td>21 (11.4)</td> <td>20 (10.9)</td> <td>15 (8.2)</td> <td></td> <td>6.4 (3.5 – 12.2)</td> <td></td> <td>39 (21)</td> <td></td>		128 (69.6)	21 (11.4)	20 (10.9)	15 (8.2)		6.4 (3.5 – 12.2)		39 (21)	
No196 (71.3)32 (11.6)25 (9.1)22 (8) $5.7 (3.1 - 10.9)$ $71 (25.6)$ Prenatal SSRI use		16 (60.6)		2 (12)	0	0.42	7506 150	0.00	2 (12)	0.00
Prenatal SSR1 use Constraint of the second seco						0.43		0.20		0.22
Yes4 (28.6)6 (42.9)04 (28.6)<0.0013.9 (2.4 - 6.7)0.145 (35.7)0.3No209 (73.3)30 (10.5)28 (9.8)18 (6.3)0.005 5.3 (3.0 - 8.7)0.145 (35.7)0.3Postnatal SSRI use		196 (71.3)	32 (11.6)	25 (9.1)	22 (8)		5.7 (5.1 – 10.9)		/1 (25.6)	
No209 (73.3) 30 (10.5) 28 (9.8) 18 (6.3) 6.0 (3.2 - 11.3) 71 (24.8)Postnatal SSRI useYes 7 (41.2) 6 (35.3) 1 (5.9) 3 (17.6) 0.005 5.3 (3.0 - 8.7) 0.59 4 (23.5) 0.8 Prenatal smoke exposure 71 (25.3) 25 (8.9) 20 (7.1) 71 (24.8) 71 (25.3) 71 (24.8)Prenatal smoke exposure 71 (25.3) 20 (7.2) 30 (10.7) 25 (8.9) 20 (7.1) 5.3 (3.0 - 8.7) 0.59 4 (23.5) 0.8 Postnatal smoke exposure 71 (25.3) 22 (12.5) 1 (6.3) 0.77 5.1 (1.8 - 12.3) 0.39 6 (37.5) 0.2 Postnatal smoke exposure 71 (24.3) 26 (9.2) 21 (7.4) 0.26 4.9 (2.8 - 9.6) 0.25 14 (30.4) 0.3 Post at home 71 (24.3) 25 (9.8) 17 (6.7) 0.089 5.1 (2.7 - 10.0) 0.045 43 (29.3) 0.1 Pets at home 71 (26.7) 19 (12.4) 85 (5.4) 15 (10.2) 0.089 5.1 (2.7 - 10.0) 0.045 43 (29.3) 0.2 Other children at home 71 (24.8) 19 (12.4) 15 (10.9) 10 (7.2) 0.33 6.4 (3.7 - 11.0) 0.16 30 (21.6) 0.21 Pets at forme 71 (24.8) 12 (7.4) 12 (7.4) 12 (7.4) 12 (7.4) 5.3 (2.7 - 11.0) 1.6 (3.6 (2.6) 7.2 (2.4.1)Pets at home 71 (25.8) 71 (24.8) 10 (7.2) 0.33 6.4 (3.7		1 (29 6)	6 (42 0)	0	1 (28 6)	<0.001	20(24 67)	0.14	5 (25 7)	0.26
Postnatal SSR1 use Indication Indication <th< td=""><td></td><td></td><td></td><td></td><td></td><td><0.001</td><td></td><td>0.14</td><td></td><td>0.50</td></th<>						<0.001		0.14		0.50
Yes7 (41.2)6 (35.3)1 (5.9)3 (17.6)0.0055.3 (3.0 - 8.7)0.594 (23.5)0.8No205 (8.9)20 (7.1)5.7 (3.1 - 11.4)71 (25.3)Prenatal smoke exposure71 (25.3)10 (62.5)3 (18.8)2 (12.5)1 (6.3)0.775.1 (1.8 - 12.3)0.396 (37.5)0.2No203 (72)32 (11.3)26 (9.2)21 (7.4)6.0 (3.2 - 11.1)0.396 (37.5)0.2Postnatal smoke exposure71 (5.2)2 (4.3)6 (13)0.264.9 (2.8 - 9.6)0.2514 (30.4)0.3No184 (72.2)29 (11.4)25 (9.8)17 (6.7)6.0 (3.2 - 11.9)6.0430.430.430.43Pets at home71 (5.2)2 (4.3)6 (13)0.264.9 (2.8 - 9.6)0.2514 (30.4)0.3Yes106 (72.1)18 (12.2)8 (5.4)15 (10.2)0.0895.1 (2.7 - 10.0)0.04543 (29.3)0.1No108 (72.1)18 (11.8)19 (12.4)8 (5.2)6.4 (3.6 - 12.3)33 (21.4)33 (21.4)Other children at home7172 (2.87)15 (10.9)10 (7.2)0.336.4 (3.7 - 11.0)0.1630 (21.6)0.20No101 (73.2)12 (8.7)15 (10.9)10 (7.2)0.336.4 (3.7 - 11.0)0.1630 (21.6)0.20No101 (73.2)12 (8.7)15 (10.9)10 (7.2)0.336.4 (3.7 - 11.0)0.1630 (21.6)0.20Reastfeeding at stool collection79		209 (75.5)	50 (10.5)	28 (9.8)	18 (0.5)		0.0 (3.2 - 11.3)		/1 (24.8)	
No $205 (73.2)$ $30 (10.7)$ $25 (8.9)$ $20 (7.1)$ $5.7 (3.1 - 11.4)$ $71 (25.3)$ Prenatal smoke exposure		7 (41 2)	6 (35 3)	1 (5.9)	3 (17.6)	0.005	53(30-87)	0.59	4 (23 5)	0.89
Prenatal smoke exposure COL						0.000		0.57		0.07
Yes $10(62.5)$ $3(18.8)$ $2(12.5)$ $1(6.3)$ 0.77 $5.1(18-12.3)$ 0.39 $6(37.5)$ 0.2 No $203(72)$ $32(11.3)$ $26(9.2)$ $21(7.4)$ $60(3.2-11.1)$ $69(24.4)$ Postnatal smoke exposure Ves $31(67.4)$ $7(15.2)$ $2(4.3)$ $6(13)$ 0.26 $4.9(2.8-9.6)$ 0.25 $14(30.4)$ 0.3 No $184(72.2)$ $29(11.4)$ $25(9.8)$ $17(6.7)$ $6.0(3.2-11.9)$ $62(24.2)$ Pets at home Ves $106(72.1)$ $18(12.2)$ $8(5.4)$ $15(10.2)$ 0.089 $5.1(2.7-10.0)$ 0.045 $43(29.3)$ 0.1 No $108(70.6)$ $18(11.8)$ $19(12.4)$ $8(5.2)$ $6.4(3.6-12.3)$ 0.16 $30(21.6)$ 0.2 Other children at home Ves $101(73.2)$ $12(8.7)$ $15(10.9)$ $10(7.2)$ 0.33 $6.4(3.7-11.0)$ 0.16 $30(21.6)$ 0.2 Preastfeeding at stool collection $Zero$ $59(72)$ $9(11)$ $8(9.8)$ $6(7.3)$ 0.60 $3.7(2.1-6.0)$ <0.001 $36(43.6)$ <0.2 Partial $79(70.5)$ $10(8.9)$ $14(12.5)$ $9(8)$ $5.6(3.3-10.3)$ $27(24.1)$ $<0(11.5)$ Infant allergic ezema $Ues(54.7)$ $12(13.8)$ $44(4.6)$ 66.9 $10.4(5.6-18.1)$ $00(11.5)$		200 (10.2)	50 (10.7)	20 (0.5)	20 (7.1)		0.7 (0.1 11.1)		,1 (20.0)	
No $203\ (72)$ $32\ (11.3)$ $26\ (9.2)$ $21\ (7.4)$ $6.0\ (3.2-11.1)$ $69\ (24.4)$ Postnatal smoke exposureYes $31\ (67.4)$ $7\ (15.2)$ $2\ (4.3)$ $6\ (13)$ 0.26 $4.9\ (2.8-9.6)$ 0.25 $14\ (30.4)$ 0.3 No $184\ (72.2)$ $29\ (11.4)$ $25\ (9.8)$ $17\ (6.7)$ $6.0\ (3.2-11.9)$ $62\ (24.2)$ Pets at home Ves $106\ (72.1)$ $18\ (12.2)$ $8\ (5.4)$ $15\ (10.2)$ 0.089 $5.1\ (2.7-10.0)$ 0.045 $43\ (29.3)$ 0.1 Other children at home Ves $100\ (7.2)$ $10\ (7.2)$ 0.33 $6.4\ (3.5-11.0)$ $0.16\ 30\ (21.6)$ 0.2 Other children at home Ves $101\ (73.2)$ $12\ (8.7)$ $15\ (10.9)$ $10\ (7.2)$ $0.33\ 6.4\ (3.7-11.0)$ $0.16\ 30\ (21.6)$ 0.2 Breastfeeding at stool collection $Zero$ $59\ (72)$ $9\ (11)$ $8\ (9.8)$ $6\ (7.3)$ $0.60\ 3.7\ (2.1-6.0)$ $<0.001\ 36\ (43.6)$ $<0.27\ (24.1)$ Zero $59\ (72.7)$ $9\ (11)$ $8\ (9.8)$ $6\ (7.3)$ $0.60\ 3.7\ (2.1-6.0)$ $<0.001\ 36\ (43.6)$ $<0.27\ (24.1)$ Infant allergic eczema $12\ (13.8)$ $4\ (4.6)$ $6\ (6.9)$ $10.4\ (5.6-18.1)$ $10\ (11.5)$		10 (62.5)	3 (18.8)	2(12.5)	1 (6.3)	0.77	5.1(1.8 - 12.3)	0.39	6 (37.5)	0.24
Yes 31 (67.4) 7 (15.2) 2 (4.3) 6 (13) 0.26 4.9 (2.8 - 9.6) 0.25 14 (30.4) 0.3 No 18 (72.2) 29 (11.4) 25 (9.8) 17 (6.7) 6.0 (3.2 - 11.9) 62 (24.2) Pets at home	No	203 (72)	32 (11.3)	26 (9.2)	21 (7.4)		6.0 (3.2 – 11.1)		69 (24.4)	
No 184 (72.2) 29 (11.4) 25 (9.8) 17 (6.7) 6.0 (3.2 - 11.9) 62 (24.2) Pets at home	Postnatal smoke exposure									
Pets at home Ves 106 (72.1) 18 (12.2) 8 (5.4) 15 (10.2) 0.089 5.1 (2.7 - 10.0) 0.045 43 (29.3) 0.1 No 108 (70.6) 18 (11.8) 19 (12.4) 8 (5.2) 6.4 (3.6 - 12.3) 33 (21.4) 0.045 43 (29.3) 0.1 Other children at home Ves 6.4 (3.6 - 12.3) 33 (21.4) 0.16 30 (21.6) 0.2 0.045 45 (27.8) 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 0.16 30 (21.6) 0.2 10 10 10 10 10 10						0.26		0.25		0.37
Yes 106 (72.1) 18 (12.2) 8 (5.4) 15 (10.2) 0.089 5.1 (2.7 - 10.0) 0.045 43 (29.3) 0.1 No 108 (70.6) 18 (11.8) 19 (12.4) 8 (5.2) 6.4 (3.6 - 12.3) 33 (21.4) Other children at home Ves 0.101 (73.2) 12 (8.7) 15 (10.9) 10 (7.2) 0.33 6.4 (3.7 - 11.0) 0.16 30 (21.6) 0.2 No 114 (70.4) 24 (14.8) 12 (7.4) 12 (7.4) 5.3 (2.7 - 11.0) 0.16 30 (21.6) 0.2 Breastfeeding at stool collection Zero 59 (72) 9 (11) 8 (9.8) 6 (7.3) 0.60 3.7 (2.1 - 6.0) <0.001 36 (43.6) <0.2 Partial 79 (70.5) 10 (8.9) 14 (12.5) 9 (8) 5.6 (3.3 - 10.3) 27 (24.1) Exclusive 65 (74.7) 12 (13.8) 4 (4.6) 6 (6.9) 10.4 (5.6 - 18.1) 10 (11.5)		184 (72.2)	29 (11.4)	25 (9.8)	17 (6.7)		6.0 (3.2 - 11.9)		62 (24.2)	
No 108 (70.6) 18 (11.8) 19 (12.4) 8 (5.2) 6.4 (3.6 - 12.3) 33 (21.4) Other children at home										
Other children at home Yes 101 (73.2) 12 (8.7) 15 (10.9) 10 (7.2) 0.33 6.4 (3.7 - 11.0) 0.16 30 (21.6) 0.2 No 114 (70.4) 24 (14.8) 12 (7.4) 12 (7.4) 5.3 (2.7 - 11.0) 0.16 30 (21.6) 0.2 Breastfeeding at stool collection 2 2 12 (7.4) 12 (7.4) 5.3 (2.7 - 11.0) 45 (27.8) Zero 59 (72) 9 (11) 8 (9.8) 6 (7.3) 0.60 3.7 (2.1 - 6.0) <0.001						0.089		0.045		0.12
Yes 101 (73.2) 12 (8.7) 15 (10.9) 10 (7.2) 0.33 6.4 (3.7 - 11.0) 0.16 30 (21.6) 0.2 No 114 (70.4) 24 (14.8) 12 (7.4) 12 (7.4) 5.3 (2.7 - 11.0) 45 (27.8) Breastfeeding at stool collection Zero 59 (72) 9 (11) 8 (9.8) 6 (7.3) 0.60 3.7 (2.1 - 6.0) <0.01 36 (43.6) <0.01 Partial 79 (70.5) 10 (8.9) 14 (12.5) 9 (8) 5.6 (3.3 - 10.3) 27 (24.1) Exclusive 65 (74.7) 12 (13.8) 4 (4.6) 6 (6.9) 10.4 (5.6 - 18.1) 10 (11.5)		108 (70.6)	18 (11.8)	19 (12.4)	8 (5.2)		6.4 (3.6 – 12.3)		33 (21.4)	
No 114 (70.4) 24 (14.8) 12 (7.4) 12 (7.4) 5.3 (2.7 - 11.0) 45 (27.8) Breastfeeding at stool collection Zero 59 (72) 9 (11) 8 (9.8) 6 (7.3) 0.60 3.7 (2.1 - 6.0) <0.001 36 (43.6) <0.001 Partial 79 (70.5) 10 (8.9) 14 (12.5) 9 (8) 5.6 (3.3 - 10.3) 27 (24.1) Exclusive 65 (74.7) 12 (13.8) 4 (4.6) 6 (6.9) 10.4 (5.6 - 18.1) 10 (11.5)		101 (72.0)	12 (0.7)	15 (10.0)	10 (7.2)	0.22	6 4 (2 7 11 0)	0.16	20 (21 ()	0.00
Breastfeeding at stool collection Zero 59 (72) 9 (11) 8 (9.8) 6 (7.3) 0.60 3.7 (2.1 - 6.0) <0.001 36 (43.6) <0. Partial 79 (70.5) 10 (8.9) 14 (12.5) 9 (8) 5.6 (3.3 - 10.3) 27 (24.1) Exclusive 65 (74.7) 12 (13.8) 4 (4.6) 6 (6.9) 10.4 (5.6 - 18.1) 10 (11.5)						0.33		0.16		0.22
Zero 59 (72) 9 (11) 8 (9.8) 6 (7.3) 0.60 3.7 (2.1 - 6.0) <0.01 36 (43.6) <0. Partial 79 (70.5) 10 (8.9) 14 (12.5) 9 (8) 5.6 (3.3 - 10.3) 27 (24.1) Exclusive 65 (74.7) 12 (13.8) 4 (4.6) 6 (6.9) 10.4 (5.6 - 18.1) 10 (11.5)		114 (70.4)	24 (14.8)	12 (7.4)	12 (7.4)		5.5(2.7 - 11.0)		45 (27.8)	
Partial 79 (70.5) 10 (8.9) 14 (12.5) 9 (8) 5.6 (3.3 - 10.3) 27 (24.1) Exclusive 65 (74.7) 12 (13.8) 4 (4.6) 6 (6.9) 10.4 (5.6 - 18.1) 10 (11.5) Infant allergic eczema		50 (72)	0(11)	8 (0.8)	6 (7 3)	0.60	37(21 60)	<0.001	36 (13 6)	<0.001
Exclusive 65 (74.7) 12 (13.8) 4 (4.6) 6 (6.9) 10.4 (5.6 - 18.1) 10 (11.5) Infant allergic eczema 10 (11.5)						0.00		~0.001		~0.001
Infant allergic eczema										
		05 (14.7)	12 (13.0)	4 (4.0)	0 (0.2)		10.4 (0.0 - 10.1)		10(11.5)	
100 - 100		20 (57 1)	4 (11 4)	4 (11 4)	7 (20)	0.025	59(32 - 112)	0.90	8 (22.9)	0.76
No 194 (73.5) 32 (12.1) 22 (8.3) 16 (6.1) 5.7 (3.1 - 11.0) 67 (25.3)						0.020		0.70		0.70

Table A2. Median comparisons of infant sIgA between maternal depressive symptoms groups when mothers are overweight/obese and normal weight (N=304)

sIgA (mg/g feces)	Refer	ence group										Interaction
Median (IQR)	Ν	Lower symptoms	Ν	Prenatal only	p-value	Ν	Postnatal only	p-value	Ν	Both	p-value	p-value
Depressive	215	6.42	36	5.27	0.22	28	5.70	0.28	23	3.70 (1.78 - 8.09)	0.027*	-
symptoms		(3.52 - 12.40)		(2.64 - 10.29)			(2.64 – 9.06)					
Pregnancy	85	5.46	19	4.77	0.35	11	5.66	0.73	9	2.30 (1.61 - 8.70)	0.047*	0.95
overweight/obese		(3.28 – 11.27)		(2.37 – 9.66)			(3.07 – 11.23)					
Normal weight	128	7.20	17	5.59	0.52	15	6.04	0.36	14	5.14 (2.54 - 8.32)	0.15	
		(3.58 – 13.14)		(3.29 - 10.74)			(2.62 - 9.23)					

*Significant at alpha of 0.05 determined a priori

Table A3. Logistic regression analyses of lowest quartile sIgA and maternal depressive symptoms (N=304)

(Ref: Less depressive	N for lowest	OR for depressive	OR for covariate	Decision
symptoms)	quartile sIgA	symptoms		
5 1)	1 0	5 1		
Crude OR	76	1.38 (0.62 - 3.05)		
		1.69 (0.72 - 3.99)		
		2.75 (1.13 - 6.67)		
Adjusted for child sex	M-43	1.39 (0.62 - 3.09)	0.91 (0.53 - 1.54)	Out
	F-33	1.70 (0.72 – 4.01)		
		2.77 (1.14 – 6.71)		
Adjusted for maternal	Y-44	1.44 (0.64 – 3.22)	0.75 (0.43 – 1.29)	In
asthma/allergy	N-31	1.67 (0.71 – 3.96)		
		3.17 (1.28 - 7.84)		
Adjusted for pregnancy	Y-35	1.31 (0.59 – 2.93)	1.32 (0.77 – 2.24)	Out
overweight	N-40	1.56 (0.64 - 3.82)		
		2.73 (1.13 - 6.64)		
Adjusted for depression	Y-16	1.41 (0.63 – 3.18)	1.17 (0.59 – 2.31)	Out
history	N-59	1.68 (0.71 – 3.98)		
		2.94 (1.18 - 7.30)		
Adjusted for gravida	Y-35	1.42 (0.63 - 3.18)	1.75 (1.02 – 3.02)	In (p<0.20)
	N-39	1.89 (0.79 – 4.51)		
		3.30 (1.32 - 8.21)		
Adjusted for birth mode	El-6	1.37 (0.61 – 3.06)	1.44 (0.73 – 2.82)	In
	Em-17	1.88 (0.79 – 4.48)	0.72 (0.28 – 1.87)	
	V-51	2.99 (1.21 – 7.41)		
Adjusted for SGA	Y-3	1.47 (0.66 – 3.28)	0.45 (0.13 – 1.58)	In (marginally
	N-71	1.80 (0.76 – 4.28)		significant)
		2.95 (1.19 - 7.28)		-
Adjusted for antibiotics	Y-42	1.54 (0.68 – 3.49)	1.29 (0.75 – 2.23)	In
exposure	N-31	1.60(0.65 - 3.90)		
	F 10	3.40 (1.35 - 8.53)		
Adjusted for	E-10	1.52(0.62 - 3.70) 1.28(0.54 - 3.50)	6.25 (2.80 - 13.96)	In (always in model)
breastfeeding at time of	P-27	1.38(0.54 - 3.50) 2.42(1.20 - 0.07)	2.45 (1.10 – 5.46)	
stool collection	Z-36 Y-43	3.43(1.30-9.07)	1.51 (0.88 - 2.58)	$I_{\rm m}$ (m < 0.20)
Adjusted for pets at		1.37 (0.61 - 3.05) 1.94 (0.81 - 4.66)	1.51 (0.88 – 2.58)	In (p<0.20)
home	N-33			
Adjusted for other	Y-30	2.59 (1.06 – 6.31) 1.32 (0.59 – 2.94)	0.72 (0.42 – 1.23)	In
children	N-45		0.72(0.42 - 1.23)	111
ciliaren	IN-43	$\begin{array}{c} 1.85 \ (0.78 - 4.39) \\ 2.48 \ (0.99 - 6.16) \end{array}$		
Adjusted for prenatal	Y-5	1.31 (0.58 - 2.97)	1.28 (0.38 - 4.27)	Out
SSRI use	N-71	1.51(0.58 - 2.97) 1.68(0.71 - 3.96)	1.28 (0.38 - 4.27)	Out
SSICI use	11-/1	2.83 (1.13 - 7.11)		
Adjusted for postnatal	Y-4	1.40(0.62 - 3.15)	0.75 (0.23 – 2.47)	Out
SSRI use	N-71	1.40(0.02 - 3.13) 1.56(0.64 - 3.82)	0.75 (0.25 - 2.47)	Out
SSICIUSE	14-71	2.78(1.14 - 6.79)		
Adjusted for prenatal	Y-6	1.42 (0.63 - 3.18)	1.83 (0.63 - 5.28)	In
smoke exposure	N-69	1.70(0.72 - 4.01)	1.05 (0.05 5.20)	
shioke exposure	11 05	3.04(1.23-7.50)		
Adjusted for postnatal	Y-14	1.36(0.61 - 3.02)	1.31 (0.65 – 2.65)	Out
smoke exposure	N-62	1.82(0.77 - 4.34)	1.51 (0.05 2.05)	
supposite enposite		2.67 (1.10 - 6.50)		
Adjusted for infant	Y-8	1.41 (0.64 - 3.15)	0.71 (0.30 - 1.70)	In
allergic eczema	N-67	1.97(0.82 - 4.73)	0.71 (0.50 1.70)	
		3.02(1.22 - 7.50)		
Adjusted for birth	73	1.59(0.69 - 3.67)		
characteristics (gravida,	, 5	1.81(0.72 - 4.51)		
birth mode, antibiotics,		3.37 (1.32 - 8.62)		
	L	5.57 (1.52 0.02)	1	1

SGA)			
Adjusted for gravida,	73	1.64 (0.72 – 3.75)	Keep birth mode out
antibiotics, SGA		1.74(0.70 - 4.32)	1
		3.43 (1.35 - 8.72)	
Adjusted for gravida and	73	1.57 (0.69 - 3.57)	Keep SGA out
antibiotics		1.71(0.69 - 4.21)	L L
		3.60 (1.42 – 9.13)	
Adjusted for home	75	1.31 (0.59 – 2.93)	In
environment (pets and		2.00(0.83 - 4.81)	
children)		2.34(0.94 - 5.87)	
Adjusted for home	72	1.40 (0.57 - 3.46)	
environment and		1.69(0.64 - 4.44)	
breastfeeding		2.88 (1.06 - 7.83)	
Adjusted for other	72	1.42 (0.58 - 3.51)	Keep pets out
children and		1.57(0.61 - 4.06)	r r r
breastfeeding		3.04 (1.12 - 8.25)	
Adjusted for pets and	73	1.49 (0.61 – 3.65)	Keep children in
breastfeeding		1.65(0.63 - 4.30)	
e		3.22 (1.21 – 8.56)	
Adjusted for	72	1.70 (0.69 - 4.18)	Keep asthma/allergy
breastfeeding and		1.34(0.52 - 3.46)	in
maternal asthma/allergy		3.76 (1.40 – 10.10)	
Adjusted for	72	1.47 (0.60 – 3.60)	Keep overweight in
breastfeeding and	, <u> </u>	1.23 (0.46 - 3.27)	
pregnancy overweight		3.40 (1.28 - 9.03)	
Adjusted for	72	1.66(0.67 - 4.11)	Keep smoke out
breastfeeding and	12	1.39(0.54 - 3.57)	reep shioke out
prenatal smoke exposure		3.53 (1.33 – 9.40)	
Adjusted for	73	1.52 (0.62 - 3.71)	In
breastfeeding and	15	1.52(0.52 - 3.92) 1.52(0.59 - 3.92)	
postnatal smoke		3.44(1.29-9.12)	
exposure		5 (I. <u>_</u>)	
Adjusted for	73	1.42 (0.57 – 3.54)	In
breastfeeding and	15	1.36(0.53 - 3.46)	111
prenatal SSRI		3.10 (1.12 – 8.57)	
Adjusted for	72	1.60 (0.65 – 3.96)	Out
breastfeeding and	12	1.24 (0.46 - 3.30)	out
postnatal SSRI		3.67 (1.36 – 9.95)	
Adjusted for	72	$\frac{1.53(0.63 - 3.74)}{1.53(0.63 - 3.74)}$	In
breastfeeding and infant	12	1.62(0.62 - 4.19)	111
allergic eczema		3.70(1.36 - 10.11)	
Adjusted for gravida,	70	1.81 (0.71 – 4.63)	In
antibiotics,	70	1.29(0.48 - 3.48)	111
breastfeeding		4.82(1.68 - 13.84)	
Adjusted for gravida,	69	2.02 (0.78 - 5.24)	In
antibiotics,	0)	1.28(0.47 - 3.49)	111
breastfeeding, maternal		5.18(1.79 - 15.03)	
asthma/allergy		5.18 (1.79 - 15.05)	
Adjusted for previous	68	1.95 (0.76 – 5.09)	Keep overweight out
and overweight	00	1.95(0.76 - 5.09) 1.13(0.39 - 3.26)	Keep overweight out
and overweight			
Adjusted for gravida,	69	5.15 (1.77 – 15.03) 2.01 (0.77 – 5.23)	In
antibiotics,	07	2.01(0.77 - 5.23) 1.45(0.52 - 4.02)	111
breastfeeding,		4.56(1.53 - 13.62)	
asthma/allergy, other		4.50 (1.55 - 15.02)	
children	60	1.02 (0.75 4.00)	V and and 1 and
Adjusted for antibiotics,	68	1.93 (0.75 - 4.99) 1.28 (0.50 - 2.80)	Keep gravida out
breastfeeding,		1.38(0.50 - 3.80) 4.24(1.46 - 12.02)	
asthma/allergy, other		4.34 (1.46 – 12.93)	
children	(0)		· ·
Adjusted for previous &	68	1.73 (0.65 – 4.59)	In

prenatal SSRI		1.37 (0.50 - 3.77) 3.55 (1.13 - 11.13)	
Adjusted for previous & postnatal smoke	68	$\begin{array}{c} 1.75 & (0.66 - 4.66) \\ 1.35 & (0.49 - 3.73) \\ 3.56 & (1.14 - 11.16) \end{array}$	Keep smoke out
Adjusted for previous & prenatal smoke	69	$\begin{array}{c} 1.69 \ (0.63 - 4.49) \\ 1.34 \ (0.48 - 3.71) \\ 3.54 \ (1.12 - 11.14) \end{array}$	Keep smoke out
Adjusted for antibiotics, breastfeeding, other children, maternal asthma/allergy, prenatal SSRI, infant allergic eczema	67	1.75 (0.66 - 4.62) 1.47 (0.53 - 4.08) 3.83 (1.19 - 12.36)	Keep infant eczema out
Adjusted for antibiotics, breastfeeding, other children, maternal asthma/allergy, prenatal SSRI	68	1.73 (0.65 - 4.59) 1.37 (0.50 - 3.77) 3.55 (1.13 - 11.13)	Final Model (No significant interactions)

Appendix B: Preliminary analyses with perceived stress cut-off at mean score of CHILD cohort (12.96)

sIgA (mg/g feces)	All infants	Lower depressive	Higher depressive	Lower perceived stress	Above CHILD mean
Median (IQR)		symptoms	symptoms (above 16 for		stress (for prenatal,
			prenatal, postnatal or both)		postnatal or both)
1 st quartile (lowest)	1.88 (1.15 – 2.55)	1.89 (1.16 – 2.60)	1.87 (1.11 – 2.48)	1.57 (1.07 – 2.10)	2.01 (1.18 - 2.58)
2 nd quartile	4.46 (3.80 - 5.12)	4.33 (3.81 - 5.04)	4.81 (3.74 - 5.59)	4.33 (3.61 – 4.90)	4.73 (3.86 - 5.29)
3 rd quartile	8.00 (6.62 - 9.67)	8.00 (6.61 - 9.82)	7.99 (6.87 – 9.34)	8.00 (6.64 - 9.62)	8.00 (6.62 - 9.80)
4 th quartile	15.82 (13.44 - 21.64)	15.71 (13.21 – 20.08)	18.03 (13.81 – 22.94)	15.43 (13.15 – 25.17)	15.97 (13.48 – 21.32)
1 st tertile (lowest)	2.32 (1.42 - 3.24)	2.46 (1.50 - 3.35)	2.23 (1.27 - 3.00)	2.09 (1.27 - 3.35)	2.33 (1.51 – 3.21)
2 nd tertile	5.97 (4.97 - 7.11)	5.79 (4.87 - 6.83)	6.19 (5.45 - 7.60)	6.10 (4.86 - 7.20)	5.96 (5.08 - 7.07)
3 rd tertile	14.06 (11.02 - 19.41)	14.01 (11.11 - 19.03)	14.14 (10.80 - 21.94)	14.26 (11.03 - 19.58)	13.95 (11.01 – 19.22)

Table B1. Quartile and tertile medians for sIgA (N=403)

Note: There is a suggestion that a greater difference between greater distress and lower distress groups with the tertiles. Logistic

regression analyses were tested for both lowest tertile and lowest quartile (yes/no).

	Lower stress (28.4% overall)	Prenatal only stress (19.5% overall)	Postnatal only stress (14.7% overall)	Both (37.4% overall)		Fecal sIgA		Low sIgA	
		N (%)	N (%)	N (%)	p-value (χ^2)	median (IQR)	p-value	N (%)	p-value
Child sex									
Male	66 (29.5)	45 (20.1)	33 (14.7)	80 (35.7)	0.88	6.1 (3.3 – 11.7)	0.58	54 (24)	0.67
Female	48 (27.1)	33 (18.6)	26 (14.7)	70 (39.5)		5.6 (3.2 - 10.4)		46 (25.8)	
Depression History									
Yes	19 (24.1)	15 (19)	6 (7.6)	29 (49.4)	0.056	6.1 (3.3 – 11.4)	0.04	25 (31.6)	0.13
No	93 (29.5)	62 (19.7)	51 (16.2)	109 (34.6)		5.1 (2.3 - 9.0)		74 (23.4)	
Aaternal asthma/allergy during									
oregnancy									
Yes	78 (29.3)	50 (18.8)	32 (12)	106 (39.8)	0.17	6.0 (3.2 - 12.0)	0.49	66 (24.7)	0.99
No	34 (26.6)	27 (21.1)	25 (19.5)	42 (32.8)		5.7(3.2 - 9.8)		32 (24.8)	
Delivery mode									
Vaginal	85 (30.1)	52 (18.4)	43 (15.2)	102 (36.2)	0.64	5.6 (3.6 - 10.4)	0.50	70 (24.6)	0.56
C-elective	12 (25.5)	11 (23.4)	4 (8.5)	20 (42.6)		5.3(2.9 - 10.4)		19 (27.9)	
C-emergent	15 (22.1)	15 (22.1)	12 (17.6)	26 (38.2)		6.2(3.2 - 11.4)		9 (19.1)	
Antibiotics exposure up to 3 months									
Yes	62 (28.2)	43 (19.5)	33 (15)	82 (37.3)	0.99	5.6 (3.1 - 11.0)	0.28	58 (26.2)	0.43
No	50 (29.4)	34 (20)	24 (14.1)	62 (36.5)		6.1(3.4 - 12.0)		39 (22.8)	
Gravida	50 (2).1)	5.(20)	2.()	02 (00.0)		0.1 (0.1 12.0)		57 (22.0)	
Primigravida	46 (30.3)	27 (17.8)	25 (16.4)	54 (35.5)	0.70	5.2(3.0 - 10.0)	0.077	43 (28.3)	0.18
Multigravida	66 (26.9)	51 (20.8)	34 (13.9)	94 (38.4)	0.70	6.4(3.5-12.0)	0.077	55 (22.3)	0.10
mall for gestational age	00 (20.9)	51 (20.0)	51(15.5)	91 (30.1)		0.1 (5.5 12.0)		55 (22.5)	
Yes	8 (29.6)	8 (29.6)	2 (7.4)	9 (33.3)	0.45	7.5 (4.7 – 15.7)	0.063	3 (11.1)	0.09
No	103 (28)	70 (19)	57 (15.5)	138 (37.5)	0.45	6.0(3.2 - 10.9)	0.005	95 (25.7)	0.07
Prenatal SSRI use	105 (20)	70(1))	57 (15.5)	150 (57.5)		0.0 (5.2 10.7))5 (25.1)	
Yes	0	3 (16.7)	0 (0)	15 (83.3)	<0.001	3.8(2.7-6.7)	0.12	6 (33)	0.42
No	109 (29.1)	75 (20.1)	59 (15.8)	131 (35)	<0.001	6.0(3.2 - 11.0)	0.12	93 (24.8)	0.42
Postnatal SSRI use	107 (27.1)	75 (20.1)	57 (15.6)	151 (55)		0.0 (5.2 11.0))) (24.0)	
Yes	2 (8.7)	5 (21.7)	2 (8.7)	14 (60.9)	0.056	4.7 (3.2 – 7.1)	0.18	6 (26.1)	0.89
No	109 (29.5)	72 (19.5)	56 (15.1)	133 (35.9)	0.050	6.1(3.2 - 11.4)	0.18	92 (24.8)	0.09
Prenatal smoke exposure	109 (29.5)	72 (19.5)	50 (15.1)	155 (55.9)		0.1 (3.2 - 11.4)		92 (24.0)	
Yes	3 (12.5)	9 (37.5)	3 (12.5)	9 (37.5)	0.087	5.1 (1.8 – 12.3)	0.39	6 (37.5)	0.24
No	109 (29.4)	68 (18.3)	54 (14.6)	140 (37.7)	0.087	6.0(3.2 - 11.1)	0.59	69 (24.4)	0.24
	109 (29.4)	08 (18.5)	54 (14.0)	140 (37.7)		0.0(5.2 - 11.1)		09 (24.4)	
Postnatal smoke exposure Yes	12 (18.8)	13 (20.3)	7 (10.9)	32 (50)	0.084	4.9 (2.8 - 9.6)	0.25	14 (30.4)	0.37
					0.084	()	0.23		0.57
No	102 (30.4)	65 (19.3)	52 (15.5)	117 (34.8)		6.1 (3.2 – 11.5)		62 (24.4)	
Pets at home	40 (2(5)	26 (10.5)	20 (15 7)	71 (20 4)	0.97	5 2 (2 7 10 4)	0.021	55 (20.7)	0.045
Yes	49 (26.5)	36 (19.5)	29 (15.7)	71 (38.4)	0.86	5.3(2.7-10.4)	0.031	55 (29.7)	0.045
No	64 (30)	42 (19.7)	30 (14.1)	77 (36.2)		6.4 (3.7 – 12.0)		45 (21)	
Children at home	54 (00 ()	20 (20 ()	29 (14.9)	(0.(2())	0.05	6.5 (2.6 11.0)	0.11	56 (06 0)	0.24
Yes	54 (28.6)	39 (20.6)	28 (14.8)	68 (36)	0.95	6.5 (3.6 – 11.9)	0.11	56 (26.8)	0.34
No	59 (28.2)	39 (18.7)	31 (14.8)	80 (38.3)	_	5.3 (3.1 – 10.7)	_	43 (22.6)	
creastfeeding at stool collection	22 (22)	21 (21)	14 (14)	12 (12)	0.57	22/20 (0)		15 (15)	.0.001
Zero	23 (23)	21 (21)	14 (14)	42 (42)	0.57	3.3 (2.0 – 6.0)	< 0.001	47 (47)	< 0.001
Partial	50 (33.3)	28 (18.7)	19 (12.7)	53 (35.3)		5.4 (3.2 – 9.8)		39 (26)	
Exclusive	40 (30.5)	23 (17.6)	23 (17.6)	45 (34.4)		10.0 (5.6 – 16.1)		11 (8.4)	
nfant allergic eczema									
Yes	10 (23.3)	5 (11.6)	9 (20.9)	19 (44.2)	0.28	5.7 (3.4 – 11.4)	0.84	9 (20.9)	0.55
No	103 (29.2)	72 (20.4)	50 (14.2)	128 (36.3)		6.0(3.2 - 11.2)		89 (25.1)	

Table B2. Infant fecal sIgA levels and percentage distribution of potential covariates in relation to maternal perceived stress above the CHILD cohort mean and lowest quartile sIgA (n=403)

	Refer	ence group										
sIgA (mg/g feces) Median (IQR)	N	Less stress (1.8% SSRI use)	N	Prenatal only (6.4% SSRI use)	p-value	Ν	Postnatal only (3.4% SSRI use)	p-value	N	Both pre and postnatal stress (12% SSRI use)	p-value	p-value (K-W)
Perceived stress	114	6.17 (3.48 - 12.01)	78	6.16 (3.43-11.02)	0.92	59	5.53 (3.16 - 11.40)	0.59	150	5.68 (2.71 - 10.78)	0.43	0.78
Depression history	19	6.48 (3.53 - 12.06)	15	5.19 (1.99 - 10.84)	0.64	6	2.01 (0.54 - 4.18)	0.009**	39	5.07 (2.31 - 7.96)	0.15	0.052
No depression history	93	6.10 (3.30 – 11.31)	62	6.69 (3.43 - 11.52)	0.57	51	6.62 (3.86 - 12.70)	0.61	109	6.07 (3.10 – 11.13)	0.97	0.88
Maternal asthma or allergy	78	6.30 (3.28 - 13.14)	50	7.90 (3.43 - 11.19)	0.68	32	5.51 (3.47 - 12.50)	0.94	106	5.64 (2.92 - 11.74)	0.69	0.86
No maternal asthma or allergy	34	6.12 (3.75 - 10.53)	27	5.19 (3.02 - 9.80)	0.77	25	6.42 (1.92 - 10.63)	0.60	42	5.70 (2.47 - 9.87)	0.62	0.95
Breastfed	90	7.82 (4.16 – 13.43)	51	5.66 (2.49 - 10.35)	0.32	42	6.50 (3.22 – 9.73)	0.62	98	5.66 (3.22 - 9.73)	0.32	0.55
Not breastfed	23	4.04 (2.22 - 6.09)	21	2.78 (1.54 - 4.77)	0.03	14	2.71 (1.07 - 6.39)	0.23	42	2.46 (1.63 - 2.97)	0.032*	0.25
Male	66	6.17 (3.48 - 12.21)	45	6.27 (3.63 - 12.30)	0.77	33	6.42 (3.04 - 12.59)	0.77	80	5.70 (2.62 - 11.45)	0.57	0.87
Female	48	6.28 (3.34 - 11.01)	33	6.06 (3.03 - 10.91)	0.89	26	5.43 (3.82 - 10.20)	0.58	70	5.64 (3.08 - 9.93)	0.56	0.92
Primigravida	46	4.92 (2.55 - 9.80)	27	5.19 (3.22 - 10.13)	0.26	25	4.34 (3.04 - 11.16)	0.93	54	5.68 (2.71 - 10.27)	0.41	0.70
Multigravida	66	7.59 (4.05 - 13.14)	51	7.26 (3.49 - 12.06)	0.35	34	6.52 (3.25 - 11.53)	0.33	94	5.71 (2.68 - 10.98)	0.09	0.38
Abx exposure	62	5.16 (3.31 - 10.53)	43	7.11 (3.22 – 11.34)	0.45	33	6.12 (3.42 - 13.86)	0.43	82	5.45 (2.49 - 9.38)	0.70	0.53
No Abx exposure	50	6.95 (3.62 - 13.38)	34	5.58 (3.39 - 10.18)	0.33	24	4.64 (2.35 - 9.12)	0.12	62	5.90 (3.22 - 11.27)	0.35	0.40
SGA	8	7.66 (4.21 - 15.88)	8	8.33 (2.23 - 40.11)	1.00	2	12.62 (4.17)	0.89 ^a	9	5.61 (5.17 - 13.29)	0.70	0.99
Not SGA	103	6.15 (3.35 - 11.99)	70	6.04 (3.53 - 10.87)	0.97	57	5.53 (3.11 - 10.71)	0.56	138	5.71 (2.63 - 10.78)	0.46	0.83
Pets at home	49	6.15 (3.28 - 11.31)	36	5.09 (3.09 - 11.79)	0.89	29	4.94 (2.51 - 9.94)	0.38	71	5.10 (2.48 - 9.01)	0.26	0.59
No pets at home	64	6.28 (3.58 - 12.23)	42	7.64 (3.57 – 10.91)	0.67	30	6.62 (3.93 - 12.97)	0.81	77	6.23 (3.56 - 12.94)	0.88	0.98
Other children	54	8.43 (4.16 - 13.38)	39	8.00 (3.54 - 11.34)	0.44	28	6.64 (3.13 - 11.43)	0.37	68	5.70 (2.84 - 9.13)	0.032*	0.15
No other children	59	4.97 (3.10 - 9.61)	39	5.14 (3.22 - 10.33)	0.45	31	4.24 (3.16 - 11.40)	0.94	80	5.64 (2.70 - 12.18)	0.34	0.77
Prenatal SSRI	0	-	3	3.54 (3.22)	-	0	-	-	15	4.15 (2.48 - 7.08)	-	0.37
No Prenatal SSRI	109	6.10 (3.34 - 11.70)	75	7.11 (3.49 – 11.14)	0.79	59	5.53 (3.16 - 11.40)	0.78	131	5.71 (2.71 - 10.90)	0.82	0.82
Postnatal SSRI	2	6.38 (5.27)	5	3.59 (3.38 - 7.54)	0.38 ^a	2	3.82 (2.12)	0.67 ^a	14	4.44 (2.46 - 8.12)	0.50 ^a	0.70
No postnatal SSRI	109	6.19 (3.44 - 12.19)	72	6.69 (3.17 – 11.29)	0.97	56	6.27 (3.37 - 11.78)	0.75	133	5.69 (2.71 - 10.97)	0.55	0.85
Infant allergic eczema	10	8.27 (4.18 - 16.23)	5	9.16 (5.79 - 17.39)	0.62	9	6.62 (2.64 - 13.86)	0.62	19	4.74 (3.21 - 8.00)	0.13	0.31
No infant allergic eczema	103	6.15 (3.52 - 12.06)	72	5.98 (3.10 - 10.95)	0.98	50	5.51 (3.25 - 10.36)	0.62	128	5.84 (2.70 - 11.18)	0.73	0.96
Breastfed and other children	43	10.20 (4.67 - 14.40)	26	9.85 (7.40 - 12.42)	0.99	21	6.91 (4.83 - 16.51)	0.73	42	6.20 (4.06 - 10.87)	0.21	0.27
Not breastfed and other children	10	4.53 (2.99 - 8.84)	11	2.56 (1.54 - 3.76)	0.02***	5	4.94 (1.51 - 8.22)	0.62	21	2.71 (1.61 - 6.06)	0.18	0.19
Breastfed and no pets	51	7.18 (3.62 – 13.15)	29	9.16 (5.16 - 10.99)	0.38	23	6.77 (4.17 – 15.82)	0.52	54	7.12 (4.55 – 13.85)	0.66	0.81
Not breastfed and no pets	12	4.29 (3.33 - 6.44)	9	2.56 (1.32 - 3.43)	0.013***	6	3.89 (1.93 - 7.09)	0.71	18	4.60 (1.79 - 6.41)	0.64	0.15

Table B3. Median sIgA depending on maternal perceived stress above CHILD cohort mean after stratifying for potential covariates

*Significant at 0.05 a priori; **Significant at Bonferroni adjusted p-value 0.05/1; *** Significant at Bonferroni adjusted p-value 0.05/2; aFisher's Exact test

Table B4. Logistic regression analyses of lowest quartile sIgA and perceived stress above CHILD mean (N=403)

(Ref: lower stress)	OR for prenatal	OR for postnatal	OR for both
Crude OR	1.21 (0.61 – 2.40)	1.28 (0.61 - 2.68)	1.46 (0.82 – 2.59)
Adjusted for birth characteristics	1.35 (0.67 – 2.73)	1.19 (0.55 – 2.55)	1.60 (0.88 - 2.89)
(gravida, birth mode, SGA,			
antibiotics)			
Adjusted for breastfeeding at time of	1.19 (0.57 – 2.48)	1.27 (0.57 – 2.85)	1.34 (0.72 – 2.48)
stool collection			
Adjusted for home environment	1.17 (0.59 – 2.33)	1.20 (0.56 – 2.57)	1.34 (0.75 – 2.40)
(children and pets at home)			
Adjusted for postnatal SSRI	1.19 (0.60 – 2.37)	1.15 (0.54 – 2.45)	1.40 (0.78 – 2.51)
Adjusted for maternal asthma or	1.20 (0.61 – 2.40)	1.18 (0.55 – 2.51)	1.45 (0.82 – 2.58)
allergy			
Adjusted for maternal depression	1.17 (0.58 – 2.33)	1.10 (0.51 – 2.38)	1.33 (0.74 – 2.39)
characteristics (depression history			
and postnatal SSRI)			
Adjusted for breastfeeding at stool	1.15 (0.55 – 2.40)	1.26 (0.56 – 2.82)	1.34 (0.72 – 2.50)
collection and pets at home			
Adjusted for postnatal SSRI, pets at	1.19 (0.56 – 2.52)	1.10 (0.47 – 2.58)	1.39 (0.73 – 2.64)
home, breastfeeding, antibiotics			

Note: Significant odds ratios were not found; therefore, a new cut-off of 17 for the highest

quartile of PSS scores was used to indicate higher stress.

Appendix C: Logistic regression analyses on below median sIgA, highest quartile and lowest tertile sIgA as outcome variables (yes/no)

(Ref: less depressive symptoms/below cut-off)	OR (95% CI) for prenatal	OR (95% CI) for postnatal	OR (95% CI) for both
Crude	1.70(0.92 - 3.14)	1.39 (0.69 – 2.81)	1.92 (0.95 - 3.89)
Adjusted for antibiotics use	1.96 (1.03 – 3.73)	1.33 (0.65 – 2.72)	1.76 (0.86 – 3.61)
Adjusted for gravida (primiparity)	1.74 (0.94 - 3.24)	1.45 (0.71 – 2.96)	2.02 (0.98 - 4.15)
Adjusted for birth characteristics (gravida, birth mode, SGA, antibiotics)	2.02 (1.05 - 3.88)	1.45 (0.70 - 3.00)	1.74 (0.83 – 3.64)
Adjusted for pets at home	1.71 (0.92 - 3.19)	1.54 (0.75 – 3.16)	1.81 (0.89 - 3.68)
Adjusted for home environment (pets and children at home)	1.68 (0.90 - 3.13)	1.57 (0.76 – 3.24)	1.76 (0.86 – 3.63)
Adjusted for prenatal SSRI	1.49 (0.79 – 2.82)	1.46 (0.71 – 2.99)	1.91 (0.92 – 3.97)
Adjusted for depression history	1.50 (0.79 – 2.86)	1.27 (0.62 – 2.60)	1.89 (0.91 – 3.91)
Adjusted for maternal depression characteristics (prenatal SSRI and depression history)	1.34 (0.69 – 2.60)	1.35 (0.66 – 2.80)	1.76 (0.84 – 3.69)
Adjusted for breastfeeding	1.94 (0.94 – 4.01)	0.91 (0.42 – 1.97)	2.15 (0.96 - 4.80)
Adjusted for antibiotics, gravida, pets, breastfeeding	2.57 (1.17 - 5.65)	1.02 (0.45 – 2.31)	2.06 (0.88 - 4.80)
Adjusted for antibiotics, gravida, pets, depression history, breastfeeding	2.44 (1.07 – 5.56)	0.90 (0.39 - 2.08)	1.92 (0.80 - 4.59)

Table C1. Odds of below median sIgA after exposure to pre and postnatal depressive symptoms

Note: No significant odd ratios were found with perceived stress.

(Ref: less depressive symptoms/below cut-off)	OR (95% CI) for prenatal	OR (95% CI) for postnatal	OR (95% CI) for both
Crude	0.48 (0.22 – 1.07)	0.51 (0.20 – 1.27)	0.48 (0.19 – 1.18)
Adjusted for antibiotics use	0.43 (0.19 – 1.01)	0.51 (0.20 - 1.28)	0.49 (0.20 – 1.24)
Adjusted for gravida (primiparity)	0.47 (0.21 – 1.04)	0.49 (0.19 – 1.22)	0.46 (0.18 – 1.14)
Adjusted for SGA	0.47 (0.21 – 1.04)	0.50 (0.20 - 1.26)	0.51 (0.20 – 1.28)
Adjusted for birth characteristics (gravida, birth mode, SGA, antibiotics)	0.43 (0.18 – 1.00)	0.47 (0.19 – 1.20)	0.51 (0.20 – 1.29)
Adjusted for home environment (pets and children at home)	0.48 (0.22 – 1.08)	0.52 (0.21 – 1.29)	0.51 (0.21 – 1.29)
Adjusted for breastfeeding	0.32 (0.12 - 0.88)	0.77 (0.29 – 2.08)	0.43 (0.15 – 1.22)
Adjusted for prenatal SSRI	0.52 (0.23 – 1.18)	0.54 (0.21 – 1.35)	0.41 (0.15 – 1.12)
Adjusted for maternal depression history and prenatal SSRI	0.59 (0.25 – 1.36)	0.56 (0.22 – 1.41)	0.44 (0.16 - 1.18)

Table C2. Odds of high sIgA in infants after exposure to pre and postnatal depressive symptoms

(Ref: lower stress/below cut-off)	OR (95% CI) for	OR (95% CI) for	OR (95% CI) for both
	prenatal	postnatal	
Crude OR	0.86 (0.45 – 1.67)	0.91 (0.45 – 1.87)	0.85 (0.48 - 1.48)
Adjusted for breastfeeding at time of stool collection	0.75 (0.35 – 1.59)	0.91 (0.42 – 1.99)	0.86 (0.47 – 1.59)
Adjusted for prenatal SSRI	0.95 (0.48 - 1.86)	0.99 (0.48 - 2.04)	0.95 (0.53 – 1.71)
Adjusted for maternal depression characteristics (depression history, prenatal SSRI)	0.98 (0.50 – 1.92)	1.00 (0.48 - 2.08)	1.01 (0.56 – 1.81)

Table C3. Odds of high sIgA in infants after exposure to pre and postnatal stress

Table C4. Odds of lowest tertile in infants after exposure to pre and postnatal depressive symptoms

(Ref: Less depressive symptoms)	OR for depression	OR for covariate	Decision
Crude OR	1.81 (0.97 – 3.37)		
	1.61 (0.78 - 3.32)		
	2.05 (1.02 - 4.12)		
Adjusted for child age at stool	1.61 (0.85 - 3.04)	1.60 (0.83 - 3.08)	In (p<0.20)
collection (quartiles)	1.50 (0.71 - 3.16)	2.65 (1.39 - 5.06)	
	2.35 (1.15 - 4.82)	3.19 (1.67 - 6.09)	
Adjusted for antibiotics exposure	2.00 (1.06 - 3.79)	1.24 (0.80 - 1.91)	In (confounding)
	1.54 (0.73 – 3.23)		
	2.08 (1.02 - 4.25)		
Adjusted for gravida (primigravida)	1.85 (0.99 - 3.45)	1.14 (0.74 – 1.77)	Out
	1.66 (0.80 - 3.42)		
	2.01 (0.99 - 4.10)		
Adjusted for maternal asthma and	1.74 (0.93 – 3.27)	0.81 (0.52 - 1.26)	Out
allergy	1.57 (0.76 – 3.24)		
	2.21 (1.09 - 4.48)		
Adjusted for breastfeeding at time of	1.83 (0.89 – 3.75)	9.18 (4.78 - 17.63)	In (always in model)
stool collection	1.21 (0.55 – 2.66)	3.75 (2.02 - 6.95)	
	2.14 (0.98 - 4.70)		
Adjusted for pets at home	1.84 (0.99 – 3.45)	1.57 (1.03 – 2.41)	In (p<0.20)
	1.77 (0.85 – 3.70)		
	1.93 (0.96 – 3.91)		
Adjusted for other children at home	1.76 (0.94 – 3.29)	0.70 (0.46 - 1.06)	Out
	1.70 (0.82 – 3.53)		
	1.92 (0.95 - 3.91)		
Adjusted for prenatal smoke exposure	1.58 (0.83 - 3.00)	3.39 (1.43 - 8.06)	In (p<0.20)
	1.60 (0.77 – 3.32)		
	2.09 (1.03 – 4.27)		
Adjusted for postnatal smoke	1.77 (0.95 – 3.31)	1.25 (0.71 – 2.19)	Out
exposure	1.70 (0.82 – 3.53)		
	2.01 (1.00 - 4.04)		
Adjusted for prenatal SSRI	1.50 (0.78 - 2.88)	2.08 (0.77 - 5.63)	In (confounding)
	1.66 (0.80 – 3.45)		
	1.92 (0.94 - 3.94)		
Adjusted for postnatal SSRI	1.81 (0.96 – 3.42)	1.32 (0.55 – 3.18)	In (confounding)
	1.45 (0.68 – 3.10)		
	2.08 (1.00 - 4.04)		
Adjusted for antibiotics, infant age,	1.71 (0.77 – 3.80)		Final model
pets, breastfeeding, prenatal SSRI	1.33 (0.57 – 3.12)		
	2.16 (0.91 - 5.17)		

Appendix D: Sensitivity analyses for home births, age, gestational age and SSRI use

(Ref: less depressive symptoms/below	OR (95% CI)	
cut-off)	Includes home births	Excludes home births
	N=415	N=403
Crude OR	2.26 (1.19 – 4.33)	2.08 (1.08 - 4.01)
	2.11 (1.01 – 4.40)	2.31 (1.10 – 4.87)
	2.03 (0.98 - 4.20)	2.12(1.02 - 4.42)
Adjusted for presence of pets at home	2.26 (1.18 – 4.34)	2.08 (1.08 – 4.03)
	2.27 (1.08 – 4.77)	2.51 (1.18 - 5.35)
	1.88 (0.90 - 3.93)	1.96 (0.93 – 4.12)
Adjusted for breastfeeding at time of	2.45 (1.17 – 5.14)	2.21 (1.05 – 4.65)
stool collection	1.66 (0.74 – 3.69)	1.78 (0.79 – 4.01)
	2.32 (1.04 – 5.20)	2.39 (1.06 – 5.39)
Adjusted for antibiotics	2.52 (1.30 – 4.88)	2.30 (1.18 – 4.49)
	2.00 (0.94 – 4.25)	2.22(1.03-4.78)
	2.35 (1.12 – 4.96)	2.39 (1.13 – 5.05)
Adjusted for birth characteristics	2.66 (1.36 – 5.22)	2.46 (1.25 – 4.87)
	2.04 (0.95 – 4.39)	2.26 (1.03 – 4.91)
	2.55 (1.18 - 5.48)	2.59 (1.20 – 5.59)
Adjusted for birth characteristics and	3.02 (1.39 – 6.58)	2.68 (1.23 – 5.86)
breastfeeding	1.56 (0.67 – 3.63)	1.70 (0.73 – 3.99)
	2.99 (1.24 – 7.21)	3.00 (1.24 - 7.24)
Adjusted for birth characteristics,	3.05 (1.40 - 6.67)	2.71 (1.23 – 5.94)
breastfeeding, pets at home	1.76 (0.75 – 4.16)	1.93 (0.81 – 4.59)
	2.86 (1.19 – 6.89)	2.87 (1.19 – 6.94)

Although home births were meant to be excluded from the CHILD study, mothers that went

through home delivery were still in our sample. The sensitivity analyses show that including home births in our sample change our odds ratios for low IgA in the three depressive symptom categories. Since home births seems to be a major confounding variable, we excluded the infants born from home (n=12) from this study.

(Ref: less depressive symptoms/below	OR (95% CI)	
cut-off)	Includes greater than 6mo	Excludes greater than
	N=403	6mo
		N=372
Crude OR	2.08 (1.08 – 4.01)	2.15 (1.07 – 4.31)
	2.31 (1.10 – 4.87)	2.18 (0.99 – 4.81)
	2.12 (1.02 – 4.42)	2.25 (1.07 – 4.73)
Adjusted for presence of pets at home	2.08 (1.08 - 4.03)	2.18 (1.08 - 4.39)
	2.51 (1.18 - 5.35)	2.38 (1.06 - 5.32)
	1.96 (0.93 – 4.12)	2.06 (0.97 - 4.37)
Adjusted for breastfeeding at time of	2.21 (1.05 – 4.65)	2.14 (0.98 - 4.69)
stool collection	1.78 (0.79 – 4.01)	1.75 (0.75 – 4.09)
	2.39 (1.06 - 5.39)	2.40 (1.06 - 5.44)
Adjusted for antibiotics	2.30 (1.18 – 4.49)	2.34 (1.16 – 4.75)
	2.22 (1.03 – 4.78)	2.06 (0.91 – 4.67)
	2.39 (1.13 – 5.05)	2.54 (1.19 – 5.40)
Adjusted for birth characteristics	2.46 (1.25 – 4.87)	2.51 (1.22 - 5.16)
(antibiotics use, birth mode, gravida,	2.26 (1.03 – 4.91)	2.08 (0.91 - 4.78)
SGA)	2.59 (1.20 - 5.59)	2.77 (1.27 – 6.05)
Adjusted for birth characteristics and	2.68 (1.23 – 5.86)	2.44 (1.08 - 5.50)
breastfeeding	1.70 (0.73 – 3.99)	1.64 (0.67 – 3.99)
	3.00 (1.24 – 7.24)	2.99 (1.23 - 7.29)
Adjusted for birth characteristics,	2.71 (1.23 – 5.94)	2.50 (1.11 - 5.67)
breastfeeding, pets at home	1.93 (0.81 – 4.59)	1.90 (0.77 – 4.72)
	2.87 (1.19 – 6.94)	2.85 (1.17 - 6.96)

Table D2. Sensitivity analyses for age at stool collection greater than 6 months in 403 subsample

30 (8%) infants in our sample had their stool collected at greater than six months of infant age

even though the stool was meant to be collected at 3 months. Percent differences between OR's were under 10%, and so we kept this portion of infants in our sample. This portion of infants having sample collected after six months will help our analyses with postnatal stress and

depressive symptoms.

(Ref: less depressive symptoms/below	OR (95% CI)	
cut-off)	Includes less than 37	Excludes less than 37
	weeks gestation	weeks gestation
	N=403	N=382
Crude OR	2.08 (1.08 - 4.01)	2.18 (1.09 – 4.35)
	2.31 (1.10 – 4.87)	2.38 (1.10 – 5.21)
	2.12(1.02 - 4.42)	2.18(0.99 - 4.79)
Adjusted for presence of pets at home	2.08 (1.08 - 4.03)	2.20 (1.09 – 4.41)
	2.51 (1.18 - 5.35)	2.58 (1.18 - 5.64)
	1.96 (0.93 – 4.12)	2.05(0.93 - 4.54)
Adjusted for breastfeeding at time of	2.21 (1.05 - 4.65)	2.50 (1.16 - 5.37)*
stool collection	1.78 (0.79 – 4.01)	1.81 (0.79 – 4.15)
	2.39 (1.06 - 5.39)	2.69 (1.13 – 6.41)*
Adjusted for antibiotics	2.30 (1.18 - 4.49)	2.38 (1.18 - 4.82)
-	2.22(1.03 - 4.78)	2.18(0.99 - 4.79)
	2.39 (1.13 - 5.05)	2.27 (1.03 – 5.02)
Adjusted for postnatal smoke exposure	1.96 (1.01 – 3.81)	2.05 (1.02 – 4.12)
	2.48 (1.16 – 5.26)	2.57 (1.18 - 5.60)
	2.00 (0.95 – 4.21)	2.05(0.93 - 4.54)
Adjusted for birth characteristics	2.46 (1.25 – 4.87)	2.51 (1.23 – 5.15)
(antibiotics use, birth mode, gravida,	2.26 (1.03 – 4.91)	2.25 (1.01 – 5.01)
SGA)	2.59 (1.20 - 5.59)	2.29(1.02 - 5.15)
Adjusted for birth characteristics and	2.68 (1.23 - 5.86)	2.96 (1.33 - 6.59)
breastfeeding	1.70 (0.73 – 3.99)	1.69 (0.71 – 4.05)
	3.00 (1.24 - 7.24)	2.80 (1.12 - 6.95)
Adjusted for antibiotics, gravida,	2.66 (1.21 - 5.82)	2.87 (1.28 - 6.40)
breastfeeding, pets at home, postnatal	1.94 (0.82 - 4.58)	1.89 (0.78 - 4.58)
smoke exposure	2.80 (1.19 - 6.59)	2.86 (1.17 - 7.00)

Table D3. Sensitivity analyses for late preterm infants included in 403 sub-sample

Sixteen (4%) infants in our sample were born late preterm at less than 37 weeks gestation (34-36

weeks). Percent differences between ORs were mostly under 10%, except for the OR's after adjusting for breastfeeding status. However, the multivariate logistic regression models that include breastfeeding status did not show greater than 10% difference after excluding the late preterm infants, and so we kept the preterm infants in our sample.

Table D4. Median infant sIgA levels with maternal SSRI use during pre and postnatal time period

sIgA (mg/g feces)	No SSRI use (N=374)	Prenatal use only (N=4)	Postnatal use only (N=9)	SSRI use during both pre and postnatal time period (N=14)
Median	6.06	2.25	5.27	4.07
(IQR)	(3.28 – 11.36)	(1.19 – 10.17)	(1.99 – 8.74)	(3.46 - 6.72)

Note: None of the sIgA levels were significantly different when SSRIs were used than the sIgA

levels with no SSRI use.

(Ref: less depressive symptoms/below	OR (95% CI)	
cut-off)	Includes mothers that used	Excludes mothers that
	SSRIs	used SSRIs
	N=403	N=374
Crude OR	2.08 (1.08 – 4.01)	1.82 (0.88 - 3.76)*
	2.31 (1.10 – 4.87)	2.34 (1.11 – 4.97)
	2.12 (1.02 – 4.42)	1.55 (0.68 - 3.55)*
Adjusted for presence of pets at home	2.08 (1.08 - 4.03)	1.86 (0.90 - 3.86)*
	2.51 (1.18 - 5.35)	2.53 (1.18 - 5.43)
	1.96 (0.93 – 4.12)	1.40 (0.61 – 3.25)*
Adjusted for breastfeeding at time of	2.21 (1.05 – 4.65)	2.14 (0.97 – 4.71)
stool collection	1.78 (0.79 – 4.01)	1.82 (0.81 – 4.12)
	2.39 (1.06 - 5.39)	1.81 (0.74 – 4.46)*
Adjusted for antibiotics	2.30 (1.18 – 4.49)	1.98 (0.95 – 4.13)*
	2.22 (1.03 – 4.78)	2.28 (1.05 - 4.94)
	2.39 (1.13 – 5.05)	1.66 (0.72 – 3.83)*
Adjusted for postnatal smoke exposure	1.96 (1.01 – 3.81)	1.72 (0.83 – 3.58)*
	2.48 (1.16 – 5.26)	2.51 (1.17 – 5.37)
	2.00 (0.95 - 4.21)	1.48 (0.64 - 3.42)*
Adjusted for birth characteristics	2.46 (1.25 – 4.87)	2.08 (0.98 - 4.42)*
(antibiotics use, birth mode, gravida,	2.26 (1.03 – 4.91)	2.33 (1.06 - 5.14)
SGA)	2.59 (1.20 - 5.59)	1.72 (0.73 – 4.06)*
Adjusted for antibiotics, gravida,	2.66 (1.21 – 5.82)	2.44 (1.07 – 5.54)*
breastfeeding, pets at home, postnatal	1.94 (0.82 – 4.58)	1.98 (0.83 - 4.69)
smoke exposure	2.80 (1.19 – 6.59)	1.92 (0.75 - 4.93)*

Table D5. Ser	nsitivity analyse	s for SSRI us	e included in 40.	3 sub-sample
---------------	-------------------	---------------	-------------------	---------------------

In the sub-sample, 27 (6.7%) mothers took SSRIs during the pre or postnatal period. Although

median sIgA were not significantly different with SSRI use in Table D4, the sensitivity analyses indicate excluding the mothers that took SSRIs causes significant changes to the odds ratios for low IgA. Greater than 10% change was seen for odd ratios for prenatal only depressive symptoms and both pre and postnatal groups depressive symptoms with the exclusion of SSRI

use.

Appendix E: Maternal distress and sIgA median comparisons and logistic regression models

sIgA (mg/g feces)	Refer	ence group										Interaction ^b
Median (IQR)	Ν	Less symptoms (3.9% SSRI use)	Ν	Prenatal Only (18.8% SSRI use)	p-value	Ν	Postnatal Only (3.2% SSRI use)	p-value	N	Both (16.7% SSRI use)	p-value	
Depressive symptoms	280	6.30 (3.58 - 12.38)	49	5.19 (2.02 - 9.78)	0.057	35	5.69 (2.62 - 8.56)	0.11	37	4.43 (2.39 - 8.03)	0.033*	-
Depression history	40	5.46 (3.58 - 11.76)	18	3.68 (1.36 - 6.59)	0.06	10	3.54 (2.21 - 5.70)	0.13	11	5.54 (2.35 - 9.01)	0.65	0.41
No depression history	236	6.37 (3.58 - 12.38)	29	5.71 (3.00 - 12.09)	0.68	25	6.39 (2.84 - 9.56)	0.47	25	3.72 (2.37 - 7.39)	0.021*	0.05 0.71
Maternal asthma or allergy	175	6.55 (3.77 – 12.66)	33	4.86 (1.70 – 12.21)	0.15	18	4.81 (2.69 - 7.18)	0.041***	27	4.43 (2.43 - 7.96)	0.032*	0.53 0.13
No maternal asthma or allergy	101	6.07 (3.21 – 11.73)	15	5.59 (2.78 - 8.31)	0.33	17	6.39 (2.31 - 12.73)	0.93	9	3.72 (1.82 - 9.69)	0.26	0.13
Primigravida	112	5.32 (3.19 - 11.11)	19	5.05 (2.37 - 8.11)	0.22	12	5.00 (2.64 - 7.54)	0.30	9	5.74 (2.06 - 9.06)	0.58	0.74
Multigravida	165	7.11 (3.89 – 13.14)	30	5.66 (1.88 - 10.80)	0.11	23	6.04 (2.31 – 9.23)	0.15	27	4.43 (2.43 - 7.96)	0.022*	0.74
Prenatal SSRI	5	4.15 (3.76 - 9.63)	8	4.89 (2.59 - 12.30)	0.77	0	-	-	5	2.71 (2.13 – 4.17)	0.095 ^a	
No prenatal SSRI	269	6.19 (3.53 – 12.36)	40	5.27 (1.95 - 9.84)	0.12	34	5.68 (2.54 - 8.14)	0.11	31	5.54 (235 - 8.09)	0.12	-
Prenatal smoke exposure	13	5.07 (2.39 - 13.69)	6	2.54 (1.43 - 5.32)	0.14	2	1.70 (0.68 – none)	0.11ª	3	2.43 (1.74 – none)	0.093	1.00
No prenatal smoke exposure	263	6.27 (3.64 - 12.06)	42	5.61 (2.32 - 10.56)	0.26	33	5.71 (2.84 - 8.89)	0.23	33	4.74 (2.42 - 8.03)	0.068	1.00
Male	159	6.62 (3.53 - 13.15)	25	5.35 (2.48 - 10.78)	0.31	20	4.56 (2.00 - 7.99)	0.027***	20	4.05 (2.34 - 7.82)	0.07	0.90 0.10
Female	121	6.07 (3.63 - 11.55)	24	4.98 (1.27 – 9.84)	0.11	15	6.23 (3.23 – 9.89)	0.86	17	4.74 (2.42 - 8.49)	0.26	0.53
Caesarean section	79	5.49 (3.53 - 10.85)	19	5.19 (2.16 - 14.04)	0.47	5	6.39 (4.18 – 11.12)	0.83ª	12	4.05 (1.91 - 6.92)	0.057	0.83
Vaginal	198	6.62 (3.76 - 12.67)	30	5.32 (1.80 - 9.05)	0.052	30	5.46 (2.54 - 8.72)	0.058	24	5.24 (2.44 - 11.14)	0.24	0.068 0.53
SGA	20	7.66 (3.72 – 15.67)	4	5.60 (4.97 - 34.36)	1.00 ^a	2	15.15 (9.23)	0.36ª	1	-	0.76 ^a	
Not SGA	256	6.23 (3.63 - 12.25)	45	5.05 (1.87 - 9.78)	0.035	33	5.66 (2.47 - 7.99)	0.052	34	4.55 (2.34 - 8.32)	0.049*	-
Breastfed	207	7.82 (4.16 - 13.43)	28	5.66 (2.49 - 10.35)	0.15	22	6.50 (3.22 - 9.73)	0.31	24	5.66 (3.22 - 9.73)	0.17	0.75
Not breastfed	64	4.04 (2.22 - 6.09)	15	2.78 (1.54 - 4.77)	0.15	11	2.71 (1.07 - 6.39)	0.61	10	2.46 (1.63 - 2.97)	0.033*	0.75
Antibiotics exposure	155	6.15 (3.53 - 11.41)	31	3.80 (1.54 - 10.50)	0.046	16	6.13 (2.39 - 10.42)	0.36	18	4.40 (1.77 - 7.38)	0.027*	
No Antibiotics exposure	120	6.42 (3.78 - 13.03)	15	5.71 (3.23 - 8.85)	0.23	18	5.68 (2.98 - 8.72)	0.29	17	5.74 (2.46 - 12.85)	0.51	0.50
Other children	135	6.87 (4.05 - 12.31)	20	7.44 (1.94 – 13.28)	0.76	17	6.04 (3.15 - 8.61)	0.32	17	3.70 (2.28 - 7.69)	0.038*	0.17
No other children	144	5.59 (3.37 - 12.42)	29	4.77 (2.13 – 7.15)	0.039	17	3.86 (2.31 - 8.27)	0.11	19	5.57 (2.43 - 9.01)	0.53	0.72 0.098
Pets at home	127	6.06 (3.31 - 12.06)	22	3.22 (1.36 - 7.73)	0.027**	13	3.86 (1.85 - 7.99)	0.14	23	3.70 (2.35 - 5.74)	0.022*	0.91
No pets at home	151	6.60 (3.93 - 12.70)	27	5.59 (3.49 - 10.50)	0.54	21	5.71 (2.76 - 8.89)	0.30	14	7.39 (2.56 – 12.41)	0.81	0.88 0.04
Postnatal smoke exposure	39	5.07 (3.54 - 9.80)	12	5.24 (1.52 - 10.92)	0.72	4	1.78 (0.54 - 11.43)	0.15 ^a	9	2.43 (1.91 - 3.73)	0.005*	0.70 0.57

 Table E1. Medians of infant fecal sIgA depending on maternal depressive symptoms after stratifying for potential covariates

No postnatal smoke exposure	241	6.42 (3.60 - 12.68)	37	5.19 (2.27 - 8.58)	0.055	30	5.70 (2.95 - 8.14)	0.19	28	5.66 (3.62 - 9.54)	0.38	0.15
Postnatal SSRI	10	5.19 (3.88 - 6.83)	8	4.89 (2.59 - 12.30)	0.93	1	-	0.18 ^a	4	3.04 (1.78 - 4.46)	0.076 ^a	
No postnatal SSRI	266	6.42 (3.56 - 12.67)	40	5.12 (1.62 - 9.84)	0.057	31	5.69 (2.62 - 8.00)	0.13	33	5.54 (2.39 - 8.55)	0.096	-
Infant allergic eczema	28	7.06 (3.89 - 11.78)	4	9.10 (3.88 - 20.41)	0.68ª	4	2.76 (0.91 - 6.80)	0.062ª	7	4.74 (2.30 - 8.09)	0.32	
No infant allergic eczema	251	6.27 (3.57 – 12.66)	44	5.12 (1.87 - 9.46)	0.029**	28	5.70 (2.80 - 8.91)	0.33	30	4.40 (2.41 - 8.22)	0.052	0.74
Not breastfed and multigravida	38	4.04 (2.48 - 6.16)	8	1.70 (1.14 - 3.50)	0.026**	6	4.18 (1.64 - 8.38)	0.76	7	2.43 (1.55 - 2.71)	0.013*	-
Breastfed and multigravida	122	9.10 (4.87 – 14.11)	18	7.11 (3.16 - 10.80)	0.18	16	6.84 (3.21 - 10.73)	0.20	17	5.57 (4.03 - 9.74)	0.095	-
Not breastfed and not SGA	59	3.93 (2.36 - 6.07)	14	2.69 (1.46 - 4.40)	0.11	11	2.71 (1.07 - 6.39)	0.63	10	2.46 (1.63 - 2.97)	0.03*	-
Breastfed and pets at home	89	7.82 (4.06 – 13.71)	12	5.28 (1.09 - 10.47)	0.07	6	5.92 (2.95 - 11.54)	0.43	15	4.43 (2.35 - 7.02)	0.047*	-
Not breastfed and other children	32	4.20 (2.22 - 6.38)	6	1.70 (0.99 - 5.45)	0.07	4	5.87 (2.23 - 6.31)	0.75 ^a	5	1.66 (1.53 – 2.60)	0.028*, a	-
Not breastfed and female	31	3.64 (2.08 - 6.07)	5	2.78 (0.75 - 4.00)	0.21 ^a	3	6.39 (6.04 - none)	0.11ª	5	2.48 (1.53 - 2.70)	0.078 ^a	-
Not breastfed and no depression history in mother	55	4.20 (2.52 - 6.27)	7	3.22 (2.37 - 8.11)	0.60	8	4.18 (0.91 - 9.02)	0.64	9	2.48 (1.65 - 3.22)	0.04*	-
Not breastfed and mother has asthma/allergy	43	4.20 (2.36 - 6.12)	11	2.37 (1.23 – 4.77)	0.10	5	2.71 (0.76 - 7.97)	0.47	8	2.46 (1.68 – 2.71)	0.055*	-
Primigravida and pets at home	51	5.37 (2.71 - 12.40)	10	2.58 (0.87 - 3.97)	0.025**	7	2.71 (1.07 – 4.34)	0.058	6	2.52 (1.77 - 6.81)	0.15	-
Multigravida and not SGA	151	6.87 (3.86 - 12.66)	29	5.71 (1.87 - 10.86)	0.13	21	5.71 (2.31 - 8.28)	0.068	25	4.37 (2.37 - 8.49)	0.031*	-
Multigravida and has asthma/allergy	101	7.26 (4.18 – 12.25)	22	5.64 (1.50 - 13.59)	0.15	10	5.49 (2.66 - 7.49)	0.10	17	4.43 (2.46 - 7.67)	0.030*	-
Multigravida and antibiotics exposure	85	7.23 (3.86 - 12.06)	19	3.80 (1.54 - 14.04)	0.17	12	6.22 (2.31 - 10.42)	0.20	14	4.98 (2.14 - 7.55)	0.053*	-
Antibiotics exposure and pets at home	75	6.15 (3.52 - 10.85)	10	2.13 (1.00 - 4.02)	0.003**	6	4.95 (2.18 - 9.58)	0.45	13	3.72 (1.76 - 5.56)	0.017*	-
Antibiotics exposure and mother has asthma/allergy	103	6.33 (3.62 - 12.06)	22	2.80 (1.34 - 15.26)	0.10	5	2.62 (1.14 - 4.95)	0.020*** ^{,a}	14	4.40 (1.77 - 7.52)	0.047*	-
Antibiotics exposure and no other children	91	5.57 (3.35 - 13.04)	20	3.70 (1.35 - 5.55)	0.023**	9	3.86 (2.31 - 14.28)	0.45	11	5.54 (2.30 - 7.37)	0.24	-
Antibiotics exposure and male infant	95	6.19 (3.53 – 13.56)	17	5.05 (1.87 - 17.68)	0.54	11	3.86 (2.31 - 11.23)	0.19	11	3.72 (1.74 – 5.57)	0.02*	-
No antibiotics exposure and male infant	59	7.83 (4.01 – 13.15)	7	5.71 (3.49 - 7.79)	0.27	9	5.27 (1.49 - 6.84)	0.037***	8	7.56 (3.62 – 13.29)	0.92	-
Antibiotics exposure and female infant	60	5.81 (3.66 - 10.41)	14	3.19 (1.12 - 6.29)	0.024**	5	6.23 (4.63 - 11.76)	0.86ª	7	7.37 (2.70 – 7.96)	0.57	-

Antibiotics exposure and mother had depression history	24	5.88 (3.99 - 10.85)	11	1.87 (1.17 – 5.19)	0.006**	4	3.54 (2.54 - 13.81)	0.43ª	8	5.56 (2.42 - 8.75)	0.63	-
Antibiotics exposure and mother did not have depression history	130	6.24 (3.53 - 12.30)	18	6.68 (2.32 - 19.52)	0.99	12	6.31 (2.39 - 10.42)	0.51	10	3.21 (1.69 - 6.69)	0.015*	-
Pets and no depression history	99	6.07 (3.27 – 12.06)	13	3.23 (1.68 - 8.37)	0.084	11	4.34 (1.07 - 8.00)	0.26	14	3.65 (2.40 - 6.06)	0.059	-
Pets and no other children	66	5.51 (3.01 - 11.50)	12	2.58 (0.99 - 4.45)	0.008**	8	3.28 (1.29 - 7.07)	0.14	13	3.72 (2.33 - 8.01)	0.18	-
Pets and mother has asthma/allergy	80	5.76 (3.52 - 12.47)	16	4.04 (1.52 - 9.83)	0.12	9	3.07 (1.73 - 5.19)	0.014***	19	3.70 (2.35 - 7.02)	0.042*	-
Pets and male infant	67	6.20 (3.31 - 12.66)	10	4.04 (2.25 - 6.25)	0.054	9	2.71 (0.96 - 7.99)	0.076	16	3.68 (1.91 - 5.56)	0.043*	-

*Significant at 0.05 a priori; **Significant at Bonferroni adjusted p-value 0.05/2; *** Significant at Bonferroni adjusted p-value 0.05/1; aFisher's Exact test

 b Interactions between the covariate and depressive symptoms group variable were tested. When interaction terms with p<0.20 were found, the interaction term for each depressive symptoms group was reported.

Note: Stratifications with two covariates that did not have significant differences were not reported in this table.

Table E2. Logistic regression analyses for low sIgA in infants after exposure to pre and postnatal depressive symptoms

(Ref: Less depressive	N for lowest	OR for depression	OR for covariate	Decision
symptoms/below cut-off)	quartile of			Deelsion
	sIgA			
Crude OR	100	2.08 (1.08 - 4.01)		
	100	2.31(1.10 - 4.87)		
		2.12(1.02 - 4.42)		
Adjusted for child sex	M-54	2.07 (1.02 - 1.12) 2.07 (1.07 - 3.99)	1.09 (0.69 - 1.72)	Out
Adjusted for ening sex	F-46	2.31(1.10 - 4.87)	1.07(0.0) - 1.72)	Out
	1-40	2.12(1.01 - 4.41)		
Adjusted for child age at stool	1 st -18	1.88(0.96 - 3.67)	0.99 (0.48 - 2.04)	In (p<0.20)
collection (quartiles)	2 nd -19	2.12(0.99 - 4.55)	1.65 (0.83 - 3.28)	III (p<0.20)
conection (quartities)	3 rd -27	2.35(1.11 - 4.97)	2.51 (1.28 - 4.91)	
	4 th -36	2.55 (1.11 - 4.97)	2.31 (1.26 - 4.91)	
Adjusted for birth mode	V-70	2.16 (1.12 – 4.20)	1.18 (0.64 - 2.18)	Out
Adjusted for birth mode	EC-19			Out
		2.45(1.15-5.21)	0.70 (0.32 – 1.54)	
	SC-9	2.23 (1.06 – 4.70)		
Adjusted for antibiotics exposure	Y-58	2.30 (1.18 – 4.49)	1.21 (0.75 – 1.95)	In (confounding)
	N-39	2.22 (1.03 – 4.78)		
		2.39 (1.13 - 5.05)		
Adjusted for gravida	Y-43	2.17 (1.12 – 4.20)	1.47 (0.91 – 2.36)	In (p<0.20)
(primigravida)	N-55	2.46 (1.16 – 5.21)		
		2.44 (1.15 - 5.16)		
Adjusted for SGA	Y-3	2.17 (1.12 – 4.20)	0.37 (0.11 – 1.26)	In (p<0.20)
5	N-95	2.36(1.12 - 5.00)		u ,
		2.31 (1.09 – 4.87)		
Adjusted for maternal asthma and	Y-61	2.18 (1.12 – 4.21)	0.85(0.53 - 1.38)	Out
allergy	N-38	2.28(1.08 - 4.82)	0.00 (0.00 1.00)	out
unorgy	11 50	2.27(1.08 - 4.76)		
Adjusted for breastfeeding at time	E-11	2.21 (1.05 - 4.65)	9.40 (4.48 - 19.71)	In (always in model)
of stool collection	P-39	1.78(0.79-4.01)	3.87 (1.87 - 8.01)	III (always III III0uci)
of stool confection	Z-47		5.07(1.07 - 0.01)	
A directed for motor of home	Y-55	2.39 (1.06 - 5.39) 2.08 (1.08 - 4.03)	1.59 (1.00 - 2.53)	$I_{\rm m}$ (m < 0.20)
Adjusted for pets at home			1.59 (1.00 – 2.53)	In (p<0.20)
	N-45	2.51(1.18 - 5.35)		
		1.96 (0.93 – 4.12)		
Adjusted for other children at	Y-43	2.04 (1.06 – 3.94)	0.81 (0.51 – 1.30)	Out
home	N-56	2.42 (1.14 – 5.14)		
		1.95 (0.92 – 4.13)		
Adjusted for prenatal smoke	Y-13	1.96 (1.00 – 3.96)	3.63 (1.54 - 8.52)	In (p<0.20)
exposure	N-86	2.33 (1.10 – 4.95)		
		2.15 (1.01 – 4.55)		
Adjusted for postnatal smoke	Y-23	1.96 (1.01 - 3.81)	1.79(1.00 - 3.21)	In (p<0.20)
exposure	N-77	2.48 (1.16 - 5.26)	. , ,	
		2.00(0.95 - 4.21)		
Adjusted for prenatal SSRI	Y-6	1.86 (0.94 – 3.70)	1.16 (0.40 - 3.34)	In (confounding)
	N-93	2.36(1.12-5.01)	(
	1, 25	2.11(1.00 - 4.48)		
Adjusted for postnatal SSRI	Y-6	2.15(1.10-4.21)	0.85 (0.32 - 2.30)	In (confounding)
rugustu iti posulatal SSICI	N-92	2.13(1.10-4.21) 2.01(0.92-4.41)	0.05 (0.52 - 2.50)	In (contounding)
	11-92			
A directed for donression histor	V 25	2.11(1.01 - 4.41)	1.26 (0.72 - 2.20)	Out
Adjusted for depression history	Y-25	2.11(1.07 - 4.15) 2.25(1.06 - 4.77)	1.26 (0.72 – 2.20)	Out
	N-74	2.25(1.06 - 4.77)		
		2.14 (1.02 – 4.52)		
Adjusted for infant allergic eczema	Y-9	2.18 (1.12 – 4.21)	0.74 (0.34 – 1.63)	Out
	N-89	2.41 (1.11 – 5.23)		
	1	1222(10(-4(5)))		1
		2.22 (1.06 - 4.65)		
Adjusted for birth characteristics (gravida, birth mode, SGA,	97	2.22 (1.06 - 4.05) 2.46 (1.25 - 4.87)		In (confounding)

antibiotics)		2.59 (1.20 - 5.59)	
Adjusted for SGA, gravida,	97	2.41 (1.22 – 4.75)	Keep birth mode out
antibiotics		2.26 (1.04 – 4.89)	
	~-	2.59 (1.21 - 5.56)	
Adjusted for gravida and	97	2.36(1.20 - 4.62)	Keep SGA out
antibiotics		2.29 (1.06 – 4.94)	
		2.56 (1.20 - 5.45)	
Adjusted for gravida and SGA	98	2.20 (1.13 – 4.28)	Keep antibiotics in
		2.43 (1.14 – 5.16)	
	99	2.45 (1.51 - 5.23)	
Adjusted for home environment	99	2.05(1.06 - 3.96)	In (confounding)
(pets and children at home)		2.50(1.17 - 5.34)	
Adjusted for breastfeeding and	96	1.81 (0.85 – 3.86) 2.46 (1.16 – 5.21)	In (confounding)
maternal asthma/allergy	90	1.74(0.77 - 3.94)	in (contounding)
maternar astima/anergy			
Adjusted for maternal depression	98	2.61 (1.14 – 5.95) 1.94 (0.95 – 3.93)	Out
characteristics (depression history,	98	2.33(1.10 - 4.97)	Out
prenatal SSRI)		2.09(0.98 - 4.46)	
Adjusted for maternal depression	97	2.10 (1.05 - 4.19)	In
characteristics (depression history,	21	1.93(0.87 - 4.27)	111
postnatal SSRI)		2.12(1.00 - 4.49)	
Adjusted for prenatal SSRI and	96	2.05 (0.95 - 4.42)	Out
breastfeeding	70	1.81(0.80 - 4.10)	Out
breastreeding		2.39(1.04 - 5.49)	
Adjusted for postnatal SSRI and	95	2.37(1.10-5.10)	In
breastfeeding	,,,	1.52(0.64 - 3.58)	
		2.41(1.06 - 5.48)	
Adjusted for breastfeeding and	96	2.26 (1.05 – 4.83)	Keep smoke in
prenatal smoke		1.81(0.80 - 4.09)	(p<0.20)
r · · · · · · · · ·		2.42 (1.06 - 5.52)	4
Adjusted for breastfeeding and	97	2.17 (1.03 - 4.60)	Keep smoke out
postnatal smoke		1.94(0.86 - 4.41)	p
1		2.37(1.05 - 5.36)	
Adjusted for age, antibiotics,	91	2.49 (1.06 - 5.82)	
gravida, depression history,		1.42 (0.56 - 3.59)	
postnatal SSRI, pets, prenatal		3.04 (1.23 - 7.55)	
smoking, breastfeeding			
Adjusted for age, antibiotics,	91	2.61 (1.13 - 6.03)	Keep depression
gravida, postnatal SSRI, pets,		1.48 (0.59 – 3.72)	history out
prenatal smoking, breastfeeding		3.17 (1.28 - 7.82)	
Adjusted for age, antibiotics,	92	2.53 (1.12 – 5.75)	Keep smoking out
gravida, postnatal SSRI, pets,		1.46 (0.59 – 3.64)	
breastfeeding		3.16 (1.30 - 7.69)	
Adjusted for age, antibiotics,	91	2.82 (1.23 – 6.47)	Final model for
maternal asthma and allergy,		1.43 (0.57 – 3.60)	postnatal
postnatal SSRI, pets, breastfeeding,		3.33 (1.36 - 8.19)	depressive
gravida			symptoms
Adjusted for age, antibiotics,	93	2.72 (1.21 – 6.10)	
maternal asthma and allergy, pets,		1.88 (0.78 – 4.52)	
breastfeeding, gravida		3.33 (1.37 - 8.08)	
Adjusted for age, antibiotics,	92	2.44 (1.07 – 5.57)	Final model for
maternal asthma and allergy, pets,		1.93 (0.80 – 4.68)	prenatal only and
breastfeeding, gravida, prenatal SSRI		3.07 (1.25 – 7.55)	both
Final model for postnatal with	91	0.69 (0.16 - 3.03)	
interaction term for antibiotics	71	2.01 (0.52 - 7.80)	
interaction term for antibiotics		5.84 (1.53 - 22.35)	
Final model for prenatal and both	92	0.70 (0.16 - 3.06)	
with interaction term for antibiotics		3.13(0.83 - 11.78)	

Variable	Interactions with depressive symptoms (p-value)
Antibiotics exposure	0.04
	0.34
	0.39
Maternal asthma and allergy	0.46
Gravida	0.79
Breastfeeding	0.77
Prenatal SSRI	1.00
Postnatal SSRI	0.91
Pets at home	0.68
Age	0.79

Table E3. Interactions between depressive symptoms and covariates in final model

Note: Interactions between the covariate and depressive symptoms group variable were tested.

When interaction terms with p < 0.05 were found, the interaction term for each depressive

symptoms group was reported.

sIgA (mg/g feces)	Refer	ence group										Interaction ^b
Median (IQR)	N	Less stress (3.7% SSRI use)	N	Prenatal Only (11.9% SSRI use)	p-value	Ν	Postnatal Only (4.9% SSRI use)	p-value	N	Both (12% SSRI use)	p-value	
Highest quartile of stress	218	6.12 (3.54 - 11.36)	67	5.61 (3.05 - 10.33)	0.68	41	5.66 (2.89 - 18.70)	0.97	75	5.57 (2.59 - 9.89)	0.17	-
Depression history	31	5.35 (2.79 - 10.85)	17	5.11 (1.66 - 8.32)	0.32	8	4.44 (2.10 - 17.44)	0.70	23	5.07 (2.35 - 7.96)	0.61	
No depression history	184	6.12 (3.63 – 11.31)	49	7.82 (3.36 - 12.62)	0.62	32	6.20 (3.10 - 18.81)	0.83	50	5.72 (2.59 - 10.20)	0.26	-
Maternal asthma/allergy	136	6.12 (3.55 – 11.76)	44	6.22 (3.09 - 10.09)	0.56	24	6.96 (3.23 - 21.28)	0.26	49	5.27 (2.55 - 9.45)	0.12	
No maternal asthma/allergy	79	6.10 (3.53 - 11.03)	22	6.84 (2.66 - 12.50)	0.76	16	3.96 (1.89 - 10.12)	0.15	25	5.71 (2.45 - 11.94)	0.76	-
Primigravida	85	5.14 (3.19 - 10.58)	27	5.05 (2.63 - 9.90)	0.66	16	5.64 (2.09 - 14.76)	0.84	24	5.96 (2.77 - 9.50)	0.93	
Multigravida	131	6.67 (3.93 - 12.06)	40	7.58 (3.62 – 11.73)	0.82	24	6.84 (3.21 - 20.57)	0.89	50	5.55 (2.40 - 10.78)	0.066] -
Prenatal SSRI	4	5.30 (3.65 - 11.15)	6	3.41 (2.22 - 4.88)	0.26 ^a	1	-	-	7	3.60 (2.48 - 6.19)	0.41 ^a	
No prenatal SSRI	208	6.08 (3.52 - 11.29)	61	7.37 (3.27 – 10.74)	0.82	39	5.62 (2.71 - 18.48)	0.79	66	5.65 (2.61 - 9.93)	0.37] -
Prenatal smoke exposure	7	2.79 (1.77 – 9.05)	7	7.83 (3.22 – 13.95)	0.28	3	2.71 (0.68 – none)	0.67ª	7	2.43 (1.74 - 5.07)	0.48	
No prenatal smoke exposure	208	6.14 (3.68 - 16.60)	59	5.61 (2.71 - 10.10)	0.56	37	5.69 (3.14 - 19.00)	0.67	67	5.71 (2.70 - 10.02)	0.35	-
Male	125	6.42 (3.53 - 11.70)	33	7.79 (3.61 – 12.62)	0.73	22	5.66 (2.21 - 19.65)	0.73	44	5.17 (2.47 - 11.37)	0.16	
Female	93	6.06 (3.60 - 10.99)	34	5.05 (2.38 - 9.94)	0.36	19	5.66 (3.21 - 18.48)	0.76	31	6.04 (2.70 - 9.66)	0.63	-
Caesarean section	59	5.10 (3.53 - 10.35)	26	5.27 (2.56 - 10.15)	0.78	9	21.35 (2.81 - 31.40)	0.064	21	5.57 (3.02 - 6.92)	0.20	0.81
Vaginal	157	6.48 (3.81 - 12.06)	41	7.11 (3.54 – 10.66)	0.80	31	5.62 (3.07 - 11.23)	0.34	53	5.71 (2.46 - 10.82)	0.76	0.057 0.41
SGA	14	6.81 (3.49 - 11.88)	7	9.16 (4.77 – 43.94)	0.21	3	15.55 (9.23 -none)	0.091 ^a	3	5.59 (5.57 -none)	0.77ª	
Not SGA	201	6.12 (3.54 - 11.40)	0	5.27 (2.79 - 10.09)	0.35	37	5.62 (2.89 - 18.70)	0.75	70	5.64 (2.47 - 9.93)	0.15	
Breastfed	164	7.36 (4.14 – 12.66)	41	7.82 (3.63 – 10.41)	0.56	26	8.12 (3.86 - 21.55)	0.24	50	5.72 (3.69 - 12.00)	0.42	_
Not breastfed	49	3.93 (2.11 - 6.11)	17	3.22 (2.37 - 4.30)	0.36	13	2.71 (1.48 - 5.49)	0.24	21	2.70 (1.70 - 6.43)	0.68	
Antibiotics exposure	123	6.12 (3.53 – 11.34)	41	4.77 (2.38 - 9.89)	0.20	16	9.00 (3.32 - 22.79)	0.16	40	5.30 (2.31 - 7.82)	0.077	
No antibiotics exposure	92	6.11 (3.59 – 11.97)	25	8.07 (4.50 - 12.62)	0.38	22	5.64 (2.51 - 10.02)	0.34	31	5.71 (2.59 - 13.73)	0.86	
Other children	105	6.72 (4.11 – 12.06)	28	7.95 (3.86 – 11.73)	0.83	21	6.91 (3.15 – 21.21)	0.64	35	4.37 (2.08 - 6.42)	0.004*	0.21
No other children	112	5.14 (3.28 - 10.71)	39	4.91 (2.63 - 9.98)	0.56	20	4.72 (2.41 - 11.06)	0.60	38	6.21 (2.92 - 13.64)	0.37	0.65 0.015
Pets at home	96	6.11 (3.52 – 11.78)	31	3.76 (2.37 - 9.98)	0.17	18	3.71 (1.79 – 14.03)	0.25	40	4.40 (2.44 - 7.59)	0.041*	0.39
No pets at home	121	6.12 (3.63 - 11.09)	36	7.58 (4.22 – 10.46)	0.46	23	6.77 (3.23 - 19.08)	0.32	33	6.39 (2.85 – 14.14)	0.72	0.21 0.08
Postnatal smoke exposure	30	4.80 (3.03 - 8.68)	11	7.83 (3.22 – 13.43)	0.39	5	4.85 (1.58 - 37.98)	0.74	18	3.43 (2.03 - 9.17)	0.32	-
No postnatal smoke exposure	188	6.23 (3.68 - 11.78)	56	5.48 (2.79 - 10.09)	0.43	36	5.68 (3.10 - 17.75)	0.84	55	5.74 (3.35 - 11.52)	0.49	-
Postnatal SSRI use	7	5.27 (3.54 - 6.60)	7	4.15 (3.22 - 7.08)	0.85	2	15.08 (11.23 -none)	0.056*** ^{,a}	7	3.60 (1.87 - 6.19)	0.41	0.99
No postnatal SSRI	208	6.17 (3.54 - 11.78)	59	7.11 (2.71 – 10.98)	0.81	38	5.64 (2.61 - 18.63)	0.77	65	5.59 (2.66 - 10.38)	0.29]

 Table E4. Medians of infant fecal sIgA depending on maternal perceived stress after stratifying for potential covariates

	r	1		1	r		1		1	1		
use												
Infant allergic eczema	20	8.00 (4.30 - 11.78)	9	4.77 (3.68 - 8.63)	0.28	4	2.76 (0.91 - 30.92)	0.24 ^a	10	5.15 (3.15 - 14.01)	0.40	
No infant allergic eczema	196	6.06 (3.54 - 11.31)	57	7.08 (2.67 – 10.74)	0.88	37	5.69 (3.15 - 18.70)	0.68	63	5.59 (2.59 - 9.66)	0.27	-
Breastfed and other children	77	8.56 (5.22 – 13.35)	17	9.16 (7.45 - 12.24)	0.93	16	10.23 (3.63 – 21.94)	0.48	22	5.71 (3.69 - 8.39)	0.025*	-
Breastfed and antibiotics exposure	99	7.23 (3.99 – 12.06)	24	5.48 (2.22 - 9.90)	0.21	11	18.48 (4.85 – 29.01)	0.05***	25	5.57 (3.17 - 8.13)	0.16	-
Pets and antibiotics exposure	56	6.91 (3.54 – 11.11)	19	3.05 (2.36 - 9.80)	0.061	7	4.85 (3.31 - 18.92)	0.60	22	4.26 (1.72 - 5.69)	0.006*	-
Antibiotics exposure and no pets	66	5.24 (3.22 - 11.36)	22	5.27 (3.72 - 10.15)	0.95	9	18.48 (5.00 - 31.40)	0.024***	18	6.43 (2.82 - 13.59)	0.76	-
Primigravida and no pets	48	5.04 (3.33 - 9.63)	12	6.72 (4.94 - 12.53)	0.08	7	6.62 (5.66 - 18.48)	0.032***	9	8.11 (5.91 - 16.62)	0.054	-
Other children and pets	47	6.62 (4.05 - 12.06)	11	7.83 (3.76 - 13.50)	0.96	8	4.08 (1.89 - 20.75)	0.32	18	4.26 (2.56 - 6.14)	0.052	-
Other children and no pets	57	6.87 (4.18 - 12.15)	17	8.07 (3.96 - 11.04)	0.76	13	9.23 (4.46 - 22.17)	0.65	17	5.27 (1.71 - 7.69)	0.037*	-
No other children and no pets	64	5.04 (3.41 - 10.26)	19	5.61 (4.77 - 10.50)	0.26	10	5.64 (2.98 - 16.28)	0.73	16	10.87 (5.75 – 16.78)	0.015*	-
No antibiotics and no other children	35	5.08 (3.10 - 9.64)	13	5.11 (3.10 - 11.80)	0.55	13	4.07 (2.31 - 6.82)	0.51	12	9.07 (3.97 - 19.60)	0.057	-
Other children and no antibiotics	57	6.43 (4.11 - 13.14)	12	8.46 (7.26 - 13.12)	0.38	9	6.91 (2.28 - 20.08)	0.96	18	4.48 (2.38 - 9.93)	0.05*	-
Other children and antibiotics	46	7.36 (4.20 – 11.36)	15	7.11 (2.16 – 10.03)	0.51	10	15.08 (4.46 - 25.90)	0.09	16	4.40 (1.71 - 6.42)	0.019*	-

*Significant at 0.05 a priori; **Significant at Bonferroni adjusted p-value 0.05/2; *** Significant at Bonferroni adjusted p-value 0.05/1; aFisher's Exact test

^bInteractions between the covariate and perceived stress group variable were tested. When significant interactions were found, the interaction term for each perceived stress group was reported.

Note: Stratifications with two covariates that did not have significant differences were not reported in this table.

 Table E5. Logistic regression analyses for low sIgA in infants after exposure to pre and postnatal highest quartile of perceived stress

(Ref: lower stress)	N for lowest quartile of sIgA	OR for stress	OR for covariate	Decision
Crude OR	100	1.37 (0.73 – 2.58)		
	100	1.74 (0.83 - 3.62)		
		1.65(0.92 - 2.98)		
Adjusted for child sex	M-54	1.36(0.73 - 2.56)	1.10 (0.70 –	Out
Aujusteu for clina sex	F-46	1.30(0.73 - 2.50) 1.73(0.83 - 3.61)	1.74)	Out
	г-40		1.74)	
	1 et 10	1.66(0.92 - 2.99)	0.04 (0.46	L (-0.00)
Adjusted for child age at stool collection	1 st -18	1.38 (0.73 – 2.62)	0.94 (0.46 -	In (p<0.20)
(quartiles)	2 nd -19	1.70 (0.80 – 3.61)	1.94)	
	3 rd -27	1.60 (0.88 – 2.91)	1.67 (0.85 –	
	4 th -36		3.29)	
			2.43 (1.26 –	
			4.71)	
Adjusted for breastfeeding at stool	E-11	1.72 (0.86 – 3.44)	9.51 (4.56 –	Always in model
collection	P-39	1.70 (0.77 – 3.78)	19.84)	
	Z-47	1.44 (0.76 – 2.74)	3.91 (1.90 -	
			8.05)	
Adjusted for birth mode	SC-9	1.36 (0.72 - 2.58)	0.71 (0.33 -	Out
	EC-19	1.63 (0.77 - 3.46)	1.55)	
	V-70	1.03(0.95 - 3.12)	1.17 (0.64 –	
		1.15 (0.75 5.12)	2.14)	
Adjusted for antibiotics exposure	Y-58	1.40 (0.74 - 2.65)	1.22 (0.76 –	In
Aujusted for antibiotics exposure	N-39	1.40(0.74 - 2.03) 1.59(0.73 - 3.46)	1.96)	111
	IN-39		1.90)	
	N/ 40	1.82(1.00 - 3.30)	1.20 (0.07	I (10.00)
Adjusted for gravida (primigravida)	Y-43	1.39 (0.74 – 2.63)	1.39 (0.87 –	In (p<0.20)
	N-55	1.63 (0.77 – 3.46)	2.22)	
		1.76 (0.97 – 3.19)		
Adjusted for SGA	Y-3	1.44 (0.76 – 2.71)	0.36 (0.11 –	In (p<0.20)
	N-95	1.64 (0.77 – 3.49)	1.22)	
		1.71 (0.94 – 3.10)		
Adjusted for depression history	Y-25	1.36 (0.72 – 2.58)	1.39 (0.80 -	Out
	N-74	1.79(0.85 - 3.75)	2.42)	
		1.64(0.90 - 3.00)		
Adjusted for maternal asthma and allergy	Y-66	1.42 (0.75 - 2.67)	0.98 (0.60 -	Out
	N-32	1.82 (0.87 - 3.81)	1.61)	
		1.63 (0.90 – 2.97)	,	
Adjusted for prenatal SSRI	Y-6	1.30(0.69 - 2.45)	1.37 (0.49 -	Out
Adjusted for prenatal SSKI	N-93	1.50(0.0) - 2.43) 1.73(0.83 - 3.63)	3.82)	Out
	11-75	1.75(0.83 - 5.05) 1.52(0.83 - 2.78)	5.62)	
Adjusted for postnatal SSRI	Y-6	1.32(0.83 - 2.78) 1.38(0.73 - 2.61)	0.97 (0.37 –	In
Aujusted for postilatal SSKI				111
	N-92	1.58(0.74 - 3.34)	2.55)	
		1.62 (0.89 - 2.96)	0.01 (0.51	
Adjusted for prenatal smoke exposure	Y-26	1.43 (0.76 – 2.71)	0.91 (0.54 -	Out
	N-73	1.82 (0.87 – 3.81)	1.53)	
		1.75 (0.97 – 3.17)		
	T 12	1.35(0.72 - 2.55)	1.86 (1.05 –	In (p<0.20)
Adjusted for postnatal smoke exposure	T-23			1
Adjusted for postnatal smoke exposure	N-76	1.77 (0.84 – 3.69)	3.32)	
			3.32)	
Adjusted for postnatal smoke exposure Adjusted for pets at home		1.77 (0.84 – 3.69)	3.32)	In (p<0.20)
	N-76 Y-55	$\begin{array}{c} 1.77 \ (0.84 - 3.69) \\ 1.51 \ (0.82 - 2.76) \\ 1.36 \ (0.72 - 2.56) \end{array}$	1.55 (0.98 –	In (p<0.20)
	N-76	$\begin{array}{c} 1.77\ (0.84-3.69)\\ \hline 1.51\ (0.82-2.76)\\ \hline 1.36\ (0.72-2.56)\\ \hline 1.74\ (0.83-3.63)\\ \end{array}$		In (p<0.20)
Adjusted for pets at home	N-76 Y-55 N-45	$\begin{array}{c} 1.77\ (0.84-3.69)\\ 1.51\ (0.82-2.76)\\ 1.36\ (0.72-2.56)\\ 1.74\ (0.83-3.63)\\ 1.64\ (0.90-2.98)\end{array}$	1.55 (0.98 – 2.45)	
	N-76 Y-55 N-45 Y-43	$\begin{array}{c} 1.77\ (0.84-3.69)\\ 1.51\ (0.82-2.76)\\ 1.36\ (0.72-2.56)\\ 1.74\ (0.83-3.63)\\ 1.64\ (0.90-2.98)\\ 1.35\ (0.72-2.53)\\ \end{array}$	1.55 (0.98 – 2.45) 0.80 (0.51 –	In (p<0.20) Out
Adjusted for pets at home	N-76 Y-55 N-45	$\begin{array}{c} 1.77\ (0.84-3.69)\\ 1.51\ (0.82-2.76)\\ 1.36\ (0.72-2.56)\\ 1.74\ (0.83-3.63)\\ 1.64\ (0.90-2.98)\\ 1.35\ (0.72-2.53)\\ 1.74\ (0.83-3.63)\\ \end{array}$	1.55 (0.98 – 2.45)	
Adjusted for pets at home	N-76 Y-55 N-45 Y-43	$\begin{array}{c} 1.77\ (0.84-3.69)\\ 1.51\ (0.82-2.76)\\ 1.36\ (0.72-2.56)\\ 1.74\ (0.83-3.63)\\ 1.64\ (0.90-2.98)\\ 1.35\ (0.72-2.53)\\ \end{array}$	1.55 (0.98 – 2.45) 0.80 (0.51 –	

Adjusted for antibiotics and gravida	97	<u>1.66 (0.91 - 3.02)</u> <u>1.40 (0.74 - 2.65)</u>	In
Adjusted for antibiotics and gravida	97	1.40(0.74 - 2.65) 1.59(0.73 - 3.48)	In
		1.39(0.73 - 3.48) 1.87(1.03 - 3.41)	
Adjusted for antibiotics and SGA	97	1.45 (0.77 – 2.75)	Keep SGA out
regusted for antibioties and 5674	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	1.60(0.77 - 2.79) 1.60(0.73 - 3.49)	Reep Son out
		1.82(1.00 - 3.32)	
Adjusted for antibiotics and breastfeeding	94	1.77 (0.87 – 3.60)	In
		1.56 (0.67 – 3.64)	
		1.62 (0.84 – 3.13)	
Adjusted for postnatal SSRI and	95	1.70 (0.84 – 3.44)	In
breastfeeding		1.46(0.65 - 3.31)	
A divised for note and broastfooding	97	$\frac{1.45 (0.74 - 2.81)}{1.65 (0.82 - 3.34)}$	V con note out
Adjusted for pets and breastfeeding	97	1.05(0.82 - 3.34) 1.73(0.78 - 3.84)	Keep pets out
		1.75(0.78 - 5.84) 1.47(0.77 - 2.82)	
Adjusted for pets and postnatal SSRI	98	$\frac{1.47(0.77-2.52)}{1.37(0.72-2.59)}$	Keep pets out
regusted for pets and postnatal 551Cl	20	1.57(0.72 - 2.57) 1.59(0.75 - 3.37)	Reep pers our
		1.59(0.87 - 2.92)	
Adjusted for pets and other children	99	1.33(0.70-2.51)	Out
5 1		1.74 (0.83 – 3.63)	
		1.57 (0.86 – 2.86)	
Adjusted for pets and postnatal smoking	99	1.34 (0.71 – 2.53)	In
		1.76 (0.84 – 3.69)	
		1.48 (0.80 – 2.72)	
Adjusted for breastfeeding and postnatal	96	1.69 (0.84 – 3.41)	Keep smoking out
smoking		1.71 (0.77 – 3.79)	
		1.38 (0.71 – 2.67)	
Adjusted for breastfeeding and prenatal	96	1.87 (0.93 - 3.79) 1.76 (0.70 - 2.02)	Out
smoking		1.76(0.79 - 3.93) 1.53(0.80 - 2.94)	
Adjusted for maternal asthma/allergy and	96	$\frac{1.35(0.80-2.94)}{1.44(0.76-2.71)}$	Out
gravida	90	1.44(0.76 - 2.71) 1.72(0.80 - 3.67)	Out
gravida		1.72(0.00-3.07) 1.74(0.95-3.18)	
Adjusted for maternal characteristics	95	1.36 (0.72 – 2.59)	Out
(asthma/allergy, prenatal SSRI, gravida)		1.72(0.80 - 3.69)	
		1.60 (0.86 – 2.97)	
Adjusted for depression history and	98	1.31 (0.69 - 2.49)	Out
prenatal SSRI		1.81 (0.86 – 3.82)	
		1.51 (0.82 – 2.80)	
Adjusted for depression history and	97	1.36 (0.72 – 2.60)	Out
postnatal SSRI		1.61 (0.76 – 3.45)	
		1.56 (0.85 – 2.89)	
Adjusted for breastfeeding, pets at home,	94	1.72(0.84 - 3.51)	Keep pets out
antibiotics		1.57(0.67 - 3.67)	
A directed for house the direction	93	1.67 (0.86 - 3.25)	Kaan amalaina an
Adjusted for breastfeeding, antibiotics, prenatal smoking	93	1.94 (0.95 - 3.97) 1.61 (0.69 - 3.77)	Keep smoking ou
prenatal shoking		1.01(0.09 - 3.77) 1.72(0.88 - 3.35)	
Adjusted for breastfeeding, antibiotics,	94	1.82 (0.89 – 3.71)	Keep SGA out
SGA		1.62(0.69 - 3.74) 1.60(0.68 - 3.74)	Reep Son out
		1.60(0.82 - 3.09)	
Adjusted for breastfeeding, antibiotics,	94	1.77 (0.87 – 3.60)	Keep gravida out
gravida		1.54(0.66 - 3.61)	
-		1.68 (0.87 – 3.25)	
Adjusted for breastfeeding, antibiotics,	92	1.74 (0.85 – 3.56)	In
postnatal SSRI		1.32 (0.55 – 3.13)	
		1.62 (0.82 – 3.19)	
	01		
Adjusted for breastfeeding, antibiotics,	91	1.91 (0.93 - 3.95) 1.22 (0.55 - 2.19)	In
postnatal SSRI, maternal asthma/allergy	1	1.33 (0.55 – 3.18)	

		1.65 (0.84 - 3.28)	
Adjusted for breastfeeding, antibiotics,	91	1.86 (0.90 – 3.87)	Final Model
postnatal SSRI, maternal asthma/allergy,		1.38 (0.56 – 3.37)	
age		1.72 (0.86 – 3.42)	
Final Model with interaction for	91	0.57 (0.14 – 2.33)	
antibiotics exposure		1.68 (0.53 – 5.34)	
_		2.46 (0.85 - 7.11)	

Table E6. Interactions between depressive symptoms and covariates in final model

Variable	Interactions with depressive symptoms (p-value)
Antibiotics exposure	0.04
	0.57
	0.41
Maternal asthma and allergy	0.79
Breastfeeding	0.99
Postnatal SSRI	0.80
Age	0.55

Note: Interactions between the covariate and depressive symptoms group variable were tested.

When interaction terms with p < 0.05 were found, the interaction term for each depressive

symptoms group was reported.